

# Polyphenolic contents and antimicrobial activity of different extracts of *Padina boryana* Thivy and *Enteromorpha* sp marine algae

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## ABSTRACT

Marine algae are potential sources of biologically active compounds with antiviral, antimicrobial and anticancer activities. Two species of marine algae, *Padina boryana* Thivy and *Enteromorpha* sp were collected from the Red Sea. Both species were extracted with ethanol, acetone, and hexane. The extracts of each sample were examined for, total phenolic, total flavonoid contents and antimicrobial activities. The results showed that solvents with different polarities have various effects on phenolic and flavonoid contents. Among the tested solvents, ethanolic extract of *Padina boryana* Thivy and *Enteromorpha* sp showed the highest phenolic content. Results also indicated that the extracts of *Padina boryana* Thivy showed higher antimicrobial activity compared to *Enteromorpha* sp. The present study demonstrates that both algae species exhibited excellent antimicrobial properties which may use as natural food preservative for possible application in food for health promotion.

## INTRODUCTION

The marine environment is a good source of bioactive secondary metabolites, many of which exhibit structural features not found in terrestrial natural products (Cantillo *et al.*, 2010). Marine seaweeds contains many valuable components that could potentially be exploited as functional ingredients and therapeutic agents. (Gupta and Abu-Ghannam, 2011). Recently, researchers have described a wide range of biological activities for algal compounds including antibiotic, anti-inflammatory, anti-fungal anticancer, and antineoplastic (Ayyad *et al.*, 2003; Lincolon *et al.*, 1991). Seaweeds are of nutritional interest as they are low calorie food and are rich in vitamins, antioxidant enzymes, proteins, polyphenolic compounds, sulphated polysaccharides and dietary fibers (Burtin 2003; MacArtain *et al.*, 2007). Several *in vitro* studies have demonstrated that algal derived some

phytochemicals such as polyphenols and flavonoids showed antioxidant and antimicrobial activity (Chandini *et al.*, 2008; Zaragoza *et al.*, 2008; Abd El-Aty *et al.*, 2014). Phenolic compounds are diverse plant secondary metabolites comprised of aromatic rings, phenolic compounds from medicinal herbs and dietary plants include phenolic acids, flavonoids, tannins, stilbenes, coumarins, lignans, quinones, and others. Traditionally, many substances obtained from seaweeds are considered as a source of bioactive compounds which they have been used for decades in medicine and pharmacotherapy, whereas some of the isolated substances have bacteriostatic and bactericidal properties (Nair *et al.*, 2007; O'Sullivan 2010).

The extracts and active constituents of various marine algae have been shown to antimicrobial potential (Lima-Filho *et al.*, 2002; Paul *et al.*, 2006). The antimicrobial compounds derived from the marine algae consist of a diverse group of chemical compounds (Nor Afifah *et al.*, 2010). The ability of algae to produce antimicrobial substances could be used not only as a defense agent (against pathogens) but also as pharmaceutical bioactive natural compounds.

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Though much is known about the chemistry and the antimicrobial action of several phytochemicals, very few reports are available on the possible mechanism of action. Algal antimicrobial activity has been recognized based on existence of compounds belonging to numerous chemical classes including phenols, fatty acids, indoles, terpenes. Each of the various types of antibiotics kill microorganisms in a unique way. Some disturb the structure of the bacterial cell wall; others interfere with the production of essential proteins; and still others interfere with the transformation (metabolism) of nucleic acid (Flodin, and Whitfield 2000; Jin *et al.*, 2006). The aim of this study is to evaluate the antimicrobial activity of ethanol, acetone, and hexane extracts of *Padina boryana* Thivy and *Enteromorpha sp.* Besides, phenolic and flavonoid content were determined as well.

## MATERIALS AND METHODS

### Chemicals

Folin-ciocalteu reagent, gallic acid and ethanol were purchased from Merck Company (Darmstadt, Germany). All other chemicals and reagents such as aluminum chloride and sodium hydroxide were obtained from BDH, Dorset, UK. The HPLC grade organic solvents were obtained from Merck. All other chemicals were of analytical-grade purity

### Algae collection and extraction

Two marine algae were collected by hand picking from the Red Sea in Al-Leith Provence, Saudia during June 2015 (Fig. 1). Algal samples were cleaned from epiphytes, extraneous matter and necrotic were removed. Samples were washed thoroughly, air dried, cut into fine pieces and then ground in a tissue grinder until reach fine powder shape.



**Fig. 1:** (A). *Padina boryana* Thivy and (B). *Enteromorpha sp.*

The pulverized algal material (10 g) was macerated separately with (200 ml) of ethanol, acetone, and hexane at room

temperature for 48 h. After filtration through Whatman No.4 filter paper, algae residue was re-extracted twice with the same solvents. The pooled extracts were concentrated to remove the solvents completely from the extract. The dry crude extract was re-dissolved in 5 ml using the same solvents (Abd El -Aty *et al.*, 2014).

### Estimation of total phenolic content

Total phenolic (TP) contents were determined by the spectrophotometric method (Slinkard and Singleton, 1977). In brief, a 0.5 ml of each extract was made up to 3 ml with distilled water, and then mixed with 0.5 ml of Folin-Ciocalteu's phenol reagent. After 5 min, 2 ml of a 2 %  $\text{Na}_2\text{CO}_3$  solution were added to the mixture and thoroughly mixed. The mixture was kept at 30 °C for 60 min in dark place, and then the absorbance was recorded at 650 nm. The measurement was compared to a standard curve prepared with gallic acid solution. The total phenolic content was expressed as milligrams of gallic acid equivalents per gram of dried weight.

### Determination of total flavonoid content

Total flavonoid (TF) was determined by a colorimetric method as described by Zhishen *et al.*, (1999). A 0.5 ml of each extract was made up to 1 ml with methanol. Afterwards 0.4 ml of distilled water was added followed by 0.3 ml of 5 %  $\text{NaNO}_2$  solution and the mixture was left for 5 min.

Thereafter, 0.3 ml of (10%)  $\text{AlCl}_3$  solution was added and allowed to stand for 6 min. Two ml of (1 M) NaOH solution was added to the mixture and the final volume was adjusted to 10 ml with distilled water. The mixture was thoroughly shaken and allowed to stand for 15 min. Absorbance of the reaction mixture was read at 510 nm. The concentrations of total flavonoids were determined as quercetin equivalents (mg/g of dry weight).

### Antimicrobial activity

#### Microbial strains

The algal extracts were tested for their antimicrobial activity against seven Gram-negative strains (i.e. *Escherichia coli* ATCC 11229, *Moraxella catarrhalis* ATCC 25238, *Klebsiella oxytoca* ATCC 49131, *Klebsiella pneumoniae* ATCC 700603, *Neisseria gonorrhoeae* ATCC 49226, *Salmonella typhimurium* ATCC 14028, *Serratia sp.* ATCC 39006 and five Gram positive strains (i.e. Methicillin-resistant *Staphylococcus aureus* (MRSA) ATCC 43330, *Micrococcus luteus* ATCC 49732, *Staphylococcus epidermidis* ATCC 12228 *Enterococcus faecalis* ATCC 29212, *Staphylococcus aureus* ATCC 25923 and a pathogen fungus (i.e. *Candida neoformans* ATCC 90112).

The bacterial pathogens were maintained on Mueller Hinton Agar medium while *Candida neoformans* was maintained on Potato Dextrose Agar at 4 °C. Overnight nutrient broth subculture of the test organisms were done before use (Elbeshehy *et al.*, 2015).

### Agar well diffusion method

The antimicrobial activity of three algal crude extracts derived from *Padina boryana* Thivy and *Enteromorpha* sp were tested by using agar diffusion technique. The Mueller Hinton Agar for bacterial and PDA for fungal strain were poured into Petri plates then inoculated with microbial inocula (containing  $10^5$ - $10^6$  CFU/mL) that spread onto the agar surface of the plates using a sterile cotton swabs in order to get a uniform microbial growth on both control and test plates. Algal crude extracts were dissolved in dimethyl sulfoxide (DMSO) to a final concentration 1mg/mL, sterilized by filtration using sintered glass filter, and stored at 4°C. 6mm diameter wells were dug on the inoculated nutrient agar medium using sterilized cork borer and 50 µl of each extract were transferred into the wells. In each plate one well loaded with DMSO was used as a negative control. Bacterial strains were incubated at 37 °C overnight while plates inoculated with *C. neoformans* were incubated at 28°C for 48 h. After incubation, the diameter of inhibition zone of bacterial growth around the agar wells was calculated in mm.

### Minimum inhibitory concentration (MIC)

The standard micro-dilution method in 96-well plates from the Clinical Laboratory Standards Institute was carried out to evaluate the MIC. The effective algal extracts were selected and dissolved in water containing 4% DMSO and the initial extract concentration was 500 µg/ml. 50µL of each extract was serially diluted two-fold with MH broth (Difco, MD, USA), ranging the concentrations between 500 to 7.8 µg/ml. Each well was inoculated with (50 µL) of four sensitive tested bacterial strains at  $10^6$ - $10^7$  (CFU/ml). The plates were incubated overnight at 37 °C, and then the optical density at 600 nm was measured using a Bio-Rad Microplate Reader Model-550 (Bio-Rad, USA). Wells without inoculum added were used as controls, and positive controls (tetracycline and ciprofloxacin) were added to the inoculated growth medium without the substances. The MIC was recorded as the lowest concentration of antimicrobials extract with no visible growth of bacteria (Elazzazy *et al.*, 2015).

### Statistical analysis

All data are presented as means  $\pm$  SD; the mean values were calculated based on the data taken from at least three independent experiments conducted on separate days using freshly prepared reagents.

## RESULTS AND DISCUSSION

### Total phenolic (TP) and total flavonoid (TF) contents of two marine algae

Phenolic compounds in particular are considered as one of the most important classes of natural products. Phenolics act as antioxidants by inhibiting enzymes involved in radical generation and exhibited anticancer, antibacterial, anti-allergic, anti-diabetes, anti-aging, and anti-HIV activities, (Fresco *et al.*, 2006 and Li, *et al.*, 2011). Thus, the total phenolic and total flavonoid contents of

three different solvent extracts (ethanol, acetone and hexane) of *Padina boryana* Thivy and *Enteromorpha* were evaluated and the results are presented in table (1). The total phenolic contents ( $21.81 \pm 0.11$  and  $15.22 \pm 0.13$  mg/g gallic acid equivalent) were found to be higher in ethanol and acetone extracts of *Padina boryana* Thivy followed by ( $15.31 \pm 0.02$  and  $5.17 \pm 0.11$  mg/g gallic acid equivalent) in ethanol and acetone extracts of *Enteromorpha* sp., respectively. The ethanol and acetone solvents presented the highest values of total flavonoid contents ( $13.11 \pm 0.14$  and  $12.09 \pm 0.02$  mg/g quercetin equivalent) in *Padina boryana* Thivy and ( $6.13 \pm 0.01$  and  $4.91 \pm 0.08$  mg/g quercetin equivalent) in *Enteromorpha* sp., respectively (Table 1). In the present results *Padina boryana* Thivy showed TP and TF contents higher than *Enteromorpha* sp., and this difference perhaps could be attributed to genetic factors.

It was realized that within both algae species differences in TP and TF existed with the use of different solvents. This could be attributed to the polarity of the solvents (Hemalatha *et al.*, 2011). Ethanol was considered to be the best solvent for extraction of TP and TF. Flavonoids are widely group of natural compounds and also the most important natural phenolics. In most algae, phenols are important antioxidants because of their ability to scavenge free radicals such as singlet oxygen, superoxide and hydroxyl radicals (Shanab *et al.*, 2011). Results also show that, among all the solvent; ethanol and acetone were better solvents for effective extraction of phenolic compounds as compared to other solvent like hexane. This could be attributed to the polarity of the solvents (Hemalatha *et al.*, 2011). It is very important to point out that; there is a positive relationship between antimicrobial activity potential and amount of phenolic compounds of the crude extracts (Mohamed *et al.*, 2015).

### Antimicrobial activity:

#### *Padina boryana* Thivy and *Enteromorpha*

Seaweeds such as *Padina boryana* Thivy and *Enteromorpha* sp are broadly screened to isolate drugs or bioactive substances all over the world (Rao, 1991), they are able to produce a great variety of secondary metabolites characterized by a broad spectrum of antimicrobial activities (Cox *et al.*, 2010). Accordingly, the present study was focused to screen ethanol, acetone and hexane extracts of *Padina boryana* Thivy and *Enteromorpha* sp for the potential of antimicrobial activity on thirteen human pathogen as presented in Figures 2 and 3. Among the two marine algae screened *Padina boryana* Thivy was observed to be active than *Enteromorpha* sp. Three solvents tested, acetone exhibited more inhibitory effect followed by ethanol then hexane. For instance, the acetone extract of *Padina boryana* Thivy had strong antibacterial inhibition against gram negative bacteria *Neisseria gonorrhoeae* and *Moraxella catarrhalis* (22 and 21 mm, respectively) followed by gram positive *Methicillin-resistant Staphylococcus aureus* (20 mm). The ethanol extracts of *Padina boryana* Thivy and *Enteromorpha* sp showed a broad spectrum of antimicrobial activity against Gram (+) and Gram (-) bacteria in addition to Candida.

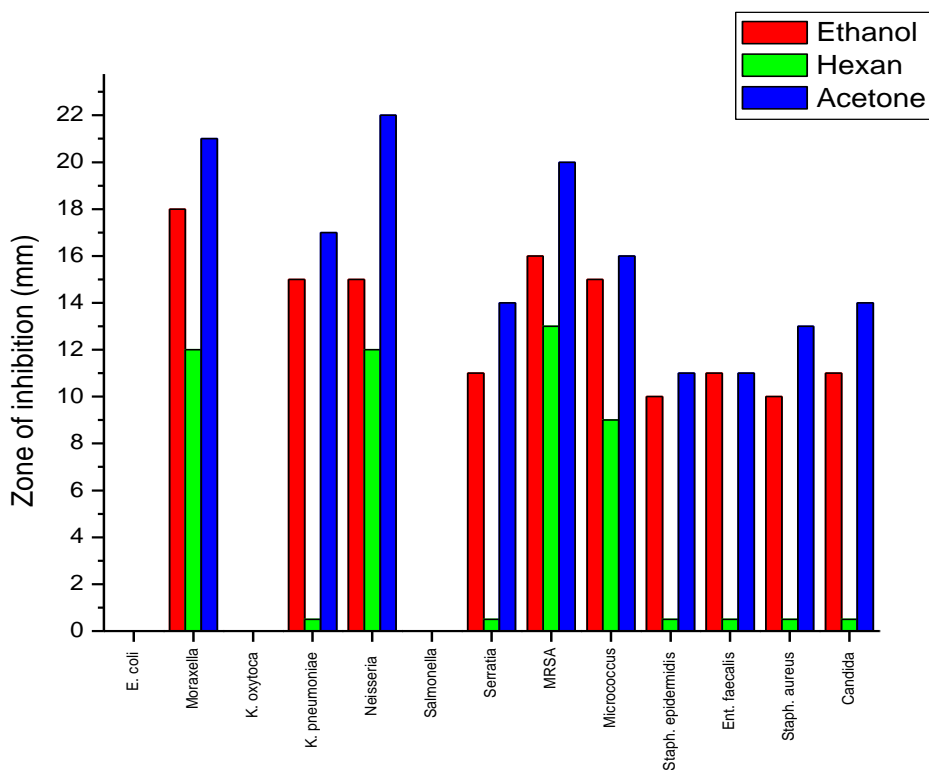


Fig. 2: Antimicrobial activity of *Padina boryana* Thivy against human pathogen. expressed by the inhibition zone (mm).

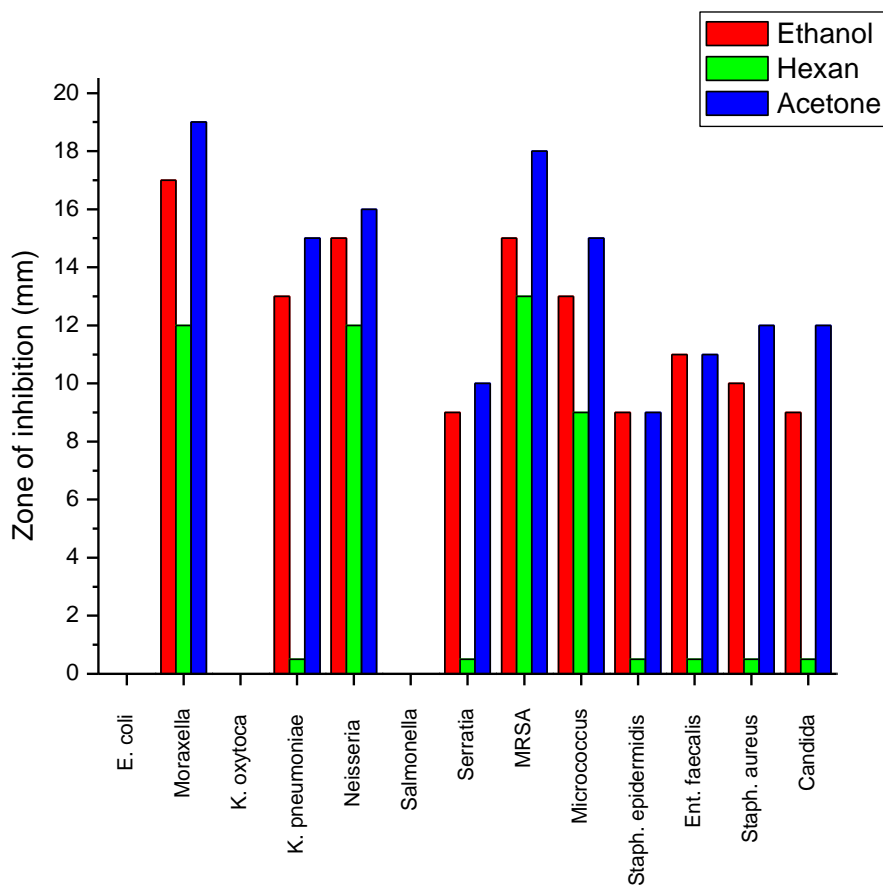


Fig. 3: Antimicrobial activity of *Enteromorpha sp* against human pathogen expressed by the inhibition zone (mm).

However, hexane extracts of the two algal strains were only effective against *MRSA*, *M. catarrhalis*, *N. gonorrhoeae* and *M. luteus* among the bacteria tested. All the three solvent extracts of the two algae were not displayed any activity against three bacterial strains *Salmonella*, *E. coli* and *Kleb. oxytoca*.

The solvent system used for the extraction played a major role in displaying the anti-bacterial activity. In the present study, among the different solvent extract, acetone and ethanol extract of *Padina boryana* Thivy and *Enteromorpha sp* displayed highest inhibitory activity against the test pathogens as compared to hexane. This result are parallel to the highest content of TP and TF in acetone and ethanol extract (Table 1), thus the antimicrobial activity of *Padina boryana* Thivy and *Enteromorpha sp* might be attributed to their phenolic and flavonoid content. In this concern, a positive relationship between antimicrobial activity potential and amount of phenolic compounds of the crude extracts was reported (Mohamed *et al.*, 2015).

**Table 1:** Total phenolic and flavonoid contents of *Padina boryana* Thivy and *Enteromorpha* in different solvent extracts.

Algal species	Solvents extract		
	Ethanol	Acetone	Hexane
<b>Total phenolic (mg/g d.wt)</b>			
<i>Padina boryana</i> Thivy	21.81 ±0.11	15.22±0.13	9.04±0.02
<i>Enteromorpha</i>	15.31±0.02	5.17±0.11	4.31±0.05
<b>Total flavonoids (mg/g d.wt)</b>			
<i>Padina boryana</i> Thivy	13.11±0.14	12.09±0.02	5.15±0.07
<i>Enteromorpha</i>	6.13±0.01	4.91±0.08	3.11±0.23

Ethanol was found to be the best solvent for extracting the active principles in almost all species of seaweeds (Rebecca *et al.*, 2013). Some studies concerning the effectiveness of extraction methods highlight that methanol extraction yields higher antimicrobial activity than n-hexane and ethyl acetate (Tuney *et al.*, 2006). It is clear that using organic solvents always provides a higher efficiency in extracting compounds for antimicrobial activities compared to water-based methods (Lima-Filho *et al.*, 2002). The minimum inhibitory concentration of *Padina boryana* Thivy and *Enteromorpha sp* were carried out using the acetone and ethanol extract, which showed the maximum activity, against most sensitive bacterial strains, as presented in Table 2. As the MIC value decreased the antibacterial increase. The acetone extract of *Padina boryana* Thivy and *Enteromorpha sp* showed highest activity against *M. catarrhalis*, *MRSA* and *N. gonorrhoeae* with the lowest MIC values (31, 62 and 125 µg/ml), respectively. Whereas, the ethanol extract of *Padina boryana* Thivy showed the lowest activity against *MRSA* and *N. gonorrhoeae* with MIC value (125 µg/ml), while, the ethanol extract of *Enteromorpha sp* exhibited lowest activity against *M. catarrhalis*, with MIC value (125µg/ml). Sahgal *et al.*, (2011) attributed the differences of the MIC value could be due to the morphological structure of the bacterial cells and their composition in the cells. The reduction in growth possibly occurred due to interference by active compounds in the extract (Beatrice *et al.*, 2010). Similarly, Lim *et al.*, (2011) and Darah *et al.*, (2013) reported that the higher concentration of

the extract was needed to kill the microorganism cells than to inhibit the growth of these cells on time-kill profile study.

**Table 2:** MIC (µg/ml) values of *Padina boryana* Thivy and *Enteromorpha sp* Against Human Pathogens in M-H Broth.

Pathogenic organisms	<i>Padina boryana</i> Thivy MIC (ug/ml)		<i>Enteromorpha sp</i> MIC (ug/ml)	
	Ethanol	Acetone	Ethanol	Acetone
<i>N. gonorrhoeae</i>	125±2	125±4	250±1	125±0
<i>MRSA</i>	125±1	62±3	500±2	<b>62±2</b>
<i>M. catarrhalis</i>	250±0	31±1	125±3	<b>31±4</b>
<i>Serratia sp</i>	500±0	250±0	500±3	250±3
<i>C. neoformans</i>	500±0	500±0	250±0	500±5
<i>Klebsiella pneumoniae</i>	250±0	250±0	500±0	500±0
<i>Micrococcus luteus</i>	250±0	250±0	500±0	500±0

## CONCLUSION

Seaweeds contained high levels of hydrophilic components, such as polyphenols which were easily extracted by polar solvents. Overall results of this study showed that ethanol and acetone solvents were effective for polyphenols extraction from *Padina boryana* Thivy and *Enteromorpha sp*. As well as the two extracts exhibited strong antimicrobial activity against the tested microorganisms. *MRSA*, *M. catarrhalis* and *N. gonorrhoeae* were the most sensitive bacterial strains for the acetone extract of *Padina boryana* Thivy and *Enteromorpha sp*. This suggests that algal polyphenols including flavonoids may be the principal constituents responsible for the antimicrobial properties of extracts from this species. These findings suggested that there may be a potential to utilize such seaweed extracts in food products to act as antimicrobial which would have promising applications in enhancing the food safety.

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