

Basic Study

Aldehyde dehydrogenase activity helps identify a subpopulation of murine adipose-derived stem cells with enhanced adipogenic and osteogenic differentiation potential

Harumichi Itoh, Shimpei Nishikawa, Tomoya Haraguchi, Yu Arikawa, Shotaro Eto, Masato Hiyama, Toshie Iseri, Yoshiki Itoh, Munekazu Nakaichi, Yusuke Sakai, Kenji Tani, Yasuho Taura, Kazuhito Itamoto

Harumichi Itoh, Shimpei Nishikawa, Tomoya Haraguchi, Yu Arikawa, Kazuhito Itamoto, Department of Small Animal Clinical Science, Joint Faculty of Veterinary Medicine, Yamaguchi University, Yamaguchi 753-8511, Japan

Shotaro Eto, Masato Hiyama, Kenji Tani, Yasuho Taura, Department of Veterinary Surgery, Joint Faculty of Veterinary Medicine, Yamaguchi University, Yamaguchi 753-8511, Japan

Toshie Iseri, Yoshiki Itoh, Munekazu Nakaichi, Department of Veterinary Radiology, Joint Faculty of Veterinary Medicine, Yamaguchi University, Yamaguchi 753-8511, Japan

Yusuke Sakai, Department of Veterinary Pathology, Joint Faculty of Veterinary Medicine, Yamaguchi University, Yamaguchi 753-8511, Japan

ORCID number: Harumichi Itoh (0000-0003-1957-0129); Shimpei Nishikawa (0000-0002-7788-9376); Tomoya Haraguchi (0000-0003-4692-9264); Yu Arikawa (0000-0002-7824-6356); Shotaro Eto (0000-0002-7183-1528); Masato Hiyama (0000-0001-7117-0355); Toshie Iseri (0000-0003-0306-9639); Yoshiki Itoh (0000-0002-8129-448X); Munekazu Nakaichi (0000-0002-6274-7228); Yusuke Sakai (0000-0003-0531-8256); Kenji Tani (0000-0001-5822-710X); Yasuho Taura (0000-0002-2946-0241); Kazuhito Itamoto (0000-0002-6820-8903).

Author contributions: Itoh H performed the majority of experiments and wrote the paper; Itoh H, Arikawa Y, Eto S, Iseri T and Itoh Y contributed for data analysis and acquisition of unpublished essential data and reagents; Itoh H, Nishikawa S, Haraguchi T, Nakaichi M, Tani K, Taura Y and Itamoto K designed research and analyzed data; all the authors contributed to this article.

Supported by JSPS KAKENHI (Grants-in-Aid for Scientific Research) for Nishikawa S, No. 26893172.

Institutional review board statement: The study was performed

without any human sample or any kind of human patient involvement.

Institutional animal care and use committee statement: The animal experiments were approved by the institutional animal experiment ethics committee of Yamaguchi University (Approval No. 245).

Conflict-of-interest statement: Authors have no conflicts of interest.

Data sharing statement: Data are available from the corresponding author (sn2007@yamaguchi-u.ac.jp).

Open-Access: This article is an open-access article which was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution Non Commercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Unsolicited manuscript

Correspondence to: Shimpei Nishikawa, BvetMed, PhD, Department of Small Animal Clinical Science, Joint Faculty of Veterinary Medicine, Yamaguchi University, 1677-1 Yoshida, Yamaguchi 753-8511, Japan. sn2007@yamaguchi-u.ac.jp
Telephone: +81-83-9335931
Fax: +81-83-9335878

Received: April 24, 2017

Peer-review started: April 26, 2017

First decision: June 6, 2017

Revised: August 28, 2017

Accepted: September 12, 2017

Abstract

AIM

To identify and characterize functionally distinct subpopulation of adipose-derived stem cells (ADSCs).

METHODS

ADSCs cultured from mouse subcutaneous adipose tissue were sorted fluorescence-activated cell sorter based on aldehyde dehydrogenase (ALDH) activity, a widely used stem cell marker. Differentiation potentials were analyzed by utilizing immunocytofluorescence and its quantitative analysis.

RESULTS

Approximately 15% of bulk ADSCs showed high ALDH activity in flow cytometric analysis. Although significant difference was not seen in proliferation capacity, the adipogenic and osteogenic differentiation capacity was higher in ALDH^{hi} subpopulations than in ALDH^{lo}. Gene set enrichment analysis revealed that ribosome-related gene sets were enriched in the ALDH^{hi} subpopulation.

CONCLUSION

High ALDH activity is a useful marker for identifying functionally different subpopulations in murine ADSCs. Additionally, we suggested the importance of ribosome for differentiation of ADSCs by gene set enrichment analysis.

Key words: Adipose-derived stem/stromal cell; Aldehyde dehydrogenase activity; Flow cytometry; Subpopulation; Ribosome

© **The Author(s) 2017.** Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: Aldehyde dehydrogenase (ALDH) activity is widely used as a stem cell marker in several types of normal or malignant tissues. However, there was no report of ALDH activity in murine adipose-derived stem cells (ADSCs). Here, our study demonstrated a subpopulation defined by high ALDH activity within murine ADSCs. The subpopulation with high ALDH activity (ALDH^{hi}) showed enhanced differentiation potentials into adipocyte and osteocyte. Furthermore, gene set enrichment analysis revealed that ribosome-related gene sets were enriched in ALDH^{hi} of murine ADSCs. We showed relationship between ALDH^{hi} and ribosome biosynthesis, providing a novel insight of mesenchymal stem cell biology.

Itoh H, Nishikawa S, Haraguchi T, Arikawa Y, Eto S, Hiyama M, Iseri T, Itoh Y, Nakaichi M, Sakai Y, Tani K, Taura Y, Itamoto K. Aldehyde dehydrogenase activity helps identify a subpopulation of murine adipose-derived stem cells with enhanced adipogenic and osteogenic differentiation potential. *World J Stem Cells* 2017; 9(10): 179-186 Available from: URL: <http://www.wjgnet.com/1948-0210/>

INTRODUCTION

Stem cells can self-renew and differentiate into specialized cells of various tissues^[1]. Therefore, these cells, for example, embryonic stem cells (ESCs), induced pluripotent stem cells (iPSCs), hematopoietic stem cells, and mesenchymal stem cells (MSCs), have been the object of basic research and clinical applications. Among these types of stem cells, MSCs, as represented by adipose-derived stem/stromal cells (ADSCs) and bone marrow-derived stem/stromal cells (BMSCs), have been recognized as useful material for cell-based therapy^[2]. MSCs have been isolated from various tissues, including adipose tissue, the bone marrow, peripheral blood, cord blood, the liver, dental pulp, and fetal tissue; of these, adipose tissue is one of the most abundant source of MSCs^[3]. ADSCs possess multipotency and have the potential to differentiate into cell types such as adipocytes, osteocytes, chondrocytes, neurons, vascular endothelial cells, cardiomyocytes, myoblasts, and islet β -cells under appropriate conditions^[4].

The researches have suggested that ADSCs are heterogeneous and comprise phenotypically and/or functionally different subpopulations^[5-7]. For example, the cluster of differentiation (CD)73⁺ subpopulation of murine ADSCs possesses increased potential for cardiomyocyte differentiation^[6]. The CD90⁺ subpopulation of murine ADSCs has higher tube-forming ability than the CD90⁻ subpopulation, which has high adipogenic potential^[8]. The CD90⁺ subpopulation also exhibits higher efficiency of iPSC induction than the CD90⁻ subpopulation^[5]. Human ADSCs also include the CD105^{lo} subpopulation, which has high osteogenic potential^[7]. Some studies have identified different subpopulations in ADSCs on the basis of surface antigen markers^[5-7]. However, it is unclear how these markers (*e.g.*, CD90 and CD105) are functionally related to cell differentiation.

In mice, aldehyde dehydrogenase (ALDH) is a superfamily comprising 20 intracellular enzymes and is responsible for the oxidization of various aldehydes^[9]. High ALDH activity has been shown in normal hematopoietic stem cells, neural stem cells, and cancer stem cells in various types of neoplastic diseases^[10]. Therefore, high ALDH activity is considered to be a common marker for normal and malignant stem cells. In human ADSCs, however, only one study has been performed on the ALDH^{hi} subpopulation, whose significance in differentiation potential is unclear^[11]. Moreover, to our knowledge, the existence of the ALDH^{hi} subpopulation within murine ADSCs has not yet been reported.

In the current study, the ALDH^{hi} and ALDH^{lo} subpopulations of murine ADSCs were sorted using flow cytometry. The differentiation potential and proliferation of the sorted ALDH^{hi} and ALDH^{lo} subpopulations were analyzed. Furthermore, we analyzed the transcriptional profiles of the ALDH^{hi} and ALDH^{lo} subpopulations by utilizing gene set enrichment analysis (GSEA).

MATERIALS AND METHODS

Animals and ADSC isolation

C57BL/6J mice were purchased from Kyudo Co., Ltd (Saga, Japan). All animal experiments were carried out in accordance with the National Institutes of Health guide for the care and use of Laboratory animals (NIH Publications No. 8023, revised 1978) and the institutional guidelines of Yamaguchi University. The animal experiments were approved by the institutional animal experiment ethics committee of Yamaguchi University.

Murine ADSCs were isolated from twenty of C57BL/6J female mice of 4- to 6-wk-old, as previously described^[8]. Briefly, the subcutaneous adipose tissue was resected, washed with Dulbecco's phosphate-buffered saline (DPBS; Wako, Osaka, Japan), and cut into small pieces. The adipose tissue pieces were digested in high glucose Dulbecco's modified Eagle's medium (DMEM; Wako, Osaka, Japan) containing 1.0 mg/mL collagenase type I (Sigma-Aldrich, St. Louis, MO, United States), 10% fetal bovine serum (FBS; Sigma-Aldrich), and antibiotic-antimycotic agents (PSM; penicillin: 100 U/mL, streptomycin: 100 µg/mL, and amphotericin B: 0.25 µg/mL, final concentrations; Nacalai Tesque, Kyoto, Japan), using a shaking incubator at 37.5 °C and 250 rpm for 1 h. The digested tissue was filtered through a sterile ø100-µm nylon mesh (EASYstrainer, 100 µm; Greiner Bio-One Japan, Tokyo, Japan), followed by centrifugation at 400 × *g* for 5 min in DPBS supplemented with 1% FBS and 1 mmol/L EDTA-3Na (Wako, Osaka, Japan). The pellet was resuspended in DMEM supplemented with 10% FBS and antibacterial/antimycotic agent and was cultured at 37.0 °C in a 5% CO₂ atmosphere, using ø10 cm dish (Corning, NY, United States). When the cultures reached 80%-90% confluence, the ADSCs were dissociated from the dish by using Accutase solution (Innovative Cell Technologies, San Diego, United States), and seeded into new dishes.

Flow cytometry analysis

Adherent ADSCs from passage 4 were dissociated using Accutase solution; 1×10^6 cells were resuspended and incubated for 5 min on ice with 2 µL of anti-mouse CD16/32 rat monoclonal antibody (BioLegend, San Diego, CA, United States). Cells were stained with 1 µL viability probe (Zombie NIR, Biolegend) for 20 min at room temperature to stain dead cells. ALDH activity was assessed by utilizing the ALDEFUOR kit (Stemcell Technologies, Vancouver, Canada) according to the manufacturer's instructions. Briefly, 1×10^6 cells were resuspended in 1 mL assay buffer and 5 µL ALDEFUOR reagent was added after thorough mixing; then, 0.5 mL of the cell suspension was transferred to a new tube with 5 µL diethylaminobenzaldehyde (DEAB) reagent (ALDH inhibitor) for negative control of ALDH activity. Flow cytometric analysis and cell sorting were performed using Accuri C6 (BD Bioscience, San Jose, CA, United States) and the SH800 cell sorter (Sony, Tokyo, Japan). Flow cytometric data were analyzed with the FlowJo (Tree Star,

Ashland, OR, United States) software.

Measurement of proliferation potential

To assess the viability of the ADSC subpopulations, we used a cell WST-8 assay (Cell Counting Kit-8; Dojindo Laboratories, Kumamoto, Japan) according to the manufacturer's instructions. Briefly, sorted ALDH^{hi} or ALDH^{lo} murine ADSCs were seeded in 96-well plates at a density of 3×10^3 cells/well. After 12, 24, 48, and 72 h, 100 µL fresh medium containing 10 µL CCK-8 solution was added to each well, followed by incubation at 37 °C for 1 h. The absorbance of each well at 450 nm was measured using an Epoch microplate spectrophotometer (BioTek Instruments, Winooski, VT, United States). Six replicates were prepared for each group.

Cell differentiation and immunofluorescence staining

The adipogenic and osteogenic differentiations of ADSCs were characterized using a Mouse Mesenchymal Stem Cell Functional Identification Kit (R and D Systems, Minneapolis, MN, United States) according to the manufacturer's instructions. Briefly, for adipogenic differentiation, cells (3×10^3 /well) were cultured at 37 °C in a 5% CO₂ atmosphere in a 96-well plate in 100 µL adipogenic differentiation medium composed of α -minimal essential medium (α MEM) supplemented with 10% FBS, 1% PSM, L-glutamine, and 50 µL adipogenic supplement containing hydrocortisone, isobutylmethylxanthine, and indomethacin for 15 d in 37 °C and a 5% CO₂ atmosphere.

For osteogenic differentiation, cells were cultured in osteogenic differentiation medium composed of 5 mL α -MEM basal medium and 250 µL osteogenic supplement containing ascorbate-phosphate, β -glycerolphosphate, and recombinant human bone morphogenetic protein-2 for 15 d in 37 °C and a 5% CO₂ atmosphere. The medium was replaced every 2-3 d.

To assess adipogenic and osteogenic differentiation by immunocytochemistry, cultured cells were fixed in 4% paraformaldehyde phosphate buffer solution (Wako, Osaka, Japan) for 20 min. After the cells were washed with DPBS, they were permeabilized and blocked with DPBS supplemented with 0.3% Triton X-100 (Sigma-Aldrich), and 10% FBS for 45 min. The cells were subsequently incubated for 1 h in DPBS containing 10 µg/mL goat anti-mouse fatty acid binding protein (FABP) 4 polyclonal antibody to label adipocytes or were incubated with 10 µg/mL goat anti-mouse osteopontin polyclonal antibody to label osteocytes. They were then washed with DPBS and incubated for 1 h in DPBS containing phycoerythrin (PE)-conjugated rabbit anti-goat IgG antibody [rabbit F(ab')₂ anti-goat IgG-H and L (PE), pre-adsorbed, Abcam, Cambridge, United Kingdom]. Nuclei were stained with 5 µg/mL Hoechst 33342 (Dojindo Laboratories, Kumamoto, Japan). Photographs were obtained and analyzed using a fluorescence microscope (BZ-9000; Keyence, Osaka, Japan) and its analysis software.

Gene expression array analysis and GSEA

Gene expression array analysis and GSEA were performed

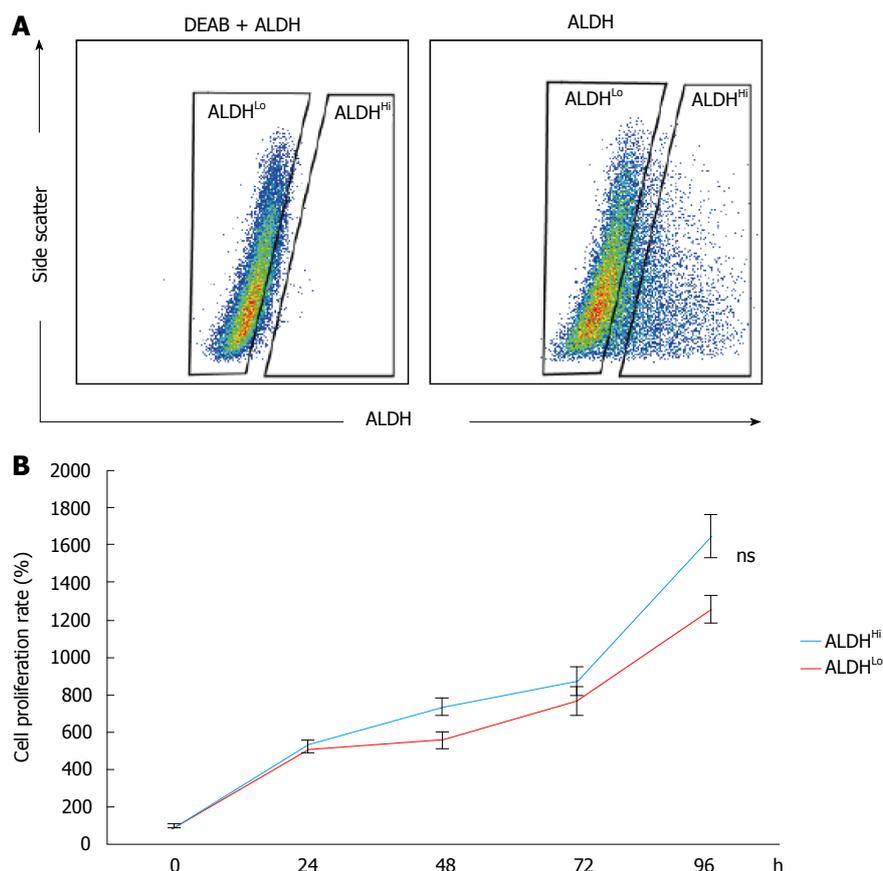


Figure 1 Detection of aldehyde dehydrogenase-positive subpopulations of murine adipose-derived stem cells and evaluation of proliferation rates. A: Flow cytometric analysis of murine ADSCs. Baseline fluorescence was established by adding the ALDH inhibitor diethylaminobenzaldehyde; B: Cell proliferation rates were not significantly different between the ALDH^{Hi} and ALDH^{Lo} subpopulations. Values have been expressed in terms of mean \pm SE ($n = 5$). ns: Not significant; ALDH: Aldehyde dehydrogenase; ADSCs: Adipose-derived stem cells; DEAB: Diethylaminobenzaldehyde.

on the published gene expression profile of C57BL/6 mice divided by ALDH^{Hi} and ALDH^{Lo} subpopulations of ADSCs. About 3×10^6 cells from each subpopulation were lysed and total RNA was isolated using the RNeasy Mini Kit (Qiagen, Hilden, Germany). Cyanine-3 (Cy3)-labeled cRNA was prepared from 0.1 μ g total RNA by using the Low Input Quick Amp Labeling Kit (Agilent Technologies, Santa Clara, CA, United States) according to the manufacturer's instructions; this was followed by RNeasy column purification (Qiagen). Dye incorporation and cRNA yield were checked with the NanoDrop ND-2000 Spectrophotometer (Thermo Fisher Scientific, Waltham, MA, United States). Cy3-labelled cRNA (0.6 μ g) was fragmented at 60 °C for 30 min in a reaction volume of 25 μ L containing 1 \times Agilent fragmentation buffer and 2 \times Agilent blocking agent following the manufacturer's instructions. On completion of the fragmentation reaction, 25 μ L of 2 \times Agilent hybridization buffer was added to the fragmentation mixture and hybridized to SurePrint G3 Mouse GE 8 \times 60 K Ver1.0 (Agilent Technologies) for 17 h at 65 °C in a rotating Agilent hybridization oven. After hybridization, the microarrays were washed for 1 min at room temperature with GE Wash Buffer 1 (Agilent Technologies) and 1 min with 37 °C GE Wash buffer 2 (Agilent Technologies). The slides were scanned immediately after washing on the Agilent SureScan

Microarray Scanner (G2600D), using one color scan setting for 8 \times 60 k array slides (scan area, 61 \times 21.6 mm; scan resolution, 3 μ m; dye channel set for Green PMT was set to 100%). The scanned images were analyzed with Feature Extraction Software 11.5.1.1 (Agilent Technologies), using default parameters to obtain the subtracted background and spatially detrended Processed Signal intensities.

Statistical analysis

Statistical analysis was performed using GraphPad Prism version 6.01 for Windows (GraphPad Software, La Jolla, CA, United States). The results have been expressed in terms of mean \pm SE. Comparisons of two groups were performed with the independent *t*-test. Multiple comparisons were performed with one-way analysis of variance. Data were considered statistically significant when the *P* value was ≤ 0.05 .

RESULTS

ALDH activity of murine ADSCs

To identify the subpopulation defined by ALDH activity in murine ADSCs, single-cell suspensions of cultured murine ADSCs were stained using the ALDEFUOR kit and analyzed with flow cytometry. A small subpopulation

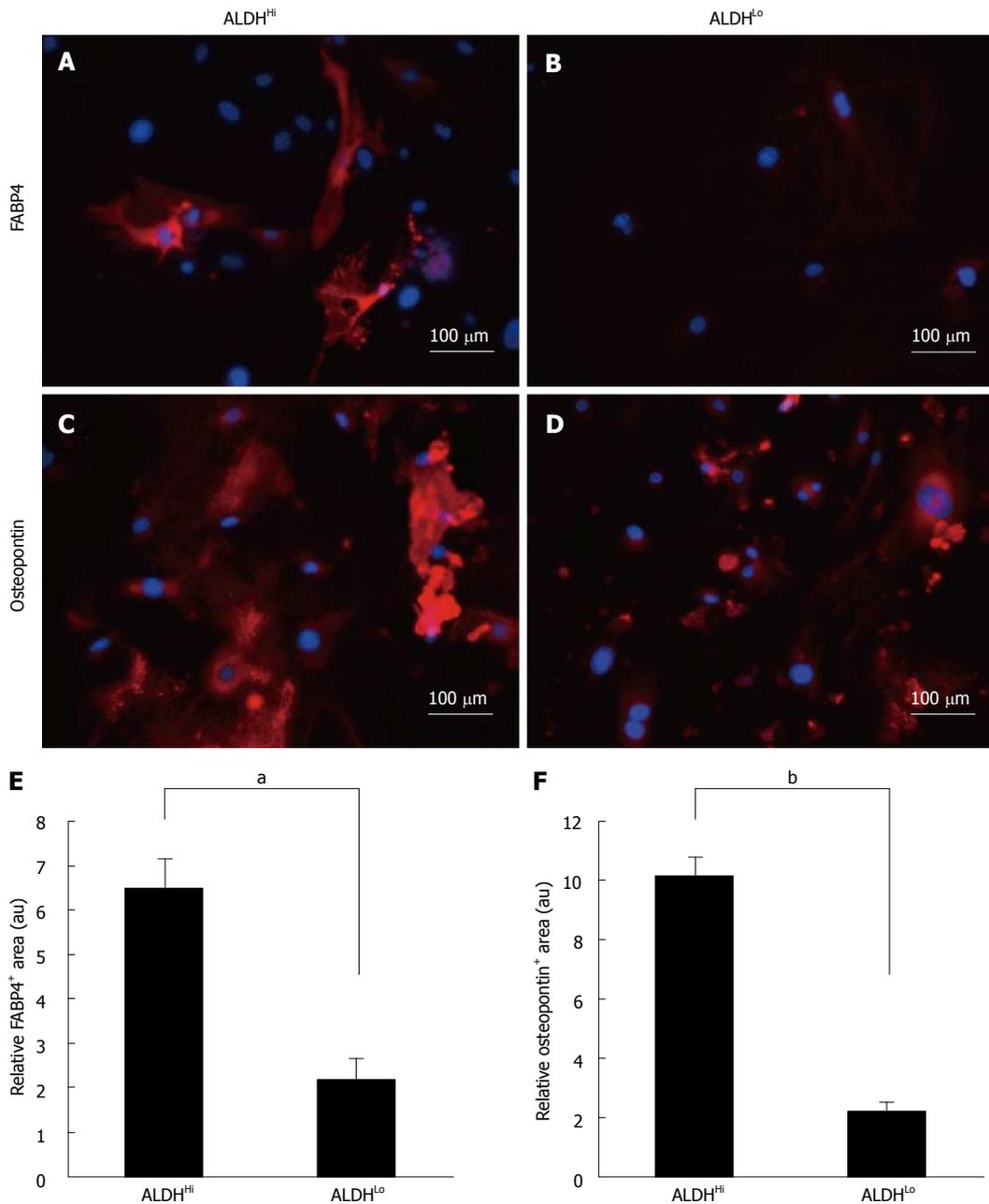


Figure 2 Marker analysis of differentiation potentials. Differentiation potential of ALDH^{hi} (A, C) and ALDH^{lo} (B, D) subpopulations of ADSCs. FABP4 (A, B) and osteopontin (C, D) expression in ADSCs (red) following adipogenic (A, B) and osteogenic (C, D) differentiation of ALDH^{hi} and ALDH^{lo} subpopulations, as determined by immunocytochemistry. The nuclei were stained with Hoechst 33342 (blue). Quantitative analysis of differentiation-marker-positive areas in differentiated ADSCs. FABP4-positive (E) and osteopontin-positive (F) area ratios relative to respective areas of nuclear staining for ALDH^{hi} and ALDH^{lo} subpopulations of ADSCs after adipogenic (E) and osteogenic (F) induction. Values have been expressed in terms of mean \pm SE ($n = 5$). ^a $P < 0.05$, ^b $P < 0.0001$, au: Arbitrary unit. ALDH: Aldehyde dehydrogenase; ADSCs: Adipose-derived stem cells; FABP: Fatty acid binding protein.

with distinctively high ALDH activity (ALDH^{hi} cells) was detected within the bulk populations of ADSCs (Figure 1A). The percentage of ALDH^{hi} cells was approximately 15% of the bulk murine ADSC population (Figure 1A). However, on adding the ALDH inhibitor *N,N*-diethylaminobenzaldehyde (DEAB), a distinct ALDH^{hi} subpopulation was not detected (Figure 1A). To assess the difference in the proliferation potentials of the ALDH^{hi} and ALDH^{lo} subpopulations, we measured the proliferation rate of each subpopulation by using the WST assay. The proliferation potential of the ALDH^{hi} subpopulation of

ADSCs was not significantly different compared to the ALDH^{lo} subpopulation (Figure 1B).

Cell differentiation to adipocytes and osteocytes

To assess the adipogenic and osteogenic differentiation potential of the two subpopulations, sorted ALDH^{hi} and ALDH^{lo} cells were cultured under adipogenic or osteogenic differentiation conditions. After *in vitro* differentiation, immunofluorescence staining for FABP4 (marker of adipocytes) and immunofluorescence staining for osteopontin (marker of osteocytes) were performed

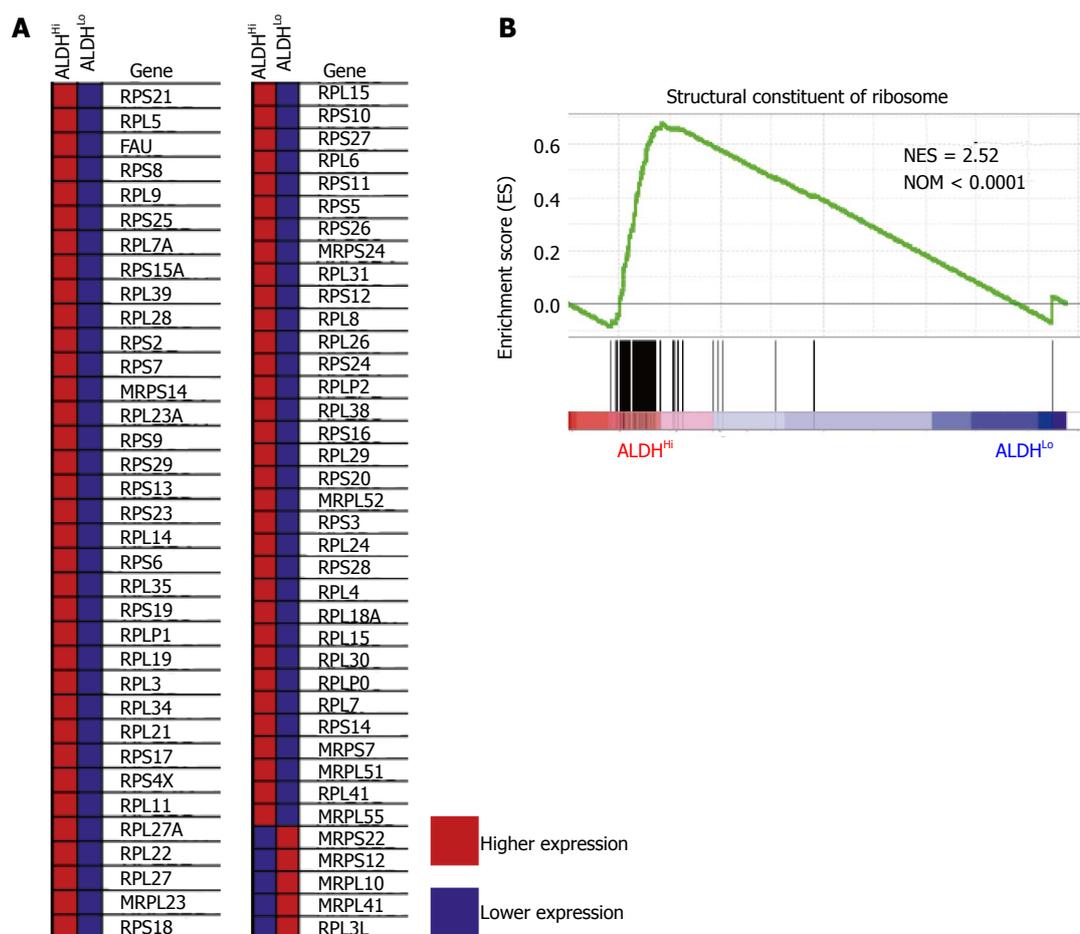


Figure 3 Gene set enrichment analysis of each ALDH^{hi} and ALDH^{lo} subpopulation. A: Heat map of enrichment profile of ribosomal protein mRNAs for ALDH^{hi} and ALDH^{lo} subpopulations; B: Gene set enrichment analysis of transcription data related to structural constituents of ribosomes for the ALDH^{hi} and ALDH^{lo} subpopulations. Normalized enrichment score and nominal *P* values are shown. ALDH: Aldehyde dehydrogenase.

(Figure 2A-D). ADSCs that differentiated into adipocytes appeared as accumulated lipid droplets in the cytosol in each ALDH^{hi} and ALDH^{lo} subpopulation (Figure 2A and B). Furthermore, immunofluorescence staining for osteopontin revealed that ADSCs that differentiated into osteocytes appeared as accumulated granules in the cytosol in each ALDH^{hi} and ALDH^{lo} subpopulation (Figure 2C and D).

Evaluation of differentiation

Adipogenic and osteogenic differentiation of each ALDH^{hi} and ALDH^{lo} subpopulation was quantitatively assessed using the BZ-9000 microscope and its analysis software. Ten visual fields were taken randomly for every 3 wells, and the immunofluorescence-staining positive-areas in 30 visual fields were analyzed. Subsequently, the immunofluorescence-staining-positive area was divided by the Hoechst 33342-positive area for each of the 30 visual fields. The ALDH^{hi} subpopulation was found to have significantly more adipogenic and osteogenic relative-differentiation-marker-positive areas than the ALDH^{lo} subpopulation (Figure 2E and F).

GSEA

To identify the sets of gene that were up- or down-

regulated in the ALDH^{hi} subpopulations, we performed GSEA for the published gene expression profile of C57BL/6 mice divided by the ALDH^{hi} and ALDH^{lo} subpopulations of ADSCs. Intriguingly, high gene set enrichment scores were obtained for the structural constituents of ribosomes (Figure 3).

DISCUSSION

MSCs are reported to commonly express CD29, CD73, CD90, and CD105 and to be negative for markers such as CD45 and CD56^[12,13]. There have been many studies on cell surface antigen markers of ADSCs, such as CD34 and CD44^[12,14,15]. Recently, however, studies have shown that some markers such as CD90 or CD105 are not expressed homogeneously in bulk ADSC populations but are expressed in small ADSC subpopulations, suggesting that ADSCs are phenotypically heterogeneous^[5,7,8]. In our current study, we detected ALDH activity as a stem cell marker in murine ADSCs. High ALDH activity has been reported as a marker for cells such as hematopoietic stem cells and cancer stem cells^[10]. However, not many studies have been performed on ALDH activity and cell differentiation potential in MSCs. In one of these studies, Estes *et al.*^[11] showed the presence of a subpopulation

with high ALDH activity in human ADSCs; however, no difference was found in terms of differentiation potential. In our present study, the cultured murine bulk ADSC population contained approximately 15% of the ALDH^{hi} subpopulation. Additionally, in the induction experiment for adipogenic and osteogenic differentiation for each sorted ALDH^{hi} and ALDH^{Lo} subpopulation, significantly higher adipogenic and osteogenic potentials were found in the ALDH^{hi} subpopulation. The ALDH^{hi} subpopulation had higher cell differentiation potential than the ALDH^{Lo} subpopulation. To the best of our knowledge, this is the first report on the functionally distinguishable subpopulation defined by ALDH activity within murine ADSCs.

Relationships between ribosome biogenesis and stem cells have been described only recently. For example, it was reported that the transition from self-renewal to differentiation depends on the enhancement of ribosome biogenesis accompanied by increased protein synthesis in female *Drosophila* germline stem cells^[16]. Slow growth, low biosynthesis and markedly reduced ribosome biogenesis were observed in hematopoietic stem cells that lacked RUNX1, which is known to promote the transcription of essential ribosome-related proteins^[17]. We have few reports about relationship between ribosome biogenesis and MSCs. One of these reports presented one of core proteins of 60S ribosome is necessary for differentiation of osteocyte from MSCs^[18]. In our current study, GSEA revealed the significant enrichment of ribosome-related genes in the ALDH^{hi} subpopulation compared to that in the ALDH^{Lo} subpopulation, suggesting that ribosome biogenesis is part of the mechanism underlying the higher differentiation potential of the ALDH^{hi} subpopulation.

ADSCs can be obtained in a less invasive manner from adipose tissue. Therefore, ADSCs are considered to be a promising source of cell-based therapy in the clinical setting. ADSCs have already been used in clinical studies for cardiovascular disease, breast reconstruction after mastectomy, spinal cord injury, cirrhosis, renal insufficiency, skin fistula after surgery, and skin fistula with Crohn's disease^[4,19,20]. Some of those trials reported the therapy to be safe and effective; however, there is obvious room for improvement. For instance, in a phase 3 trial for therapy with allogeneic expanded ADSCs for treatment-refractory complex perianal fistulas in patients with Crohn's disease (ADMIRE-CD trial), approximately 50% of patients who received ADSC-therapy experienced remissions^[21]. Although this is a significant achievement, further research and development are required in relation to the patients who did not respond to this trial.

Purification of specific subpopulations and engineering of ADSCs into cells that are highly efficient in differentiating into specific tissues might help obtain basic knowledge for cell-based therapy, which is more specific to individual disease conditions of each organ for which ADSCs are used. Further investigation is required to identify the underlying mechanisms that regulate ribosome biogenesis and differentiation in ALDH^{hi} ADSCs.

In conclusion, we demonstrated that murine ADSCs

have a distinct subpopulation defined by ALDH activity. Furthermore, the ALDH^{hi} subpopulation had higher osteogenic and adipogenic differentiation potential than the ALDH^{Lo} subpopulation. Ribosome biosynthesis is suggested to be a remarkable difference between ALDH^{hi} and ALDH^{Lo} subpopulations.

COMMENTS

Background

Adipose-derived stem cells (ADSCs) are recognized as useful materials for regenerative therapy. Recent study revealed the existence of subpopulations in ADSCs by surface antigen markers. However, functions of these markers remain elusive. Aldehyde dehydrogenase (ALDH) activity is commonly used as functional marker to identify human and mouse hematopoietic stem cell, though there has been no report about identification of a subpopulation(s) in murine ADSCs using ALDH.

Research frontiers

Several surface antigen markers are reported to be capable of prospectively identifying distinct ADSCs subpopulations in human and murine. However, the function(s) of those reported markers are poorly understood. ALDH has its known function, such as a protective effect to hematopoietic stem cells through acetaldehyde detoxification, although it is not known in ADSCs.

Innovations and breakthroughs

The authors suggest a novel area of research consisted of ALDH, stem cell, and ribosome biosynthesis, by reporting here ALDH^{hi} murine ADSCs are highly capable of differentiation, and have enriched ribosome-related gene sets.

Applications

The authors current findings of ALDH^{hi} subpopulation of ADSCs might provide future application for enrichment of more useful cells which is applicable to an efficient cell-based therapy. Moreover, by elucidating mechanisms of the higher differentiation potentials shown in ALDH^{hi} subpopulation of ADSCs might provide knowledge of a key regulator(s) of differentiation, and links between ribosome biosynthesis.

Terminology

ADSCs: Adipose-derived stem cells can be obtained from adipose tissues and induced to differentiate into adipocytes, osteocytes and chondrocytes; ALDH: ALDH is a superfamily comprising 20 intracellular enzymes and is responsible for the oxidization of various aldehydes. Some reports identified ALDH is a marker that detect hematopoietic stem cells and cancer stem cells; GSEA: Gene set enrichment analysis is a comprehensive analysis of gene expression by a computational method.

Peer-review

This is a very interesting and well executed piece of work, with suitable controls.

ACKNOWLEDGMENTS

The authors thank the staff of Yamaguchi University Animal Medical Center for their support.

REFERENCES

- 1 **Li L, Xie T.** Stem cell niche: structure and function. *Annu Rev Cell Dev Biol* 2005; **21**: 605-631 [PMID: 16212509 DOI: 10.1146/annurev.cellbio.21.012704.131525]
- 2 **Keating A.** Mesenchymal stromal cells: new directions. *Cell Stem Cell* 2012; **10**: 709-716 [PMID: 22704511 DOI: 10.1016/j.stem.2012.05.015]

- 3 **Levi B**, Longaker MT. Concise review: adipose-derived stromal cells for skeletal regenerative medicine. *Stem Cells* 2011; **29**: 576-582 [PMID: 21305671 DOI: 10.1002/stem.612]
- 4 **Konno M**, Hamabe A, Hasegawa S, Ogawa H, Fukusumi T, Nishikawa S, Ohta K, Kano Y, Ozaki M, Noguchi Y, Sakai D, Kudoh T, Kawamoto K, Eguchi H, Satoh T, Tanemura M, Nagano H, Doki Y, Mori M, Ishii H. Adipose-derived mesenchymal stem cells and regenerative medicine. *Dev Growth Differ* 2013; **55**: 309-318 [PMID: 23452121 DOI: 10.1111/dgd.12049]
- 5 **Kawamoto K**, Konno M, Nagano H, Nishikawa S, Tomimaru Y, Akita H, Hama N, Wada H, Kobayashi S, Eguchi H, Tanemura M, Ito T, Doki Y, Mori M, Ishii H. CD90- (Thy-1-) high selection enhances reprogramming capacity of murine adipose-derived mesenchymal stem cells. *Dis Markers* 2013; **35**: 573-579 [PMID: 24282338 DOI: 10.1155/2013/392578]
- 6 **Li Q**, Qi LJ, Guo ZK, Li H, Zuo HB, Li NN. CD73+ adipose-derived mesenchymal stem cells possess higher potential to differentiate into cardiomyocytes in vitro. *J Mol Histol* 2013; **44**: 411-422 [PMID: 23456425 DOI: 10.1007/s10735-013-9492-9]
- 7 **Yamamoto M**, Nakata H, Hao J, Chou J, Kasugai S, Kuroda S. Osteogenic Potential of Mouse Adipose-Derived Stem Cells Sorted for CD90 and CD105 In Vitro. *Stem Cells Int* 2014; **2014**: 576358 [PMID: 25302065 DOI: 10.1155/2014/576358]
- 8 **Takahashi H**, Haraguchi N, Nishikawa S, Miyazaki S, Suzuki Y, Mizushima T, Nishimura J, Takemasa I, Yamamoto H, Mimori K, Ishii H, Doki Y, Mori M. Biological and clinical availability of adipose-derived stem cells for pelvic dead space repair. *Stem Cells Transl Med* 2012; **1**: 803-810 [PMID: 23197692 DOI: 10.5966/sctm.2012-0085]
- 9 **Marchitti SA**, Brocker C, Stagos D, Vasilioi V. Non-P450 aldehyde oxidizing enzymes: the aldehyde dehydrogenase superfamily. *Expert Opin Drug Metab Toxicol* 2008; **4**: 697-720 [PMID: 18611112 DOI: 10.1517/17425255.4.6.697]
- 10 **Balber AE**. Concise review: aldehyde dehydrogenase bright stem and progenitor cell populations from normal tissues: characteristics, activities, and emerging uses in regenerative medicine. *Stem Cells* 2011; **29**: 570-575 [PMID: 21308868 DOI: 10.1002/stem.613]
- 11 **Estes BT**, Wu AW, Storms RW, Guilak F. Extended passaging, but not aldehyde dehydrogenase activity, increases the chondrogenic potential of human adipose-derived adult stem cells. *J Cell Physiol* 2006; **209**: 987-995 [PMID: 16972251 DOI: 10.1002/jcp.20808]
- 12 **Troyer DL**, Weiss ML. Wharton's jelly-derived cells are a primitive stromal cell population. *Stem Cells* 2008; **26**: 591-599 [PMID: 18065397 DOI: 10.1634/stemcells.2007-0439]
- 13 **Jones RJ**, Barber JP, Vala MS, Collector MI, Kaufmann SH, Ludeman SM, Colvin OM, Hilton J. Assessment of aldehyde dehydrogenase in viable cells. *Blood* 1995; **85**: 2742-2746 [PMID: 7742535]
- 14 **Pittenger MF**, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; **284**: 143-147 [PMID: 10102814]
- 15 **Kuroda Y**, Dezawa M. Mesenchymal stem cells and their subpopulation, pluripotent muse cells, in basic research and regenerative medicine. *Anat Rec (Hoboken)* 2014; **297**: 98-110 [PMID: 24293378 DOI: 10.1002/ar.22798]
- 16 **Sanchez CG**, Teixeira FK, Czech B, Preall JB, Zamparini AL, Seifert JR, Malone CD, Hannon GJ, Lehmann R. Regulation of Ribosome Biogenesis and Protein Synthesis Controls Germline Stem Cell Differentiation. *Cell Stem Cell* 2016; **18**: 276-290 [PMID: 26669894 DOI: 10.1016/j.stem.2015.11.004]
- 17 **Cai X**, Gao L, Teng L, Ge J, Oo ZM, Kumar AR, Gilliland DG, Mason PJ, Tan K, Speck NA. Runx1 Deficiency Decreases Ribosome Biogenesis and Confers Stress Resistance to Hematopoietic Stem and Progenitor Cells. *Cell Stem Cell* 2015; **17**: 165-177 [PMID: 26165925 DOI: 10.1016/j.stem.2015.06.002]
- 18 **Lin YM**, Wu CC, Chang YC, Wu CH, Ho HL, Hu JW, Chang RC, Wang CT, Ouyang P. Target disruption of ribosomal protein pNO40 accelerates aging and impairs osteogenic differentiation of mesenchymal stem cells. *Biochem Biophys Res Commun* 2016; **469**: 903-910 [PMID: 26721440 DOI: 10.1016/j.bbrc.2015.12.003]
- 19 **Illouz Y-Gr**, Sterodimas A. Adipose stem cells and regenerative medicine. Heidelberg; New York: Springer, 2011
- 20 **Hong SJ**, Traktuev DO, March KL. Therapeutic potential of adipose-derived stem cells in vascular growth and tissue repair. *Curr Opin Organ Transplant* 2010; **15**: 86-91 [PMID: 19949335 DOI: 10.1097/MOT.0b013e328334f074]
- 21 **Panés J**, García-Olmo D, Van Assche G, Colombel JF, Reinisch W, Baumgart DC, Dignass A, Nachury M, Ferrante M, Kazemi-Shirazi L, Grimaud JC, de la Portilla F, Goldin E, Richard MP, Leselbaum A, Danese S; ADMIRE CD Study Group Collaborators. Expanded allogeneic adipose-derived mesenchymal stem cells (Cx601) for complex perianal fistulas in Crohn's disease: a phase 3 randomised, double-blind controlled trial. *Lancet* 2016; **388**: 1281-1290 [PMID: 27477896 DOI: 10.1016/S0140-6736(16)31203-X]

P- Reviewer: Goebel WS, Kan L, Kiselev SL, Lee Y, Ramirez M, Shawcross SG, Tanabe S, Wakao H **S- Editor:** Ji FF
L- Editor: A **E- Editor:** Lu YJ

