

Oxidative stress, renal infiltration of immune cells, and salt-sensitive hypertension: all for one and one for all

Bernardo Rodríguez-Iturbe,¹ Nosratola D. Vaziri,² Jaime Herrera-Acosta,³ and Richard J. Johnson⁴

¹Servicio de Nefrología, Hospital Universitario, Universidad del Zulia, Instituto de Inmunobiología (Fundacite-Zulia), Maracaibo 400-A, Venezuela; ²Division of Nephrology and Hypertension, Department of Medicine, Physiology, and Biophysics, University of California, Irvine, California 92697; ³Departamento de Nefrología, Instituto Nacional de Cardiología “Ignacio Chávez,” 14080 Mexico City, Mexico; and ⁴Renal Division, University of Florida, Gainesville, Florida 32610

Rodríguez-Iturbe, Bernardo, Nosratola D. Vaziri, Jaime Herrera-Acosta, and Richard J. Johnson. Oxidative stress, renal infiltration of immune cells, and salt-sensitive hypertension: all for one and one for all. *Am J Physiol Renal Physiol* 286: F606–F616, 2004; 10.1152/ajprenal.00269.2003.—Recent evidence indicates that interstitial infiltration of T cells and macrophages plays a role in the pathogenesis of salt-sensitive hypertension. The present review examines this evidence and summarizes the investigations linking the renal accumulation of immune cells and oxidative stress in the development of hypertension. The mechanisms involved in the hypertensive effects of oxidant stress and tubulointerstitial inflammation, in particular intrarenal ANG II activity, are discussed, focusing on their potential for sodium retention. The possibility of autoimmune reactivity in hypertension is raised in the light of the proinflammatory and immunogenic pathways stimulated by the interrelationship between oxidant stress and inflammatory response. Finally, we present some clinical considerations derived from the recognition of this interrelationship.

interstitial nephritis; autoimmunity; reactive oxygen species; angiotensin

THE RELATIONSHIP BETWEEN INCREASED blood pressure and oxidative stress has been recognized for some time, but it is only recently that the role of the renal infiltration of immune cells has been made evident as the third “musketeer” in this association. The purpose of this review is to summarize the work documenting the association between oxidative stress and interstitial accumulation of immune cells in the kidney in the pathogenesis of salt-sensitive hypertension. We shall consider the mechanisms that are involved in the prohypertensive effects of these conditions and identify some pathways that may be responsible for their interrelationship. Finally, we will review the conflicting results obtained with antioxidant medications in the treatment of hypertension and discuss why further studies are needed to explore the potential clinical usefulness of treatments directed to reduce oxidative stress and renal inflammation.

FREE RADICALS: REACTIVE OXYGEN AND NITROGEN SPECIES

The pathogenic role of oxygen free radicals in diseased states was first recognized by Harman (54, 55), who hypothesized that they were generated *in vivo* where they played a role in cell injury, cancer, and the process of aging. Subsequently, free radicals have been shown not only to be a cause of cell damage but are also involved in a variety of mechanisms that ensure cellular physiological equilibrium, such as regulation of vascular tone, sensing of oxygen tension, and signal transduc-

tion (35). Biologically active free radicals are of two kinds, reactive oxygen species (ROS) and reactive nitrogen species (RNS), and their generation, chemical reactions, and *in vivo* effects are closely interrelated.

The major ROS are superoxide ($O_2^{\cdot-}$), hydrogen peroxide (H_2O_2), and hydroxyl radical ($\cdot OH$). These ROS are generated as intermediate products in the reduction of oxygen to water (redox reactions).

One of the most important biological mechanisms of ROS generation results from the generation of $O_2^{\cdot-}$ from O_2 by the enzyme NAD(P)H oxidase (48). The best characterized NAD(P)H oxidase is that present in neutrophils where it has a critical role in the “oxidative burst” that is the first line of defense against bacteria. NADPH oxidase isoforms have also been identified in vascular smooth muscle cells (47, 108), fibroblasts (69), endothelial cells (68), and mesangial cells (67). Hydrogen peroxide is produced from $O_2^{\cdot-}$ by the enzyme superoxide dismutase and in the presence of iron-containing molecules, H_2O_2 is reduced to $\cdot OH$. The hydroxyl radical is highly reactive and hence is short-lived, with its toxicity exerted locally.

RNS, such as nitrosonium cation (NO^+), nitroxyl anion (NO^-), and peroxynitrate ($ONOO^-$) represent the second major type of free radicals. Many of these are generated by reaction of ROS with nitric oxide (NO) or NO-related products. NO generated by endothelial cells has a critical role in mediating endothelial cell viability and vascular smooth muscle vasodilation in a reaction of low-NO flux and very fast kinetics (71). Interestingly, the reaction of oxidants with NO may scavenge $O_2^{\cdot-}$ and H_2O_2 and hence provide some cellular protection by preventing ROS-mediated lipid peroxidation of cell membranes (62, 133), a benefit that is offset by the

Address for reprint requests and other correspondence: B. Rodríguez-Iturbe, Servicio de Nefrología, Hospital Universitario, Universidad del Zulia, Instituto de Inmunobiología (Fundacite-Zulia), Maracaibo 400-A, Venezuela (E-mail: bri@iamnet.com).

generation of RNS that act by themselves to impose additional oxidative stress on the cell (Fig. 1).

Under normal circumstances, the host is protected from the toxic effect of ROS and RNS by extra- and intracellular antioxidants and oxygen radical scavengers; however, when these defense systems are overwhelmed, the cell is placed under "oxidative stress" and may be injured, activated, or even die.

OXIDATIVE STRESS AND HYPERTENSION

Oxidative stress has been documented in both experimental and human hypertension (120, 150). A number of investigations have shown that hypertension results from stimulating systemic ROS generation (85, 111, 143, 145, 146), and a variety of antioxidant treatments reduce blood pressure in genetic and experimentally induced models of hypertension (23, 26, 36, 40, 77, 78, 102, 104, 127, 128, 144, 146, 147, 152).

Increased ROS not only have a critical role in the initiation of hypertension but they may be generated by the hypertension itself, suggesting a vicious cycle (151). Originally, it was assumed that elevation of the blood pressure per se does not induce oxidative stress in the vascular endothelium because norepinephrine-induced hypertension does not increase superoxide generation (81); however, this assumption was challenged by the studies of Barton et al. (8), who showed that in experimental aortic coarctation, the organs in the hypertensive upper body had evidence of oxidative stress, whereas the organs in the lower normotensive body did not. Because the findings could not be attributed to hormonal or humoral factors, they were considered to be due to differences in baromechanical stress.

Stimulation of NAD(P)H oxidase is the primary source of oxidants in the systemic arterial vessels in ANG II-induced hypertension, DOCA-salt hypertension, renovascular hypertension, chronic renal insufficiency, and in the spontaneously hypertensive rat (SHR) (108, 120, 143, 154, 155). In humans, NAD(P)H oxidase is the source of basal O_2^- production in the vascular smooth muscle cells (12), and it is increased in patients with essential hypertension (141).

Prohypertensive Mechanisms of Oxidative Stress

The mechanisms whereby systemic oxidative stress plays a role in the pathogenesis of hypertension involve both hemo-

dynamic (vasoconstrictive) and structural (vascular remodeling) mechanisms. ROS can activate signaling cascades in vascular smooth muscle cells (6, 28, 34, 52, 79) that induce remodeling in resistance arteries, resulting in increased wall rigidity and narrowing of the lumen. These changes have been assumed to cause or maintain hypertension. However, the real contribution of these modifications to the elevation of the blood pressure has been questioned by the lack of correlation between vascular remodeling and blood pressure (56, 97) and by recent observations in one-kidney one-clip renovascular hypertension which have demonstrated that removal of the clip normalized the blood pressure while the structural alterations in peripheral arteries remained unchanged (80).

A separate issue is the role of hypertrophic modifications in the afferent arteriole of the glomerulus. ROS-induced vascular remodeling in these arterioles may impair the vasomotor responses that protect glomeruli from systemic hypertension and induce distal tubulointerstitial ischemia. Both of these effects likely have a pathogenetic role in hypertension (65).

As indicated in Fig. 1, vasoconstriction of both systemic and intrarenal vessels may also result from both direct and indirect actions of ROS (125). ROS can inactivate endothelial NO, resulting in impaired vasodilatation (24, 82, 122, 152), and recent studies in humans with renovascular hypertension have validated this postulate (61). In addition, there are direct effects of ROS on vascular tone. Whereas ROS can induce vasoconstriction or vasodilation, depending on the amount produced and the vascular bed (35), the more common response to O_2^- is vasoconstriction (125). Other mechanisms of ROS-induced vasoconstriction include oxidation of arachidonic acid with formation of vasoconstrictive eicosanoids (such as prostaglandin $F_{2\alpha}$) (138) and inhibition of the synthesis of vasodilatory PGI_2 (160). Furthermore, O_2^- induces increments in intracellular calcium in smooth muscle and endothelial cells (84), thereby mediating the actions of other vasoconstrictors such as ANG II, thromboxane (TXA_2), endothelin-1 (ET-1), and norepinephrine (125).

In addition to systemic effects of ROS, recent evidence suggests that oxidant stress within the kidney plays a central role in the pathophysiology of sodium retention because it results in tubulointerstitial accumulation of ANG II-positive cells (114, 116, 117). The sodium-retaining mechanisms resulting from intrarenal ANG II activity will be discussed in the next section. The prohypertensive role of intrarenal ROS is suggested by the strong correlation between renal superoxide-positive cells and the severity of hypertension in the SHR (Fig. 2).

Tubulointerstitial Inflammation and Hypertension

Evidence for an immune mechanism in the pathogenesis of hypertension was first advanced by Svendsen (135), who observed, more than a quarter century ago, that the late salt-dependent phase of the DOCA-salt model of hypertension required an intact thymus with the infiltration in the kidney of perivascular lymphocytes displaying "delayed-type immune reactivity." These observations were largely ignored despite the findings that cyclophosphamide therapy (9), anti-thymocyte serum (11), neonatal thymectomy (73), and thymic implants from normotensive donors (5a) could ameliorate hypertension in various models in rats. These early findings were

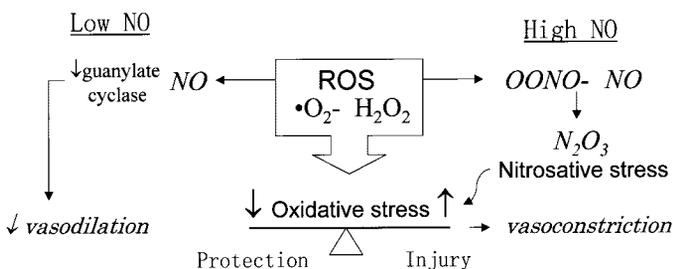


Fig. 1. Balance between oxidative stress and nitrosative stress depends on nitric oxide (NO) flux. Low-NO fluxes ($1-2 \mu M$) reduce cellular injury caused by O_2^- and H_2O_2 with termination of lipid peroxidation reactions. Reactive oxygen species (ROS) inactivate NO and thereby reduce the binding of NO to the ferrous heme moiety of guanylate cyclase. As a result, vessel relaxation depending on the guanylate cyclase-induced conversion of GTP to cGMP is decreased by oxidative stress. Direct effects of oxidative stress induce vasoconstriction.

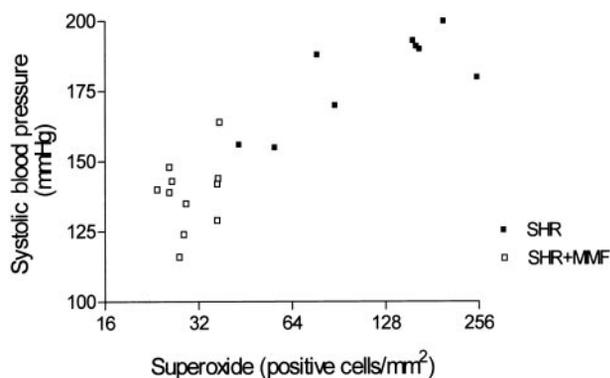


Fig. 2. Direct correlation ($r = 0.82$, $P < 0.001$) between oxidative stress, represented here as the number of superoxide positive cells in the renal tubulointerstitium, and the systolic blood pressure of spontaneously hypertensive rats (SHR) treated for 3 wk with mycophenolate mofetil (SHR + MMF) or untreated rats (SHR). Data are from Ref. 116 with permission.

interpreted by the investigators as evidence that the hypertension resulted from autoimmune vasculitis. In fact, the immune dysfunction observed in SHR (reviewed in Refs. 38 and 72) was considered to be an adaptive defense mechanism against otherwise life-threatening hypertension (11). In contrast, recent work has provided evidence that immune cells accumulating in the kidney may be responsible for mediating sodium retention and, thereby, for the development of hypertension. First, tubulointerstitial infiltration of lymphocytes and macrophages appears to be universally present in experimental models of salt-sensitive hypertension. These include DOCA-salt hypertension, post-ANG II infusion salt-sensitive hypertension, post-catecholamine infusion salt-sensitive hypertension, hyperuricemia-induced salt sensitivity, hypertension after chronic NO synthesis inhibition, hypertension associated with protein overload proteinuria, hypokalemic nephropathy-associated salt sensitivity, two-kidney one-clip hypertension (persisting after clip removal), aging nephropathy, and cyclosporine nephropathy, as well as genetic models of hypertension such as the SHR, the stroke-prone SHR, and the double transgenic rat harboring the human renin and angiotensinogen genes (reviewed in Ref. 115). Second, several investigations have demonstrated a direct correlation between the number of infiltrating cells and the severity of hypertension (3, 116, 117). An example of this correlation is shown in Fig. 3.

Finally, and most importantly, a number of studies have shown that treatment strategies that result in a reduction in the renal inflammatory cell infiltrate also prevent the development of salt-sensitive hypertension (4, 107, 114) or improve established hypertension in genetically prone strains of hypertensive rats (88, 94, 95, 99, 116, 117). These studies are summarized in Table 1. While these studies strongly suggest that the immune infiltrate is mediating the salt sensitivity, a caveat is that mycophenolate mofetil (MMF) and the other therapies may also be affecting resident cell populations (reviewed in Ref. 156), which may also have a contributory role in the prevention of salt-driven hypertension in these experimental models.

Prohypertensive Effects of Tubulointerstitial Inflammation

The mechanism(s) by which the immune infiltrate contributes to the pathogenesis of hypertension is incompletely de-

finer but may relate to the sodium-retaining effects of intrarenal ANG II activity induced by the accumulation of immune cells. As shown by double-immunostaining studies, ANG II is expressed by infiltrating T cells and macrophages in experimental models of hypertension (4, 107, 116). Both of these cells are known to express angiotensin-converting enzyme (31), and macrophages are capable of synthesizing ANG II (148). Interstitial accumulation of ANG II-positive cells has also been postulated as the reason for primary sodium retention in patients with the nephrotic syndrome (112).

As shown in Fig. 4, well-known renal effects of ANG II include a decreased glomerular filtration rate (GFR; which will reduce the filtered sodium load), an increase in tubular sodium reabsorption, and an impairment of pressure-natriuresis. Franco et al. (42) studied the glomerular hemodynamic findings in the model of salt-sensitive hypertension induced by ANG II infusion and the changes associated with MMF treatment. ANG II infusion, as expected, caused an increase in afferent and efferent arteriolar resistances with a decrease in single-nephron GFR and filtration coefficient K_f . In the weeks that follow ANG II infusion, in which a high-salt diet induces hypertension, the hemodynamic alterations remained essentially unchanged, consistent with a persistent intrarenal vasoconstriction. Treatment with MMF did not change the glomerular vasoconstriction induced during the exogenous ANG II administration but prevented glomerular vasoconstriction in the subsequent salt-sensitive period (42). These studies suggested a role for ANG II-like intrarenal activity that was related to the interstitial immune infiltrate. Investigations from Nishiyama et al. (101) have convincingly shown increased endogenous production of intrarenal ANG II in the ANG II infusion model and that interstitial ANG II functions as a separate compartment that is not modified by the systemic hemodynamic changes known to modulate plasma ANG II concentrations (100, 101). An example of ANG II-positive tubular cells and infiltrating cells is shown in Fig. 5.

In addition to the sodium-retaining effects, intrarenal ANG II activity has other potential consequences, including the activation of signaling cascades and transcription factors that could further increase interstitial inflammation (123) and stimulation of NAD(P)H oxidase-mediated superoxide production.

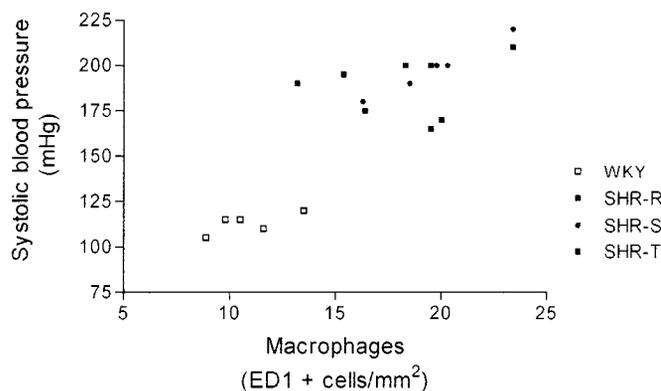


Fig. 3. Relationship ($r = 0.87$, $P < 0.001$) between macrophage infiltration and systolic blood pressure in spontaneously hypertensive rats given a regular diet (SHR), an antioxidant test diet (SHR-T), or switched from a regular to an antioxidant-rich diet (SHR-S). Control Wistar-Kyoto rats (WKY) are included. Data are reproduced from Ref. 117 with permission.

Table 1. *Effects of reduction of interstitial immune cell infiltration in experimental models of hypertension*

Experimental Model	Treatment	Renal Findings (Primary/Additional)	Results in Blood Pressure	Reference No.
dTGF rats	PDTC	↓ NF-κB/ ↓ Mφ	Improvement in HBP (improvement in end-organ damage)	94
dTGF rats	DEXA MMF	↓ Interstitial L, ↓ MHC II, ↓ Oxidative stress,	Effects independent of BP (improvement in end-organ damage)	95
dTGF rats	Lipoic acid	↓ Oxidative stress/ ↓ NF-κB, AP-1/ ↓ Interstitial L and Mφ	Improvement in HBP (improvement in end-organ damage)	88
ANG II infusion	MMF	↓ Interstitial L & Mφ/ ↓ oxidative stress	Prevention of post-ANG II SSHBP	114
L-NAME-induced NOS inhibition	MMF	↓ Interstitial L and Mφ	Prevention of post-L-NAME SSHBP	107
DOCA-salt hypertension	Tempol	↓ Oxidative stress/ ↓ interstitial L and Mφ	Improvement of HBP	13
ANG II infusion	MMF	Abrogation of glomerular hemodynamic changes in the post-ANG II SSHBP	Prevention of post-ANG II SSHBP	42
Oxonic acid-induced hyperuricemia	MMF	↓ Interstitial L and Mφ	Prevention of posthyperuricemia SSHBP	3
Protein overload proteinuria	MMF	↓ Interstitial L and Mφ/ ↓ oxidative stress	Prevention of post-protein overload SSHBP	4
SHR	MMF	↓ Interstitial L and Mφ/ ↓ oxidative stress	Improvement of HBP	116
SHR	Antioxidant-rich diet	↓ Oxidative stress/ ↓ interstitial L and Mφ	Improvement of HBP	117
SHR	Melatonin	↓ Oxidative stress/ ↓ NF-κB/ ↓ interstitial L and Mφ	Improvement of HBP	99

HBP, hypertension; SSHBP, salt-sensitive HBP; dTGF, double transgenic rats harboring both human renin and angiotensinogen genes; AP-1, activator protein 1; DEXA, dexamethasone; MHC II, major histocompatibility complex II; PDTC, pyrrolidine dithiocarbamate; NF-κB, nuclear factor-κB; L, lymphocytes; Mφ, macrophages; NOS, nitric oxide synthesis; MMF, mycophenolate mofetil; L-NAME, *N*^ω-nitro-L-arginine-methyl ester; SHR, spontaneously hypertensive rat. The reduction of oxidative stress shown represents a reduction of urinary or renal malondialdehyde content, plasma H₂O₂ concentration, renal nitrotyrosine abundance, or the number of superoxide-positive cells.

These changes likely contribute to the maintenance of the low-grade renal injury and peritubular capillary loss (65) that participate in the pathophysiology of sodium balance in hypertension (51, 53).

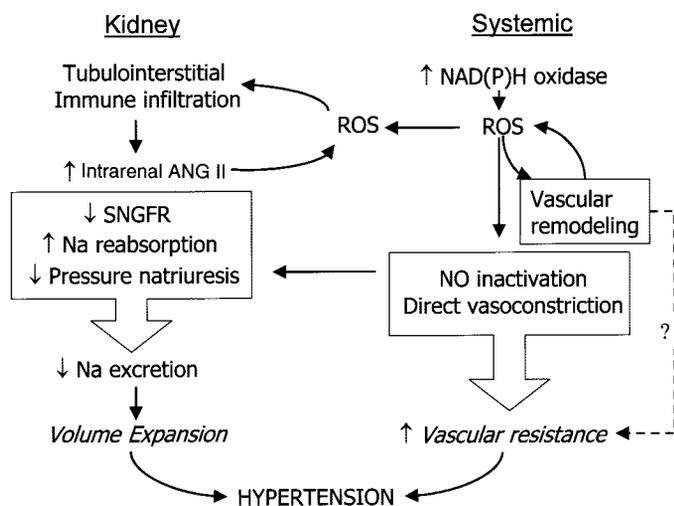


Fig. 4. Intrarenal ANG II activity resulting, at least in part, from ANG II-positive interstitial mononuclear cells and tubular cells induces sodium retention by the combined effects of reducing filtered sodium, increasing proximal tubular sodium reabsorption, and impairing pressure-natriuresis. Increased intrarenal ANG II in association with oxidative stress constitutes a feedback loop for the maintenance of interstitial renal inflammation. Systemic prohypertensive effects of oxidative stress include vasoconstriction resulting from both NO consumption and direct effects of ROS and, questionably (85), the consequences of long-term vascular remodeling. The feedback loops between systemic effects and renal effects involve the generation of ROS.

Interrelationship Between Renal Oxidative Stress and Interstitial Inflammation

Interstitial accumulation of lymphocytes and macrophages in the kidney is a consequence of the complex and intimate relationship between inflammatory reactivity and oxidative stress that stimulates mechanisms of cell death and cell survival. As an example of this relationship, Fig. 6 shows the close correlation between the renal infiltration of macrophages and oxidative stress in the SHR.

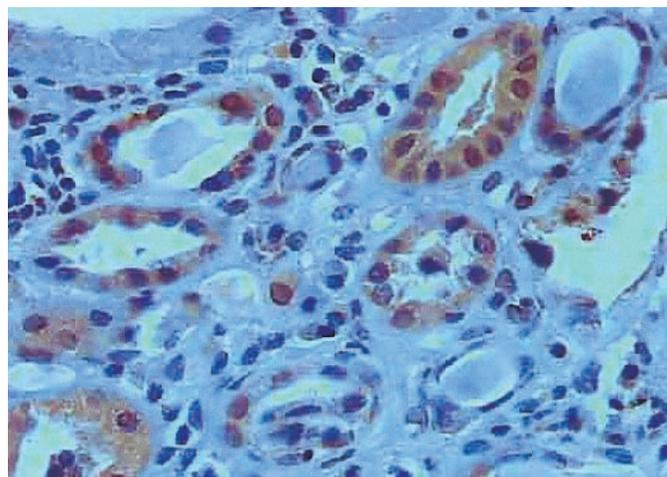


Fig. 5. Tubular cells and infiltrating mononuclear cells in tubulointerstitium staining positive for ANG II in a renal biopsy of a patient with nephrotic syndrome (immunoperoxidase technique). (Courtesy of Dr. Sergio Mezzano).

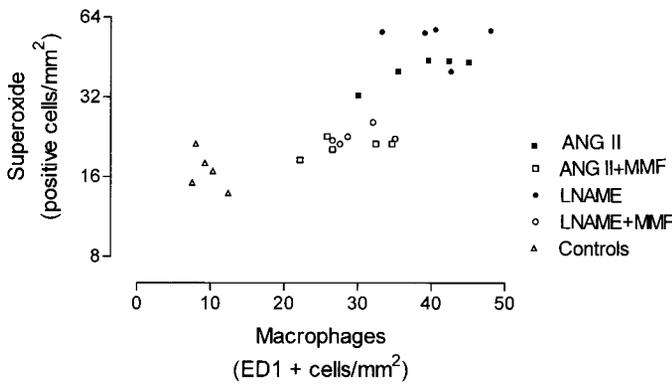


Fig. 6. Relationship ($r = 0.76, P < 0.001$) between the tubulointerstitial infiltration of macrophages (ED1-positive cells) and the intensity of oxidative stress (superoxide-positive cells). Data were obtained from kidneys harvested from rats that were given 2 wk of subcutaneous infusion of ANG II (ANG II group) or 3 wk of oral administration of *N*^ω-nitro-L-arginine methyl ester (L-NAME) to inhibit NO synthase (L-NAME group). Additional groups of rats received mycophenolate mofetil during ANG II infusion (ANG II + MMF group) and during L-NAME administration (L-NAME + MMF group). Data are from studies described in Refs. 19 and 106.

ROS may induce a wide range of cellular responses, ranging from proliferation to cell apoptosis (21, 50, 63). Certain transcription factors, such as NF- κ B and activator protein-1, are redox sensitive and will be activated by oxidants (35). MAP kinases, such as ERK 1 and 2 are activated by O_2^- in vascular smooth muscle cells (6), and JNK and p38 are activated by H_2O_2 (28, 29).

The different pathways involved in signal transduction stimulated by oxidative stress are highly interconnected and modulate each other's activities so that the outcome depends on the dose of oxidant stress and the physiological context in which they are examined. Figure 7 indicates some of the biological pathways that interrelate oxidative stress and inflammatory reactivity that have already been found to be stimulated in models of salt-sensitive hypertension.

In general, low doses of ROS induce mitogenic responses, intermediate doses induce growth arrest, and severe oxidant stress causes apoptosis or necrosis (86). Mitogenic, predominantly cell survival responses include activation of the ERK pathway, phosphatidylinositol 3-kinase/Akt (protein kinase B), and phospholipase C- γ 1 signaling (51, 153, 158). In addition, oxidative stress induces activation of NF- κ B (17, 83, 129), which is a rapid-response transcription factor of proinflammatory genes. The activation of NF- κ B is the result of peroxide-induced phosphorylation of the inhibitory binding protein I κ B (130, 157). NF- κ B mediates the synthesis of a variety of cytokines and, in addition, promotes leukocyte infiltration because it increases the expression of adhesion molecules E-selectin, VCAM-1, and ICAM-1 (26, 91, 137). As a result of these effects, oxidative stress is capable of inducing nonspecific inflammation, which would be maintained as long as oxidant stress is sustained (Fig. 7). In models of salt-sensitive hypertension, interstitial inflammation is, in fact, associated with increased apoptosis and activation of NF- κ B (Fig. 8A) (106). Furthermore, inhibition of NF- κ B reduces the interstitial accumulation of inflammatory cells and lowers the blood pressure in hypertensive rat strains (88, 94).

An additional mechanism for ROS-mediated inflammation may be the expression of heat shock proteins (HSPs), a well-

preserved response of living organisms to stressful situations such as heat, ATP depletion, and oxidants. HSPs act as chaperones that guide the assembly, folding, and location of various proteins in cells. HSPs are grouped in six families classified according to their molecular mass (HSP 100, 90, 70, 60, 40, and smaller HSPs). ROS is a well-known inducer of HSPs, and prior treatment with antioxidants inhibits their expression (46). In turn, HSPs protect proteins against oxidative damage by decreasing intracellular levels of ROS by maintaining glutathione in a reduced state (5, 7), suppressing apoptotic pathways, such as the JNK pathway (103), and inhibiting cytochrome *c* release and caspase activation (22, 32). However, HSPs' effects on cell survival are complex and at times seemingly contradictory. If inflammation follows the stimulation of HSPs, they exert a protective effect; in contrast, when inflammation precedes the induction of HSPs, the resulting effect is frequently cell death by apoptosis, a phenomenon called the "heat shock paradox," in which the participation of NF- κ B activity has been postulated (33).

As shown in Fig. 7, HSPs are part of the vicious cycle that results in renal interstitial inflammation in circumstances of sustained ROS production. HSPs induce the production of proinflammatory cytokines and overexpression of adhesion molecules E-selectin, ICAM-1, and VCAM-1 (45, 76, 105) and, therefore, facilitate the accumulation of immune cells characteristic of experimental models of salt-sensitive hypertension.

The distribution and function of HSPs in the kidney have recently been reviewed (10), and several studies have demonstrated that experimental manipulations known to be associated with the subsequent development of salt-sensitive hypertension stimulate expression of HSPs. Infusion of ANG II induces renal overexpression of HSP70, HSP60, HSP25, HSP32, and heme oxygenase (HO-1) (1, 19, 64) that is mediated by ANG II type 1 receptor activation (114). As demonstrated in Fig. 8B, overexpression of HSP70 is also induced by NO synthase (NOS) inhibition. In both of these models, increased oxidative stress has been postulated as the likely stimulus for HSP production.

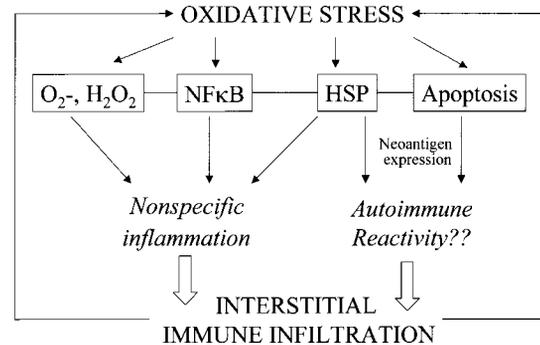


Fig. 7. Mechanisms interrelating oxidative stress and interstitial infiltration of immune cells that have been demonstrated in experimental models of salt-sensitive hypertension. Apoptosis, heat shock protein expression (HSP), activation of NF- κ B, and generation of ROS are interconnected by multiple signaling pathways and, depending on the pathophysiological context, may stimulate or inhibit one another, as reviewed recently by Martindale and Holbrook (86). Evidence of epithelial/mesenchymal transdifferentiation (shown with neoverexpression of vimentin) suggests the possibility of autoantigen expression resulting from cell injury. Autoantigenic reactivity may result from HSP expression and intense apoptosis and could be amplified by oxidative stress (see text), but this possibility is, at present, entirely speculative.

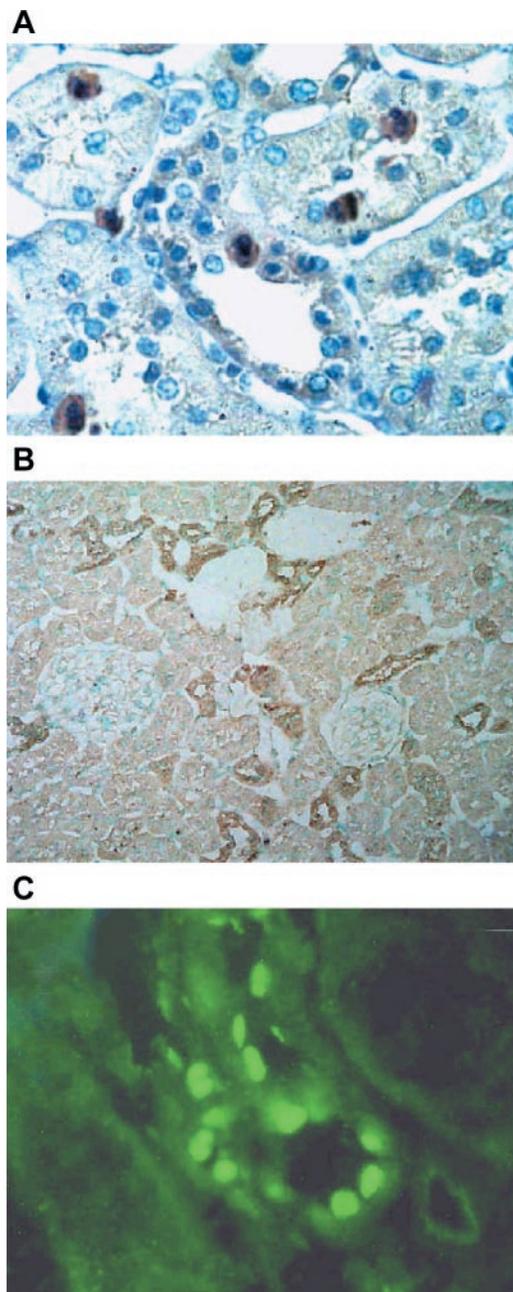


Fig. 8. Microphotographs demonstrating activated NF- κ B, shown as staining for its p65 subunit (A; reproduced from Ref. 106 with permission), and neoexpression of HSP70 in the renal cortex of Sprague-Dawley rats after 3 wk of NO synthesis inhibition (B; reproduced from Ref. 19 with permission). Apoptotic tubular cells, shown as TUNEL-positive cells, are present in the kidney of rats treated with exogenous ANG II during 2 wk (C; reproduced from Ref. 106 with permission).

Last, interstitial inflammation and oxidative stress may participate jointly in the development and maintenance of hypertension by the reduction of the number of nephron units, which thereby limits sodium filtration (20). It is well recognized that the severity of tubulointerstitial damage correlates with renal functional deterioration (15, 16, 98), and immune cell infiltration is a final common pathway to end-stage renal disease (113). The reduction in tubulointerstitial inflammation by a variety of treatment modalities prevents or retards the devel-

opment of end-stage renal disease (43, 44, 109, 119). The link among interstitial immune infiltration and oxidative stress and hypertension is obvious in these circumstances. In addition, recent work links inflammatory reactivity and oxidative stress in the tubulointerstitium with the development of glomerular arteriolopathy, a process that may lead to impaired autoregulatory responses, resulting in increased transmission of systemic pressures to the glomeruli where they may predispose the animal to the development of glomerulosclerosis (14, 139).

DOES T CELL INFILTRATION IN THE RENAL INTERSTITIUM REFLECT AN AUTOIMMUNE REACTION?

The coexistence of HSPs, increased apoptosis, and oxidative stress brings up the possibility that autoimmunity reactivity could be involved in the maintenance of low-grade, self-sustained interstitial inflammation and thereby participate in the pathogenesis of salt-sensitive hypertension. While evidence in favor of this possibility is lacking at the present time, certain aspects make this speculation worth considering. Models of salt-sensitive hypertension require an induction phase of 2- to 3-wk duration (4, 107, 114), and this induction phase is characterized by cellular injury that could result in the expression of neoantigens or altered self-antigens that are viewed as "foreign" by the host. For example, tubular epithelial cell transdifferentiation, as demonstrated by vimentin neoexpression, is a feature of some of these models (19, 34). This finding is not unexpected because activated macrophages (92) and ANG II (75) can induce vimentin expression. This raises the possibility that immune reactivity to vimentin or related proteins may be involved in the development of autoimmunity, as has been shown in rejection episodes of human heart transplantation and in autoimmune myocarditis (70, 124).

HSPs may also have a direct role in the development of autoimmunity. HSPs have a role in antigen presentation (159) and may act as activators of innate immunity (100). HSPs themselves can be the cause of autoimmune disease. For example, Weiss et al. (149) showed that T cells reactive against HSPs can induce interstitial nephritis. In addition, HSPs are known to bind peptides in damaged tissue to form HSP-peptide complexes with strong immunogenicity (27, 142).

Apoptosis is another potential cause of antigen-specific inflammatory reactivity (Fig. 7). While prompt phagocytosis of apoptotic cells is not associated with inflammation, apoptotic cells have intracellular antigens translocated to the cell surface, and recent evidence indicates that an excess load or abnormal processing of apoptotic cells can generate autoantibody formation. Hypergammaglobulinemia, anti-DNA, and anti-cardiolipin antibodies can be generated by exposure to syngeneic apoptotic cells (30, 89, 90). Of note, apoptosis is markedly increased in the kidney in models of experimental hypertension such as ANG II infusion and NOS synthesis inhibition (106). An example of apoptotic tubulointerstitial cells induced by ANG II is shown in Fig. 8C.

Another influence that could favor the development of local autoimmunity is oxidative stress itself. Functional activation of lymphocytes is stimulated by ROS as a shift in the intracellular redox state can amplify the responses after relatively weak receptor stimulation (59). An example of this amplified response is the generation of cytotoxic T cells after immunization

in mice with cells expressing foreign minor histocompatibility antigens (121).

CLINICAL CONSIDERATIONS

The experimental evidence supporting the role of oxidative stress and infiltration of immune cells in the renal interstitium in the pathogenesis of arterial hypertension is compelling. In contrast, clinical studies have failed to yield conclusive results. Some studies have shown that the administration of antioxidant vitamins reduces blood pressure (18, 36, 41, 93), which is consistent with earlier studies that showed that local infusion of ascorbic acid improves endothelial-dependent vasodilatation (136) and with reports of an inverse correlation between serum carotene and vitamin C and blood pressure (25). In contrast, as reviewed recently (140), other large series failed to show any blood pressure-lowering effect (74), and studies designed to evaluate the modification of cardiovascular risk by antioxidant therapy did not report significant effects on blood pressure (49, 57, 58, 132).

There are several aspects worth considering with regard to the lack of uniformity in the results of the clinical trials that examined the effects of antioxidant therapy in hypertension. These aspects may also serve as potential guidelines in future clinical studies. First, there is the problem of defining the severity of systemic oxidative stress and the intensity of the antioxidant treatment that would be required. Most studies determine levels of antioxidant vitamins but not the baseline levels of ROS or the changes in ROS levels with treatment which would indicate that a therapeutic goal has been achieved. It may be important to adjust the dose of antioxidant therapy based on the hydrogen peroxide or malondialdehyde plasma levels. While establishing target levels is relatively easy for drug dosages in acute studies, such as when intravenous iron or erythropoietin is given (60), this may be considerably more difficult in the long-term follow-up of patients given the variability introduced by diet, hemoglobin levels, and physical activity, among others. Second, a separate assessment of systemic vs. intrarenal oxidative stress may be useful. High urinary malondialdehyde excretion may reflect intrarenal ROS with active interstitial inflammation in the kidney (118, 119) and may suggest a potential benefit of antioxidants and, indeed, sodium restriction. Third, there is the choice of antioxidant treatment for a specific patient. It is possible that patients with obesity, hyperinsulinemia, and hypertension may benefit from dietary modifications and exercise that induce reduction in oxidative stress with improvement in the metabolic profile and blood pressure (110), whereas patients with acute increments of oxidative stress, such as from a hypertensive crisis, may require an antioxidant that acts rapidly after oral administration, such as melatonin (60), in addition to standard emergency antihypertension treatment.

Finally, while antioxidants may block some of the inflammatory response, it is possible that the concomitant use of other anti-inflammatory agents may help to prevent the infiltration of ANG II- and oxidant-producing cells. While nonsteroidal anti-inflammatory agents would be contraindicated because of the potential deterioration of renal function, angiotensin-converting enzyme inhibitors and statins have considerable anti-inflammatory actions (131, 134). Experimental studies also suggest the possibility of using uric acid-lowering drugs as a

means of reducing renal microvascular and tubulointerstitial inflammation and blood pressure (66, 87).

Future clinical studies evaluating treatment strategies for salt-sensitive hypertension should consider focusing not only on drugs that lower blood pressure but additionally on the control of oxidative stress, intrarenal ANG II activity, and interstitial inflammation in the kidney. The combined approach may be more effective because these four elements support one another, all for one and one for all, like Alexander Dumas' three musketeers, who, after all, also ended up numbering four. Studies directed to gain insight into the intimate relationship that binds these elements may lead to a better understanding of the pathogenesis of essential hypertension and its treatment.

GRANTS

Studies in our laboratories were done with the financial support of Fondo Nacional de Ciencia y Tecnología Créase el Fondo Nacional de Ciencia y Tecnología Grant S1-2001001097 and Asociación de Amigos del Riñón (B. Rodríguez-Iturbe), Thomas Yuen (N. D. Vaziri), Consejo Nacional de Ciencia y Tecnología Grant 37275 (J. Herrera-Acosta), and US Public Health Service Grants DK-43422, DK-52121, and DK-47659 (R. J. Johnson).

REFERENCES

1. Aizawa T, Ishizaka N, Taguchi J, Nagai R, Mori I, Tang SS, Ingelfinger JR, and Ohno M. Heme-oxygenase 1 is upregulated in the kidney of angiotensin II-induced hypertensive rats: possible role in renoprotection. *Hypertension* 35: 800–806, 2000.
2. Allison AC and Eugui EM. Mycophenolate mofetil and its mechanism of action. *Immunopharmacology* 47: 85–118, 2000.
3. Alvarez V, Nava M, Quiroz Y, Chavez M, Herrera-Acosta J, Johnson RJ, and Rodriguez-Iturbe B. Hyperuricemia induces salt sensitive hypertension (SHTA) that may be prevented by reduction of tubulointerstitial inflammatory infiltrate (Abstract). *J Am Soc Nephrol* 13: 328A, 2002.
4. Alvarez V, Quiroz Y, Nava M, and Rodriguez-Iturbe B. Overload proteinuria is followed by salt-sensitive hypertension caused by renal infiltration of immune cells. *Am J Physiol Renal Physiol* 283: F1132–F1141, 2002.
5. Arrigo AP. Small stress proteins: chaperones that act as regulators of intracellular redox state and programmed cell death. *J Biol Chem* 379: 19–26, 1998.
- 5a. Ba D, Takeichi N, Kodama T, and Kobayashi H. Restoration of T cell depression and suppression of blood pressure in spontaneously hypertensive rats (SHR) by thymus grafts or thymus extracts. *J Immunol* 128: 1211–1216, 1982.
6. Baas AS and Berk BC. Differential activation of mitogen-activated protein kinases by H₂O₂ and O₂⁻ in vascular smooth muscle cells. *Circ Res* 77: 29–36, 1995.
7. Baek SH, Min JN, Park EM, Han MY, Lee YS, and Park YM. Role of small heat shock protein HSP25 in radioresistance and glutathione-redox cycle. *J Cell Physiol* 183: 100–107, 2000.
8. Barton CH, Ni Z, and Vaziri ND. Enhanced nitric oxide inactivation in aortic coarctation-induced hypertension. *Kidney Int* 60: 1083–1087, 2001.
9. Bataillard A, Vincent M, Sassard J, and Touraine JL. Antihypertensive effect of an immunosuppressive agent, cyclophosphamide, in genetically hypertensive rats of the Lyon strain. *Int J Immunopharmacol* 11: 377–384, 1989.
10. Beck FX, Neuhofer W, and Müller E. Molecular chaperones in the kidney: distribution, putative roles and regulation. *Am J Physiol Renal Physiol* 279: F203–F215, 2000.
11. Bendich A, Belisle EH, and Strausser HR. Immune system modulation and its effect on blood pressure of the spontaneously hypertensive male and female rat. *Biochem Biophys Res Comm* 99: 600–607, 1981.
12. Berry C, Hamilton CA, Brosnan MJ, Magill FG, Berg GA, McMurray JJV, and Dominiczak AF. Investigation into the sources of superoxide in human vessels: angiotensin II increases superoxide production in human internal mammary arteries. *Circulation* 101: 2206–2212, 2000.
13. Beswick RA, Zhang H, Marable D, Catravas JD, Hill WD, and Webb RC. Long-term antioxidant administration attenuates mineralocorticoid

- hypertension and renal inflammatory response. *Hypertension* 37: 781–786, 2001.
14. Bobadilla NA, Tack I, Tapia E, Sanchez-Lozada LG, Santamaria J, Jimenez F, Striker LJ, Striker GE, and Herrera-Acosta J. Pentosan polysulfate prevents glomerular hypertension and structural injury despite persisting hypertension in 5/6 nephrectomized rats. *J Am Soc Nephrol* 12: 2080–2087, 2001.
 15. Bohle A, Kresse LG, Müller CA, and Müller GA. The pathogenesis of chronic renal failure. *Pathol Res Pract* 185: 421–440, 1989.
 16. Bohle A, Muller GA, Wehrmann M, Mackensen-Haen S, and Xiao JC. Pathogenesis of chronic renal failure in the primary glomerulopathies, renal vasculopathies and chronic interstitial nephritides. *Kidney Int* 49, Suppl 54: S2–S9, 1996.
 17. Bonizzi G, Oiette J, Merville MP, and Bours V. Cell type-specific role for reactive oxygen species in nuclear factor κ -B activation by interleukin 1. *Biochem Pharmacol* 59: 7–11, 2000.
 18. Boshtam M, Rafiei M, Sadeghi K, and Sarraf-Zadegan N. Vitamin E can reduce blood pressure in mild hypertensives. *Int J Vitamin Nutr Res* 72: 309–314, 2002.
 19. Bravo J, Quiroz Y, Pons H, Parra G, Herrera-Acosta J, Johnson RJ, and Rodríguez-Iturbe B. Tubulointerstitial injury resulting from angiotensin II and inhibition of nitric oxide synthesis: neoexpression of vimentin and heat shock proteins. *Kidney Int* 64, Suppl 86: S46–S51, 2003.
 20. Brenner BM, Garcia DL, and Anderson S. Glomeruli and blood pressure: less of one, more of the other? *Am J Hypertens* 1: 335–347, 1988.
 21. Buttke TM and Sandstrom PA. Oxidative stress as a mediator of apoptosis. *Immunol Today* 15: 77–110, 1994.
 22. Buzzard KA, Giaccia AJ, Killender M, and Anderson RL. Heat shock protein 72 modulates pathways of stress-induced apoptosis. *J Biol Chem* 273: 17147–17153, 1998.
 23. Cabassi A, Bouchard JF, Dumont EC, Girouard H, Le Jossec M, Lamontagne D, Besner JG, and de Champlain J. Effect of antioxidant treatments on nitrate tolerance development in normotensive and hypertensive rats. *J Hypertens* 18: 187–196, 2000.
 24. Cai H and Harrison DG. Endothelial dysfunction in cardiovascular diseases: role of oxidant stress. *Circ Res* 87: 840–844, 2000.
 25. Chen J, He J, Hamm L, Batuman V, and Whelton PK. Serum antioxidant vitamins and blood pressure in the United States population. *Hypertension* 40: 810–816, 2002.
 26. Chen X, Touyz RM, Park JB, and Schiffrin EL. Antioxidant effects of vitamin C and E are associated with altered activation of vascular NADPH oxidase and superoxide dismutase in stroke-prone SHR. *Hypertension* 38: 606–611, 2000.
 27. Cho BK, Palliser D, Guillen E, Wisniewski J, Young RA, Chen J, and Eisen HN. A proposed mechanism for the induction of cytotoxic T lymphocyte production by heat shock fusion proteins. *Immunity* 12: 263–272, 2000.
 28. Clerk A, Fuller SJ, Michael A, and Sugden PH. Stimulation of “stress-activated” mitogen-activated protein kinases/c-Jun N-terminal kinases, and p38 mitogen-activated protein kinases in perfuse rat hearts by oxidative and other stresses. *J Biol Chem* 273: 7228–7234, 1998.
 29. Clerk A, Michael A, and Sugden PH. Stimulation of multiple mitogen-activated kinase subfamilies by oxidative stress and phosphorylation of the small heat shock protein, HSP 25/27, in neonatal ventricular myocytes. *Biochem J* 333: 581–583, 1998.
 30. Cocca BA, Seal SN, D’Agnillo P, Mueller IM, Katsikis PD, Rauch J, Weigert M, and Radic MC. Structural basis for autoantibody recognition of phosphatidylserine- β 2 glycoprotein I and apoptotic cells. *Proc Natl Acad Sci USA* 98: 13826–13831, 2001.
 31. Costerousse O, Allegrini J, Lopez M, and Alhene-Gelas F. Angiotensin I-converting enzyme in human circulating mononuclear cells: genetic polymorphism of expression in T-lymphocytes. *Biochem J* 290: 333–340, 1993.
 32. Creagh EM, Carmody RJ, and Cotter TG. Heat shock protein 70 inhibits caspase-dependent, and -independent apoptosis in Jurkat T cells. *Exp Cell Res* 257: 58–66, 2000.
 33. DeMeester SL, Buchman TG, and Cobb JP. The heat shock paradox: does NF κ B determine cell fate? *FASEB J* 15: 270–274, 2001.
 34. Denu JM and Tanner KG. Specific and reversible inactivation of protein tyrosine phosphatases by hydrogen peroxide: evidence for a sulfenic acid intermediate and implications for redox regulation. *Biochemistry* 7: 5633–5642, 1998.
 35. Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev* 82: 47–95, 2002.
 36. Duffy SJ, Gokce N, Holbrook M, Huang A, Frei B, Keaney JF Jr, and Vita JA. Treatment of hypertension with ascorbic acid. *Lancet* 354: 2048–2049, 1999.
 37. Duffy SJ, Gokce N, Holbrook M, Huang A, Frei B, Keaney JF Jr, and Vita JA. Prevention of hypertension, insulin resistance, and oxidative stress by α -lipoic acid. *Hypertension* 39: 303–307, 2002.
 38. Dzielak DJ. The immune system and hypertension. *Hypertension* 19, Suppl 1: 36–44, 1992.
 39. Eddy AA. Interstitial nephritis induced by protein-overload proteinuria. *Am J Pathol* 135: 719–731, 1989.
 40. El Midaoui A and de Champlain J. Prevention of hypertension, insulin resistance and oxidative stress by α -lipoic acid. *Hypertension* 39: 303–307, 2002.
 41. Fotherby MD, Williams JC, Forster LA, Craner P, and Ferns GA. Effect of vitamin C on ambulatory blood pressure and plasma lipids in older patients. *J Hypertens* 18: 411–415, 2000.
 42. Franco M, Tapia E, Santamaria J, Zafra I, García-Torres R, Gordon KL, Rodríguez-Iturbe B, Pons H, Johnson RJ, and Herrera-Acosta J. Renal cortical vasoconstriction contributes to the development of salt-sensitive hypertension after angiotensin II exposure. *J Am Soc Nephrol* 12: 2263–2271, 2001.
 43. Fujihara CK, Malheiros DM, Zatz R, and Noronha IL. Mycophenolate mofetil attenuates renal injury in the rat remnant kidney. *Kidney Int* 54: 1510–1519, 1998.
 44. Fujihara CK, Noronha IL, Malheiros Antunes GR, de Oliveira IB, and Zatz R. Combined mycophenolate mofetil and losartan therapy arrests established injury in the remnant kidney. *J Am Soc Nephrol* 11: 283–290, 2000.
 45. Galdiero M, de Léro GC, and Marcatili A. Cytokine and adhesion molecule expression in human monocytes and endothelial cells stimulated with bacterial heat shock proteins. *Infect Immun* 65: 699–707, 1997.
 46. Gorman AM, Heavy B, Creagh E, Cotter TG, and Samali A. Antioxidant-mediated inhibition of the heat shock response leads to apoptosis. *FEBS Lett* 445: 98–102, 1999.
 47. Griendling KK, Minieri CA, Ollerenshaw JD, and Alexander RW. Angiotensin II stimulates NADH and NADPH oxidase activity in cultures vascular smooth muscle cells. *Circ Res* 74: 1141–1148, 1994.
 48. Griendling KK, Sorescu D, and Ushio-Fukai M. NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ Res* 86: 494–501, 2000.
 49. Gruppo. Italiano per lo Studio della Sopravvivenza nell’infarto miocardico (GISSI). Dietary supplementation with n-3 polyunsaturated fatty acids and vitamin E after myocardial infarction: results of the GISSI-Prevenzione trial. *Lancet* 354: 447–455, 1999.
 50. Gulbins E, Jekle A, Ferlinz H, Grassme H, and Lang F. Physiology of apoptosis. *Am J Physiol Renal Physiol* 279: F605–F615, 2000.
 51. Guyton AC, Coleman TG, Cowley AV Jr, Scheel KW, Manning RD Jr, and Norman RA Jr. Arterial pressure regulation. Overriding dominance of the kidneys in long-term regulation and in hypertension. *Am J Med* 52: 584–594, 1972.
 52. Guyton KC, Liu Y, Gorospe M, Xu Q, and Holbrook NJ. Activation of mitogen-activated protein kinase by H₂O₂. Role in cell survival. *J Biol Chem* 271: 4138–4142, 1996.
 53. Hall JE. The kidney, hypertension and obesity. *Hypertension* 41: 625–633, 2003.
 54. Harman D. Ageing: a theory based on free radical and radiation chemistry. *J Gerontol* 11: 298–300, 1956.
 55. Harman D. The ageing process. *Proc Natl Acad Sci USA* 78: 7124–7128, 1981.
 56. Heagerty AM and Izzard AS. Small-artery changes in hypertension. *J Hypertens* 13: 1560–1565, 1995.
 57. Heart. Outcomes Prevention Evaluation (HOPE) Investigators. Vitamin E supplementation and cardiovascular events in high risk patients. *N Engl J Med* 342: 154–160, 2000.
 58. Heart. Protection Study Collaborative Group. MRC/BHF heart protection study of antioxidant vitamin supplementation in 20,536 high-risk individuals: a randomized placebo-controlled trial. *Lancet* 360: 23–33, 2002.
 59. Hehner SP, Breitkreutz R, Shubinsky G, Unsoeld H, Shulze-Osthoff K, Schmitz ML, and Dröge W. Enhancement of T cell receptor signaling by a mild oxidative shift in the intracellular thiol pool. *J Immunol* 165: 4319–4328, 2000.

60. **Herrera J, Nava M, Romero F, and Rodriguez-Iturbe B.** Melatonin prevents oxidative stress from iron and erythropoietin administration. *Am J Kidney Dis* 37: 750–757, 2001.
61. **Higashi Y, Sasaki S, Nakagawa K, Matsuura H, Oshima T, and Chayama K.** Endothelial dysfunction and oxidative stress in renovascular hypertension. *N Engl J Med* 346: 1954–1962, 2002.
62. **Hogg N, Struck SP, Goss N, Santanam J, Joseph J, Parthasarathy S, and Kalyanaraman B.** Inhibition of macrophage-dependent low density lipoprotein oxidation by nitric-oxide donors. *J Lipid Res* 36: 1756–1762, 1995.
63. **Irani K.** Oxidant signaling in vascular cell growth, death and survival. A review of the roles of reactive oxygen species in smooth muscle and endothelial cell mitogenic and apoptotic signaling. *Circ Res* 87: 179–183, 2000.
64. **Ishizaka N, Aizawa T, Ohno M, Usui Si S, Mori I, Tang SS, Ingelfinger JR, Kimura S, and Nagai R.** Regulation and localization of HSP70 and HSP25 in the kidney of rats undergoing long-term administration of angiotensin II. *Hypertension* 39: 122–128, 2002.
65. **Johnson RJ, Herrera J, Schreiner G, and Rodríguez-Iturbe B.** Acquired and subtle renal injury as a mechanism for salt-sensitive hypertension: bridging the hypothesis of Goldblatt and Guyton. *N Engl J Med* 346: 913–923, 2002.
66. **Johnson RJ, Kang DH, Feig D, Kivlighn S, Kanellis J, Watanabe S, Tuttle KR, Rodríguez-Iturbe B, Herrera-Acosta J, and Mazzali M.** Is there a pathogenetic role for uric acid in hypertension and cardiovascular and renal disease? *Hypertension* 41: 1183–1190, 2003.
67. **Jones SA, Hancock J, Jones OTG, Neubauer A, and Topley N.** The expression of NADPH oxidase components in human glomerular mesangial cells: detection of protein and mRNA for p47phox, p67phox, and p22 phox. *J Am Soc Nephrol* 5: 1483–1491, 1995.
68. **Jones SA, O'Donnell VB, Wood JD, Broughton JP, Hughes EJ, and Jones OT.** Expression of phagocyte NADPH oxidase components in human endothelial cells. *Am J Physiol Cell Physiol* 271: C626–C634, 1996.
69. **Jones SA, Wood JD, Coffey MJ, and Jones OT.** The functional expression of p47-phox and p67-phox may contribute to the generation of superoxide by NADPH oxidase-like system in human fibroblasts. *FEBS Lett* 355: 178–182, 1994.
70. **Jurcevic S, Ainsworth ME, Pomerance A, Smith JD, Robinson DR, Dun MJ, Yacoub MH, and Rose ML.** Antivimentin antibodies are an independent predictor of transplant-associated coronary artery disease after cardiac transplantation. *Transplantation* 71: 886–892, 2001.
71. **Kharitonov VVG, Sharma VS, Magde D, and Koesling D.** Kinetics of nitric oxide dissociation from five- and six-coordinate nitrosyl hemes and heme proteins, including soluble guanylate cyclase. *Biochemistry* 36: 6814–6818, 1997.
72. **Khraibi AA.** Associations between disturbances of the immune system and hypertension. *Am J Hypertens* 4: 635–641, 1991.
73. **Khraibi AA, Smith TL, Hutchins PM, Lynch CD, and Dusseau JW.** Thymectomy delays the development of hypertension in Okamoto spontaneously hypertensive rats. *J Hypertens* 5: 537–541, 1987.
74. **Kim MK, Sasaki S, Sasazuki S, Okubo S, Hayashi M, and Tsugane S.** Lack of long-term effect of vitamin C supplementation on blood pressure. *Hypertension* 40: 797–803, 2002.
75. **Kobayashi S, Ishida A, Moriya H, Mori N, Fukuda T, and Takamura T.** Angiotensin II receptor blockade limits kidney injury in two-kidney one-clip Goldblatt hypertensive rats with special reference to phenotypic changes. *J Lab Clin Med* 133: 134–143, 1999.
76. **Kol A, Bourcier T, Lichtman A, and Libby P.** Chlamydial and human heat shock protein 60 activate human vascular endothelium, smooth muscle cells and macrophages. *J Clin Invest* 103: 571–577, 1999.
77. **Koo JR, Oveisi F, Ni Z, and Vaziri ND.** Antioxidant therapy potentiates antihypertensive action of insulin in diabetes. *Clin Exp Hypertension* 24: 333–344, 2002.
78. **Koo JR and Vaziri ND.** Effect of diabetes, insulin and antioxidants on NO synthase abundance and NO interaction with reactive oxygen species. *Kidney Int* 63: 195–201, 2003.
79. **Kunsch C and Medford RM.** Oxidative stress as a regulator of gene expression in the vasculature. *Circ Res* 85: 753–766, 1999.
80. **Kvist S and Mulvany MJ.** Contrasting regression of blood pressure and cardiovascular structure in decliped renovascular hypertensive rats. *Hypertension* 41: 540–545, 2003.
81. **Laursen JB, Rjagopalan S, Galis Z, Tarpey M, Freeman BA, and Harrison DG.** Role of superoxide in angiotensin II-induced but not in catecholamine-induced hypertension. *Circulation* 95: 588–593, 1997.
82. **Leclercq B, Jaimes EA, and Raij L.** Nitric oxide synthase and hypertension. *Curr Opin Nephrol Hypertens* 11: 185–189, 2002.
83. **Los M, Schenk H, Hexel K, Baeurle PA, Dröge W, and Schulze-Osthoff K.** IL-2 gene expression and NF- κ B activation through CD28 requires reactive oxygen production by 5-lipoxygenase. *EMBO J* 14: 3731–3740, 1995.
84. **Lounsbury KM, Hu Q, and Ziegelstein RC.** Calcium signaling and oxidant stress in the vasculature. *Free Rad Biol Med* 28: 1362–1369, 2000.
85. **Makino A, Skelton MM, Zou AP, Roman RJ, and Cowley AW Jr.** Increased renal medullary oxidative stress produces hypertension. *Hypertension* 39: 667–672, 2002.
86. **Martindale JL and Holbrook NJ.** Cellular response to oxidative stress: signaling for suicide and survival. *J Cell Physiol* 192: 1–15, 2002.
87. **Mazzali M, Hughes J, Kim YG, Jefferson JA, Kang DH, Gordon KL, Lan HY, Kivlighn S, and Johnson RJ.** Elevated uric acid increases blood pressure in the rat by a novel crystal-independent mechanism. *Hypertension* 38: 1101–1106, 2001.
88. **Mervaala E, Finckenberg P, Lapatto R, Muller DN, Park JK, Dechend R, Ganten D, Vapaatalo H, and Luft FC.** Lipoic acid supplementation prevents angiotensin II-induced renal injury. *Kidney Int* 64: 501–508, 2003.
89. **Mevorach D.** Systemic exposure to irradiated apoptotic cells induces autoantibody production. *J Exp Med* 188: 387–392, 1998.
90. **Mevorach D.** The immune response to apoptotic cells. *Ann NY Acad Sci* 887: 191–198, 1999.
91. **Miagkov AV, Kovalenko DV, Brown CE, Didsbury JR, Cogswell JP, Stimpson SA, Baldwin AS, and Makarov SS.** NF- κ B activation provides the potential link between inflammation and hyperplasia in the arthritic joint. *Proc Natl Acad Sci USA* 95: 13859–13864, 1998.
92. **Mor-Vaknin N, Punturieri A, Sitwala K, and Markovitz DM.** Vimentin is secreted by activated macrophages. *Nat Cell Biol* 5: 59–63, 2003.
93. **Mullan B, Young IS, Fee H, and McCance DR.** Ascorbic acid reduces blood pressure and arterial stiffness in type 2 diabetes. *Hypertension* 40: 804–809, 2002.
94. **Müller DN, Dechend R, Mervaala EM, Park JK, Schmidt F, Fiebeler A, Theuer J, Breu V, Ganten D, Haller H, and Luft FC.** NF κ B inhibition ameliorates angiotensin II-induced inflammatory damage in rats. *Hypertension* 35: 193–201, 2000.
95. **Müller DN, Shagdarsuren E, Park JK, Dechend R, Mervaala E, Hampic F, Fiebeler A, Ju X, Finckenberg P, Theuer J, Viedt C, Kreuzer J, Heidecke H, Haller H, Zenke M, and Luft FC.** Immunosuppressive treatment protects against angiotensin II-induced renal damage. *Am J Pathol* 161: 1679–1693, 2002.
96. **Multhoff G and Botzler C.** Heat shock response and the immune response. *Ann NY Acad Sci* 851: 86–93, 1998.
97. **Mulvany MJ.** Resistance vessel growth and remodeling: cause or consequence in cardiovascular disease. *J Hum Hypertension* 9: 479–485, 1995.
98. **Nath KA.** Tubulointerstitial changes as a major determinant in the progression of renal damage. *Am J Kidney Dis* 20: 1–17, 1992.
99. **Nava M, Quiroz Y, Vaziri N, and Rodríguez-Iturbe B.** Melatonin reduces renal interstitial inflammation and improves hypertension in spontaneously hypertensive rats. *Am J Physiol Renal Physiol* 284: F447–F454, 2003.
100. **Navar LG, Harrison-Bernard LM, Nishiyama A, and Kobori H.** Regulation of intrarenal angiotensin II in hypertension. *Hypertension* 39: 316–322, 2002.
101. **Nishiyama A, Seth DM, and Navar LG.** Renal interstitial fluid angiotensin I and angiotensin II concentrations during local angiotensin-converting enzyme inhibition. *J Am Soc Nephrol* 13: 2207–2212, 2002.
102. **Noguchi T, Ikeda K, Sasaki Y, Yamamoto J, Seki J, Yamagata K, Nara I, Hara H, Kakuta H, and Yamori Y.** Effects of vitamin E and sesamin on hypertension and cerebral thrombogenesis in stroke-prone spontaneously hypertensive rats. *Hypertens Res* 24: 735–742, 2001.
103. **Park HS, Lee JS, Huh SH, Seo JS, and Coi EJ.** Hsp72 functions as a natural inhibitory protein of c-Jun N-terminal kinase. *EMBO J* 20: 446–456, 2001.
104. **Park JB, Touyz RM, Chen X, and Schiffrin EL.** Chronic treatment with a superoxide dismutase mimetic prevents remodeling, and progres-

- sion of hypertension in salt-loaded stroke-prone spontaneously hypertensive rats. *Am J Hypertens* 15: 78–84, 2002.
105. Polla BS, Bachelet M, Elia G, and Santoro MG. Stress proteins in inflammation. *Ann NY Acad Sci* 851: 75–85, 1998.
 106. Quiroz Y, Bravo J, Herrera-Acosta J, Johnson RJ, and Rodríguez-Iturbe B. Increased apoptosis and NF- κ B activation are simultaneously induced in renal tubulointerstitium in experimental models of hypertension. *Kidney Int* 64, Suppl 86: S27–S32, 2003.
 107. Quiroz Y, Pons H, Gordon KL, Rincón J, Chávez M, Parra G, Herrera-Acosta J, Gómez-Garre D, Largo R, Egido J, Johnson RJ, and Rodríguez-Iturbe B. Mycophenolate mofetil prevents salt-sensitive hypertension resulting from nitric oxide synthesis inhibition. *Am J Physiol Renal Physiol* 281: F38–F47, 2001.
 108. Ragopalan S, Kurz S, Münzel T, Tarpey M, Freeman BA, Griendling KK, and Harrison DG. Angiotensin II-mediated hypertension in the rat increases vascular superoxide production via membrane NADH/NADPH oxidase activation: contribution to alterations to vasomotor tone. *J Clin Invest* 97: 1916–1923, 1996.
 109. Remuzzi G, Zoja C, Gagliardini E, Corna D, Abbate M, and Benigni A. Combining an antiproteinuric approach with mycophenolate mofetil fully suppresses progressive nephropathy of experimental animals. *J Am Soc Nephrol* 10: 1542–1549, 1999.
 110. Roberts CK, Vaziri ND, and Barnard RJ. Effect of diet and exercise intervention on blood pressure, insulin resistance, oxidative stress and nitric oxide availability. *Circulation* 106: 2530–2532, 2002.
 111. Roberts CK, Vaziri ND, Wang XQ, and Barnard RJ. Enhanced NO inactivation induced by a high fat, refined-carbohydrate diet. *Hypertension* 36: 423–429, 2000.
 112. Rodríguez-Iturbe B, Herrera-Acosta J, and Johnson RJ. Interstitial inflammation, sodium retention, and the pathogenesis of nephrotic edema: a unifying hypothesis. *Kidney Int* 62: 1379–1384, 2002.
 113. Rodríguez-Iturbe B, Pons H, Herrera-Acosta J, and Johnson RJ. The role of immunocompetent cells in non-immune renal diseases. *Kidney Int* 59: 1626–1640, 2001.
 114. Rodríguez-Iturbe B, Pons H, Quiroz Y, Gordon K, Rincón J, Chavez M, Parra G, Herrera-Acosta J, Gomez-Garre D, Largo R, Egido J, and Johnson RJ. Mycophenolate mofetil prevents salt-sensitive hypertension resulting from angiotensin II exposure. *Kidney Int* 59: 2222–2232, 2001.
 115. Rodríguez-Iturbe B, Quiroz Y, Herrera-Acosta J, Johnson RJ, and Pons HA. The role of immune cells infiltrating the kidney in the pathogenesis of salt-sensitive hypertension. *J Hypertens* 20, Suppl 3: S9–S14, 2002.
 116. Rodríguez-Iturbe B, Quiroz Y, Nava M, Bonet L, Chavez M, Herrera-Acosta J, Johnson RJ, and Pons HA. Reduction of renal immune cell infiltration results in blood pressure control in genetically hypertensive rats. *Am J Physiol Renal Physiol* 282: F191–F201, 2002.
 117. Rodríguez-Iturbe B, Zhan CD, Quiroz Y, Sindhu RK, and Vaziri ND. Antioxidant-rich diet relieves hypertension and reduces renal immune infiltration in spontaneously hypertensive rats. *Hypertension* 41: 341–346, 2003.
 118. Romero F, Herrera J, Nava M, and Rodríguez-Iturbe B. Oxidative stress in renal transplantation with uneventful postoperative course. *Transplant Proc* 31: 2315–2316, 1999.
 119. Romero F, Rodríguez-Iturbe B, Parra G, Gonzalez L, Herrera-Acosta J, and Tapia E. Mycophenolate mofetil prevents the progressive renal failure induced by $\frac{2}{3}$ renal ablation in rats. *Kidney Int* 55: 945–955, 1999.
 120. Romero JC and Reckelhoff JF. Role of angiotensin and oxidative stress in arterial hypertension. *Hypertension* 34: 943–949, 1999.
 121. Roth S and Dröge W. Glutathione reverses the inhibition of T cell responses by superoptimal numbers of “nonprofessional” antigen presenting cells. *Cell Immunol* 155: 183–194, 1994.
 122. Rubanyi GM. Vascular effects of oxygen-derived free radicals. *Free Radic Biol Med* 4: 107–120, 1988.
 123. Ruiz-Ortega M, Lorenzo O, Rupérez M, Köning S, Wittig B, and Egido J. Angiotensin II activates nuclear transcription factor κ B through AT₁ and AT₂ in vascular smooth muscle cells: molecular mechanisms. *Circ Res* 86: 1266–1272, 2000.
 124. Sato Y, Matsumori A, and Sasayama S. Autoantibodies against vimentin in a murine model of myocarditis. *Autoimmunity* 18: 145–148, 1994.
 125. Schnackenberg CG. Physiological and pathophysiological roles of oxygen radicals in the renal microvasculature. *Am J Physiol Regul Integr Comp Physiol* 282: R335–R342, 2002.
 126. Schnackenberg CG, Welch WJ, and Wilcox CB. TP receptor-mediated vasoconstriction in microperfused afferent arterioles: roles of O₂⁻ and NO. *Am J Physiol Renal Physiol* 279: F302–F305, 2000.
 127. Schnackenberg CG, Welch WJ, and Wilcox CS. Normalization of blood pressure and renal vascular resistance in SHR with a membrane permeable superoxide dismutase mimetic: role of nitric oxide. *Hypertension* 32: 59–64, 1998.
 128. Schnackenberg CG and Wilcox CS. Two-week administration of tempol attenuates both hypertension and renal excretion of 8-iso prostaglandin F_{2 α} . *Hypertension* 33: 424–428, 1999.
 129. Schreck R, Rieber P, and Baeuerle PA. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF- κ B transcription factor and HIV. *EMBO J* 10: 2247–2258, 1991.
 130. Shoonbroodt S, Ferreira V, Best-Belpomme M, Boelaert JR, Legrand-Poels S, Korner M, and Piette J. Crucial role of the amino-terminal tyrosine residue 42 and the carboxy-terminal PEST domain of I κ B α in NF- κ B activation by an oxidative stress. *J Immunol* 164: 4292–4300, 2000.
 131. Shovman O, Levy Y, Gilburd B, and Shoenfeld Y. Antiinflammatory and immunomodulatory properties of statins. *Immunol Res* 25: 271–285, 2002.
 132. Stephens NG, Parsons A, Schofield PM, Kelly F, Cheeseman K, and Mitchinson MJ. Randomized controlled trial of vitamin E in patients with coronary disease: Cambridge Heart Antioxidant Study (CHAOS). *Lancet* 347: 781–786, 1996.
 133. Struck AT, Hogg N, Thomas JP, and Kalyanaraman B. Nitric oxide donor compounds inhibit the toxicity of oxidized low-density lipoprotein to endothelial cells. *FEBS Lett* 361: 291–294, 1995.
 134. Suzuki Y, Ruiz-Ortega M, Lorenzo O, Rupérez M, Esteban V, and Egido J. Inflammation and angiotensin II. *Int J Biochem Cell Biol* 35: 881–900, 2003.
 135. Svendsen UG. Evidence for an initial, thymus independent and a chronic, thymus dependent phase of DOCA and salt hypertension in mice. *Path Microbiol Scand Acta* 84: 523–528, 1976.
 136. Taddei S, Virdis A, Ghiadoni L, Magagna A, and Salvetti A. Vitamin C improves endothelium-dependent vasodilatation by restoring nitric oxide activity in essential hypertension. *Circulation* 97: 2222–2229, 1998.
 137. Tak PP and Firestein GS. NF- κ B: a key role in inflammatory diseases. *J Clin Invest* 107: 7–11, 2001.
 138. Takahashi K, Nammour TM, Fukunaga M, Ebert J, Morrow JD, Roberts LJ, Hoover RL, and Badr KF. Glomerular actions of a free radical-generated novel prostaglandin, 8-epiprostaglandin F_{2 α} . Evidence for interaction with thromboxane A₂ receptors. *J Clin Invest* 90: 136–141, 1992.
 139. Tapia E, Franco M, Sanchez-Lozada LG, Soto V, Avila-Casado C, Santamaria J, Quiroz Y, Rodríguez-Iturbe B, and Herrera-Acosta J. Mycophenolate mofetil prevents arteriopathy and renal injury in subtotal renal ablation. *Kidney Int* 63: 994–1002, 2003.
 140. Touyz RM. Reactive oxygen species in vascular biology: role in arterial hypertension. *Expert Rev Cardiovasc Ther* 1: 91–106, 2003.
 141. Touyz RM and Schiffrin EL. Increased generation of superoxide by angiotensin II in smooth muscle cells from resistance arteries of hypertensive patients: role of phospholipase D-dependent NAD(H)P oxidase-sensitive pathways. *J Hypertens* 19: 1245–1254, 2001.
 142. Udono H, Yamano T, Kawabata Y, Ueda M, and Yui K. Generation of cytotoxic T lymphocytes by MHC class I ligands fused to heat shock cognate protein 70. *Int Immun* 13: 1233–1242, 2001.
 143. Vaziri ND, Dicus M, Ho N, and Sindhu RK. Oxidative stress, and dysregulation of superoxide dismutase, NAD(P)H oxidase and xanthine oxidase in chronic renal insufficiency. *Kidney Int* 63: 179–185, 2003.
 144. Vaziri ND, Ding Y, and Ni Z. Compensatory up-regulation of nitric oxide synthase isoforms in lead-induced hypertension: reversal by a superoxide dismutase mimetic drug. *J Pharmacol Exp Ther* 298: 679–685, 2001.
 145. Vaziri ND, Liang K, and Ding Y. Increased nitric oxide inactivation by reactive oxygen species in lead-induced hypertension. *Kidney Int* 56: 1492–1498, 1999.
 146. Vaziri ND, Ni Z, Oveisi F, and Trnavsky-Hobbs DL. Effect of antioxidant therapy on blood pressure, and NO synthase expression in hypertensive rats. *Hypertension* 36: 957–964, 2000.

147. **Vaziri ND, Wang XQ, Oveisi F, and Rad B.** Induction of oxidative stress by glutathione depletion causes severe hypertension in normal rats. *Hypertension* 36: 142–146, 2000.
148. **Weinstock JV and Blum AM.** Synthesis of angiotensins by cultured granuloma macrophages in murine schistosomiasis mansoni. *Cell Immunol* 1107: 273–280, 1987.
149. **Weiss RA, Madaio MP, Tomaszewski JE, and Kelly CJ.** T cells reactive to an inducible heat shock protein induce disease in toxin-induced interstitial nephritis. *J Exp Med* 180: 2239–2250, 1994.
150. **Wilcox CS.** Reactive oxygen species: roles in blood pressure and kidney function. *Curr Hypertens Rep* 4: 160–166, 2002.
151. **Wilcox CS and Welch WJ.** Oxidative stress: cause or consequence of hypertension. *Exp Biol Med* 226: 619–620, 2001.
152. **Wu R, Lamontagne D, and de Champlain J.** Antioxidative properties of acetylsalicylic acid on vascular tissues from normotensive and spontaneously hypertensive rats. *Circulation* 105: 387–392, 2002.
153. **Xia Z, Dickens M, Raungeaud J, Davies RJ, and Greenberg ME.** Opposing effects of ERK and JNK-p38 MAP kinases on apoptosis. *Science* 270: 1326–1331, 1995.
154. **Zalba G, Beaumont FJ, San José G, Fortuño A, Fortuño MA, Etayo JC, and Diez J.** Vascular NADH/NADPH oxidase is involved in enhanced superoxide production in spontaneously hypertensive rats. *Hypertension* 35: 1055–1061, 2000.
155. **Zalba G, San Jose G, Moreno MU, Fortuno MA, Fortuno A, Beaumont FJ, and Diez J.** Oxidative stress in arterial hypertension: role of NAD(P)H oxidase. *Hypertension* 38: 1395–1399, 2001.
156. **Zatz R, Noronha IL, and Fujihara CK.** Experimental and clinical rationale for use of MMF in nontransplant progressive nephropathies. *Am J Physiol Renal Physiol* 283: F1167–F1175, 2002.
157. **Zhang J, Johnston G, Stebler B, and Keller ET.** Hydrogen peroxide activates NF- κ B-inducing kinase. *Antiox Redox Signal* 3: 493–504, 2001.
158. **Zhou H, Summer SA, Birnbaum MJ, and Pittman RN.** Inhibition of Akt kinase by cell-permeable ceramide and its implications for ceramide-induced apoptosis. *J Biol Chem* 273: 16568–16575, 1998.
159. **Zihai L, Menoret A, and Srivastava P.** Roles of heat shock proteins in antigen presentation and cross-presentation. *Curr Opin Immunol* 14: 45–51, 2002.
160. **Zou MH and Ulrich V.** Peroxynitrite formed by simultaneous generation of nitric oxide and superoxide selectively inhibits bovine aortic prostacyclin synthase. *FEBS Lett* 382: 101–104, 1996.

