

# Function and Pharmacology of Spinally-Projecting Sympathetic Pre-Autonomic Neurones in the Paraventricular Nucleus of the Hypothalamus

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**Abstract:** The paraventricular nucleus (PVN) of the hypothalamus has been described as the "autonomic master controller". It co-ordinates critical physiological responses through control of the hypothalamic-pituitary-adrenal (HPA)-axis, and by modulation of the sympathetic and parasympathetic branches of the central nervous system. The PVN comprises several anatomical subdivisions, including the parvocellular/mediocellular subdivision, which contains neurones projecting to the medulla and spinal cord. Consensus indicates that output from spinally-projecting sympathetic pre-autonomic neurones (SPANs) increases blood pressure and heart rate, and dysfunction of these neurones has been directly linked to elevated sympathetic activity during heart failure. The influence of spinally-projecting SPANs on cardiovascular function highlights their potential as targets for future therapeutic drug development. Recent studies have demonstrated pharmacological control of these spinally-projecting SPANs with glutamate, GABA, nitric oxide, neuroactive steroids and a number of neuropeptides (including angiotensin, substance P, and corticotrophin-releasing factor). The underlying mechanism of control appears to be a state of tonic inhibition by GABA, which is then strengthened or relieved by the action of other modulators. The physiological function of spinally-projecting SPANs has been subject to some debate, and they may be involved in physiological stress responses, blood volume regulation, glucose regulation, thermoregulation and/or circadian rhythms. This review describes the pharmacology of PVN spinally-projecting SPANs and discusses their likely roles in cardiovascular control.

**Keywords:** Blood pressure, penile erection, GABA, PVN, paraventricular nucleus, hypothalamus, cardiovascular, sympathetic, parvocellular mediocellular, neuropeptides, oxytocin, substance P, tachykinin, vasopressin, angiotensin, pharmacology.

## 1. INTRODUCTION

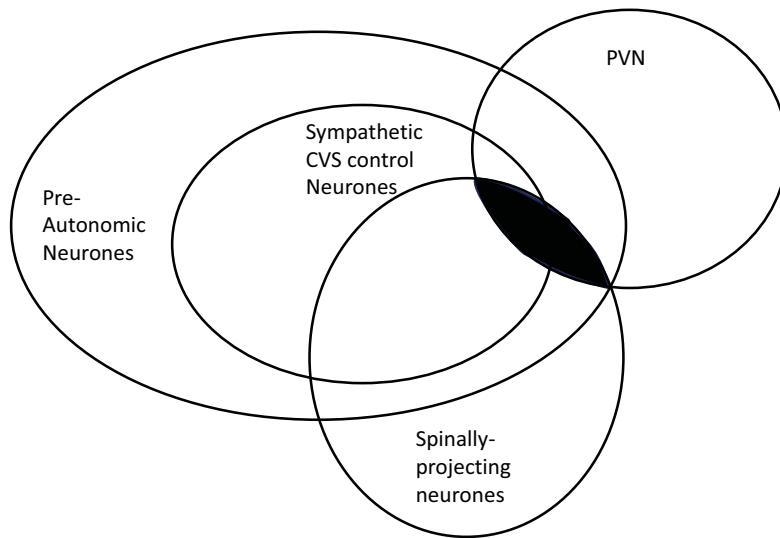
The paraventricular nucleus (PVN) of the hypothalamus is a critical regulator of numerous endocrine and autonomic functions [1-5]; Loewy [6] referred to it as being the "autonomic master controller". Many of these autonomic functions are served by spinally-projecting neurones (Fig. 1), a number of which have been shown to be involved in cardiovascular regulation [7]. These and other spinally-projecting sympathetic pre-autonomic neurones (SPANs) in the PVN are therefore attractive targets for pharmacological manipulation of both the cardiovascular system and potentially, other autonomic systems.

The cardiovascular system is subject to numerous levels of control. Although the basic functions of the heart and blood vessels are controlled locally, the whole system is centrally regulated by the brain [8, 9]. Part of the central regulation comes from the medulla, which allows rapid adaptation to changing homeostatic challenges. Anticipatory regulation, however, allows for an increase in cardiac output just prior to increased systemic oxygen demand. This was termed the "preparatory reflex" [10] and appears to be the highest level of cardiovascular control. The process is largely attributed to an increase in sympathetic activity, and was first identified

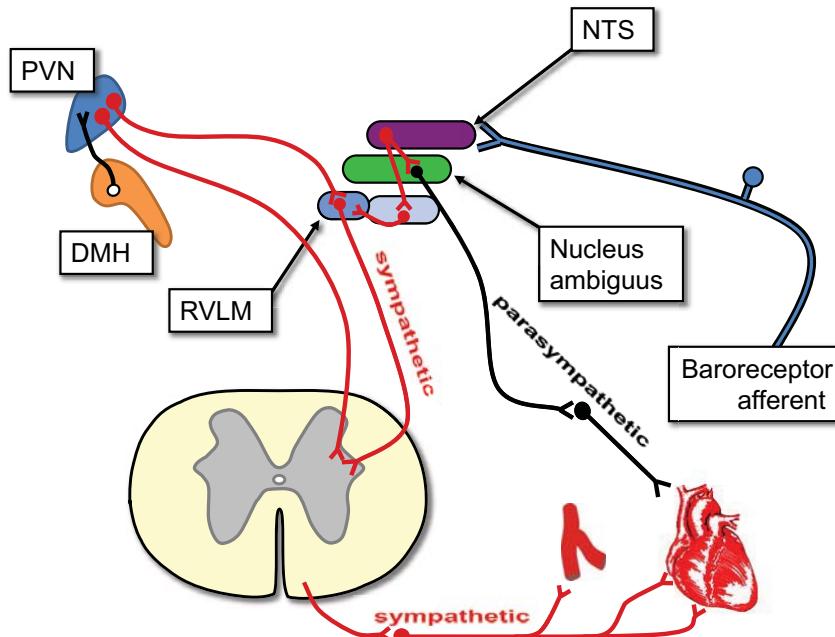
by Walter Cannon [11]. This reflex is now more commonly referred to as the "fight-or-flight" response, the "stress response", or the "defence reaction". Inappropriate activation of this system may lead to hypertension and could account for the increased cardiac death resulting from stressors such as exercise [12], anger [13] or emotional shock [14]. Furthermore, some aspects of this pathway appear to be activated in chronic heart failure [15]. Forebrain regions involved in cardiovascular regulation were discovered early in the 20<sup>th</sup> century by stimulation or ablation of specific areas [16, 17]. Later, several areas of the hypothalamus (Fig. 2) were shown to be of particular importance [e.g., 18, 19, 20]. These include areas we now know as the PVN and the dorsomedial hypothalamus (DMH), although both were poorly defined at the time. The PVN comprises approximately 21,500 neurones, approximately three quarters of which make up the parvocellular subdivision [21], which includes both spinally-projecting SPANs and hypothalamic-pituitary-adrenal (HPA) axis control neurones [3, 6, 22].

Estimates suggest that up to 2000 neurones project directly from the PVN to the intermediolateralis (IML) of the spinal cord [23-25], and terminate close to sympathetic pre-ganglionic neurones [26, 27] and are, in this review, collectively termed spinally-projecting sympathetic pre-autonomic neurones (spinally-projecting SPANs). SPAN cell bodies are found most densely in a region of the PVN termed the parvocellular subnucleus, particularly in a subdivision referred to by some authors as the mediocellular region [21].

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**Fig. (1). Spinally-projecting sympathetic pre-autonomic neurones of the paraventricular nucleus of the hypothalamus (PVN).** The PVN consists of several sub-populations of neurones some of which project to the spinal cord. There are likely to be each combination indicated in the Venn diagram. This review focuses on the shaded area; spinally-projecting sympathetic pre-autonomic neurones (spinally-projecting SPANs) of the PVN. Many of these target the cardiovascular system (CVS).



**Fig. (2). Central cardiovascular control in the rat.** The cardiovascular system is largely controlled by areas of the brainstem; however, there is also strong evidence that centres in the forebrain, including the PVN, exert “top-level” control *via* both medulla-projecting and spinally-projecting SPANs. Afferents from carotid baroreceptors ascend *via* cranial nerve IX (the glossopharyngeal nerve), while afferents from the aortic bodies ascend *via* cranial nerve X (the vagus nerve). These afferents are processed by the nucleus of the solitary tract (NTS). Parasympathetic drive emanates from the nucleus ambiguus, whilst sympathetic drive emanates from a number of locations, in particular the rostroventrolateral medulla. The role of the medulla in cardiovascular control is reviewed by [9].

Spinally-projecting SPANs are strongly implicated in cardiovascular control [7, 28-30], but their primary role is unknown (Fig. 3). Several hypotheses have been proposed such as; circadian regulation of blood pressure [31], blood-volume regulation [24, 25] and the cardiovascular response to psychological stress [32]. Furthermore, elevated sympathetic activity is associated with congestive heart failure [15, 33, 34], and may also be associated with diminished GABA

sensitivity of PVN neurones [15, 35, 36]. This review will discuss the neuropharmacology and function of spinally-projecting SPANs of the PVN. In the majority of cases these experiments have focussed on control of the rat cardiovascular system, although recently, study of spinally-projecting SPANs has broadened to entirely new systems such as glucose regulation, thermoregulation and penile erection.

## 2. ELECTROPHYSIOLOGY OF SPINALLY-PROJECTING SPANS

A number of *in vitro* studies have investigated the electrophysiological properties of PVN neurones [37-40]. They show that parvocellular neurones (termed “PVN type II” neurones) express a slowly inactivating delayed rectifier potassium conductance. Conversely, the neurosecretory magnocellular neurones of the PVN (termed “PVN type I” neurones) appear to express a rapidly inactivating (A-type) potassium conductance. Fewer studies have been conducted on *identified* spinally-projecting SPANs; medulla-projecting neurones show strong inward rectification and “A-type” potassium conductance [41, 42] and spinally-projecting SPANs show a slowly inactivating potassium conductance [43]. More recent studies have also identified ATP dependent potassium channels [44, 45], which may serve to couple glucose levels to sympathetic activity. Pharmacological characterisation of the potassium channels involved is possible using potassium channels inhibitors [46-53], although confirmation requires immunohistochemical or RT-PCR approaches since most of these inhibitors lack high selectivity. When recorded from *in vitro* most spinally-projecting SPANs fire action potentials spontaneously [41, 54], but they are apparently quiescent *in vivo* [55-58]. This implies that the tonic inhibition of spinally-projecting SPANs may be, in part at least, lost in the preparation of brain-slices for *in vitro* recording.

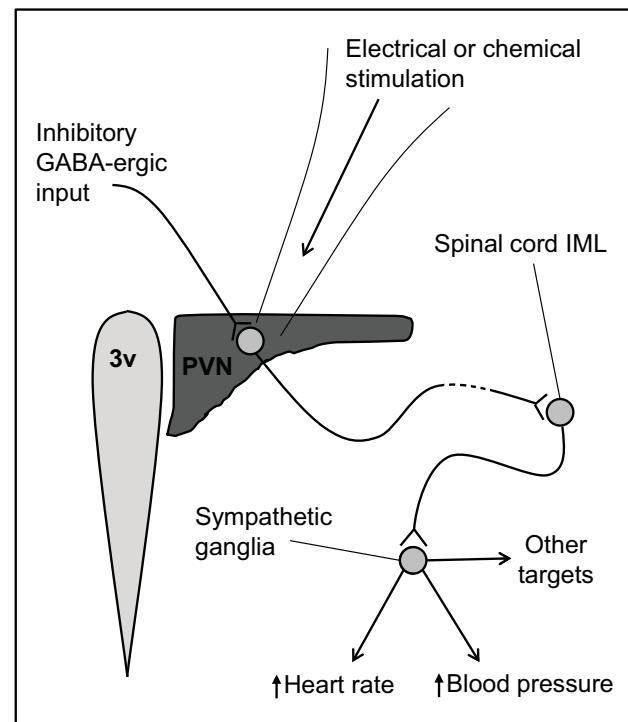
## 3. NEUROTRANSMITTERS RELEASED BY SPANS

Discussion of the neuropharmacology of SPANs can include neurotransmitters released by the neurones and neurotransmitters acting upon them. The first of these questions has been approached by the use of retrograde/anteroventral labelling, trans-synaptic tracing, immunohistochemistry and *in situ* hybridisation. Using these techniques it has been demonstrated that spinally-projecting SPANs express met-enkephalin (20%), dynorphin (up to 40%), oxytocin (up to 40%) and vasopressin (up to 40%) [59-61]. However, a significant proportion also produce dopamine, met-enkephalin (10%), leu-enkephalin, somatostatin, angiotensin II and atrial natriuretic peptide [62-65]. This clearly implies that some SPANs express more than one neuropeptide neurotransmitter. Interesting questions which remain unanswered include: what combinations of neurotransmitters are expressed by particular neurones, and, do different neurotransmitters serve different functions? For example, the closely related neuropeptides oxytocin and vasopressin are commonly co-expressed [see for example 66].

The anatomical evidence that vasopressin acts a SPAN neurotransmitter is supported by functional experiments in which PVN neurones were stimulated, producing an immediate increase in renal sympathetic nerve activity (rSNA) and mean arterial blood pressure [67]. However, these effects were prevented by lower thoracic intrathecal preloading with the vasopressin 1a ( $V_{1a}$ )-receptor antagonist  $d(CH_2)_5[Tyr(Me)]^2, arg^8]vasopressin$  [67]. Interestingly, since oxytocin exerts effects both through the oxytocin receptor (OT-R) and through the  $V_{1a}$ -R [68], some of the effects attributed to oxytocin may in fact be mediated by the latter. However,

oxytocin is clearly important as a neurotransmitter of SPANs, since both vasopressin and oxytocin levels increase in the spinal cord when the PVN is stimulated [69]. Additionally, the tachycardia associated with PVN stimulation is reduced by the highly selective OT-R antagonists  $d(CH_2)_5[Tyr(Me)]^2, Orn^8]$ -oxytocin and L-368,899 [70]. One explanation suggested for the apparent disparity of the effects of oxytocin and vasopressin antagonists is that SPAN neurotransmission in the upper thoracic spinal cord is mediated *via* oxytocin, but in the lower thoracic cord it is mediated by vasopressin [30].

Dopamine has also been strongly implicated as a SPAN neurotransmitter [65], but its activity is complex. Several reports demonstrate that dopamine, and its mimetic apomorphine, excite sympathetic pre-ganglionic neurones in rats (ie. the neurones that SPANs would act upon) [71-74]. However, evidence also suggests that it may serve as an inhibitor of sympathetic activity [75]. Data supporting the function of the other putative SPAN neurotransmitters described above is, so far, more disparate. For example, intrathecal glutamate antagonists reduce the increases in rSNA seen following chemical stimulation of the PVN [75] suggesting that the excitatory amino acid glutamate may act as a SPAN output neurotransmitter. However, whilst enkephalins and angio-



**Fig. (3). Actions of spinally-projecting SPANs originating in the paraventricular nucleus (PVN) of the hypothalamus.** The PVN lies alongside the 3<sup>rd</sup> ventricle (3v), and contains a number of neurones which project to the intermediolateralis (IML) of the spinal cord. When stimulated with excitatory neurotransmitters or electrical current, they activate sympathetic pre-ganglionic neurones in the IML which, in turn, increase heart rate and blood pressure and glucose secretion. They may also increase TSH release. These PVN neurones are tonically inhibited by GABA input. Please see main text for references.

tensin II are both produced by SPANs [59, 60, 62-64] and modulate sympathetic pre-ganglionic neurones [76-78] in rats, pharmacological block of their respective receptors (at the level of the spinal cord) has not been shown directly to prevent the sympathetic effects of PVN stimulation. This may be because SPANs expressing these neurotransmitters innervate pre-ganglionic neurones targeting less well characterised sympathetic effectors in the viscera [79] or simply that appropriate experiments to detect such effects have yet to be conducted. The amino acid  $\gamma$ -aminobutyric acid (GABA) is the most abundant inhibitory neurotransmitter in the central nervous system [80]. Recent analysis of both retrograde and trans-neuronal labelling data shows that whilst spinally-projecting SPANs receive GABA-ergic synapses, they do not contain GABA themselves [81].

In addition to the pathways leading from the PVN to the IML thence to the heart and blood vessels there are pathways projecting from the superchiasmatic nucleus (SCN), to the thyroid and to the liver, *via* the PVN [82-84]. Many neurones in this pathway express thyroid stimulating hormone (TSH) [82], possibly in the PVN spinally-projecting SPANs themselves.

#### 4. NEUROTRANSMITTERS ACTING ON SPANS

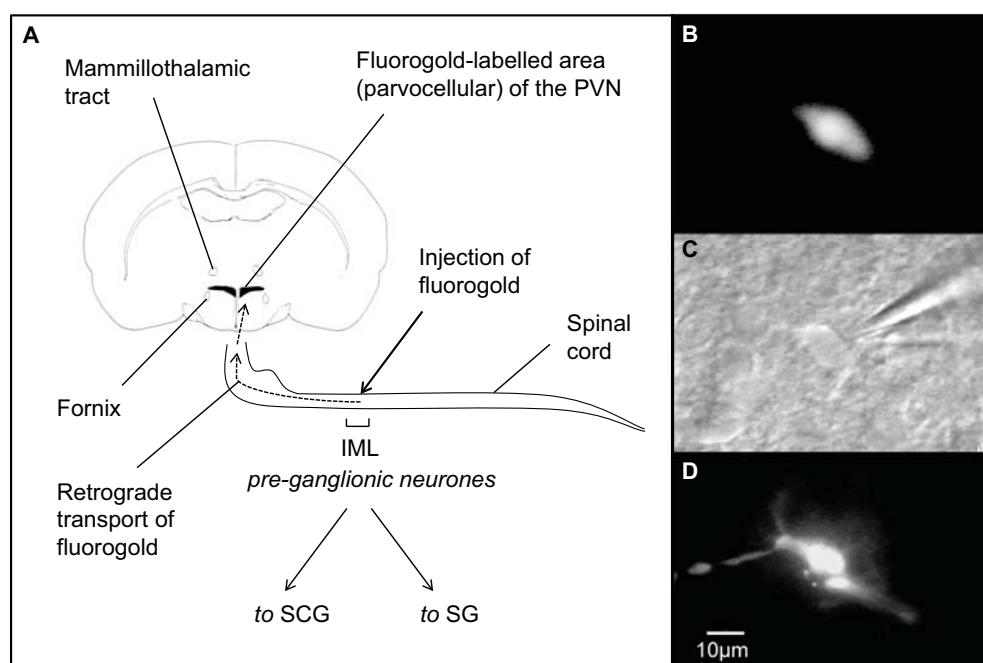
Far more studies have been published looking at the neurotransmitters expressed by SPANs than looking at neurotransmitters *acting on* SPANs. This is disappointing since, arguably, knowledge of the receptors expressed by a cell gives greater therapeutic potential than knowledge of the

transmitters released by it. A useful approach has been the combination of retrograde labelling and patch-clamp recording. In these studies, tracer is injected into the IML of the spinal cord, and a few days later brain slices are prepared. Spinally-projecting neurones are then clearly visible and can be targeted for electrophysiological study (Fig. 4).

#### 4.1. Amino Acid Neurotransmitters

A number of *in vivo* studies have investigated the neurotransmitters acting upon SPANs, or the receptors expressed by them, but there have been few *in vitro* studies on identified spinally-projecting SPANs. *In vivo* electrophysiological studies on cats confirmed an anticipated monosynaptic connection between the PVN and the spinal sympathetic motor area (the IML) [85]. Furthermore, electrical or chemical stimulation of the PVN was shown to generate a rapid rise of blood pressure and rSNA in conscious rats [86]. Further *in vivo* studies showed that PVN neurones receive tonic GABA-ergic inhibition, which maintains their spontaneous action potential discharge at relatively low rates [55, 57, 58, 87], despite additional excitatory glutaminergic input [86].

There is accumulating evidence that “tonic” inhibition is often mediated by GABA arising from extra-synaptic sources; sometimes referred to as “volume” GABA neurotransmission [88-89]. This correlates with the expression of  $\alpha_5$ -, or  $\alpha_6$ -type, and  $\delta$ -type GABA<sub>A</sub> receptor subunits [90, 91]. However, this has not yet been specifically examined with spinally-projecting SPANs. It may be an important issue to explore though, since a unique pharmacological pro-



**Fig. (4). Methods for patch-clamping retrogradely-labelled neurones.** A, the retrograde tracer fluorogold is injected into the rat intermediolateralis (IML) at level T2-T4, it is also possible to use other tracers, such as rhodamine-labelled microspheres (see Fig. 8). The IML is dense with pre-ganglionic neurones that project to the supercervical (SCG) and stellate (SG) ganglia, and from there to the heart and blood vessels [65, 227]. The appearance of a fluorogold-labelled neurone B, prior to patch clamp recording, C, during patch-clamp, under near infrared differential interference contrast microscopy, and D, when patched with Lucifer yellow (a fluorescent dye) in the patch clamp pipette. The dye fills the neurone, and this gives re-confirmation that recording was from the appropriate cell. Reproduced from [43], with permission.

file for extra-synaptic GABA receptors is emerging, [91-93] which may become therapeutically exploitable.

In the case of SPANs, loss of tonic inhibition would lead to an increase in sympathetic outflow. Excessive sympathetic outflow could potentially be reduced by selective activators of “volume” GABA receptors. Furthermore, the degree of tonic inhibition of sympathetic outflow is, in part at least, dependent upon the local uptake of GABA by transporter proteins, in particular GAT3 located on glial cells [94]. Therefore, there are a number of means by which parvocellular GABA inhibition could be targeted with the aim of reducing sympathetic outflow and blood pressure.

Excitation of the PVN by either glutamate or the GABA<sub>A</sub> receptor antagonist bicuculline increases blood pressure and heart rate, due to the tonic silence of SPANs [55, 56, 86, 87, 95]. This has led to many *in vitro* studies investigating control of SPANs to focus on the role of GABA. In fact, a number of studies have demonstrated the presence of GABA<sub>A</sub> receptor currents in the PVN [38, 87, 96-100]. The parvocellular region of the PVN, which contains the majority of SPANs, expresses a high density of GABA<sub>A</sub>  $\alpha_2$ -subunits [101]; this was also seen in retrogradely labelled spinally-projecting SPANs [102]. Additional studies have shown spinally-projecting parvocellular neurones to be inhibited by GABA [31, 54, 103-104], as predicted by the earlier *in vivo* work. Interpretation of both the *in vivo* and *in vitro* work is further complicated by some frequently overlooked variables. Firstly, there is little consideration of the role of pre-synaptic GABA receptors. These are typically of the GABA<sub>B</sub> receptor subtype and have been shown to inhibit both inhibitory and excitatory input to spinally-projecting SPANs [105]. Secondly, the primary GABA<sub>A</sub> antagonist used to investigate possible GABA involvement is bicuculline or bicuculline methiodide, both of which are potassium channel blockers [106-108]. The effect of potassium channel inhibition would appear very similar to block of GABA<sub>A</sub> receptors; excitation.

Neuroactive steroids and nitric oxide (NO) are well known modulators of GABA<sub>A</sub> receptors. In the PVN, for example, tetrahydrodeoxycorticosterone (THDOC) [103] modulates spinally-projecting SPANs and NO modulates rostral ventrolateral medulla (RVLM)-projecting SPANs [109]. Both effects are mediated by modulation of GABA<sub>A</sub> receptors [110]. However, other steroid hormone receptors, such as oestrogen  $\beta$ -receptors [111], are expressed by SPANs, which raises the possibility that some steroids may modulate sympathetic outflow by this mechanism. Other mechanisms may also be involved, for example cortisol is known to activate SPANs at high doses *via* an inhibition of potassium channels [100].

SPANs are excited by the excitatory amino acid neurotransmitter glutamate [31], mediated by the activation of at least two fast ionotropic glutamate conductances. This is carried largely by  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors (i.e., sensitive to 2,3-dihydroxy-6-nitro-7-sulfamoyl-benzo[f]quinoxaline-2,3-dione, NBQX, an AMPA receptor antagonist), but with an additional component of N-methyl D-aspartate (NMDA) conductance (i.e., sensitive to the classical NMDA antagonist amino-5-phosphonovaleric acid, AP-5, [31]).

Combined electrophysiological and microdialysis studies on RVLM-projecting PVN neurones have shown that at least some of their glutaminergic input is also tonically inhibited by GABA<sub>A</sub>-receptors. Since glutaminergic neurotransmission is ubiquitous within the central nervous system, there has been particular recent focus on non-glutaminergic excitation of SPANs. This is because antagonism of the relatively sparse neuropeptide receptors could become therapeutically useful when action of amino acid receptor antagonists proves to be too widespread.

#### 4.2. Neuropeptide Neurotransmitters

Leptin [112], angiotensin II (AGII) [113, 114], neuropeptide W [115] and each of the endogenous tachykinins [54] have all been investigated as pharmacological regulators of spinally-projecting SPANs. The strongest candidates appear to be AGII and the tachykinin substance P (SP), with both neuropeptides acting indirectly *via* GABA neurotransmission [54, 114].

##### 4.2.1. Angiotensin II

The neuropeptide AGII, expressed by PVN neurones [116], acts at two principal receptors: AT<sub>1</sub> and AT<sub>2</sub>. AT<sub>1</sub> is the more abundant of the two [117-118] and, in rodents, is encoded by two genes: AT<sub>1A</sub> and AT<sub>1B</sub> [119]. However, it is not clear whether both of these subtypes exist in humans, nor which subtype is more important for cardiovascular control in rodents. It has been suggested that AT<sub>1B</sub> is more important for control of blood pressure in mice [120], but *in situ* hybridisation studies show that AT<sub>1A</sub> is expressed more richly in the parvocellular region of the rat PVN [121], the region particularly important for autonomic control. Integrative experiments showed that AGII injection into the PVN gave an AT<sub>1</sub>-dependent increase in blood pressure [122], and single-unit recording studies of antidromically-identified neurones demonstrated that this involved spinally-projecting SPANs [123]. Later, the same group demonstrated inhibition of A-type potassium currents by AGII, acting *via* an AT<sub>1</sub> receptor [124]. Whilst AGII directly modulates medulla-projecting PVN neurones [125] to elevate blood pressure, spinally-projecting SPANs do not express AT<sub>1</sub> receptors [113]. Therefore, this implies that AGII activates SPANs indirectly, and this was investigated using whole-cell recording from retrogradely labelled spinally-projecting SPANs [114]. In these experiments, AT<sub>1</sub> receptors were found to co-localise with synaptophysin (a pre-synaptic marker protein). In addition, AGII decreased the frequency and amplitude of spontaneous inhibitory post-synaptic potentials in SPANs. This strongly suggests that AGII excites spinally-projecting SPANs by reducing the tonic GABA-ergic input.

##### 4.2.2. Tachykinins

The tachykinin family of neuropeptides, which includes substance P (SP), neurokinin A (NKA) and neurokinin B (NKB), is involved in central cardiovascular control [126]. Studies have shown high levels of tachykinin receptor expression within the PVN [127-129] and in our own studies we showed that injection of SP into the PVN increases heart rate, blood pressure (Fig. 6) and rSNA [54]. These effects correlated with our observation of a strong inhibition of SPAN GABA<sub>A</sub> receptors by SP [54]. Activation of SPANs

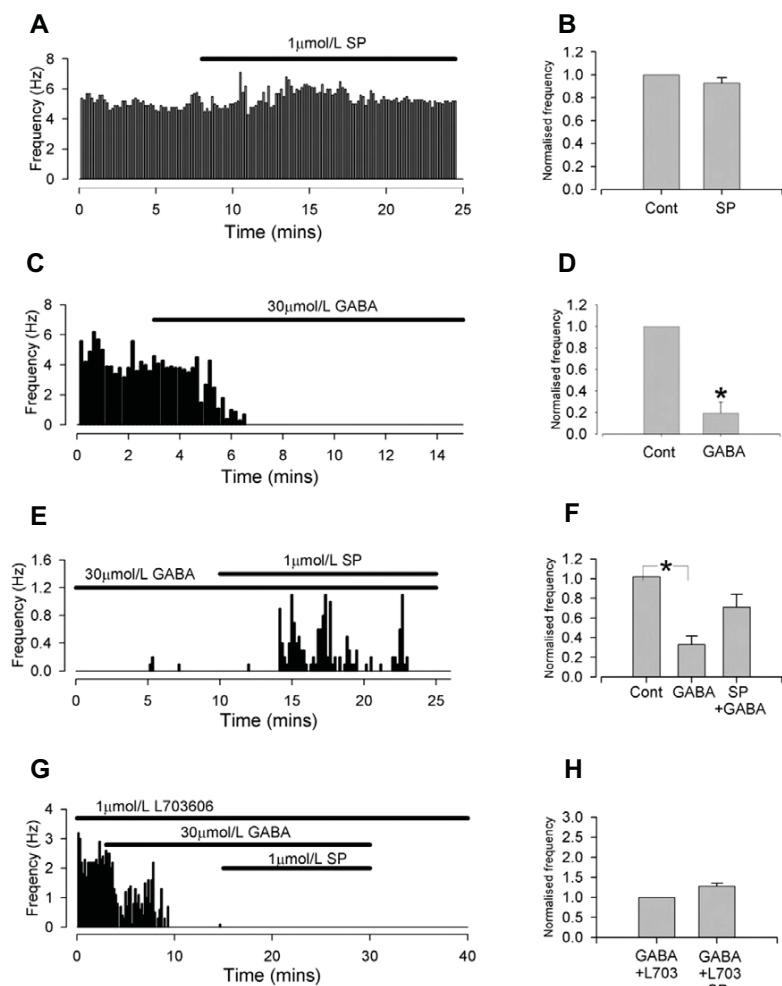
by SP in brain slice experiments was only apparent when neurones were first inhibited by GABA (Fig. 5).

All three of the principal mammalian tachykinin receptors (NK1, NK2 and NK3) are 7-transmembrane G-protein coupled receptors, although they act through a range of intracellular pathways, typically *via* pertussis toxin independent G-proteins [130, 131]. Of particular interest is that NK1 receptors activate protein kinase C (PKC) [131], and that PKC-dependent phosphorylation of GABA<sub>A</sub> can inhibit GABA<sub>A</sub> currents [132]. This was first shown using cultured cells expressing recombinant GABA<sub>A</sub> subunits. However, SPAN activation by SP in the PVN was also found to be both PKC and NK1 receptor dependent (Figs. 5 and 7) [54]. No effects of NKA or NKB agonists were observed (NKA4-10 and senktide, respectively). An interesting question remains; from where do the tachykinin inputs to SPANs arise? Of note, several intra-hypothalamic tachykinin pathways

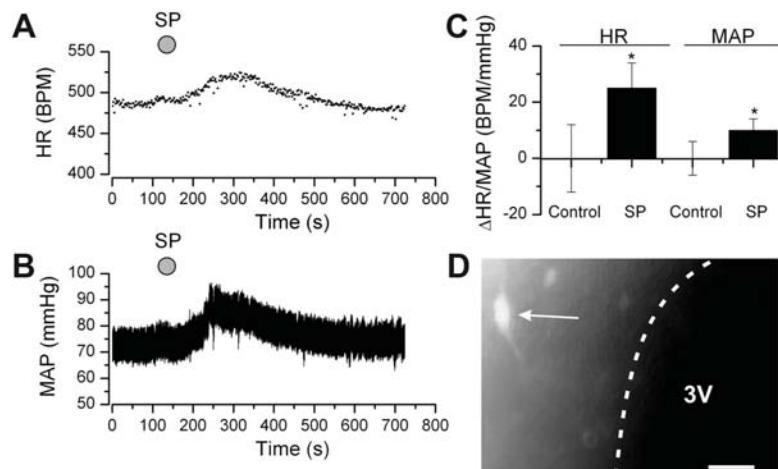
terminate in the PVN [133]. Furthermore, in brain slice experiments we found that SPAN activation following glutamate stimulation of the DMH was inhibited by tachykinin receptor antagonists [134]. This may prove significant since there is evidence that both the DMH and the PVN are important in the control of the cardiovascular system [135].

#### 4.2.3. Vasactive Intestinal Peptide (VIP) and Pituitary Adenylcyclase Activating Peptide (PACAP)

Retrograde labelling evidence has demonstrated the presence of vasactive intestinal peptide/pituitary adenylcyclase activating peptide receptors type 2 (VPAC<sub>2</sub>) on spinally-projecting SPANs [136]. PACAP<sub>(1 to 38)</sub> induced *c-fos* expression in PVN spinally-projecting neurones and this was abolished by a VPAC<sub>2</sub> receptor antagonist [136]. It seems likely, from functional and viral tracing studies that these neurones are also activated by thyroid hormone (T3) [137], although receptors have not yet been identified specifically on the



**Fig. (5). Substance P (SP) increases the rate of action potential discharge of individual neurones in brain slices in the presence of GABA.** Brain slices were taken from adult rats, and action potentials recorded from single neurones in the PVN. **A**, representative results showing SP applied to SPANs under control conditions; **B**, mean results, showing SP alone has no effect on rate of action potential discharge. **C**, representative results showing application of GABA; **D**, mean results showing that GABA inhibits spontaneous action potential frequency. **E**, representative results showing SP added during GABA inhibition; **F**, mean results showing that SP increases the rate of action potential discharge against a background of tonic GABA inhibition. **G**, Representative results showing SP interaction with NK1 antagonism; **H**, mean results showing the effects of SP are abolished by the selective NK1 antagonist L703606. Cont = control. Reproduced from [54], with permission.



**Fig. (6). Substance P (SP) injected into the PVN elicits a rise in heart rate and blood pressure.** Heart rate and blood pressure were recorded in anaesthetised adult rats. Representative records of A, heart rate and B, blood pressure during intra-PVN injection of SP/FITC-SP (solution of SP and FITC-conjugated SP). C, mean data from a number of experiments. D, epifluorescent image of the medial PVN, after SP/FITC-SP intra-PVN injection. “3V” indicates the third ventricle; the white arrow indicates a parvocellular neurone labelled with FITC-SP. Scale bar is 50 μm. “SP” indicates SP/FITC-SP injection. Reproduced from [54], with permission.

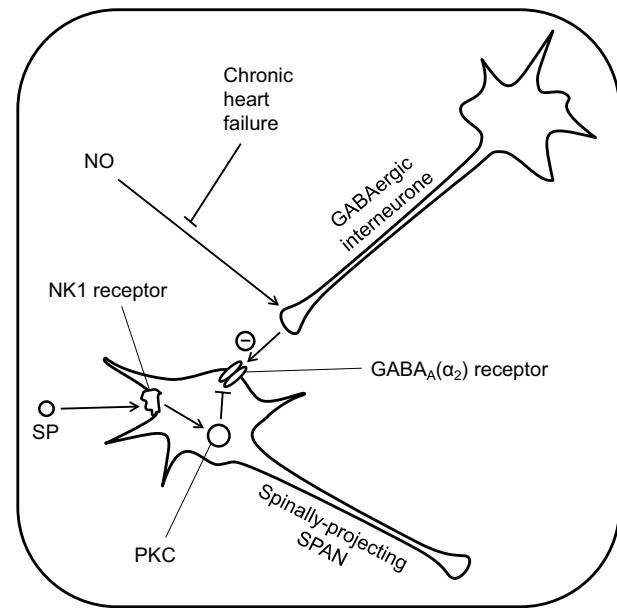
PVN spinally-projecting SPANs. Both the VPAC<sub>2</sub> receptors and T3 nuclear receptors (TR) appear to be involved with control of endogenous glucose production by the liver, see below for further discussion [136, 137].

#### 4.3. Other Receptors on Spinally-Projecting SPANs

Purinergic receptors were originally classed as either P1 or P2 subtypes [138]. The P1 receptors are now termed adenosine receptors, A<sub>x</sub> and are seven transmembrane G-protein couple receptors [139]. The P2 receptors are the ligand-gate cation channels activated by nucleotides [139]. Examples of both families of receptors have now been identified on PVN SPANs [45, 140], but only P1 on identified spinally-projecting SPANs [45]. Adenosine is inhibitory to several types of muscle cell, by acting via A<sub>1</sub> receptors and K<sub>ATP</sub> channels [141–143]. Li *et al.* [45] has now established that adenosine acts on spinally-projecting SPANs in a similar manner. P2X receptors have been identified on RLVM-projecting SPANs [140], but it remains to be seen if these receptors are also expressed on spinally-projecting SPANs. There are little data supporting the presence of other receptors on spinally-projecting SPANs. Price *et al.* [115] used multiplex single-cell RT-PCR experiments in combination with whole-cell recording in brain slices to show inhibition of SPAN neurones by neuropeptide W. This study did not use retrograde labelling to identify SPANs, but rather electrophysiological “finger-printing” and post-hoc identification of neuronal expression of vasopressin or oxytocin. Neuropeptide W is thought of as being a regulator of feeding behaviour (see for example [144]) and in this case was found to be acting through the neuropeptide W type 1 (NPBW1) receptor.

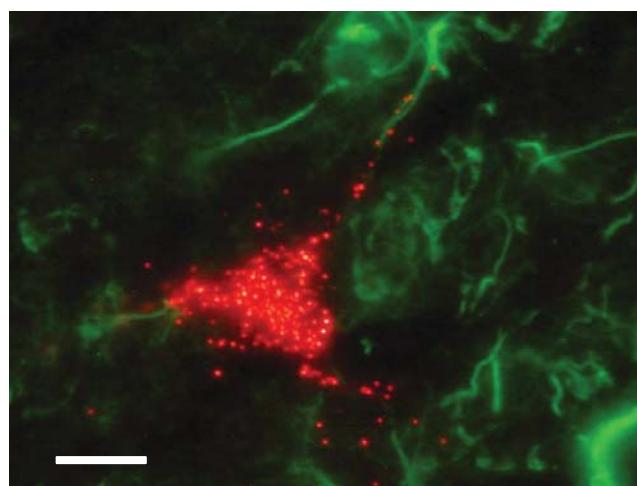
Receptors for neuropeptide FF are densely expressed in the PVN and excite both magnocellular [145] and parvocellular [146] PVN neurones. These include neurones that project to the nucleus of the solitary tract [147], but there has been no investigation of their effects specifically on spinally-projecting SPANs. Evidence also suggests that leptin [148]

150] may influence SPANs of the PVN. The leptin receptor (OB-R) is highly expressed in the PVN [151], and PVN neurones are depolarised by leptin [152]. However, leptin does not increase *c-fos* expression in spinally-projecting SPANs [112], suggesting that the PVN neurones being activated by leptin are not SPANs. Indeed, our own immunohistochemical data show that OB-R is expressed by neurones



**Fig. (7). Pharmacological control of SPANs within the PVN.** Spinally-projecting SPANs are tonically inhibited by GABA input, which is normally enhanced by nitric oxide (NO). However, during chronic heart failure, this potentiation of GABA is lost, leading to an overall increase in sympathetic activity. Substance P (SP) causes indirect activation of spinally-projecting SPANs by inhibition of the inhibitory GABA<sub>A</sub> receptors, via a protein kinase C (PKC)-dependent pathway.

immediately surrounding SPANs (unpublished data, (Fig. 8)), but not by the spinally-projecting SPANs themselves. Orexin has also been shown to depolarise neurones within the PVN, but again, this has not been shown specifically with SPANs [153]. It has been suggested that other neurotransmitters such as adrenomedullin [154] may also activate spinally-projecting SPANs, however, this has not yet been shown directly.



**Fig. (8).** Spinally-projecting SPANs do not co-localise with leptin receptor (OB-R) in rat PVN. Spinally-projecting SPANs were labelled with rhodamine beads (red) using retrograde labelling (see Fig. 2) in an adult rat. A 40 $\mu$ m cryo-slice of the PVN was counter-stained for OB-R (green), using primary goat anti-OB-R IgG, Santa Cruz biotechnology; secondary rabbit anti-goat, Jackson. Scale bar is 10 $\mu$ m. Unpublished data.

## 5. PHYSIOLOGICAL IMPLICATIONS OF SPAN ACTIVITY

Spinally-projecting SPANs of the PVN influence the cardiovascular system [7, 155] (Fig. 3), and there is clear evidence that the functioning of these neurones is altered during heart failure [15, 35]. However, the role of these neurones in normal physiology remains controversial. There is evidence PVN SPANs are involved in physiological functions as diverse as stress, thermoregulation and penile erection.

### 5.1. Involvement with Stress Response

In addition to the clear increases in blood pressure observed with physical activity [156], the simple act of anticipating exercise [157] or performing mental arithmetic [158-159] is sufficient to produce a cardiovascular response. In excess, this preparatory reflex leads to increases in heart rate and blood pressure of a pathological nature [160-162]. Duan *et al.* showed that the stress “phenotype” could be reproduced by stimulation of the PVN [163] and non-specific chemical inhibition of the PVN prevents it [164]. Certainly, the PVN is central to the integration of the hormonal response to stress [5] and it is well known that stimulation of SPANs in the PVN exerts a powerful influence on blood pressure and heart rate and sympathetic nerve activity [55-57, 86, 95, 165-167].

*In vivo* experiments suggest that tachykinins play a major role in cardiovascular control and the expression of a cardiovascular stress response [126]. For example, SP is found at high levels within the PVN [168], and central tachykinins increase efferent sympathetic nerve activity [54, 169]. Furthermore, injection of tachykinin agonists into the hypothalamus [126, 170], and specifically the PVN [54, 171, 172], leads to rapid elevation of heart rate and blood pressure (Fig. 6). Crucially, central administration of tachykinin antagonists reduces the elevation of blood pressure and heart rate resulting from psychological stress of conscious rats [173]. Trans-synaptic retrograde labelling experiments show a direct coupling between the stellate ganglion (which supplies sympathetic motor neurones to the heart) and the PVN [32], implicating PVN SPANs in control of the stress response. In fact PVN SPANs were included in the set of “central command neurones” identified by Jansen *et al.* [32], and inferred from the work of Walter B. Cannon in the early 1900s [174].

However, some scientists now reject this idea; several lines of evidence suggest that whilst the PVN is activated by stress, and stimulation of the PVN elicits increases in blood pressure and heart rate, it is the DMH which is important for the cardiovascular response to stress [135]. This conjecture is based on the observation that injection of the GABA<sub>A</sub> antagonist bicuculline into the DMH prevents the cardiovascular response to air-jet stress, whereas injection into the PVN does not [175]. Whether this PVN SPAN-independent cardiovascular response to stress is general, or stress paradigm-specific, remains to be seen. Furthermore, since the activation of SPANs by SP involves the relief of GABA<sub>A</sub> inhibition [54], inhibition of the PVN by GABA may not be the ideal means of testing this hypothesis. Non-specific PVN lesion experiments such as Busnardo and colleagues’ recent work [164], or specific block of spinally-projecting SPAN neurotransmitter receptors may resolve the controversy.

### 5.2. Control of Blood Volume

It has been suggested that spinally-projecting PVN neurones may be involved in the response to changes in blood volume [24, 176, 177]. This can, to a certain extent, be differentially regulated from blood pressure, by the cardiovascular system. The atrial/venous side of the cardiovascular system is relatively compliant and “expands” to accommodate increased blood volume with relatively little change in pressure; small decreases in volume are replenished from capacitance vessels [178]. Activation of veno-atrial stretch receptors by inflation of small balloons activates afferents and results in a reflex tachycardia (equivalent to the Bainbridge reflex) [179]. Similar results can be obtained by modest changes in blood volume, which alters veno-atrial stretch receptor activity without substantially changing blood pressure. For example, removal of 2mls of blood from adult rats (~10% blood volume, by arterial cannula) has no effect on mean blood pressure, but does increase *c-fos* expression in PVN spinally-projecting SPANs [180]. Similar results are seen in the rat water deprivation model of Toney and colleagues [181] and cardiovascular responses to water deprivation are reduced by GABA inhibition of the PVN [182]. Combined, these data strongly support the hypothesis that

spinally-projecting SPANs are involved with the control of blood volume.

### 5.3. Circadian Rhythm

Human blood pressure typically follows a distinct circadian rhythm, dipping overnight and rising in the morning [183]. It is notable that this rise occurs before waking up [184]. In some cases, hypertension is characterised by not just elevated blood pressure, but by a disturbed circadian pattern of blood pressure. Specifically, the usual night time blood pressure decrease fails to occur [185, 186]. This disturbance to the normal circadian cardiovascular pattern may contribute to the statistically higher number of cardiovascular deaths occurring first thing in the morning [187, 188]. Circadian rhythm is largely controlled at the level of the hypothalamus and, in particular, the suprachiasmatic nucleus (SCN) [189], which expresses a number of clock gene products [190, 191]. There are strong excitatory and inhibitory pathways from the SCN to the PVN of rats [99, 192], and, specifically, to spinally-projecting SPANs [31]. It is therefore possible that spinally-projecting SPANs play a role in circadian control of blood pressure.

### 5.4. Penile Erection

The PVN is involved with penile erection in rats [193]. Oxytocinergic PVN neurones, which have a positive effect on penile erection, project to the hippocampus, the medulla and, most importantly, the spinal cord (For reviews see [194, 195, 196]). A retrograde tracing study using pseudorabies virus has shown central nervous system (CNS) innervations of the penis to include sympathetic pre-ganglionic neurones and the PVN, specifically parvocellular neurones in the PVN [197]. From here, it appears likely that continuation of the sympathetic pathway involves serotonin and/or dopaminergic neurotransmission in the lower spinal cord [198, 199].

### 5.5. Glucose Control

Glucose-sensing by the brain requires input from a number of sensors, including the liver, carotid body and small intestine, as well as glucose-sensitive neurones in the brain [reviewed by 200]. Direct stimulation of the PVN by microinjection of glucose into the PVN causes an increase in sympathetic stimulation to brown adipose tissue (BAT) [201]. More indirect stimulation by peripheral glucose injection also activates the PVN, as shown by increased *c-fos* expression, particularly in identified SPANs [202, 203]. In brain slice experiments, SPANs are modulated by hypoglycaemia [44], but the mechanisms of this still need to be determined. Blood glucose levels are heavily regulated by the liver, which is under a degree of control by the autonomic nervous system. This includes sympathetic innervations projecting from the PVN [82, 84, 136, 204], which have been shown to alter blood glucose levels by influencing the liver [83, 137].

### 5.6. Regulation of Body Fat

Two papers have shown that PVN neurones project to brown adipose tissue (BAT) via spinally-projecting sympathetic nerves [205, 206]. In both cases, pseudorabies virus was injected into interscapular brown adipose tissue and, after a number of days, immunostaining for the virus could

be found in the IML and the PVN. Although this demonstrates a connection from the PVN to BAT, the mechanism remains unclear. Pharmacological stimulation of the PVN by microinjection of stimulatory amino acids has been shown to produce differing effects. Amir [207] showed an increase in sympathetic outflow to BAT, Yoshimatsu and colleagues [208] found small decreases in sympathetic activity when the anorexigenic peptide cholecystokinin (CCK) was injected into the PVN, but showed a mainly a stimulatory response to injected glutamate [209]. Conversely, Madden and Morrison [210] showed an inhibitory effect. Obviously there is a potential involvement of SPANs with regulation of body fat, but the details remain unclear.

### 5.7. Thermoregulation

Thermosensitivity of neurones in the PVN was first shown by Inenaga *et al.* [211]. Poly-synaptic viral tracing studies in rats show that PVN SPANs are directly involved in thermoregulation [212]. Since then the region has been shown to be critical in thermoregulation [213-214]. Projections from the PVN to the RVLM, which contains sympathetic pre-motor neurones, are activated following heat exposure [215]. In addition, heat exposure activates spinally-projecting PVN neurones [216]. Whether this involves a simple vasomotor responses to elevated temperature (e.g., sympathetic vasodilation of skin in humans [217] or tail in the case of rodents [212]) or some other process is unknown. The PVN also has an effect on thermogenesis, *via* innervations of brown adipose tissue (see above).

### 5.8. Involvement with Heart Failure

Whilst chronic heart failure could not be described as a function of SPANs, the relationship between NO, GABA and SPAN activity is fundamental to the development of elevated sympathetic activity seen in human patients and animal models of heart failure [15, 33-34, 218]. NO is now well established as a neuromediator in the CNS where it is synthesised by neuronal nitric oxide synthase (nNOS) [reviewed by 219, 220]. nNOS is abundantly and constitutively expressed throughout the brain, but particularly in cells of the PVN; this includes expression by pre-autonomic neurones projecting both to the IML [221] and the RVLM [222, 223]. NO appears to increase the tonic inhibition of SPANs [224] by facilitating pre-synaptic release of GABA [223]. In models where chronic heart failure has been experimentally induced in rats, there are decreased levels of nNOS expression in the PVN [225], together with evidence of reduced neuronal activity [218]. Furthermore, the degree of tonic inhibition of rSNA by GABA (albeit assessed with bicuculline) was reduced in the chronic heart failure animals [35]. When combined these data strongly suggest that the tonic inhibition of sympathetic activity is maintained by an NO-dependent local release of GABA, and that chronic heart failure leads to a loss of NO activity, and thus to a loss of inhibitory control of sympathetic neurones [36] (Fig. 7). It has recently been shown that AT<sub>1</sub> receptor mRNA expression is increased in the PVN and, consequently, intra-PVN injection of AGII is more effective at increasing rSNA in the rat chronic heart failure model [226]. Further experiments are required to determine if the increase in angiotensin

activity is causal, coincident or consequent to the decreased nNOS expression in the PVN.

## 6. CONCLUSIONS

Several studies have investigated the function and pharmacology of spinally-projecting SPANs in the PVN but, interestingly, we still know a great deal more about the expression of their neurotransmitters than anything else. There is relatively sparse information concerning which receptors are expressed by SPANs; GABA<sub>A</sub>, glutamate, adenosine A<sub>1</sub> and NK1 receptors are all strong candidates, and SPANs can be indirectly modulated by NO and AGII. In terms of the modulation of SPANs, tonic inhibition is a recurrent theme and clearly central to their regulation. For example, NO, AGII and SP all appear to act by modulating this GABA tone. These neurones modulate the cardiovascular system, and it seems likely that they have other functions too. So a big question seems to be: what is the primary role of these neurones in cardiovascular function? There is evidence they become involved in the pathological increase in sympathetic activity associated with chronic heart failure, but this still leaves the function in normal, healthy animals unknown. Putative physiological functions include: volume regulation, glucose control, circadian rhythm and response to stress. Perhaps PVN SPANs play a role in all of these. The final question would then be, do the same neurones play a role in each of these functions, or does a mixed population of SPANs exist, each with a different receptor and neurotransmitter profile and each with a separate function? A great many further experiments will be required to finally answer these fascinating questions.

## ABBREVIATIONS

3V	= 3 <sup>rd</sup> ventricle
AGII	= Angiotensin II
AMPA	= $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid
AP-5	= Amino-5-phosphonovaleric acid
AT <sub>1</sub>	= Angiotensin II type 1 receptor
AT <sub>2</sub>	= Angiotensin II type 2 receptor
BAT	= Brown adipose tissue
CCK	= Cholecystokinin
CNS	= Central nervous system
CVS	= Cardiovascular system
DMH	= Dorsomedial hypothalamus
FITC	= Fluorescein isothiocyanate
GABA	= $\gamma$ -aminobutyric acid
HPA	= Hypothalamic-pituitary-adrenal
IML	= Intermediolateralis
NBQX	= 2,3-dihydroxy-6-nitro-7-sulfamoylbenzo[f]quinoxaline-2,3-dione
NKA	= Neurokinin A

NKB	= Neurokinin B
NMDA	= N-methyl D-aspartate
nNOS	= Neuronal nitric oxide synthase
NO	= Nitric oxide
NTS	= Nucleus of the solitary tract
OB-R	= Leptin receptor
OT-R	= Oxytocin receptor
PACAP	= Pituitary adenylcyclase activating peptide
PKC	= Protein kinase C
rSNA	= Renal sympathetic nerve activity
RT-PCR	= Reverse transcriptase PCR
RVLM	= Rostral ventrolateral medulla
SCG	= Superior cervical ganglion
SCN	= Suprachiasmatic nucleus
SG	= Stellate ganglion
SP	= Substance P
SPAN	= Sympathetic pre-autonomic neurone
T3	= Thyroid hormone
THDOC	= Tetrahydrodeoxycorticosterone
TSH	= Thyroid stimulating hormone
TR	= Thyroid hormone receptor
V <sub>1a</sub> -receptor	= Vasopressin 1a receptor
VIP	= Vasoactive intestinal peptide
VPAC <sub>2</sub>	= Vasoactive intestinal peptide/pituitary adenylcyclase activating peptide receptors type 2

## REFERENCES

- [1] Swanson, L.W., Sawchenko, P.E., Wiegand, S.J., Price, J.L. Separate neurons in the paraventricular nucleus project to the median-eminence and to the medulla or spinal-cord. *Brain Res.*, **1980**, *198*, 190-195.
- [2] Palkovits, M. Interconnections between the neuroendocrine hypothalamus and the central autonomic system. *Front. Neuroendocrinol.*, **1999**, *20*, 270-295.
- [3] Swanson, L., Kuypers, H. The paraventricular nucleus of the hypothalamus: Cytoarchitectonic subdivisions and organization of projections to the pituitary, dorsal vagal complex, and spinal cord as demonstrated by retrograde fluorescence double-labeling methods. *J. Comp. Neurol.*, **1980**, *194*, 555-570.
- [4] Swanson, L.W., Sawchenko, P.E. Hypothalamic integration - organization of the paraventricular and supraoptic nuclei. *Ann. Rev. Neurosci.*, **1983**, *6*, 269-324.
- [5] Herman, J.P., Cullinan, W.E. Neurocircuitry of stress: Central control of the hypothalamo-pituitary-adrenocortical axis. *Trends Neurosci.*, **1997**, *20*, 78-84.
- [6] Loewy, A. Forebrain nuclei involved in autonomic control. *Prog. Brain Res.*, **1991**, *87*, 253.
- [7] Coote, J.H. Landmarks in understanding the central nervous control of the cardiovascular system. *Exp. Physiol.*, **2007**, *92*, 3-18.
- [8] Dampney, R.A. Functional organization of central pathways regulating the cardiovascular system. *Physiol. Rev.*, **1994**, *74*, 323-364.

- [9] Spyer, K.M. Annual-review prize lecture - central nervous mechanisms contributing to cardiovascular control. *J. Physiol. (London)*, **1994**, *474*, 1-19.
- [10] Hilton, S.M. The defense-arousal system and its relevance for circulatory and respiratory control. *J. Exp. Biol.*, **1982**, *100*, 159-174.
- [11] Cannon, W.B. *Bodily changes in pain, hunger, fear and rage*. 2nd ed Appleton, New York, **1929**.
- [12] Mittleman, M.A., Maclure, M., Tofler, G.H., Sherwood, J.B., Goldberg, R.J., Muller, J.E. Triggering of acute myocardial-infarction by heavy physical exertion - protection against triggering by regular exertion. *N. Engl. J. Med.*, **1993**, *329*, 1677-1683.
- [13] Mittleman, M.A., Maclure, M., Sherwood, J.B., Mulry, R.P., Tofler, G.H., Jacobs, S.C., Friedman, R., Benson, H., Muller, J.E. Triggering of acute myocardial-infarction onset by episodes of anger. *Circulation*, **1995**, *92*, 1720-1725.
- [14] Leor, J., Poole, W.K., Kloner, R.A. Sudden cardiac death triggered by an earthquake. *New Engl. J. Med.*, **1996**, *334*, 413-419.
- [15] Patel, K.P. Role of paraventricular nucleus in mediating sympathetic outflow in heart failure. *Heart Fail. Rev.*, **2000**, *V5*, 73-86.
- [16] Woodworth, R., Sherrington, C. A pseudaffectionate reflex and its spinal path. *J. Physiol.*, **1904**, *31*, 234.
- [17] Bard, P. A diencephalic mechanism for the expression of rage with special reference to the sympathetic nervous system. *Am. J. Physiol.*, **1928**, *84*, 490.
- [18] Bard, P. Anatomical organization of central nervous system in relation to control of heart and blood vessels. *Physiol. Rev.*, **1961**, *40*, 3-26.
- [19] Keller, A.D. Ablation and stimulation of hypothalamus - circulatory effects. *Physiol. Rev.*, **1961**, *40*, 116-120.
- [20] Delgado, J. Circulatory effects of cortical stimulation. *Physiol. Rev., Suppl.*, **1960**, *4*, 146.
- [21] Kiss, J., Martos, J., Palkovits, M. Hypothalamic paraventricular nucleus: A quantitative analysis of cytoarchitectonic subdivisions in the rat. *J. Comp. Neurol.*, **1991**, *313*, 563-573.
- [22] Koutcherov, Y., Mai, J., Ashwell, K., Paxinos, G. Organization of the human paraventricular hypothalamic nucleus. *J. Comp. Neurol.*, **2000**, *423*, 299-318.
- [23] Shafton, A., Ryan, A., Badoer, E. Neurons in the hypothalamic paraventricular nucleus send collaterals to the spinal cord and to the rostral ventrolateral medulla in the rat. *Brain Res.*, **1998**, *801*, 239-243.
- [24] Lovick, T., Malpas, S., Mahony, M. Renal vasodilatation in response to acute volume load is attenuated following lesions of parvocellular neurones in the paraventricular nucleus in rats. *J. Auton. Nerv. Syst.*, **1993**, *43*, 247-255.
- [25] Pyner, S., Coote, J. Identification of branching paraventricular neurons of the hypothalamus that project to the rostroventrolateral medulla and spinal cord. *Neuroscience*, **2000**, *100*, 549-556.
- [26] Motawei, K., Pyner, S., Ranson, R.N., Kamel, M., Coote, J.H. Terminals of paraventricular spinal neurones are closely associated with adrenal medullary sympathetic preganglionic neurones: Immunocytochemical evidence for vasopressin as a possible neurotransmitter in this pathway. *Exp. Brain Res.*, **1999**, *126*, 68-76.
- [27] Ranson, R.N., Motawei, K., Pyner, S., Coote, J.H. The paraventricular nucleus of the hypothalamus sends efferents to the spinal cord of the rat that closely appose sympathetic preganglionic neurones projecting to the stellate ganglion. *Exp. Brain Res.*, **1998**, *120*, 164-172.
- [28] Coote, J.H. Cardiovascular function of the paraventricular nucleus of the hypothalamus. *Biol. Signals*, **1995**, *4*, 142-149.
- [29] Badoer, E. Cardiovascular role of parvocellular neurons in the paraventricular nucleus of the hypothalamus. *News Physiol. Sci.*, **1996**, *11*, 43-47.
- [30] Pyner, S. Neurochemistry of the paraventricular nucleus of the hypothalamus: Implications for cardiovascular regulation. *J. Chem. Neuroanat.*, **2009**, *38*, 197-208.
- [31] Cui, L.N., Coderre, E., Renaud, L.P. Glutamate and gaba mediate suprachiasmatic nucleus inputs to spinal-projecting paraventricular neurons. *Am. J. Physiol-Regul. Integr. Comp. Physiol.*, **2001**, *281*, R1283-R1289.
- [32] Jansen, A.S., Nguyen, X.V., Karpitskiy, V., Mettenleiter, T.C., Loewy, A.D. Central command neurons of the sympathetic nervous system: Basis of the fight-or-flight response. *Science*, **1995**, *270*, 644-646.
- [33] Ferguson, D.W., Berg, W.J., Sanders, J.S. Clinical and hemodynamic correlates of sympathetic-nerve activity in normal humans and patients with heart-failure - evidence from direct microneurographic recordings. *J. Am. Coll. Cardiol.*, **1990**, *16*, 1125-1134.
- [34] Vahid-Ansari, F., Leenen, F.H.H. Pattern of neuronal activation in rats with chf after myocardial infarction. *Am. J. Physiol. Heart Circ. Physiol.*, **1998**, *44*, H2140-H2146.
- [35] Zhang, K., Li, Y.F., Patel, K.P. Reduced endogenous gaba-mediated inhibition in the pvn on renal nerve discharge in rats with heart failure. *Am. J. Physiol. Regul. Integ. Comp. Physiol.*, **2002**, *282*, R1006-R1015.
- [36] Li, Y.F., Patel, K.P. Paraventricular nucleus of the hypothalamus and elevated sympathetic activity in heart failure: The altered inhibitory mechanisms. *Acta Physiol. Scand.*, **2003**, *177*, 17-26.
- [37] Hoffman, N.W., Tasker, J.G., Dudek, F.E. Immunohistochemical differentiation of electrophysiologically defined neuronal populations in the region of the rat hypothalamic paraventricular nucleus. *J. Comp. Neurol.*, **1991**, *307*, 405-416.
- [38] Tasker, J.G., Dudek, F.E. Local inhibitory synaptic inputs to neurones of the paraventricular nucleus in slices of rat hypothalamus. *J. Physiol. (Lond)*, **1993**, *469*, 179-192.
- [39] Luther, J.A., Halmos, K.C., Tasker, J.G. A slow transient potassium current expressed in a subset of neurosecretory neurons of the hypothalamic paraventricular nucleus. *J. Neurophysiol.*, **2000**, *84*, 1814-1825.
- [40] Luther, J.A., Tasker, J.G. Voltage-gated currents distinguish parvocellular from magnocellular neurones in the rat hypothalamic paraventricular nucleus. *J. Physiol-London*, **2000**, *523*, 193-209.
- [41] Stern, J.E. Electrophysiological and morphological properties of pre-autonomic neurones in the rat hypothalamic paraventricular nucleus. *J. Physiol-London*, **2001**, *537*, 161-177.
- [42] Sonner, P.M., Stern, J.E. Role of a-type potassium currents on the excitability and firing activity of rvlm-projecting pvn neurons. *FASEB J.*, **2005**, *19*, A599-A599.
- [43] Barrett-Jolley, R., Pyner, S., Coote, J.H. Measurement of voltage-gated potassium currents in identified spinally-projecting sympathetic neurones of the paraventricular nucleus. *J. Neurosci. Methods*, **2000**, *102*, 25-33.
- [44] Lewis, R., Mills, A.F., Barrett-Jolley, R. Models of paraventricular nucleus (pvn) sympathetic neurone modulation by glucose and hypoglycaemia. *Biophys. J.*, **2010**, *98*, 140a-140a.
- [45] Li, D.P., Chen, S.R., Pan, H.L. Adenosine inhibits paraventricular pre-synaptic neurons through atp-dependent potassium channels. *J. Neurochem.*, **2010**, *113*, 530-542.
- [46] Lesage, F. Pharmacology of neuronal background potassium channels. *Neuropharmacology*, **2003**, *44*, 1-7.
- [47] Kirsch, G.E., Narahashi, T. 3, 4-diaminopyridine - potent new potassium channel blocker. *Biophys. J.*, **1978**, *22*, 507-512.
- [48] Jan, L.Y., Jan, Y.N. Annual review prize lecture - voltage-gated and inwardly rectifying potassium channels. *J. Physiol-London*, **1997**, *505*, 267-282.
- [49] Garcia, M.L., Galvez, A., Garciacalvo, M., King, V.F., Vazquez, J., Kaczorowski, G.J. Use of toxins to study potassium channels. *J. Bioenerg. Biomembr.*, **1991**, *23*, 615-646.
- [50] Edwards, G., Weston, A.H. The pharmacology of atp-sensitive potassium channels. *Ann. Rev. Pharmacol. Toxicol.*, **1993**, *33*, 597-637.
- [51] Choi, K.L., Aldrich, R.W., Yellen, G. Tetraethylammonium blockade distinguishes 2 inactivation mechanisms in voltage-activated k<sup>+</sup> channels. *Proc. Natl. Acad. Sci. USA*, **1991**, *88*, 5092-5095.
- [52] Candia, S., Garcia, M.L., Latorre, R. Mode of action of iberiotoxin, a potent blocker of the large conductance ca-2+activated k<sup>+</sup> channel. *Biophys. J.*, **1992**, *63*, 583-590.
- [53] Barrett-Jolley, R., Dart, C., Standen, N.B. Direct block of native and cloned (kir2.1) inward rectifier k<sup>+</sup> channels by chloroethylclonidine. *Br. J. Pharmacol.*, **1999**, *128*, 760-766.

- [54] Womack, M.D., Morris, R., Gent, T.C., Barrett-Jolley, R. Substance p targets sympathetic control neurons in the paraventricular nucleus. *Circ. Res.*, **2007**, *100*, 1650-1658.
- [55] Badoer, E., Ng, C.W., De Matteo, R. Tonic sympathoinhibition arising from the hypothalamic pvn in the conscious rabbit. *Brain Res.*, **2002**, *947*, 17-24.
- [56] Martin, D., Haywood, J. Hemodynamic responses to paraventricular nucleus disinhibition with bicuculline in conscious rats. *Am. J. Physiol. Heart Circ. Physiol.*, **1993**, *265*, H1727.
- [57] Martin, D.S., Segura, T., Haywood, J.R. Cardiovascular responses to bicuculline in the paraventricular nucleus of the rat. *Hypertension*, **1991**, *18*, 48-55.
- [58] Park, J.B., Stern, J.E. A tonic, gabaa receptor-mediated inhibitory postsynaptic current restrains firing activity in preautonomic and magnocellular neuroendocrine neurons of the paraventricular nucleus of the hypothalamus (pvn). *FASEB J.*, **2005**, *19*, A599-A599.
- [59] Hallbeck, M., Blomqvist, A. Spinal cord-projecting vasopressinergic neurons in the rat paraventricular hypothalamus. *J. Comp. Neurol.*, **1999**, *411*, 201-211.
- [60] Hallbeck, M., Larhammar, D., Blomqvist, A. Neuropeptide expression in rat paraventricular hypothalamic neurons that project to the spinal cord. *J. Comp. Neurol.*, **2001**, *433*, 222-238.
- [61] Hallbeck, M. Dynorphin mRNA-expressing neurons in the rat paraventricular hypothalamic nucleus project to the spinal cord. *Neurosci. Lett.*, **2000**, *285*, 161-164.
- [62] Cechetto, D.F., Saper, C.B. Neurochemical organization of the hypothalamic projection to the spinal-cord in the rat. *J. Comp. Neurol.*, **1988**, *272*, 579-604.
- [63] Jansen, A.S.P., Wessendorf, M.W., Loewy, A.D. Transneuronal labeling of cns neuropeptide and monoamine neurons after pseudorabies virus injections into the stellate ganglion. *Brain Res.*, **1995**, *683*, 1-24.
- [64] Sawchenko, P.E., Swanson, L.W. Immunohistochemical identification of neurons in the paraventricular nucleus of the hypothalamus that project to the medulla or to the spinal-cord in the rat. *J. Comp. Neurol.*, **1982**, *205*, 260-272.
- [65] Strack, A.M., Sawyer, W.B., Platt, K.B., Loewy, A.D. Cns cell groups regulating the sympathetic outflow to adrenal-gland as revealed by trans-neuronal cell body labeling with pseudorabies virus. *Brain Res.*, **1989**, *491*, 274-296.
- [66] Xi, D., Kusano, K., Gainer, H. Quantitative analysis of oxytocin and vasopressin messenger ribonucleic acids in single magnocellular neurons isolated from supraoptic nucleus of rat hypothalamus. *Endocrinology*, **1999**, *140*, 4677-4682.
- [67] Malpas, S.C., Coote, J.H. Role of vasopressin in sympathetic response to paraventricular nucleus stimulation in anesthetized rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, **1994**, *266*, R228-R236.
- [68] Sermasi, E., Coote, J.H. Oxytocin acts at v-1 receptors to excite sympathetic preganglionic neurons in neonate rat spinal-cord *in vitro*. *Brain Res.*, **1994**, *647*, 323-332.
- [69] Pittman, Q., Riphagen, C., Lederis, K. Release of immunoassayable neurohypophyseal peptides from rat spinal cord, *in vivo*. *Brain Res.*, **1984**, *300*, 321-326.
- [70] Yang, Z., Han, D.D., Coote, J.H. Cardiac sympatho-excitatory action of pvn-spinal oxytocin neurones. *Auton. Neurosci-Basic Clin.*, **2009**, *147*, 80-85.
- [71] Gladwell, S.J., Coote, J.H. Inhibitory and indirect excitatory effects of dopamine on sympathetic preganglionic neurones in the neonatal rat spinal cord *in vitro*. *Brain Res.*, **1999**, *818*, 397-407.
- [72] Gladwell, S.J., Coote, J.H. Fast excitatory post synaptic potentials and their response to catecholaminergic antagonists in rat sympathetic preganglionic neurones *in vitro*. *Neurosci. Lett.*, **1999**, *268*, 89-92.
- [73] Gladwell, S.J., Pyner, S., Barnes, N.M., Coote, J.H. D-1-like dopamine receptors on retrogradely labelled sympathoadrenal neurones in the thoracic spinal cord of the rat. *Exp. Brain Res.*, **1999**, *128*, 377-382.
- [74] Lahlou, S. Mechanisms underlying the cardiovascular responses to spinal dopamine receptor stimulation by apomorphine in anesthetized rats. *Neurosci. Lett.*, **2003**, *335*, 187-191.
- [75] Yang, Z., Wheatley, M., Coote, J.H. Neuropeptides, amines and amino acids as mediators of the sympathetic effects of paraventricular nucleus activation in the rat. *Exp. Physiol.*, **2002**, *87*, 663-674.
- [76] Gwyn, P., Logan, S.D. Direct actions of l-enkephalin on neonatal rat sympathetic preganglionic neurones via delta opioid receptors *in vitro*. *Br. J. Pharmacol.*, **1997**, *122*, 350P.
- [77] Lewis, D.I., Coote, J.H. Angiotensin-II in the spinal-cord of the rat and its sympathoexcitatory effects. *Brain Res.*, **1993**, *614*, 1-9.
- [78] Suter, C., Coote, J.H. Intrathecally administered angiotensin-II increases sympathetic activity in the rat. *J. Auton. Nervous Syst.*, **1987**, *19*, 31-37.
- [79] Llewellyn-Smith, I.J., DiCarlo, S.E., Collins, H.L., Keast, J.R. Enkephalin-immunoreactive interneurons extensively innervate sympathetic preganglionic neurons regulating the pelvic viscera. *J. Comp. Neurol.*, **2005**, *488*, 278-289.
- [80] Macdonald, R.L., Olsen, R.W. Gaba(a) receptor channels. *Ann. Rev. Neurosci.*, **1994**, *17*, 569-602.
- [81] Watkins, N.D., Cork, S.C., Pyner, S. An immunohistochemical investigation of the relationship between neuronal nitric oxide synthase, gaba and presynaptic paraventricular neurons in the hypothalamus. *Neuroscience*, **2009**, *159*, 1079-1088.
- [82] Kalsbeek, A., Fliers, E., Franke, A.N., Wortel, J., Buijs, R.M. Functional connections between the suprachiasmatic nucleus and the thyroid gland as revealed by lesioning and viral tracing techniques in the rat. *Endocrinology*, **2000**, *141*, 3832-3841.
- [83] Kalsbeek, A., La Fleur, S., Van Heijningen, C., Buijs, R.M. Suprachiasmatic gabaergic inputs to the paraventricular nucleus control plasma glucose concentrations in the rat via sympathetic innervation of the liver. *J. Neurosci.*, **2004**, *24*, 7604-7613.
- [84] La Fleur, S.E., Kalsbeek, A., Wortel, J., Buijs, R.M. Polysynaptic neural pathways between the hypothalamus, including the suprachiasmatic nucleus, and the liver. *Brain Res.*, **2000**, *871*, 50-56.
- [85] Yamashita, H., Inenaga, K., Koizumi, K. Possible projections from regions of paraventricular and supraoptic nuclei to the spinal-cord - electrophysiological studies. *Brain Res.*, **1984**, *296*, 373-378.
- [86] Kannan, H., Hayashida, Y., Yamashita, H. Increase in sympathetic outflow by paraventricular nucleus stimulation in awake rats. *Am. J. Physiol.*, **1989**, *256*, R1325-R1330.
- [87] Li, Y.F., Jackson, K.L., Stern, J.E., Rabeler, B., Patel, K.P. Interaction between glutamate and gaba systems in the integration of sympathetic outflow by the paraventricular nucleus of the hypothalamus. *Am. J. Physiol. Heart Circ. Physiol.*, **2006**, *291*, H2847-H2856.
- [88] Brickley, S.G., CullCandy, S.G., Farrant, M. Development of a tonic form of synaptic inhibition in rat cerebellar granule cells resulting from persistent activation of gaba(a) receptors. *J. Physiol-Lond*, **1996**, *497*, 753-759.
- [89] Farrant, M., Nusser, Z. Variations on an inhibitory theme: Phasic and tonic activation of gaba(a) receptors. *Nat. Rev. Neurosci.*, **2005**, *6*, 215-229.
- [90] Glykys, J., Mann, E.O., Mody, I. Which gaba(a) receptor subunits are necessary for tonic inhibition in the hippocampus? *J. Neurosci.*, **2008**, *28*, 1421-1426.
- [91] Semyanov, A., Walker, M.C., Kullmann, D.M., Silver, R.A. Tonically active gaba(a) receptors: Modulating gain and maintaining the tone. *Trends Neurosci.*, **2004**, *27*, 262-269.
- [92] Jia, F., Yue, M., Chandra, D., Keramidas, A., Goldstein, P.A., Homannics, G.E., Harrison, N.L. Taurine is a potent activator of extrasynaptic gaba(a) receptors in the thalamus. *J. Neurosci.*, **2008**, *28*, 106-115.
- [93] Mody, I., Glykys, J., Wei, W.Z. A new meaning for "Gin & tonic": Tonic inhibition as the target for ethanol action in the brain. *Alcohol*, **2007**, *41*, 145-153.
- [94] Park, J.B., Jo, J.Y., Zheng, H., Patel, K.P., Stern, J.E. Regulation of tonic gaba inhibitory function, presynaptic neuronal activity and sympathetic outflow from the paraventricular nucleus by astroglial gaba transporters. *J. Physiol. London*, **2009**, *587*, 4645-4660.
- [95] Schlenker, E., Barnes, L., Hansen, S., Martin, D. Cardiorespiratory and metabolic responses to injection of bicuculline into the

- hypothalamic paraventricular nucleus (pvn) of conscious rats. *Brain Res.*, **2001**, *895*, 33-40.
- [96] Barrett-Jolley, R. Nippecotic acid directly activates gaba(a)-like ion channels. *Br. J. Pharmacol.*, **2001**, *133*, 673-678.
- [97] Boudaba, C., Szabo, K., Tasker, J.G. Physiological mapping of local inhibitory inputs to the hypothalamic paraventricular nucleus. *J. Neurosci.*, **1996**, *16*, 7151-7160.
- [98] Fenelon, V.S., Sieghart, W., Herbison, A.E. Cellular localization and differential distribution of gabaa receptor subunit proteins and messenger rnas within hypothalamic magnocellular neurons. *Neurosci.*, **1995**, *64*, 1129-1143.
- [99] Hermes, M.L., Coderre, E.M., Buijs, R.M., Renaud, L.P. (Gaba and glutamate mediate rapid neurotransmission from suprachiasmatic nucleus to hypothalamic paraventricular nucleus in rat. *J. Physiol.(Lond)*, **1996**, *496* (Pt 3), 749-757.
- [100] Zaki, A., Barrett-Jolley, R. Rapid neuromodulation by cortisol in the rat paraventricular nucleus: An *in vitro* study. *Br. J. Pharmacol.*, **2002**, *137*, 87-97.
- [101] Fritschy, J.M., Mohler, H. Gaba(a)-receptor heterogeneity in the adult-rat brain - differential regional and cellular-distribution of 7 major subunits. *J. Comp. Neurol.*, **1995**, *359*, 154-194.
- [102] Barrett-Jolley, R., Pyner, S. Inhibition of rat spinally projecting pvn neurones by the neurosteroid tetrahydroxycorticosterone. *J. Physiol. London*, **2003**, *548*.
- [103] Womack, M.D., Pyner, S., Barrett-Jolley, R. Inhibition by [alpha]-tetrahydrodeoxycorticosterone (thdoc) of pre-sympathetic parvocellular neurones in the paraventricular nucleus of rat hypothalamus. *Br. J. Pharmacol.*, **2006**, *149*, 600-607.
- [104] Li, D.P., Chen, S.R., Pan, H.L. Nitric oxide inhibits spinally projecting paraventricular neurons through potentiation of presynaptic gaba release. *J. Neurophysiol.*, **2002**, *88*, 2664-2674.
- [105] Chen, Q., Pan, H.L. Regulation of synaptic input to hypothalamic presynaptic neurons by gaba(b) receptors. *Neuroscience*, **2006**, *142*, 595-606.
- [106] Khawaled, R., Bruening-Wright, A., Adelman, J., Maylie, J. Bicuculline block of small-conductance calcium-activated potassium channels. *Pflügers Archiv. Eur. J. Physiol.*, **1999**, *438*, 314-321.
- [107] Strobaek, D., Jorgensen, T.D., Christoffersen, P., Ahring, P.K., Olesen, S.P. Pharmacological characterization of small-conductance ca<sup>2+</sup>-activated k<sup>+</sup> channels stably expressed in hek 293 cells. *Br. J. Pharmacol.*, **2000**, *129*, 991-999.
- [108] Druzin, M., Haage, D., Johansson, S. Bicuculline free base blocks voltage-activated k<sup>+</sup> currents in rat medial preoptic neurons. *Neuropharmacology*, **2004**, *46*, 285-295.
- [109] Stern, J.E., Li, Y.J., Sonner, P. Nitric oxide modulates firing activity of preautonomic neurons in the paraventricular nucleus (pvn) of the hypothalamus. *FASEB J.*, **2002**, *16*, A501-A501.
- [110] Stern, J.E., Zhang, W. Preautonomic neurons in the paraventricular nucleus of the hypothalamus contain estrogen receptor beta. *Brain Res.*, **2003**, *975*, 99-109.
- [111] Zhang, W.F., Stern, J.E. Preautonomic neurons in the paraventricular nucleus (pvn) of the hypothalamus express estrogen receptor-b immunoreactivity. *FASEB J.*, **2002**, *16*, A501-A501.
- [112] Badoer, E., Ryan, A. Effects of leptin on spinally projecting neurons in the pvn of the hypothalamus. *Brain Res.*, **1999**, *844*, 210-215.
- [113] Oldfield, B.J., Davern, P.J., Giles, M.E., Allen, A.M., Badoer, E., McKinley, M.J. Efferent neural projections of angiotensin receptor (at1) expressing neurones in the hypothalamic paraventricular nucleus of the rat. *J. Neuroendocrinol.*, **2001**, *13*, 139-146.
- [114] Li, D.P., Chen, S.R., Pan, H.L. Angiotensin ii stimulates spinally projecting paraventricular neurons through presynaptic disinhibition. *J. Neurosci.*, **2003**, *23*, 5041-5049.
- [115] Price, C.J., Samson, W.K., Ferguson, A.V. Neuropeptide w has cell phenotype-specific effects on the excitability of different subpopulations of paraventricular nucleus neurones. *J. Neuroendocrinol.*, **2009**, *21*, 850-857.
- [116] Lind, R.W., Swanson, L.W., Bruhn, T.O., Ganten, D. The distribution of angiotensin-ii-immunoreactive cells and fibers in the paraventriculo-hypophysial system of the rat. *Brain Res.*, **1985**, *338*, 81-89.
- [117] Alexander, S.P.H., Mathie, A., Peters, J.A. Guide to receptors and channels (grac), 3rd edition. *Br. J. Pharmacol.*, **2008**, *153*, S1-S209.
- [118] Allen, A.M., MacGregor, D.P., McKinley, M.J., Mendelsohn, F.A.O. Angiotensin ii receptors in the human brain. *Regul. Peptides*, **1999**, *79*, 1-7.
- [119] de Gasparo, M., Catt, K.J., Inagami, T., Wright, J.W., Unger, T. International union of pharmacology. Xxiii. The angiotensin II receptors. *Pharmacol. Rev.*, **2000**, *52*, 415-472.
- [120] Oliverio, M.I., Best, C.F., Kim, H.S., Arendshorst, W.J., Smithies, O., Coffman, T.M. Angiotensin II responses in at(1a) receptor-deficient mice: A role for at(1b) receptors in blood pressure regulation. *Am. J. Physiol. Renal. Physiol.*, **1997**, *41*, F515-F520.
- [121] Lenkei, Z., Palkovits, M., Corvol, P., LlorensCortes, C. Expression of angiotensin type-1 (at1) and type-2 (at2) receptor mrnas in the adult rat brain: A functional neuroanatomical review. *Front. Neuroendocrinol.*, **1997**, *18*, 383-439.
- [122] Bains, J.S., Potyok, A., Ferguson, A.V. Angiotensin-ii actions in paraventricular nucleus - functional evidence for neurotransmitter role in efferents originating in subfornical organ. *Brain Res.*, **1992**, *599*, 223-229.
- [123] Bains, J.S., Ferguson, A.V. Paraventricular nucleus neurons projecting to the spinal-cord receive excitatory input from the subfornical organ. *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, **1995**, *268*, R625-R633.
- [124] Li, Z., Ferguson, A.V. Electrophysiological properties of paraventricular magnocellular neurons in rat brain slices: Modulation of I-A by angiotensin II. *Neuroscience*, **1996**, *71*, 133-145.
- [125] Cato, M.J., Toney, G.M. Angiotensin ii excites paraventricular nucleus neurons that innervate the rostral ventrolateral medulla: An *in vitro* patch-clamp study in brain slices. *J. Neurophysiol.*, **2005**, *93*, 403-413.
- [126] Culman, J., Itoi, K., Unger, T. Hypothalamic tachykinins. Mediators of stress responses? *Ann. N.Y. Acad. Sci.*, **1995**, *771*, 204-218.
- [127] Ding, Y.Q., Shigemoto, R., Takada, M., Ohishi, H., Nakanishi, S., Mizuno, N. Localization of the neuromedin k receptor (nk3) in the central nervous system of the rat. *J. Comp. Neurol.*, **1996**, *364*, 290-310.
- [128] Koutcherov, Y., Ashwell, K.W.S., Paxinos, G. The distribution of the neurokinin b receptor in the human and rat hypothalamus. *Neuroreport*, **2000**, *11*, 3127-3131.
- [129] Shults, C.W., Buck, S.H., Burcher, E., Chase, T.N., O'Donohue, T.L. Distinct binding sites for substance p and neurokinin a (substance k): Co-transmitters in rat brain. *Peptides*, **1985**, *6*, 343-345.
- [130] Nakajima, Y., Tsuchida, K., Negishi, M., Ito, S., Nakanishi, S. Direct linkage of three tachykinin receptors to stimulation of both phosphatidylinositol hydrolysis and cyclic amp cascades in transfected chinese hamster ovary cells. *J. Biol. Chem.*, **1992**, *267*, 2437-2442.
- [131] Quartara, L., Maggi, C.A. The tachykinin nk1 receptor. Part i: Ligands and mechanisms of cellular activation. *Neuropeptides*, **1997**, *31*, 537-563.
- [132] Connolly, C.N., Kittler, J.T., Thomas, P., Uren, J.M., Brandon, N.J., Smart, T.G., Moss, S.J. Cell surface stability of gamma-aminobutyric acid type a receptors - dependence on protein kinase c activity and subunit composition. *J. Biol. Chem.*, **1999**, *274*, 36565-36572.
- [133] Bittencourt, J.C., Benoit, R., Sawchenko, P.E. Distribution and origins of substance-p-immunoreactive projections to the paraventricular and supraoptic nuclei - partial overlap with ascending catecholaminergic projections. *J. Chem. Neuroanat.*, **1991**, *4*, 63-78.
- [134] Womack, M.D., Barrett-Jolley, R. Activation of paraventricular nucleus neurones by the dorsomedial hypothalamus via a tachykinin pathway in rats. *Exp. Physiol.*, **2007**, *92*, 671-676.
- [135] DiMicco, J.A., Samuels, B.C., Zaretskaia, M.V., Zaretsky, D.V. The dorsomedial hypothalamus and the response to stress part renaissance, part revolution. *Pharmacol. Biochem. Behav.*, **2002**, *71*, 469-480.

- [136] Yi, C.-X., la Fleur, S.E., Fliers, E., Kalsbeek, A. The role of the autonomic nervous liver innervation in the control of energy metabolism. *Biochim. Biophys. Acta – Mol. Basis Dis.*, **2010**, *1802*, 416-431.
- [137] Klieverik, L.P., Janssen, S.F., van Riel, A., Foppen, E., Bisschop, P.H., Serlie, M.J., Boelen, A., Ackermans, M.T., Sauerwein, H.P., Fliers, E., Kalsbeek, A. Thyroid hormone modulates glucose production via a sympathetic pathway from the hypothalamic paraventricular nucleus to the liver. *Proc. Natl. Acad. Sci. U. S. A.*, **2009**, *106*, 5966-5971.
- [138] Fredholm, B.B., Abbracchio, M.P., Burnstock, G., Daly, J.W., Harden, T.K., Jacobson, K.A., Leff, P., Williams, M. Nomenclature and classification of purinoceptors. *Pharmacol. Rev.*, **1994**, *46*, 143-156.
- [139] Ralevic, V., Burnstock, G. Receptors for purines and pyrimidines. *Pharmacol. Rev.*, **1998**, *50*, 413-492.
- [140] Cham, J.L., Owens, N.C., Barden, J.A., Lawrence, A.J., Badoer, E. P2x purinoceptor subtypes on paraventricular nucleus neurones projecting to the rostral ventrolateral medulla in the rat. *Exp. Physiol.*, **2006**, *91*, 403-411.
- [141] Kirsch, G.E., Codina, J., Birnbaumer, L., Brown, A.M. Coupling of atp-sensitive k<sup>+</sup> channels to α1-receptors by g-proteins in rat ventricular myocytes. *Am. J. Physiol.*, **1990**, *259*, H820-H826.
- [142] Dart, C., Standen, N.B. Adenosine-activated potassium current in smooth-muscle cells isolated from the pig coronary-artery. *J. Physiol-London*, **1993**, *471*, 767-786.
- [143] Barrett-Jolley, R., Comtois, A., Davies, N.W., Stanfield, P.R., Standen, N.B. Effect of adenosine and intracellular gtp on k-atp channels of mammalian skeletal muscle. *J. Membr. Biol.*, **1996**, *152*, 111-116.
- [144] Mondal, M.S., Yamaguchi, H., Date, Y., Shimbara, T., Toshinai, K., Shimomura, Y., Mori, M., Nakazato, M. A role for neuropeptide w in the regulation of feeding behavior. *Endocrinology*, **2003**, *144*, 4729-4733.
- [145] Jhamandas, J.H., MacTavish, D., Harris, K.H. Neuropeptide ff (npff) control of magnocellular neurosecretory cells of the rat hypothalamic paraventricular nucleus (pvn). *Peptides*, **2006**, *27*, 973-979.
- [146] Jhamandas, J.H., Simonin, F., Bourguignon, J.J., Harris, K.H. Neuropeptide ff and neuropeptide vf inhibit gabaergic neurotransmission in parvocellular neurons of the rat hypothalamic paraventricular nucleus. *Am. J. Physiol-Regul. Integr. Compar. Physiol.*, **2007**, *292*, R1872-R1880.
- [147] Jhamandas, J.H., MacTavish, D. Central administration of neuropeptide ff causes activation of oxytocin paraventricular hypothalamic neurones that project to the brainstem. *J. Neuroendocrinol.*, **2003**, *15*, 24-32.
- [148] Maratos-Flier, E. The long reach of leptin. *Nat. Med.*, **2008**, *14*, 604-606.
- [149] Morton, G.J., Cummings, D.E., Baskin, D.G., Barsh, G.S., Schwartz, M.W. Central nervous system control of food intake and body weight. *Nature*, **2006**, *443*, 289-295.
- [150] Cone, R.D. Anatomy and regulation of the central melanocortin system. *Nat. Neurosci.*, **2005**, *8*, 571-578.
- [151] Meister, B., Hakansson, M.L. Leptin receptors in hypothalamus and circumventricular organs. *Clin. Exp. Pharmacol. Physiol.*, **2001**, *28*, 610-617.
- [152] Powis, J.E., Bains, J.S., Ferguson, A.V. (1998) Leptin depolarizes rat hypothalamic paraventricular nucleus neurons. *Am. J. Physiol-Regul. Integr. Comp. Physiol.*, **274**, R1468-R1472.
- [153] Shirasaka, T., Miyahara, S., Kunitake, T., Jin, Q.H., Kato, K., Takasaki, M., Kannan, H. Orexin depolarizes rat hypothalamic paraventricular nucleus neurons. *Am. J. Physiol-Regul. Integr. Comp. Physiol.*, **2001**, *281*, R1114-R1118.
- [154] Shan, J., Kruckoff, T.L. Intracerebroventricular adrenomedullin stimulates the hypothalamic-pituitary-adrenal axis, the sympathetic nervous system and production of hypothalamic nitric oxide. *J. Neuroendocrinol.*, **2001**, *13*, 975-984.
- [155] Badoer, E. Hypothalamic paraventricular nucleus and cardiovascular regulation. *Clin. Exp. Pharmacol. Physiol.*, **2001**, *28*, 95-99.
- [156] Herd, J.A. Cardiovascular response to stress in man. *Annu. Rev. Physiol.*, **1984**, *46*, 177-185.
- [157] Everson, S.A., Kaplan, G.A., Goldberg, D.E., Salonen, J.T. Anticipatory blood pressure response to exercise predicts future high blood pressure in middle-aged men. *Hypertension*, **1996**, *27*, 1059-1064.
- [158] Berdina, N.A., Kolenko, O.L., Kotz, I.M., Kuznetsov, A.P., Rodionov, I.M., Savchenko, A.P., Thorevsky, V.I. Increase in skeletal muscle performance during emotional stress in man. *Circ. Res.*, **1972**, *30*, 642-650.
- [159] Gillespie, R.D. The relative influence of mental and muscular work on the pulse-rate and blood-pressure. *J. Physiol-London*, **1924**, *58*, 425-432.
- [160] Herd, J.A. Cardiovascular response to stress. *Physiol. Rev.*, **1991**, *71*, 305-330.
- [161] Folkow, B. Mental stress and its importance for cardiovascular disorders; physiological aspects, "From-mice-to-man". *Scand. Cardiovasc. J.*, **2001**, *35*, 163-172.
- [162] Rozanski, A., Blumenthal, J.A., Kaplan, J. Impact of psychological factors on the pathogenesis of cardiovascular disease and implications for therapy. *Circulation*, **1999**, *99*, 2192-2217.
- [163] Duan, Y.F., Winters, R., McCabe, P.M., Green, E.J., Huang, Y., Schneiderman, N. Cardiorespiratory components of defense reaction elicited from paraventricular nucleus. *Physiol. Behav.*, **1997**, *61*, 325-330.
- [164] Busnardo, C., Tavares, R.F., Resstel, L.B.M., Elias, L.L.K., Correa, F.M.A. Paraventricular nucleus modulates autonomic and neuroendocrine responses to acute restraint stress in rats. *Autonomic Neurosci.*, In Press.
- [165] Li, Y.F., Wang, W., Mayhan, W.G., Patel, K.P. Angiotensin-mediated increase in renal sympathetic nerve discharge within the pvn: Role of nitric oxide. *Am. J. Physiol.-Regul. Integr. Comp. Physiol.*, **2006**, *290*, R1035-R1043.
- [166] Martin, D.S., Haywood, J.R. Sympathetic nervous-system activation by glutamate injections into the paraventricular nucleus. *Brain Res.*, **1992**, *577*, 261-267.
- [167] Martin, D.S., Haywood, J.R., Thornhill, J.A. Stimulation of the hypothalamic paraventricular nucleus causes systemic vasoconstriction. *Brain Res.*, **1993**, *604*, 318-324.
- [168] Jessop, D.S., Chowdry, H.S., Biswas, S., Lightman, S.L. Substance-p and substance-k in the rat hypothalamus following monosodium glutamate lesions of the arcuate nucleus. *Neuropeptides*, **1991**, *18*, 165-170.
- [169] Unger, T., Becker, H., Petty, M., Demmert, G., Schneider, B., Ganten, D., Lang, R.E. Differential-effects of central angiotensin-ii and substance-p on sympathetic-nerve activity in conscious rats - implications for cardiovascular adaptation to behavioral-responses. *Circ. Res.*, **1985**, *56*, 563-575.
- [170] Itoi, K., Jost, N., Badoer, E., Tschope, C., Culman, J., Unger, T. Localization of the substance p-induced cardiovascular responses in the rat hypothalamus. *Brain Res.*, **1991**, *558*, 123-126.
- [171] Nakayama, Y., Takano, Y., Saito, R., Kamiya, H. Central pressor actions of tachykinin nk-3 receptor in the paraventricular nucleus of the rat hypothalamus. *Brain Res.*, **1992**, *595*, 339-342.
- [172] Takano, Y., Nakayama, Y., Matsumoto, T., Saito, R., Kamiya, H. The mechanism of central pressor actions of tachykinin nk-3 receptor in the paraventricular nucleus of the hypothalamus in rats. *Regul. Peptides*, **1993**, *46*, 360-363.
- [173] Culman, J., Klee, S., Ohlendorf, C., Unger, T. Effect of tachykinin receptor inhibition in the brain on cardiovascular and behavioral responses to stress. *J. Pharmacol. Exp. Ther.*, **1997**, *280*, 238-246.
- [174] Cannon, W.B. Bodily changes in pain hunger, fear and rage - an account of recent researches into the function of emotional excitement. New York & London, D. Appleton & Company **1927**.
- [175] Stotz-Potter, E.H., Morin, S.M., DiMicco, J.A. Effect of microinjection of muscimol into the dorsomedial or paraventricular hypothalamic nucleus on air stress-induced neuroendocrine and cardiovascular changes in rats. *Brain Res.*, **1996**, *742*, 219-224.
- [176] Pyner, S., Deering, J., Coote, J.H. Right atrial stretch induces renal nerve inhibition and c-fos expression in parvocellular neurones of the paraventricular nucleus in rats. *Exp. Physiol.*, **2002**, *87*, 25-32.
- [177] Deering, J., Coote, J.H. Paraventricular neurones elicit a volume expansion-like change of activity in sympathetic nerves to the heart and kidney in the rabbit. *Exp. Physiol.*, **2000**, *85*, 177-186.

- [178] Greenway, C.V., Lister, G.E. Capacitance effects and blood reservoir function in splanchnic vascular bed during non-hypotensive hemorrhage and blood-volume expansion in anesthetized cats. *J. Physiol-London*, **1974**, 237, 279-294.
- [179] Fater, D.C., Schultz, H.D., Sundet, W.D., Mapes, J.S., Goetz, K.L. Effects of left atrial stretch in cardiac-denervated and intact conscious dogs. *Am. J. Physiol-Heart Circ. Physiol.*, **1982**, 242, H1056-1064.
- [180] Badoer, E., McKinley, M.J., Oldfield, B.J., McAllen, R.M. A comparison of hypotensive and nonhypotensive hemorrhage on fos expression in spinally projecting neurons of the paraventricular nucleus and rostral ventrolateral medulla. *Brain Res.*, **1993**, 610, 216-223.
- [181] Stocker, S.D., Cunningham, J.T., Toney, G.M. Water deprivation increases fos immunoreactivity in pvn autonomic neurons with projections to the spinal cord and rostral ventrolateral medulla. *Am. J. Physiol-Regul. Integrat. Comp. Physiol.*, **2004**, 287, R1172-R1183.
- [182] Stocker, S.D., Keith, K.J., Toney, G.M. Acute inhibition of the hypothalamic paraventricular nucleus decreases renal sympathetic nerve activity and arterial blood pressure in water-deprived rats. *Am. J. Physiol-Regul. Integrat. Comp. Physiol.*, **2004**, 286, R719-R725.
- [183] Millar-Craig, M.W., Bishop, C.N., Raftery, E.B. Circadian variation of blood-pressure. *Lancet*, **1978**, I, 795-797.
- [184] Degaute, J.P., Vandeborne, P., Linkowski, P., Vancauter, E. Quantitative-analysis of the 24-hour blood-pressure and heart-rate patterns in young men. *Hypertension*, **1991**, 18, 199-210.
- [185] Verdecchia, P., Schillaci, G., Guerrieri, M., Gatteschi, C., Benemio, G., Boldrini, F., Porcellati, C. Circadian blood pressure changes and left ventricular hypertrophy in essential hypertension. *Circulation*, **1990**, 81, 528-536.
- [186] Pickering, T.G. The clinical significance of diurnal blood pressure variations. Dippers and nondippers. *Circulation*, **1990**, 81, 700-702.
- [187] Boari, B., Salmi, R., Gallerani, M., Malagoni, A.M., Manfredini, F., Manfredini, R. Acute myocardial infarction: Circadian, weekly, and seasonal patterns of occurrence. *Biol. Rhythm. Res.*, **2007**, 38, 155-167.
- [188] Muller, J.E., Tofler, G.H., Stone, P.H. Circadian variation and triggers of onset of acute cardiovascular-disease. *Circulation*, **1989**, 79, 733-743.
- [189] Brown, T.M., Piggins, H.D. Electrophysiology of the suprachiasmatic circadian clock. *Prog. Neurobiol.*, **2007**, 82, 229-255.
- [190] Tei, H., Okamura, H., Shigeyoshi, Y., Fukuhara, C., Ozawa, R., Hirose, M., Sakaki, Y. Circadian oscillation of a mammalian homologue of the drosophila period gene. *Nature*, **1997**, 389, 512-516.
- [191] Reppert, S.M., Weaver, D.R. Coordination of circadian timing in mammals. *Nature*, **2002**, 418, 935-941.
- [192] Hermes, M.L., Buijs, R.M., Renaud, L.P. Electrophysiology of suprachiasmatic nucleus projections to hypothalamic paraventricular nucleus neurons. *Prog. Brain Res.*, **1996**, 111, 241-252.
- [193] Chen, K.K., Chan, S.H.H., Chang, L.S., Chan, J.Y.H. Participation of paraventricular nucleus of hypothalamus in central regulation of penile erection in the rat. *J. Urol.*, **1997**, 158, 238-244.
- [194] Argiolas, A., Melis, M.R. Neuromodulation of penile erection: An overview of the role of neurotransmitters and neuropeptides. *Prog. Neurobiol.*, **1995**, 47, 235-255.
- [195] Andersson, K.E., Wagner, G. Physiology of penile erection. *Physiol. Rev.*, **1995**, 75, 191-236.
- [196] Argiolas, A., Melis, M.R. Central control of penile erection: Role of the paraventricular nucleus of the hypothalamus. *Prog. Neurobiol.*, **2005**, 76, 1-21.
- [197] Marson, L., Platt, K.B., McKenna, K.E. Central-nervous-system innervation of the penis as revealed by the transneuronal transport of pseudorabies virus. *Neuroscience*, **1993**, 55, 263-280.
- [198] Stafford, S.A., Tang, K., Coote, J.H. Activation of lumbosacral 5-HT<sub>2C</sub> receptors induces bursts of rhythmic activity in sympathetic nerves to the vas deferens in male rats. *Br. J. Pharmacol.*, **2006**, 148, 1083-1090.
- [199] Stafford, S.A., Bowery, N.G., Tang, K., Coote, J.H. Activation by p-chloroamphetamine of the spinal ejaculatory pattern generator in anaesthetized male rats. *Neuroscience*, **2006**, 140, 1031-1040.
- [200] Levin, B.E. Metabolic sensing neurons and the control of energy homeostasis. *Physiol. Behav.*, **2006**, 89, 486-489.
- [201] Sakaguchi, T., Bray, G.A. Sympathetic activity following paraventricular injections of glucose and insulin. *Brain Res Bull.*, **1988**, 21, 25-29.
- [202] Carrasco, M., Portillo, F., Larsen, P.J., Vallo, J.J. Insulin and glucose administration stimulates fos expression in neurones of the paraventricular nucleus that project to autonomic preganglionic structures. *J. Neuroendocrinol.*, **2001**, 13, 339-346.
- [203] Dunn-Meynell, A.A., Govek, E., Levin, B.E. Intracarotid glucose selectively increases fos-like immunoreactivity in paraventricular, ventromedial and dorsomedial nuclei neurons. *Brain Res.*, **1997**, 748, 100-106.
- [204] Stanley, S., Pinto, S., Segal, J., Perez, C.A., Viale, A., DeFalco, J., Cai, X.L., Heisler, L.K., Friedman, J.M. Identification of neuronal subpopulations that project from hypothalamus to both liver and adipose tissue polysynaptically. *Proc. Natl. Acad. Sci USA*, **2010**, 107, 7024-7029.
- [205] Oldfield, B.J., Giles, M.E., Watson, A., Anderson, C., Colvill, L.M., McKinley, M.J. The neurochemical characterisation of hypothalamic pathways projecting polysynaptically to brown adipose tissue in the rat. *Neuroscience*, **2002**, 110, 515-526.
- [206] Bamshad, M., Song, C.K., Bartness, T.J. Cns origins of the sympathetic nervous system outflow to brown adipose tissue. *Am. J. Physiol-Regul. Integr. Compar. Physiol.*, **1999**, 276, R1569-R1578.
- [207] Amir, S. Stimulation of the paraventricular nucleus with glutamate activates interscapular brown adipose-tissue thermogenesis in rats. *Brain Res.*, **1990**, 508, 152-155.
- [208] Yoshimatsu, H., Egawa, M., Bray, G.A. Effects of cholecystokinin on sympathetic activity to interscapular brown adipose-tissue. *Brain Res.*, **1992**, 597, 298-303.
- [209] Yoshimatsu, H., Egawa, M., Bray, G.A. Sympathetic-nerve activity after discrete hypothalamic injections of l-glutamate. *Brain Res.*, **1993**, 601, 121-128.
- [210] Madden, C.J., Morrison, S.F. Neurons in the paraventricular nucleus of the hypothalamus inhibit sympathetic outflow to brown adipose tissue. *Am. J. Physiol-Regul. Integr. Comp. Physiol.*, **2009**, 296, R831-R843.
- [211] Inenaga, K., Osaka, T., Yamashita, H. Thermosensitivity of neurons in the paraventricular nucleus of the rat slice preparation. *Brain Res.*, **1987**, 424, 126-132.
- [212] Smith, J.E., Jansen, A.S.P., Gilbey, M.P., Loewy, A.D. Cns cell groups projecting to sympathetic outflow of tail artery: Neural circuits involved in heat loss in the rat. *Brain Res.*, **1998**, 786, 153-164.
- [213] Cham, J.L., Badoer, E. Hypothalamic paraventricular nucleus is critical for renal vasoconstriction elicited by elevations in body temperature. *Am. J. Physiol-Renal Physiol.*, **2008**, 294, F309-F315.
- [214] Chen, F., Dworak, M., Wang, Y., Cham, J.L., Badoer, E. Role of the hypothalamic pvn in the reflex reduction in mesenteric blood flow elicited by hyperthermia. *Am. J. Physiol. Regul. Integr. Comp. Physiol.*, **2008**, 295, R1874-1881.
- [215] Cham, J.L., Badoer, E. Exposure to a hot environment can activate rostral ventrolateral medulla-projecting neurones in the hypothalamic paraventricular nucleus in conscious rats. *Exp. Physiol.*, **2008**, 93, 64-74.
- [216] Cham, J.L., Klein, R., Owens, N.C., Mathai, M., McKinley, M., Badoer, E. Activation of spinally projecting and nitrergic neurons in the pvn following heat exposure. *Am. J. Physiol-Regul. Integr. Comp. Physiol.*, **2006**, 291, R91-R101.
- [217] Roddie, I.C., Shepherd, J.T., Whelan, R.F. The contribution of constrictor and dilator nerves to the skin vasodilation during body heating. *J. Physiol-London*, **1957**, 136, 489-497.
- [218] Patel, K.P., Zhang, K., Kenney, M.J., Weiss, M., Mayhan, W.G. Neuronal expression of fos protein in the hypothalamus of rats with heart failure. *Brain Res.*, **2000**, 865, 27-34.
- [219] Calabrese, V., Mancuso, C., Calvani, M., Rizzarelli, E., Butterfield, D.A., Stella, A.M.G. Nitric oxide in the central nervous system:

- Neuroprotection versus neurotoxicity. *Nat. Rev. Neurosci.*, **2007**, *8*, 766-775.
- [220] Garthwaite, J. Neural nitric-oxide signaling. *Trends Neurosci.*, **1995**, *18*, 51-52.
- [221] Yamaguchi, N., Okada, S., Usui, D., Yokotani, K. Nitric oxide synthase isozymes in spinally projecting pvn neurons are involved in crf-induced sympathetic activation. *Auton. Neurosci-Basic Clin.*, **2009**, *148*, 83-89.
- [222] Kantzides, A., Badoer, E. Nnos-containing neurons in the hypothalamus and medulla project to the rvlm. *Brain Res.*, **2005**, *1037*, 25-34.
- [223] Li, Y., Zhang, W., Stern, J.E. Nitric oxide inhibits the firing activity of hypothalamic paraventricular neurons that innervate the medulla oblongata: Role of gaba. *Neuroscience*, **2003**, *118*, 585-601.
- [224] Bains, J.S., Ferguson, A.V. Nitric oxide regulates nmda-driven gabaergic inputs to type i neurones of the rat paraventricular nucleus. *J. Physiol-London*, **1997**, *499*, 733-746.
- [225] Zhang, K., Zucker, I.H., Patel, K.P. Altered number of diaphorase (nos) positive neurons in the hypothalamus of rats with heart failure. *Brain Res.*, **1998**, *786*, 219-225.
- [226] Zheng, H., Li, Y.F., Wang, W., Patel, K.P. Enhanced angiotensin-mediated excitation of renal sympathetic nerve activity within the paraventricular nucleus of anesthetized rats with heart failure. *Am. J. Physiol-Regul. Integr. Comp. Physiol.*, **2009**, *297*, R1364-R1374.
- [227] Strack, A.M., Sawyer, W.B., Hughes, J.H., Platt, K.B., Loewy, A.D. A general pattern of cns innervation of the sympathetic outflow demonstrated by trans-neuronal pseudorabies viral-infections. *Brain Res.*, **1989**, *491*, 156-162.

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