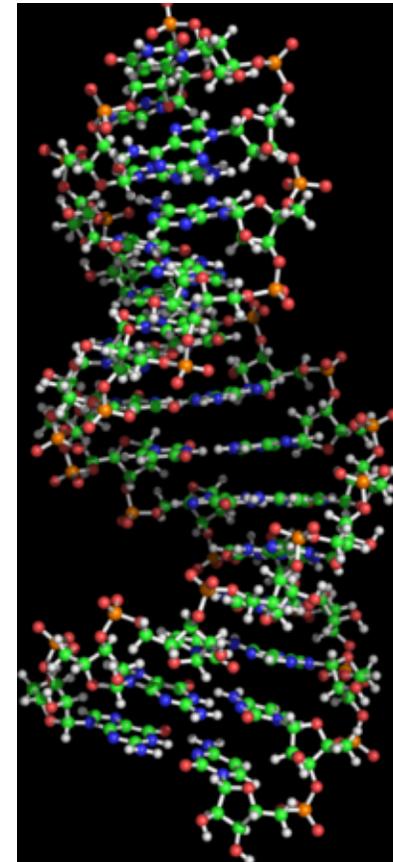
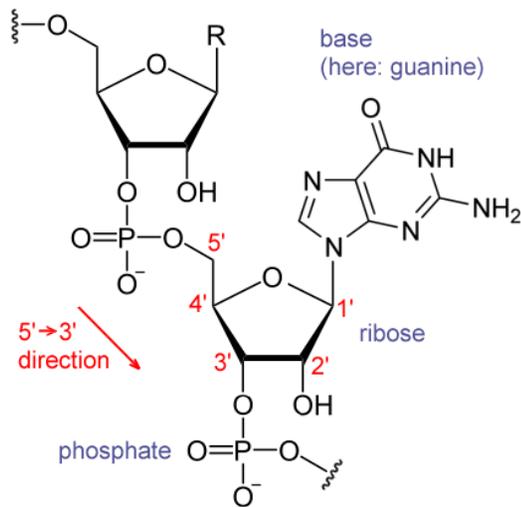


# Ribo-gnome: The Big World of Small RNAs

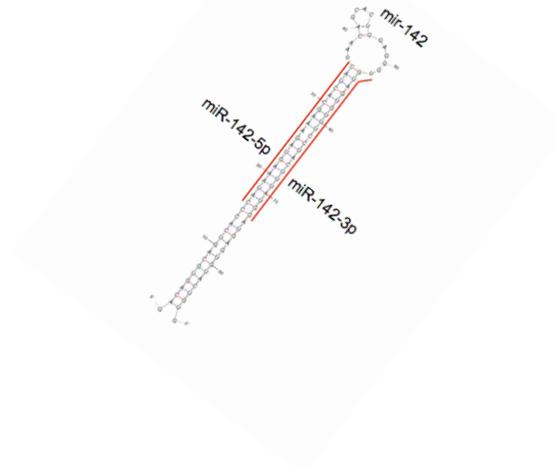
Phillip D. Zamore and Benjamin Haley



Presentation by: Christopher Jakubowski

# Three Classes of small RNAs

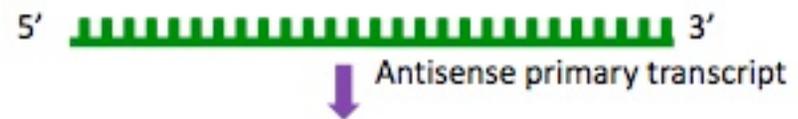
- miRNAs



- siRNAs

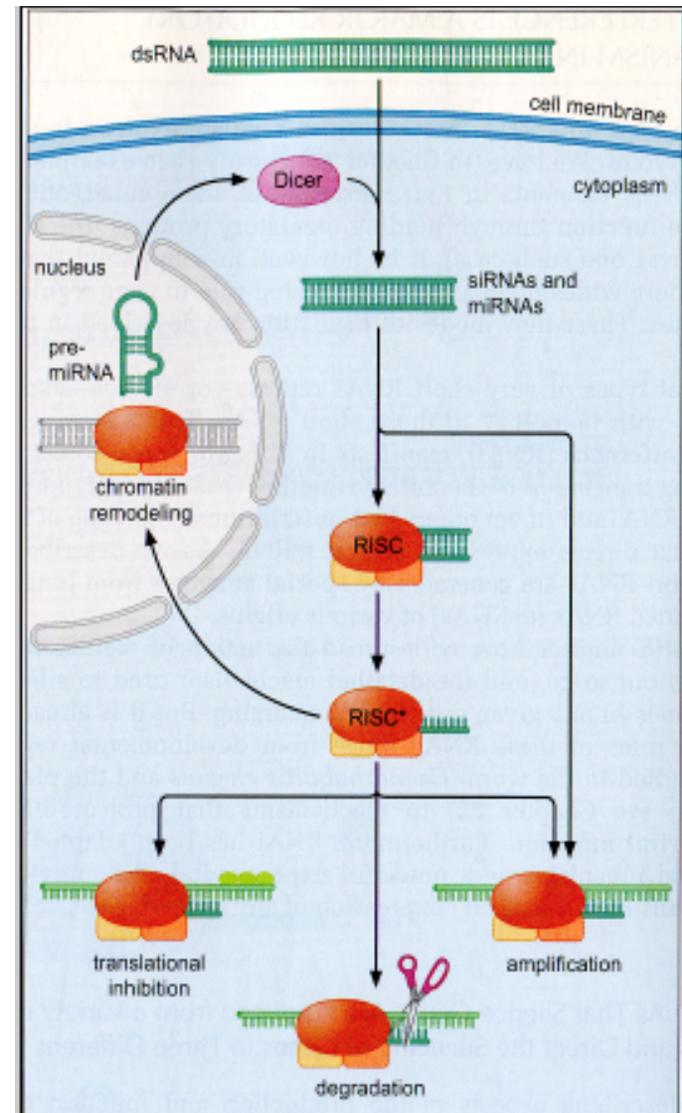


- rasiRNAs

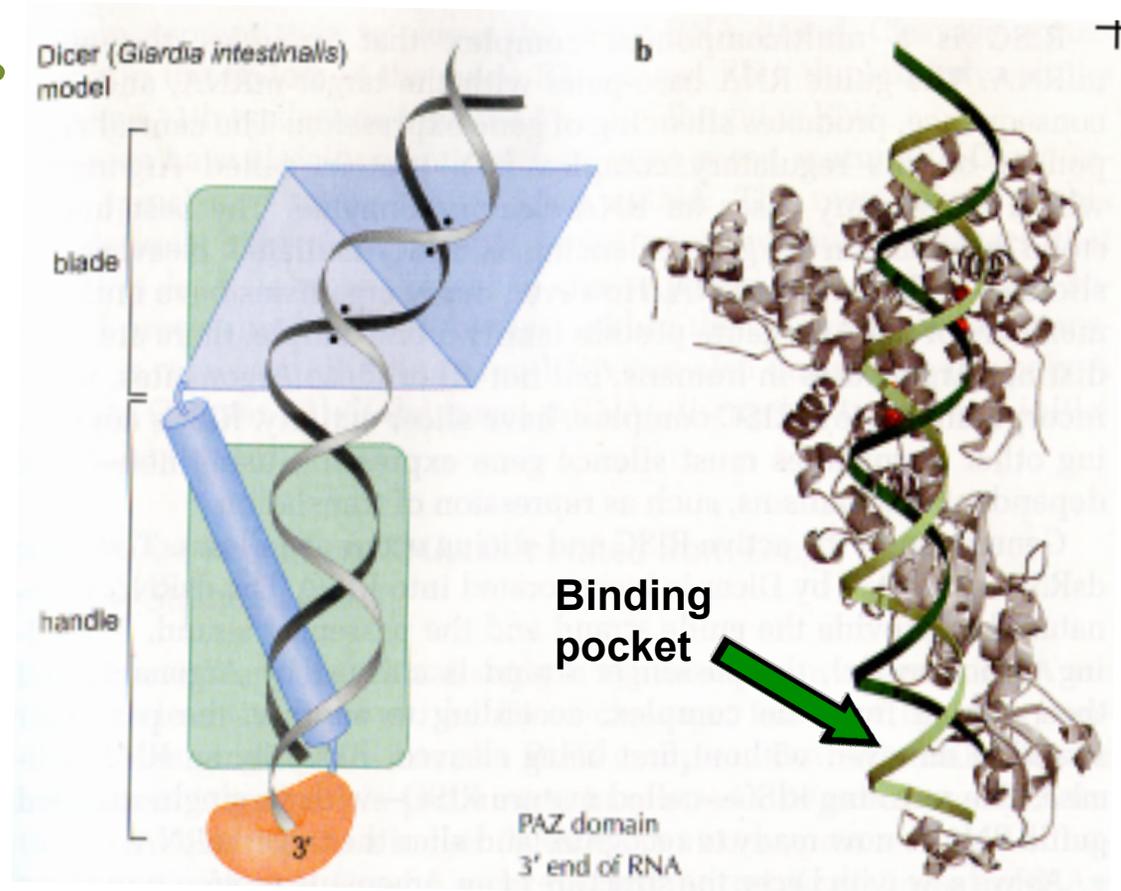


# RNA silencing pathway

- Double strand molecules must have homology
- Three methods of repression:
  1. Digest mRNA
  2. Block Translation
  3. Chromatin Modification

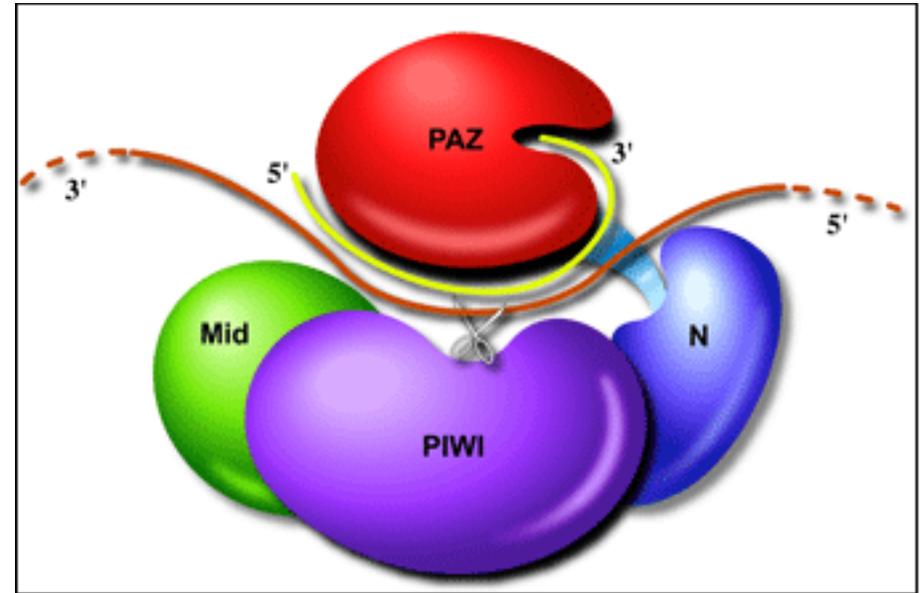
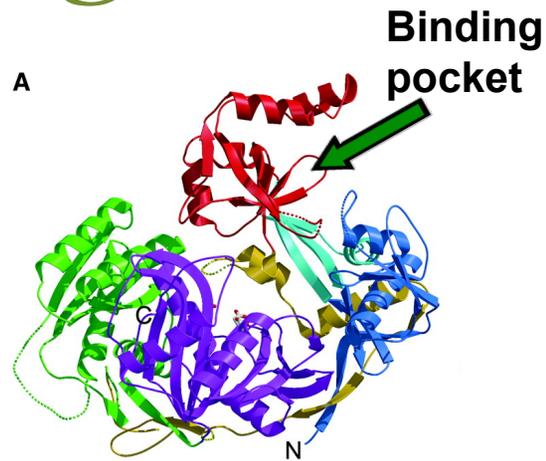


# Dicer

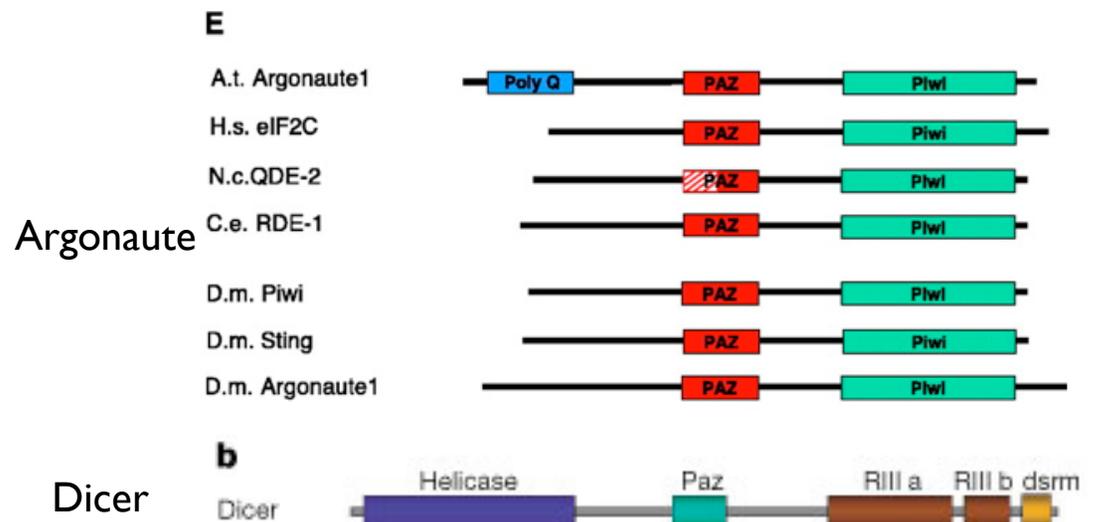


- Selects cleavage sites based on measuring
- Three modules: two RNase III domain and PAZ domain
- Cleaves 22 nucleotides from its end
- Connection between siRNA and miRNA

# Argonaute

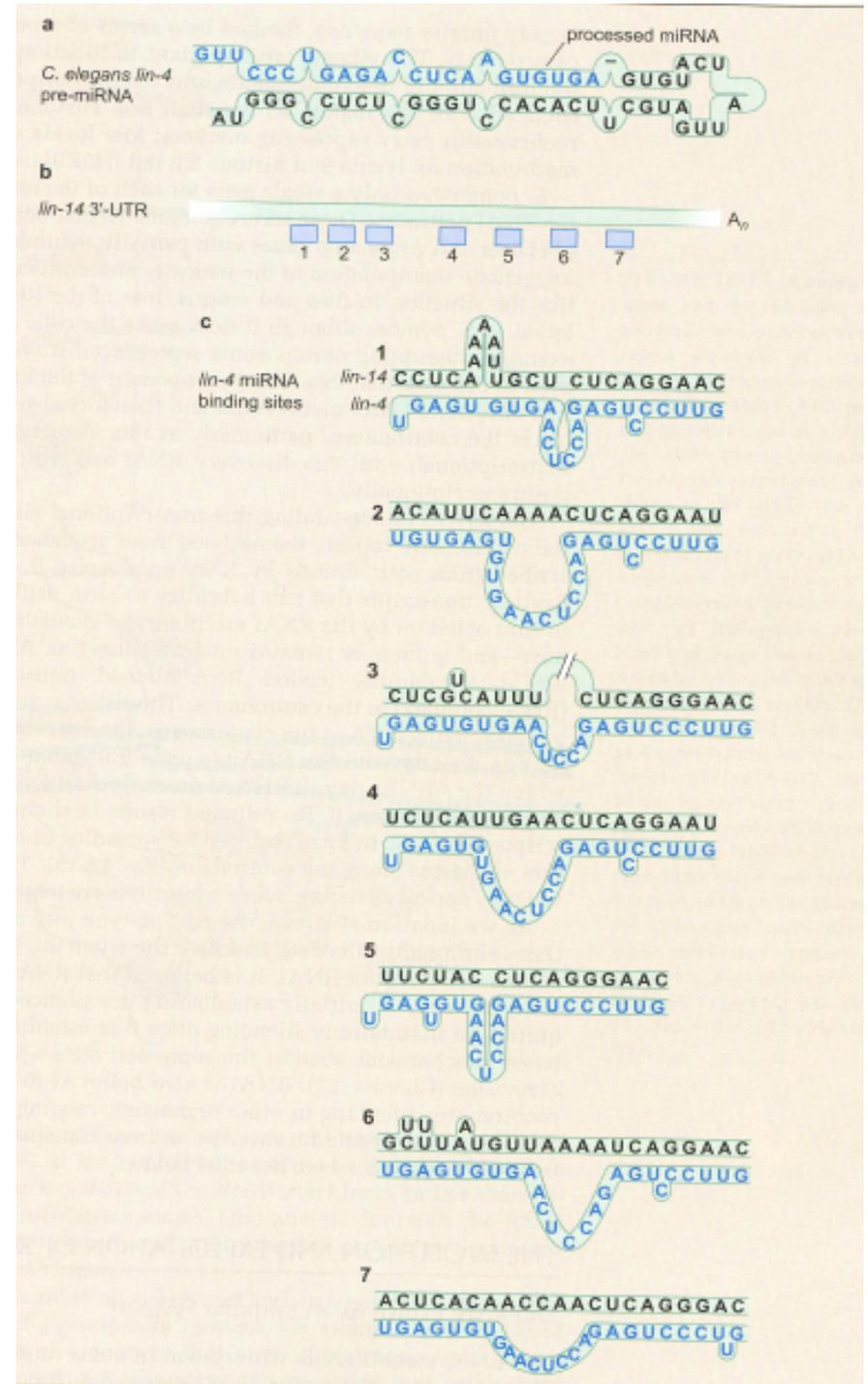


Along with  
Dicer both  
have PAZ  
domain and  
RNase Domain

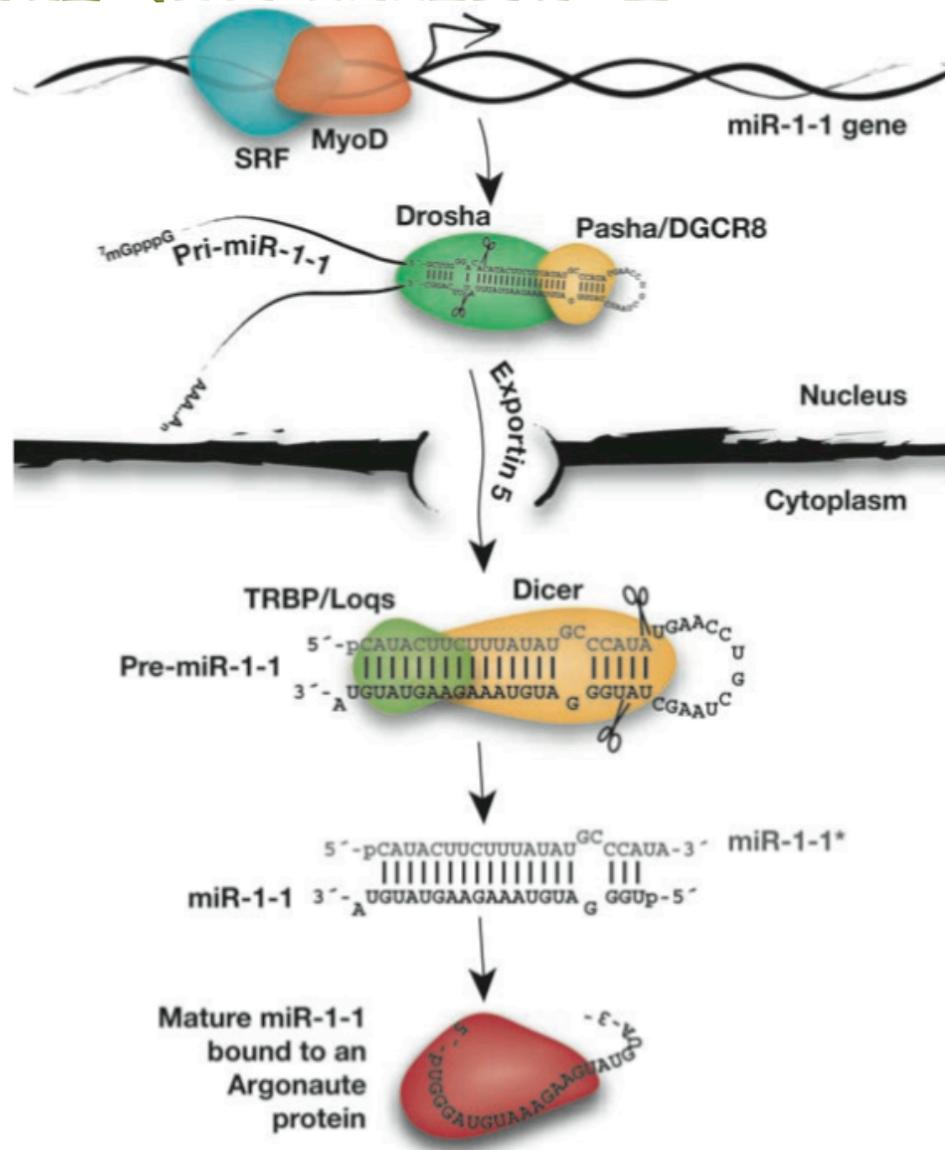


# Discovery of miRNA

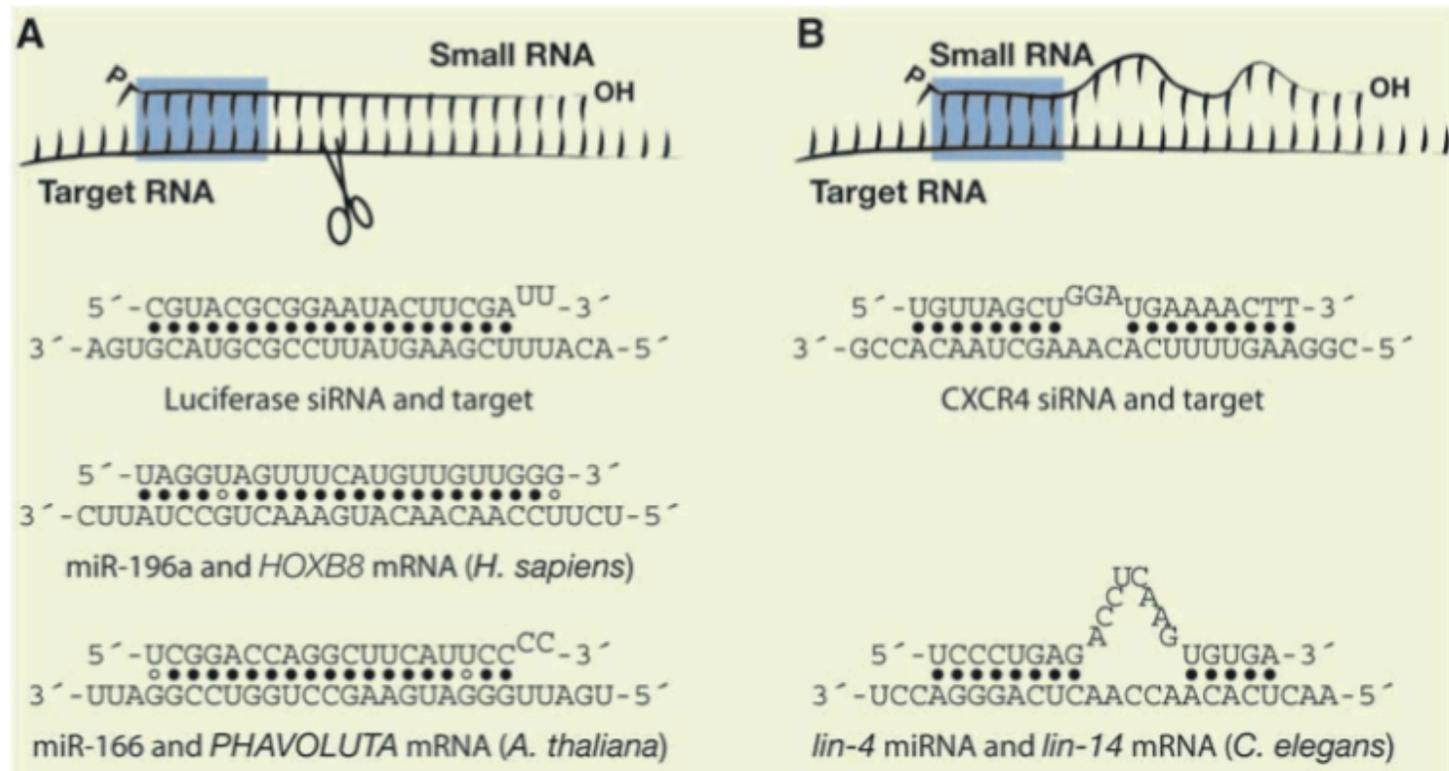
- While siRNAs are found in an assortment of eukaryotes, miRNAs have only been found in plants, animals, and their viruses
- Ambros and co-workers found two RNA transcripts from *lin-4* locus and *lin-4* could base pair sites within *lin-14*



# miRNA miR-1



# Small RNAs act in two distinct ways



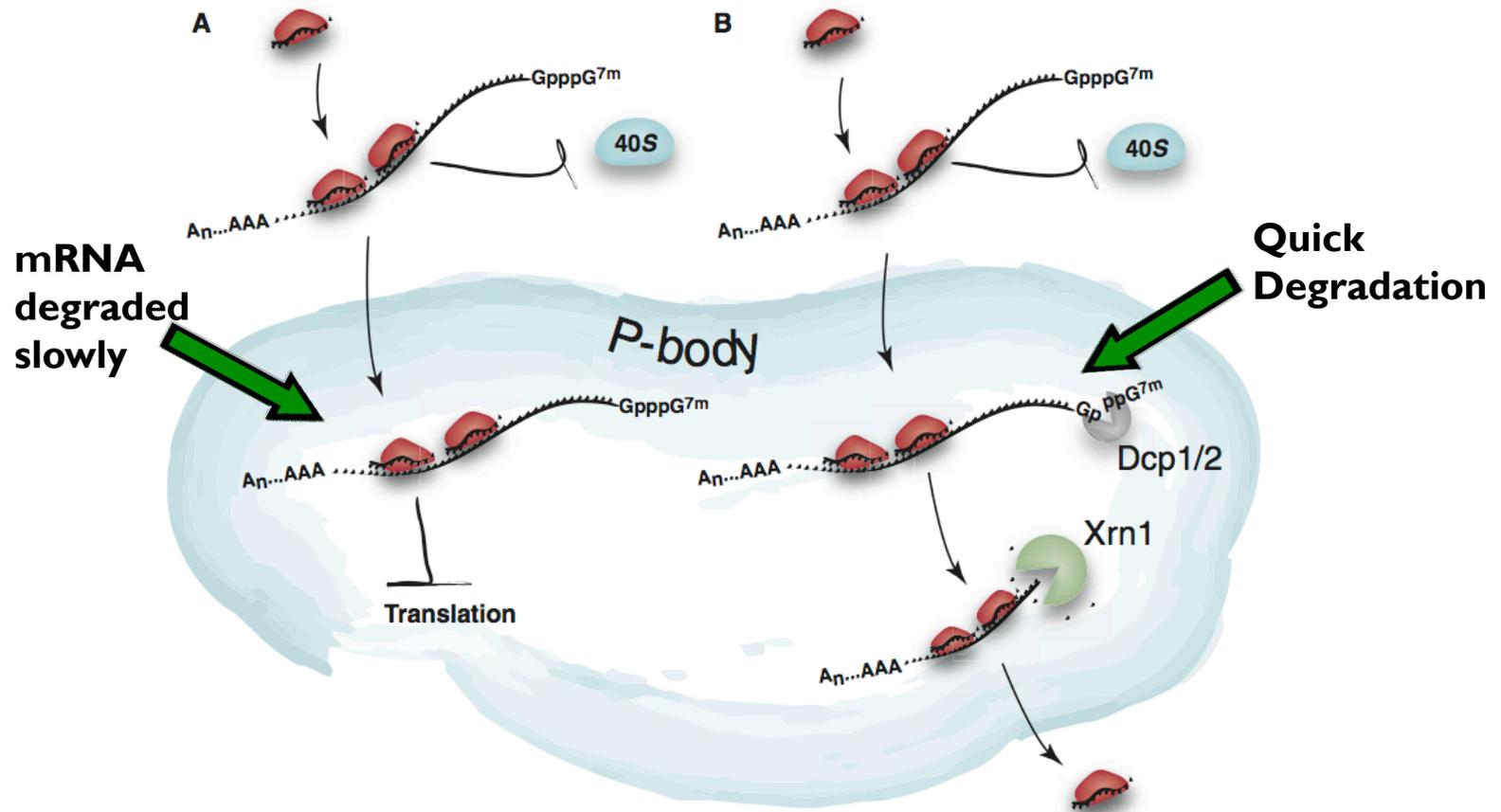
- “Seed” sequence highlighted in blue nucleates binding
- Figure (A) presents extensive RNA binding. Argonaute proteins are then directed to cut a single phosphodiester bond leading to destruction
- Figure (B) presents partial base pairing. With attached Argonaute protein translation is prevented



# Theories regarding translation inhibition

- Animal miRNAs usually act in this mode because of partial complementarity
- What happens?
  1. Direct degradation of nascent polypeptide
  2. “Freeze” ribosomes in place
- Theories called into question
  - Lim and co-workers using microarrays found miRNAs can alter stability of hundreds of mRNAs
  - Studied changes in steady-state mRNA unlikely to be due to cleavage
- **How do miRNAs make mRNA less stable then?**

# Sequestration in P-body model



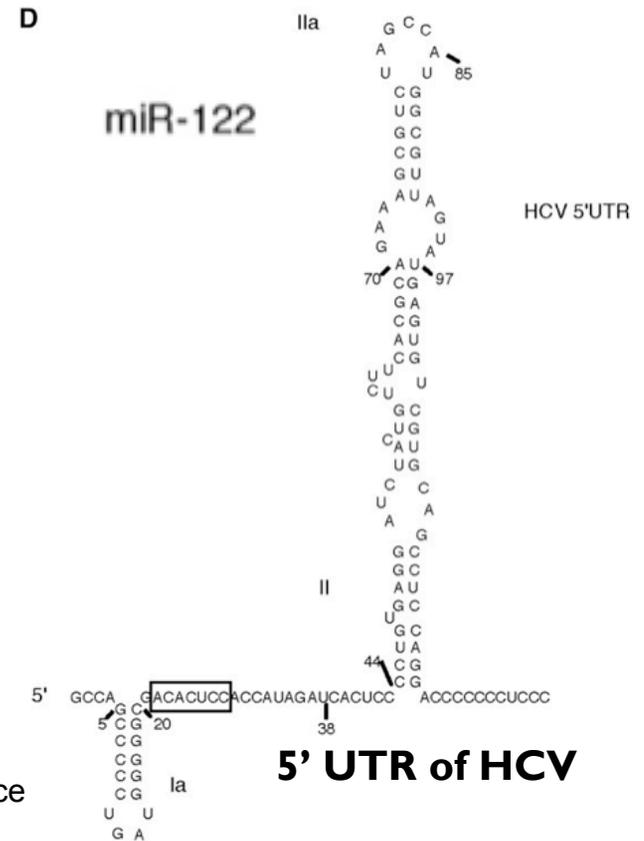
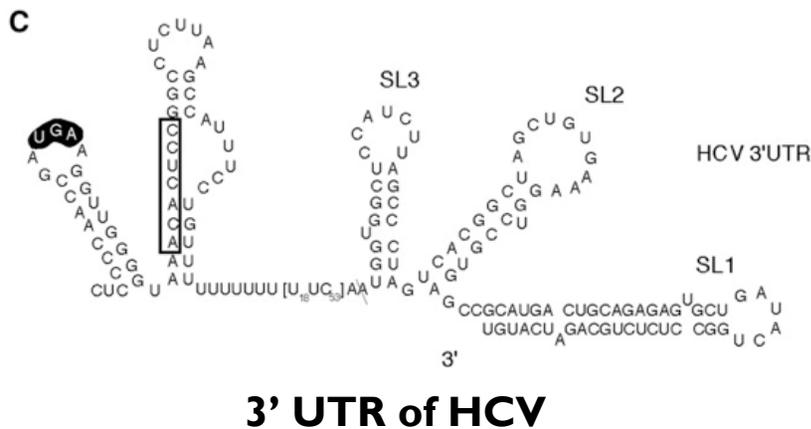
- Cytoplasmic site of mRNA decapping and degradation
- Argonaute concentrates here only when bound to miRNAs or siRNAs
- Mutant Argonautes remain in the cytosol
- Argonaute associates with decapping enzymes

# Model of miRNA gene activation

- Specific liver cells, Huh7, produce miR-122
- Hepatitis C virus (HCV) can only replicate in Huh7 cells
- **Connection between presence of miR-122 in permissive Huh7 cells?**

3' UGUUUGUGGUAACAGUGUGAGGU 5'

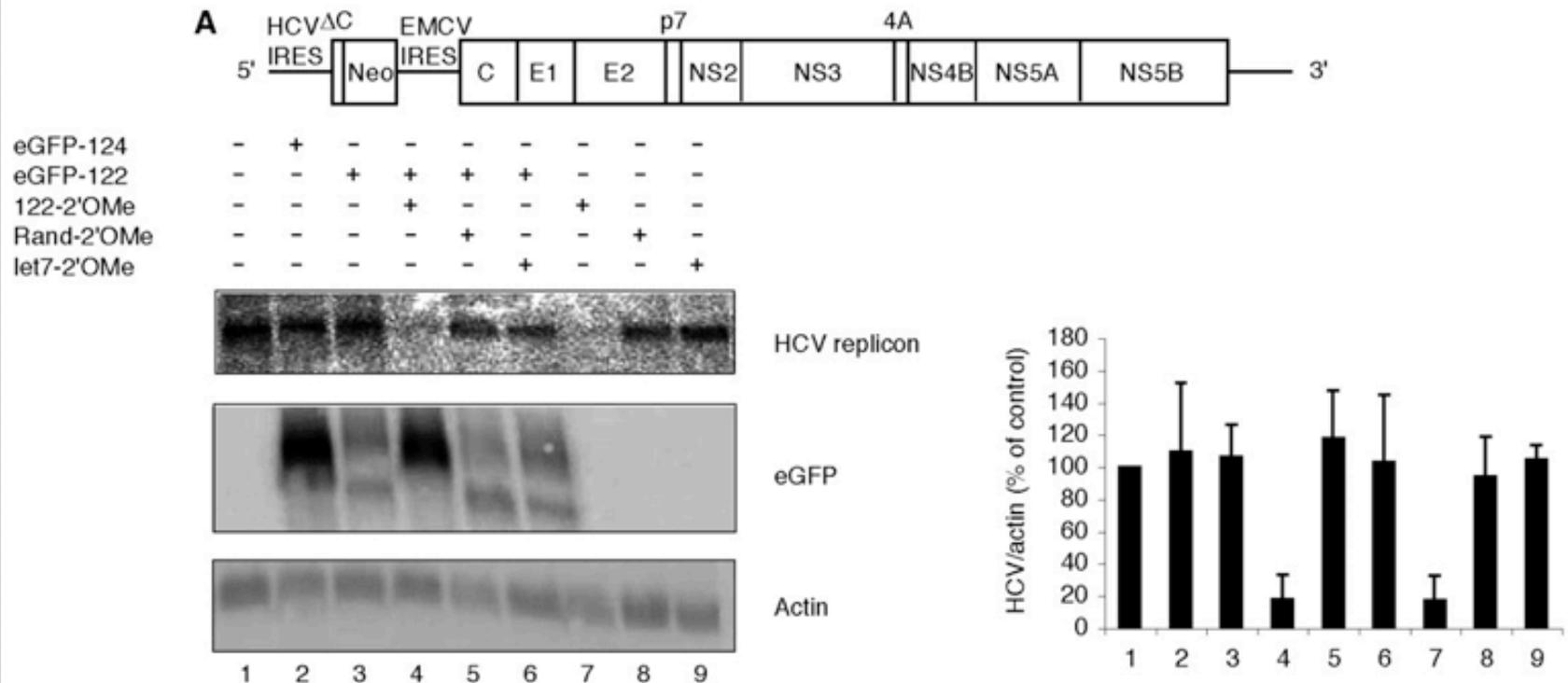
**Sequence of miR-122 with the seed sequences surrounded by a box**



Jopling, Catherine L. "Modulation of Hepatitis C Virus RNA Abundance by a Liver-Specific MicroRNA." *Science* 309 (2005): 309.

# Testing the predicted models

- Abundance of autonomously replicating, HCV RNA replicon was tested during miR-122 inactivation



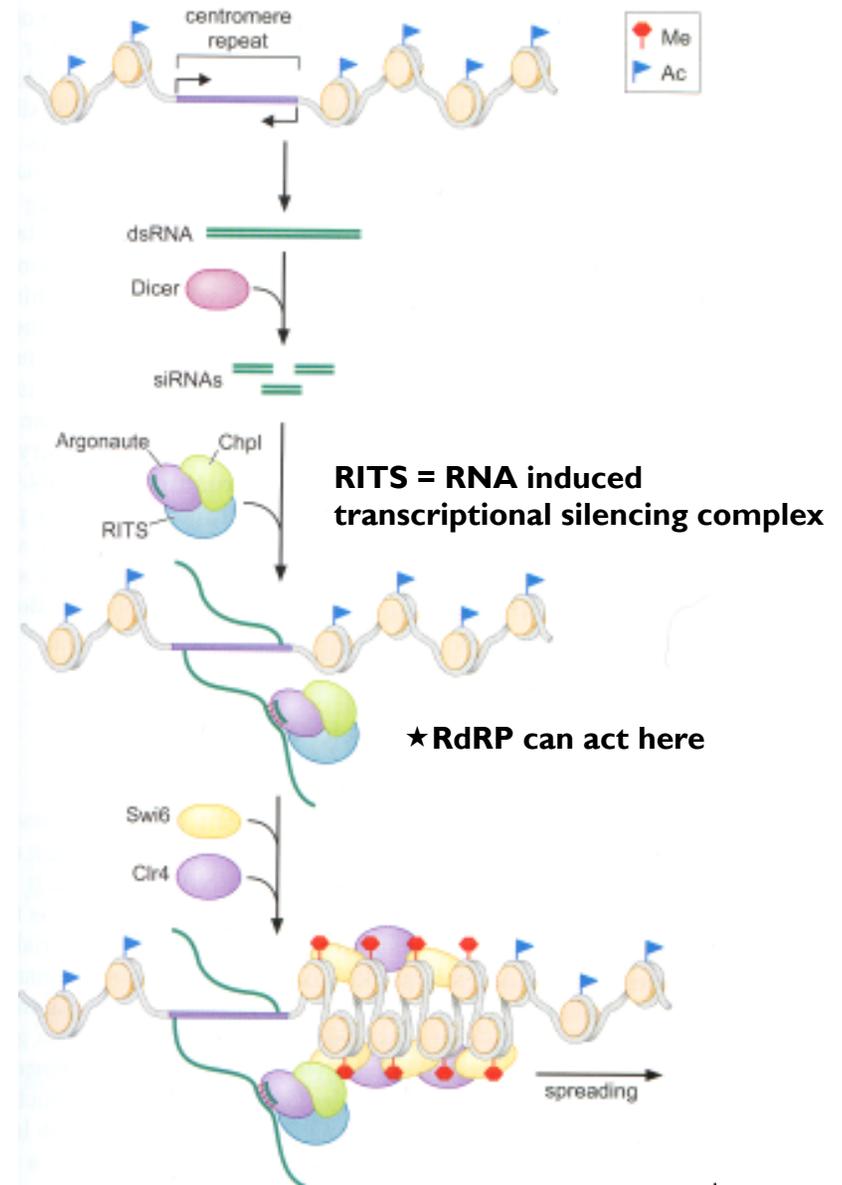


## Implications in transcriptional silencing

- Associated with heterochromatin formation
- Marked by H3K9 methylation or hypermethylation
- Topics:
  - *S. pombe*
  - RNA Polymerase IV
  - RNA Polymerase II

# si-RNA directed heterochromatin assembly

- In outer regions of centromere in *S. pombe*
- Needed for chromosome segregation
- Repetitive sequences compose out regions (similar to mammals)
- Argonaute can also slice transcripts and RdRP can make further substrates increasing efficiency



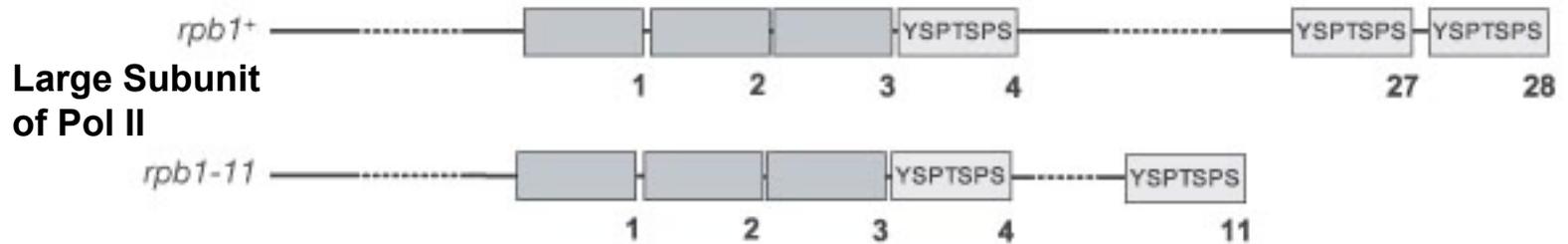


## Supplying transcripts for siRNA production

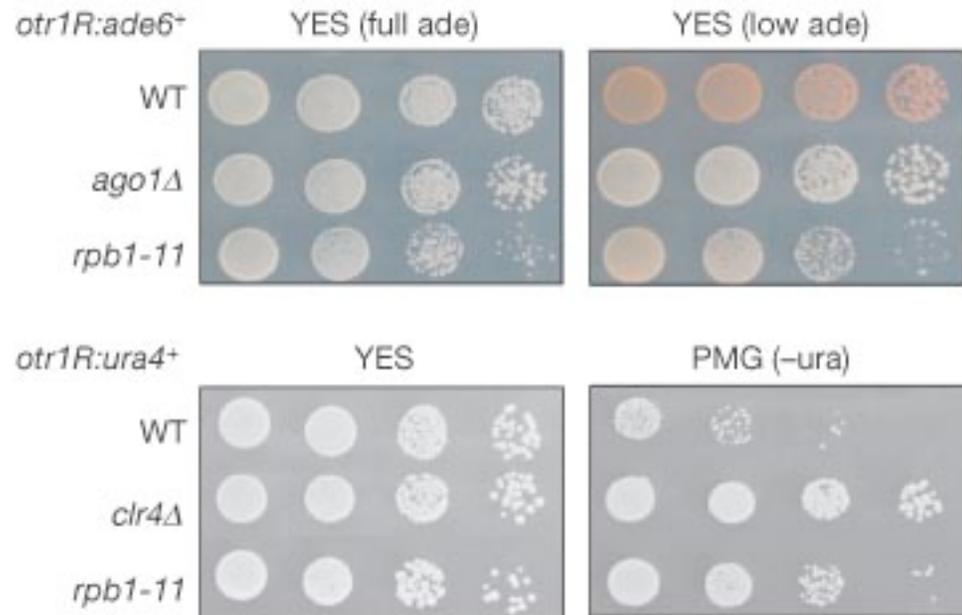
- Previous model requires transcription across silenced regions
- In plants, RNA polymerase IV transcribes silent heterochromatin
- RNA-dependent RNA polymerase (RdRP) can then make substrate for dicer
- Pol II can create targets for small RNAs as well as trigger for small RNA production
- CTD of Pol II might interact with silencing machinery, then Argonaute proteins with loaded siRNA are recruited

# Evidence for CTD interaction

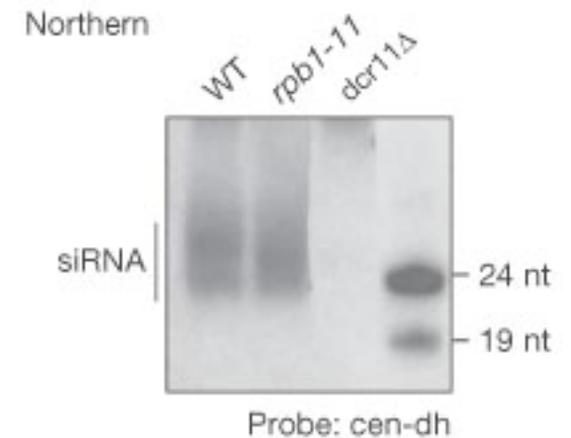
Experiments in *S. cerevisiae*



• Deletion of 16 CTD heptad repeats



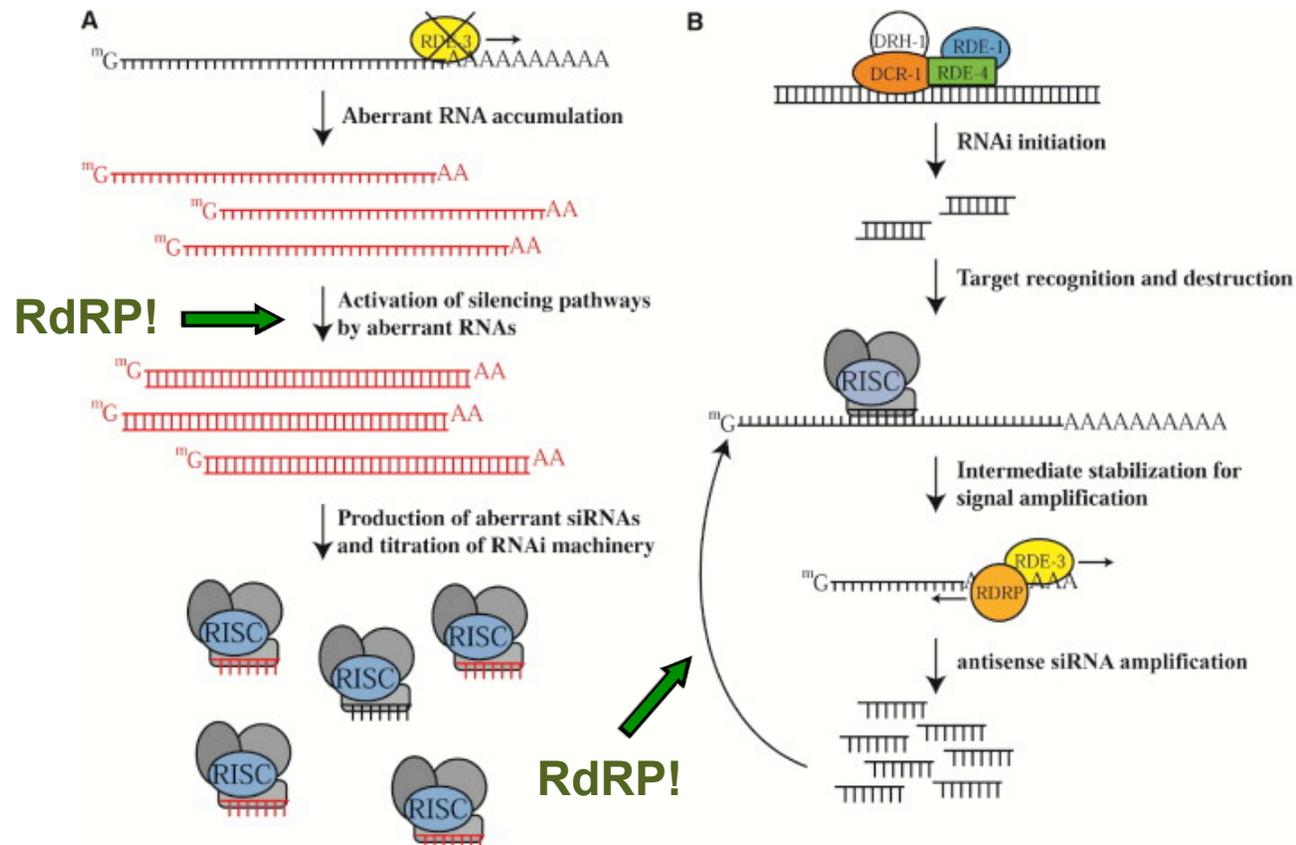
• *ura4+* and *ade6+* are centromeric markers



**Transcription but no repression!**

# Template-independent RNA polymerases

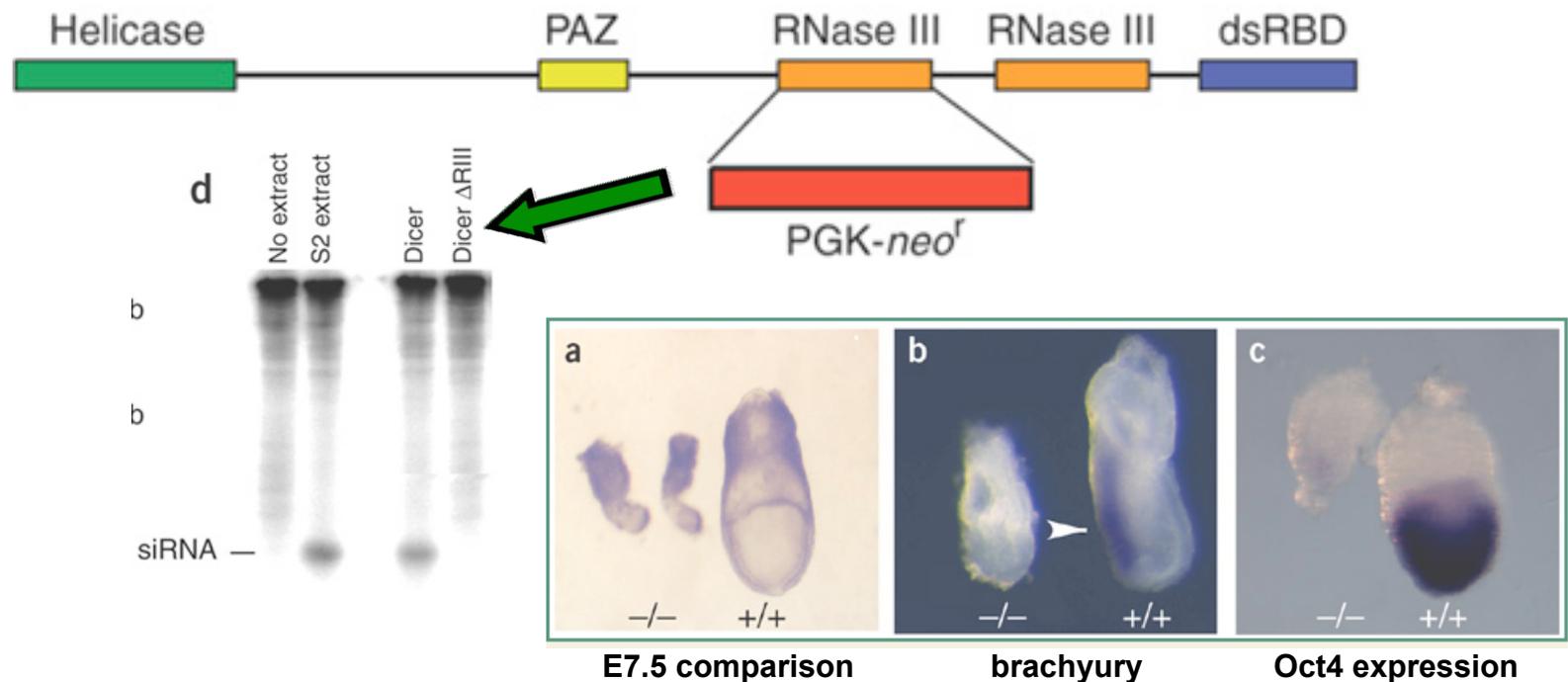
- Required for RNA silencing in worms and yeast
- Polymerase  $\beta$  nucleotidyltransferase superfamily (includes polyA polymerases)



Chen, C. C. "A Member of the Polymerase Nucleotidyltransferase Superfamily Is Required for RNA Interference in *C. elegans*." *Current Biology* 15 (2005): 378.

## Small RNAs needed for stem cell life cycle

- Embryonic stem cells lacking Dicer, Argonaute proteins, or dsRNA-binding partners die rapidly
- Defects due to either loss of miRNA or silent heterochromatin



Bernstein, E. "Dicer is essential for mouse development." *Nature Genetics* 35 (2003): 215.

# Citations

- Bernstein, E. "Dicer is essential for mouse development." Nature Genetics 35 (2003): 215.
- Bernstein, E. Nature 409 (2001): 363.
- Chen, C. C. "A Member of the Polymerase Nucleotidyltransferase Superfamily Is Required for RNA Interference in *C. elegans*." Current Biology 15 (2005): 378.
- Hammond, Scott M. "Argonaute2, a Link Between Genetic and Biochemical Analyses of RNAi." Science 293 (2001): 1146.
- Jopling, Catherine L. "Modulation of Hepatitis C Virus RNA Abundance by a Liver-Specific MicroRNA." Science 309 (2005): 309.
- Onodera, Yasuyuki, and Jeremy R. Haag. "Plant Nuclear RNA Polymerase IV Mediates siRNA and DNA Methylation-Dependent Heterochromatin Formation." Cell 120 (2005): 613.
- V. Schramke et al., Nature 435, 1275 (2005)
- Watson, James D., Tania A. Baker, Stephen P. Bell, Alexander Gann, Michael Levine, and Richard Losick. Molecular Biology of the Gene. 6th ed. Cold Spring Harbor: Cold Spring Harbor Laboratory P, 2008.
- Zamore, Phillip D., and Benjamin Haley. "Ribo-gnome: The Big World of Small RNAs." Science 309 (2005): 1519-524.