

Emerging Trends in Gas Chromatography and Mass Spectrometry Instrumentation for Analytical & Bioanalytical Techniques

Wes E Steiner^{1*} and William A English²

¹Department of Chemistry and Biochemistry, Eastern Washington University, 226 science Building, Cheney, WA 99004, USA

²Forensic Toxicology Drug Testing Laboratory, Tripler Army Medical Center, 1 Jarrett White Road, Honolulu, HI 96859, USA

*Corresponding Author: Wes E Steiner, Department of Chemistry and Biochemistry, Eastern Washington University, 226 science, Building, Cheney, USA, Tel: (509) 359-6521, Fax: (509) 359-6973; E-mail: wsteiner@ewu.edu

Received date: February 22, 2015, Accepted date: February 24, 2015, Published date: March 3, 2015

Copyright: © 2015 Steiner WE, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Description

Innovations in Gas Chromatography (GC) and Mass Spectrometry (MS) instrumentation from a variety of scientists working in the public and private sectors of research and development have driven this traditional two-dimensional hyphenated technology, and/or recently three- and four-dimensional hyphenated technologies in the case of a second GC and/or a MS equivalent being added, to become one of the dominant platforms for use in a wide assortment of analytical and bioanalytical techniques. The analysis of drugs, metabolites, pesticides, chemical warfare agents, food ingredients, medications, fuels, and etcetera [1-3] and/or in the main category of volatile and semi-volatile organic compound analysis in fields such as forensic, toxicology, environment, defense, food and beverage, pharmaceutical, petrochemical, and etcetera [4-6] are all now considered to be growing in size and in scope in the use of GC-MS instrumentation. The advancement in GC-MS instrumentation was initially and still is driven by the need for a more comprehensive analytical and bioanalytical technique that can accurately and precisely discriminate targeted and untargeted analytes from higher complexity sample mixtures in a sensitive and selective way. To that end, this review, like the preceding review on LC-MS [7], will briefly attempt to focus on the most current emerging trends in GC-MS instrumentation and their respective contributions to the field of analytical and bioanalytical techniques.

A couple of distinctions, often found in use with LC-MS, can be applied in the use and discussion of GC-MS in the forms of qualitative and quantitative analysis of higher complexity sample mixtures. With qualitative GC-MS and GCxGC-MS often taking the form of discovery types of assays employing GC coupled to mass spectrometers that can include for example quadrupole-ion trap and time-of-flight mass spectrometers (i.e. QITMS, TOFMS respectively), or GCxGC coupled to TOFMS [8-10]. Whereas with quantitative GC-MS and GCxGC-MS often taking the form of directed types of assays that can employ GC coupled to a variety of mass spectrometers that utilize high resolution magnetic sector, single quadrupole, and triple quadrupole mass spectrometers (i.e. HRMS, QMS, QqQMS), or GCxGC coupled to QqQMS [11-13]. These two types of distinctions in use in instrumentation (i.e. qualitative and quantitative) have now started to become more integrated with the emergence of a new class of GC-MS instrumentation that has the ability to a degree to do both routine qualitative as well as routine quantitative analysis of targeted and untargeted analytes of higher complexity sample mixtures. This new class of GC-MS instrumentation, for example, typically takes the form of a GC or GCxGC separation system that is interfaced to two or more mass spectrometers that are placed in series to form a hybrid mass

spectrometer such as a three-dimensional GC-QMS/TOFMS or a four-dimensional GCxGC-QMS/TOFMS system [14-17].

Before a continuation of our discussions of new class of GC-MS instrumentation a moment should be taking to briefly review the trend of the combination of serially stacking two standalone one-dimensional GC systems together to form a two-dimensional GCxGC system of separation. Here a pair of GC columns typically located in two separate oven compartments that are generally employing nonpolar and polar packed stationary phases are connected in series via a modulator that repeatedly traps and focuses the eluent from the first column before injecting it into the second column. This results in a two-dimensional chromatogram with retention times along both the x- and y-axis. Why do this and what advantages does this bring to the scientist, is common set of questions that come to mind. In addressing these two questions specifically let's take a look at what GCxGC facilitates in the form of compounds with similar boiling points that could not be separated with baseline resolution with single GC can now be separated based upon their differences in polarity. This in turn now allows for the analysis of higher complexity samples that were previously difficult to separate. Additionally, GCxGC allows for the recognition of patterns that can be correlated to groupings of compound structural classes such as alkanes, alkenes, alkynes, and etcetera. Overall the main advantages of GCxGC can often be summarized with an increased sensitivity, enhanced selectivity, increased number of compounds resolved with baseline resolution per unit of time, and as mentioned above, the formation of visualized organized patterns of compounds with the same functional class groups [18-22].

Now in focus, when pairing for example a hybrid MS system (i.e. QMS/TOFMS) as mentioned previously above, with that of a GC and/or a GCxGC system, the serial stacking of two typically independent MS instruments provides in this case a uniquely integrated qualitative and quantitative analysis workflow for the analysis of targeted and untargeted analytes of higher complexity sample mixtures of volatile and semi-volatile organic compounds into a single platform. Here the frontend of this MS system employs a QMS that is employing an Electron Ionization (EI) and/or a Chemical Ionization (CI) source that enables precursor ion scan regimes along with nominal mass EI spectral library searches over a mass range of 20-1050 Da. The middle of this MS system employs a linear hexapole Collision Induced Dissociation (CID) cell that enables precursor to product ion transitions to aid in, for example, structural elucidation for most components in higher complexity sample mixtures. With the backend of this MS system employing a TOFMS that enables precursor ion scan, product ion scan, and Multiple Reaction Monitoring (MRM) scan regimes that typically delivers acquisition speeds of 1 to 50 spectra

per second, a dynamic range of 10^3 to 10^5 , a level of sample peak resolution of 13,000 to 15,000 FWHM, a level of mass accuracy of 2 to 5 ppm, and a mass range of 20-1700 Da with 15-3000 Da extended. It should be noted that since the TOFMS continuously acquires all masses for a given mass range simultaneously the MRM scan regime mimics QqQMS instruments by using accurate mass extracted ion mass and/or masses from the product ion scan. These TOFMS attributes, with emphasis being on accurate mass identification determinations of both precursor and product ions, allows for the effective analysis of higher complexity sample mixtures thereby enabling the construction and utilization of exact mass libraries of compounds (i.e. drugs, pesticides, metabolites, and etcetera). To that end, when the combination of the QMS with that of a TOFMS takes place to form a hybrid QMS/TOFMS instrument scientists are able to utilize a rapid, sensitive, selective, higher-resolution, accurate mass hybrid style of MS that is able to help facilitate, for example, the deconvolution of coeluting GC chromatographic peaks during routine qualitative and quantitative analysis assays even if only a single GC system is used [14,15,19,23].

As scientists build upon these innovative progressions of GC-MS instrumentation that has the unique ability to a degree to do both routine qualitative and routine quantitative analysis of targeted and untargeted analytes of higher complexity sample mixtures of volatile and semi-volatile organic compounds for analytical & bioanalytical techniques we feel optimistic that this new emergent class of GC-MS instrumentation employing hybrid QMS/TOFMS technology will begin to phase out some of the traditional classes of standalone GC-MS instrumentation that are currently found in many present day laboratories. It should be noted, as was the case with that of LC-MS employing hybrid QMS/TOFMS technology, that the ability to combine routine qualitative and quantitative analysis into one instrument is not just the savings in cost of the replacement of two standalone instruments with that of one, but rather an increase in the amount of information obtained by the scientist from a single analysis. This has become especially important and advantageous to scientists working with and/or screening higher complexity samples wishing to re-investigate rich data sets of virtually unlimited numbers of compounds that were previously assayed but not analyzed for all targeted and/or untargeted component compounds of interest at that time. In comparison, conventional screening methods of higher complexity samples that employ for example standard non-hybrid QqQMS technology are limited to a narrow number of targeted compounds analyzed only and do not allow for a retrospective analysis of previously collected untargeted data. Overall, this emerging trend in GC-MS instrumentation employing hybrid QMS/TOFMS technology may just be the shift scientists have been waiting for in the main category of volatile and semi-volatile organic compound analysis in the offering of three- and/or four-dimensional, temporally resolved, information in a single GC-MS instrument for analytical & bioanalytical techniques. Lastly, as we conclude this review scientists have taken yet another innovative step forward in the research and development of an integrated five-dimensional hyphenated technology in the form of adding liquid chromatography separations to GC-MS in the form of an LC-GCxGC-QqQMS system [24]. Here, as highlighted by the authors, depending upon the analytical & bioanalytical techniques required for a given assay a number of instrumental configurations may be employed.

Conflict of Interest

The views expressed in this manuscript are those of the author(s) and do not reflect the official policy or position of the Department of Army, Department of Defense, or the U.S. Government.

References

1. Wink CSD, Meyer GMJ, Zapp J, Maurer HH (2015) Lefetamine, a controlled drug and pharmaceutical lead of new designer drugs: synthesis, metabolism, and detectability in urine and human liver preparations using GC-MS, LC-MSn, and LC-high resolution-MS/MS. *Analytical and Bioanalytical Chemistry*, 407: 1545-1557.
2. Albo RL, Valdez CA, Leif RN, Mulcahy HA, Koester C (2014) Derivatization of pinacolyl alcohol with phenyldimethylchlorosilane for enhanced detection by gas chromatography-mass spectrometry. *Anal Bioanal Chem* 406: 5231-5234.
3. Kanateva, AY, Kurganov AA, Yakubenko EE (2014) Application of two-dimensional gas chromatography-mass spectrometry to determination of biodiesel impurities in hydrocarbon fuels. *Petroleum Chemistry*, 6:459-465.
4. Welter J, Kavanagh P, Meyer MR, Maurer HH (2014) Benzofuran analogues of amphetamine and methamphetamine: studies on the metabolism and toxicological analysis of 5-APB and 5-MAPB in urine and plasma using GC-MS and LC-(HR)-MSn techniques. *Analytical and Bioanalytical Chemistry*, 407: 1371-1388.
5. Baron E, Eljarrat E, Barcelo D (2014) Gas chromatography/tandem mass spectrometry method for the simultaneous analysis of 19 brominated compounds in environmental and biological samples. *Anal Bioanal Chem*, 29:7667-7676.
6. Kächele M, Monakhova YB, Kuballa T, Lachenmeier DW4 (2014) NMR investigation of acrolein stability in hydroalcoholic solution as a foundation for the valid HS-SPME/GC-MS quantification of the unsaturated aldehyde in beverages. *Anal Chim Acta* 820: 112-118.
7. Steiner WE, English WA (2012) Emerging Trends in Liquid Chromatography and Mass Spectrometry Instrumentation for Analytical & Bioanalytical Techniques. *J. Analytical and Bioanalytical Techniques*, 4:1-2.
8. Russo MV, Notardonato I, Avino P, Cinelli G (2014) Determination of phthalate esters at trace levels in light alcoholic drinks and soft drinks by XAD-2 adsorbent and gas chromatography coupled with ion trap-mass spectrometry detection. *Analytical Methods*, 6:7030-7037.
9. Silva I, Coimbra MA, Barros AS, Marriott PJ, Rocha SM3 (2015) Can volatile organic compounds be markers of sea salt. *Food Chem* 169: 102-113.
10. Kehimkar B, Parsons BA, Hoggard JC, Billingsley MC, Bruno TJ, Synovec RE (2015) Modeling RP-1 fuel advanced distillation data using comprehensive two-dimensional gas chromatography coupled with time-of-flight mass spectrometry and partial least squares analysis. *Anal Bioanal Chem.*, 407:321-330.
11. Croes K, Colles A, Koppen G, De Galan S, Vandermarken T, et al. (2013) Determination of PCDD/Fs, PBDD/Fs and dioxin-like PCBs in human milk from mothers residing in the rural areas in Flanders, using the CALUX bioassay and GC-HRMS. *Talanta* 113: 99-105.
12. Tranchida PQ, Franchina FA, Zoccali M, Pantò S, Sciarbone D, et al. (2013) Untargeted and targeted comprehensive two-dimensional GC analysis using a novel unified high-speed triple quadrupole mass spectrometer. *J Chromatogr A* 1278: 153-159.
13. L'Homme B, Scholl G, Eppe G, Focant JF (2015) Validation of a gas chromatography-triple quadrupole mass spectrometry method for confirmatory analysis of dioxins and dioxin-like polychlorobiphenyls in feed following new EU Regulation 709/2014. *J Chromatogr A* 1376: 149-158.
14. Canellas E, Vera P, Nerin C (2014) Atmospheric pressure gas chromatography coupled to quadrupole-time of flight mass spectrometry as a tool for identification of volatile migrants from auto adhesive labels used for direct food contact. *Journal of Chromatography A*, 49:1181-1190.

15. Wylie PL, Aronova S (2014) Automated screening for hundreds of pesticide residues using a GC/Q-TOF with a new exact mass pesticide database. 248th ACS National Meeting & Exposition, California, USA.
16. Ochiai N, Mitsui K, Sasamoto K, Yoshimura Y, David F, et al. (2014) Multidimensional gas chromatography in combination with accurate mass, tandem mass spectrometry, and element-specific detection for identification of sulfur compounds in tobacco smoke. *J Chromatogr A* 1358: 240-251.
17. Zeng AX, Chin ST, Patti A, Marriott PJ (2013) Profiling of soil fatty acids using comprehensive two-dimensional gas chromatography with mass spectrometry detection. *J Chromatogr A* , 1317: 239-245.
18. Eiserbeck C, Nelson RK, Reddy CM, Grice K (2015) Advances in comprehensive two-dimensional gas chromatography (GCxGC). *Principles and Practice of Analytical Techniques in Geosciences*, 4:324-365.
19. Mitrevski B, Marriott PJ (2014) Evaluation of quadrupole-time-of-flight mass spectrometry in comprehensive two-dimensional gas chromatography. *J Chromatogr A* 1362: 262-269.
20. Silva BJ, Tranchida PQ, Purcaro G, Queiroz ME, Mondello L, et al. (2012) Evaluation of comprehensive two-dimensional gas chromatography coupled to rapid scanning quadrupole mass spectrometry for quantitative analysis. *J Chromatogr A* 1255: 177-183.
21. Parsons BA, Reaser BC, Pinkerton DK, Synovec RE (2014) Development of discovery-based techniques for the comprehensive analysis of complex samples with GC-TOFMS: Biomarker discovery for food products. 247th ACS National Meeting & Exposition, California, USA.
22. Tranchida PQ, Franchina FA, Dugo P, Mondello L (2014) Comprehensive two-dimensional gas chromatography-mass spectrometry: Recent evolution and current trends. *Mass Spectrom Rev*.
23. Chin ST, Nolvachai Y, Marriott PJ (2014) Enantiomeric separation in comprehensive two-dimensional gas chromatography with accurate mass analysis. *Chirality* 26: 747-753.
24. Zoccali M, Tranchida PQ, Mondello L (2015) On-line combination of high performance liquid chromatography with comprehensive two-dimensional gas chromatography-triple quadrupole mass spectrometry: a proof of principle study. *Anal Chem* 87: 1911-1918.