

Nano-TiO₂/PEEK bioactive composite as a bone substitute material: in vitro and in vivo studies

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Background: Compared with titanium (Ti) and other metal implant materials, poly(ether-ether ketone) (PEEK) shows outstanding biomechanical properties. A number of studies have also reported attractive bioactivity for nano-TiO₂ (n-TiO₂).

Methods: In this study, n-TiO₂/PEEK nanocomposites were prepared, taking advantage of the unique properties of both PEEK polymer and n-TiO₂. The in vitro and in vivo bioactivity of these nanocomposites was assessed against a PEEK polymer control. The effect of surface morphology or roughness on the bioactivity of the n-TiO₂/PEEK nanocomposites was also studied. n-TiO₂/PEEK was successfully fabricated and cut into disks for physical and chemical characterization and in vitro studies, and prepared as cylindrical implants for in vivo studies. Their presence on the surface and dispersion in the composites was observed and analyzed by scanning and transmission electron microscopy and X-ray photoelectron spectroscopy.

Results: Bioactivity evaluation of the nanocomposites revealed that pseudopods of osteoblasts preferred to anchor at areas where n-TiO₂ was present on the surface. In a cell attachment test, smooth PEEK showed the lowest optical density value (0.56 ± 0.07) while rough n-TiO₂/PEEK exhibited the highest optical density value (1.21 ± 0.34 , $P < 0.05$). In in vivo studies, the percent bone volume value of n-TiO₂/PEEK was approximately twice as large as that of PEEK ($P < 0.05$). Vivid three-dimensional and histologic images of the newly generated bone on the implants further supported our test results.

Conclusion: Our study demonstrates that n-TiO₂ significantly improves the bioactivity of PEEK, especially if it has a rough composite surface. A n-TiO₂/PEEK composite with a rough surface could be a novel alternative implant material for orthopedic and dental applications.

Keywords: polyether-ether-ketone, bioactivity, TiO₂, nanocomposite, polymer, implant

Introduction

Implants of titanium (Ti) and Ti alloy have been widely utilized as orthopedic and dental materials.¹⁻³ However, a number of issues associated with metal implants, including Ti and Ti alloys, have been reported in long-term clinical studies.⁴⁻⁶ Two major issues are stress shielding and local inflammation. Stress shielding is due to stress mismatch between the metal implant material and surrounding bone tissue. Local inflammation is caused by metal implant debris from wear and corrosion. These two issues are considered to be the major causes of bone loss and implant failure.^{5,6}

Poly(ether-ether-ketone) (PEEK), on the other hand, is considered to be one of the best choices to resolve stress shielding issues due to its outstanding biocompatibility and biomechanical properties,⁶ such as a low modulus, compared with a metal implant

and high strength compared with other polymers. However, the bioinert nature of PEEK is not conducive to fast bone cell attachment.⁷⁻⁹ There is a need to improve its bioactivity for orthopedic and dental applications.

The exceptional anticorrosive and biocompatibility properties of Ti and Ti alloy are due to a protective oxide layer (mainly TiO₂) which forms rapidly on the Ti surface when it is exposed to the atmosphere.^{10,11} It was reported that calcium-phosphorus mineralization tended to occur on microgrooved TiO₂ surfaces in the initial days.¹² Using an arc ion plating technique, a thin microsized TiO₂ film was deposited onto a PEEK substrate, which promoted significant adhesion, proliferation, and differentiation of osteoblast cells, compared with a PEEK substrate without TiO₂ coating.¹³

It is believed that TiO₂ nanoparticles have higher bioactivity than conventional (micron) particle sizes. When exposed to nanophase TiO₂ particles, osteoblasts and chondrocytes show a well spread morphology and increased proliferation compared with cells exposed to particles of conventional size.¹⁴ Compared with a micropit titanium surface, a micropit titanium surface with nanonodules promotes significant differentiation and proliferation of osteoblasts in *in vitro* studies.¹⁵ Further, on biomechanical testing of implants, the strength of bone-titanium integration is three times greater for implants with micropits and 300 nm nanonodules than those with micropits alone. A TiO₂ nanotube surface significantly accelerates osteoblast adhesion and shows strong bonding with bone.¹⁶ TiO₂ nanonetwork formation on the Ti surfaces significantly improves human bone marrow mesenchymal stem cell growth *in vitro* and *in vivo*.¹⁰ Therefore, various kinds of n-TiO₂ enhanced polymers have been fabricated for biomaterial applications such as g-TiO₂/poly-L-lactide acid nanocomposites¹⁷ and poly(lactic-co-glycolic acid)/TiO₂ nanoparticle-filled composites.¹⁸ It is reported that poly (D, L lactic acid) film containing 20 wt% TiO₂ could improve the formation of hydroxyapatite (HA) after 21 days' exposure to simulated body fluid and increase the relative metabolic activity of MG-63 cells after seven days of incubation.¹⁹

All the aforementioned studies suggest that the excellent biocompatibility and bioactivity of n-TiO₂ composites is due mainly to the favorable bioactivity of TiO₂ nanoparticles in composites and the surface morphology of the TiO₂ layer. The aim of this study was to utilize n-TiO₂ to improve the bioactivity of PEEK and to investigate the bioactivity of n-TiO₂/PEEK composites both *in vitro* and *in vivo*. Specific attention was also paid to the biologic effect of n-TiO₂ on

the composite surface as well as the biologic effect of the surface roughness of the n-TiO₂/PEEK composite.

Materials and methods

Sample preparation

PEEK powder was obtained from Victrex (Lancashire, UK) and the TiO₂ nanoparticle/PEEK composite (n-TiO₂/PEEK) was fabricated by powder mixing and compression molding methods²⁰ in the Key Laboratory for Ultrafine Material of Ministry of Education, School of Materials Science and Engineering, East China University of Science and Technology, Shanghai. In this study, the amount of n-TiO₂ in the n-TiO₂/PEEK composite was 40 wt% (bending modulus 3.8 GPa; bending strength 93 MPa), because a value greater than this would have interfered with the mechanical properties of the composite (data not shown).

In brief, appropriate amounts of n-TiO₂ and PEEK powder were codispersed using an electronic blender in alcohol to obtain a homogeneous powder mixture. When well dispersed, the mixture was dried in a forced convection oven at 90°C to remove the excess alcohol. The resulting powder mixture was placed in two specially designed molds, ie, disks (Φ 15 × 2 mm) for physical and chemical characterization and *in vitro* testing and cylindrical implants (Φ 4 × 7 mm) for *in vivo* testing. The moulds and powder mixtures were preheated to 150°C under a load of 35 MPa, and the temperature was increased to 400°C under a load of 15 MPa. After reaching the target temperature, the temperature and pressure were held for 10 minutes, and then the heater was turned off and the pressure was released after 10 minutes. The die and the samples were air cooled to 150°C and the samples were removed from the molds.

Disk samples of n-TiO₂/PEEK composite were polished to 2000 and tested by a mechanical profilometer (Dektak8 stylus profiler; Veeco, Plainview, NY). Disk samples with roughness average (Ra) below 0.1 μ m were collected and considered as smooth groups. Some disk samples for the smooth groups were blasted by TiO₂ particles and tested by profilometer, and the disks with Ra 1.0–2.2 μ m were collected and considered to be the rough groups. Unfilled PEEK samples were cut into the same shapes and prepared according to the same process as the control groups. Surface roughness is thought to be beneficial for the bioactivity of implants, and most implants utilized in the clinic are blasted. Therefore, all implant samples in this study were blasted for the animal experiments. In order to remove any potential free TiO₂ nanoparticles, all samples were cleaned with deionized water using an ultrasonic cleaner for 8 hours. The deionized water was changed every 20 minutes during the ultrasonic cleaning process.

Characterization

TEM analysis of n-TiO₂/PEEK composite

In order to characterize the n-TiO₂ dispersion of the n-TiO₂/PEEK composite, thin foil transmission electron microscope (TEM) specimens were prepared by microtome with a diamond knife and imaged using a FEI Tecnai F20 TEM (Philips Electron Optics, Eindhoven, the Netherlands) at 200 KeV.

SEM analysis of n-TiO₂/PEEK composites

After treatment, the samples were sputter-coated with gold by a coating device (550X; Quorum, Hampshire, UK), and the surface morphology of the samples was observed using scanning electron microscopy (SEM; S-4800, Hitachi, Tokyo, Japan).

Composite surface analysis by XPS

An X-ray photoelectron spectroscopy (XPS; Axis-Ultra, Kratos Analytical, Manchester, UK) study was carried out for surface chemical analysis using monochromatic Al K α radiation (225 W, 15 mA, 15 kV) and low-energy electron flooding for charge compensation. Binding energies were calibrated using a C 1 s hydrocarbon peak at 284.8 eV to compensate for surface charge effects. For manipulation and curve-fitting, the data were converted into a VAMAS file format and imported into CasaXPS software package.

Cytocompatibility in vitro

Cell attachment, cytotoxicity, cell morphology, and flow cytometric analysis were evaluated using MG-63 osteoblast cells obtained from the American Type Culture Collection. MG-63 was cultured at 37°C in a humidified, 5% CO₂/95% air incubator, in modified Eagle's medium with 10% fetal bovine serum (Hyclone, Logan, UT), 100 U/mL penicillin (Amresco, Cleveland, OH), and 0.1 mg/mL streptomycin (Amresco). Prior to in vitro testing, the samples were sterilized using gamma radiation at a total dose of 25 KGy. The MG-63 cells were seeded at a density of 1×10^5 cells in each well of 24-well plates for cell attachment testing and at a density of 1.05×10^4 cells/cm² for the other in vitro tests (n = 6). The culture period for cell attachment was 4 hours. There were three culture periods (3, 7, and 14 days) for cytotoxicity, cell morphology, and flow cytometric analysis.

Cell attachment

After culture on samples for 4 hours, the culture medium was removed and the specimen was rinsed with phosphate-buffered saline three times in order to remove the unattached cells.

Cell viability of the adherent cells was measured by adding 1 mL/well culture medium containing 100 μ L/well Cell Proliferation Reagent WST-1 (Roche Diagnostics, Mannheim, Germany). Adherent cells were incubated on samples at 37°C for another 4 hours, and 100 μ L of culture medium was then transferred into each well of a 96-well plate from each corresponding well of a 24-well plate. Ultraviolet absorbance was measured using an enzyme-linked immunosorbent assay reader at 450 nm with the reference wavelength at 630 nm.

Cytotoxicity

After different culture periods (3, 7, and 14 days) cytotoxicity was evaluated using the WST-1 test on the rough and smooth surfaces of PEEK and n-TiO₂/PEEK disks. At fixed detection times, the culture medium was replaced with a new 1 mL/well culture medium containing WST-1 reagent at 100 μ L/well. Cells were cultured at 37°C for another 4 hours and ultraviolet absorbance was measured as described above. The relative growth rate was calculated.

Cell morphology

To observe the cell morphology, the samples were washed with phosphate-buffered saline at fixed experimental times (3, 7, and 14 days), and cells on the various materials were fixed with 4% glutaraldehyde in phosphate-buffered saline (pH 7.3) for 30 minutes and dehydrated in a graded series of alcohols. Prior to SEM observation, the cells on the samples were sputter-coated and observed as described above.

Flow cytometric analysis

To investigate the cell cycle of MG-63 osteoblasts further, the cells were stained using propidium iodide (Invitrogen, Carlsbad, CA) and analyzed by flow cytometry (BD FACSCalibur; Becton-Dickson, Franklin Lakes, NJ). In summary, the cells were initially cultured on samples for 3, 7, and 14 days. They were then collected, washed with phosphate-buffered saline (4°C), and resuspended in phosphate-buffered saline. Next, the cells were fixed in 75% ethanol and kept in ethanol at -20°C for 16 hours. Finally, they were collected and washed again, and resuspended in 1 mL RNaseA (1 μ g/mL) at room temperature for 30 minutes. The sample cells were pelleted by centrifugation, propidium iodide (Invitrogen) was added, and the cells were incubated for 15 minutes at room temperature, collected, and suspended once again in 1 mL Tris-HCl buffer (pH 7.4), then analyzed by flow cytometry.

Biocompatibility in vivo

Surgery process

Surgical implantation was performed on three beagle dogs aged 1.5 years and weighing 11.4 ± 2.1 kg (mean \pm standard deviation). All procedures were conducted according to the ethical principles of the Peking University Institutional Animal Care and Use Committee. The sample implants were sterilized prior to surgery as described above for in vitro testing. General anesthesia was achieved using an intravenous injection of 1% pentobarbital 80 mg/kg. For every composite sample, two cylindrical implants were placed on each animal on the medial surface of each tibia in the proximal diaphyseal region. For each group, ie, PEEK and n-TiO₂/PEEK, there were six cylindrical implants placed. After surgery, three fluorochromes (Sigma, St Louis, MO), ie, calcein (100 mg/kg), calcein blue (100 mg/kg), and tetracycline (100 mg/kg) were administered to assess the osteogenic activity at weeks 1, 2, and 4, respectively. The dogs were sacrificed 4 weeks after surgery by an intracardiac injection of 10% kalium chloratum (0.5 mL/kg). The tibias were promptly fixed in 10% formalin and dehydrated in a graded series of alcohols.

Microcomputed tomography analysis

A scanner (1076, Skyscan) was utilized to evaluate all the tibias containing the implant at 40 kV (X-ray source voltage), 200 μ A (beam current), 900 msec (exposure time), 9 μ m (resolution), 0.8° (rotation step), and 180° (rotation angle). Using the CTan software package (Skyscan), percent bone volume was calculated choosing a serial polygonal region of interest in 100 slices with approximately 1 mm bonding to an implant, which represented the regenerated bone in the marrow only. Radiographs were also synthesized by the CTvol software package (Skyscan) to reconstruct three-dimensional regenerated bone of 0.5 mm width in the marrow bonding to the implants.

Histologic analysis

Following microcomputed tomographic analysis, ultrathin sections were obtained for histologic detection. In brief, the tibias were embedded in methyl methacrylate resin and sectioned after polymerization with a microtome (SP1600; Leica, Wetzlar, Germany) along the longitudinal axis of the cylindrical implants. The tissue sections were ground to a thickness of 30 μ m and detected using confocal laser scanning microscopy (LSM710 NLO; Zeiss, Oberkochen, Germany).

Statistical analysis

Data are presented as the mean \pm standard deviation ($n = 6$). Analysis of variance was utilized for calculation of the significance level of the data, and $P \leq 0.05$ was accepted as indicating statistical significance. All statistical analyses were carried out using OriginPro 8 SR3 software (OriginLab Corporation, Northampton, MA).

Results

Characterization of materials

TEM analysis of n-TiO₂/PEEK composite

Particle size and size distribution as well as dispersion play critical roles in the mechanical performance of nanocomposites as well as their biocompatibility and bioactivity. TEM imaging of our n-TiO₂/PEEK nanocomposite also showed relatively uniform dispersion of n-TiO₂ in PEEK (Figure 1).

SEM analysis of n-TiO₂/PEEK composite

In order to understand the effect of implant surface morphology on cell culture and other biologic responses of PEEK with and without n-TiO₂, SEM analysis was carried out on the smooth and rough surfaces of pure PEEK and the n-TiO₂/PEEK composite. n-TiO₂ particles were visualized on the smooth n-TiO₂/PEEK surface (Figure 2). A dramatic difference in surface morphology/topography was observed between the smooth and rough sample surface prepared for bioactivity evaluation.

Composite surface analysis by XPS

As discussed earlier, distribution of n-TiO₂ in the composite, particularly on the surface, has a significant impact on

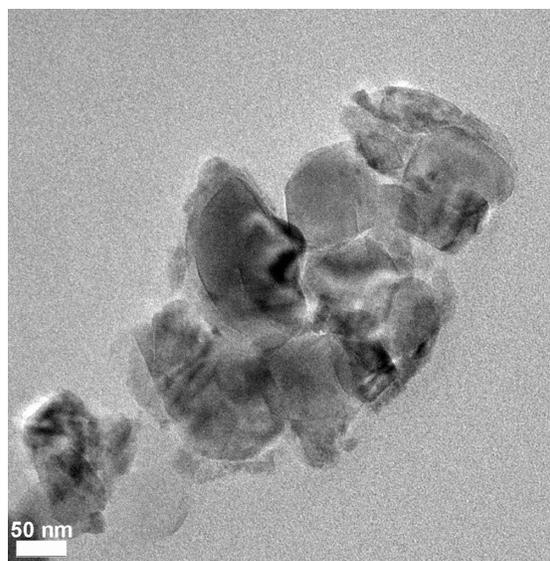


Figure 1 Transmission electron microscopy images of n-TiO₂ in TiO₂/poly(ether-ether-ketone) composite.
Note: Bar, 50 nm.

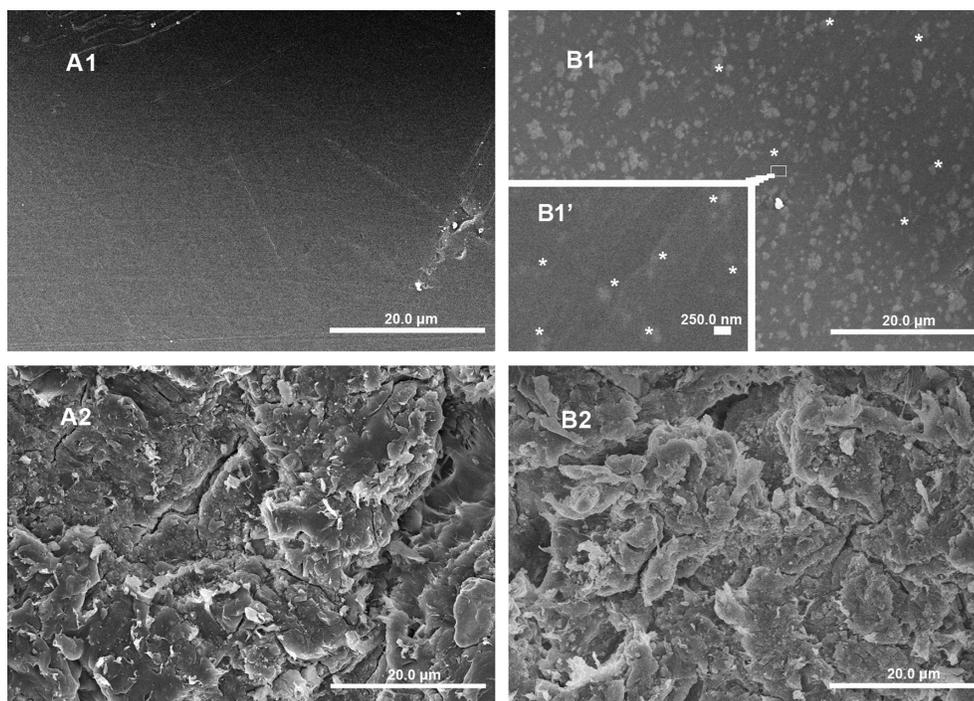


Figure 2 Scanning electron microscopy images of PEEK and n-TiO₂/PEEK before and after blasted treatment. **(A1)** Smooth PEEK, **(A2)** rough PEEK, **(B1)** smooth n-TiO₂/PEEK, and **(B2)** rough n-TiO₂/PEEK.

Notes: *n-TiO₂ particles exposed to the surface of smooth n-TiO₂/PEEK. **(A1, A2, B1, and B2)** bars, 20.0 μm; **B1** bar, 250.0 nm).

Abbreviation: PEEK, poly(ether-ether-ketone).

composite tomography as well as bioactivity. XPS analysis demonstrated the presence of Ti on the surface of the n-TiO₂/PEEK composite (Figure 3), indicating the presence of n-TiO₂ on the surface.

Cytocompatibility in vitro

Cell attachment

Cell attachment was measured by WST-1 testing after 4 hours of culture of the various samples, and the results

are shown in Figure 4. It is interesting to note that smooth PEEK showed the lowest optical density value (0.56 ± 0.07) among the groups ($P < 0.05$), while the rough n-TiO₂/PEEK group had a significantly higher optical density value (1.21 ± 0.34 , $P < 0.05$), indicating better cell attachment compared with the other groups. The study suggests the presence of n-TiO₂ in the composites, especially those with a rough surface, with marked attachment of cells to the implant surfaces.

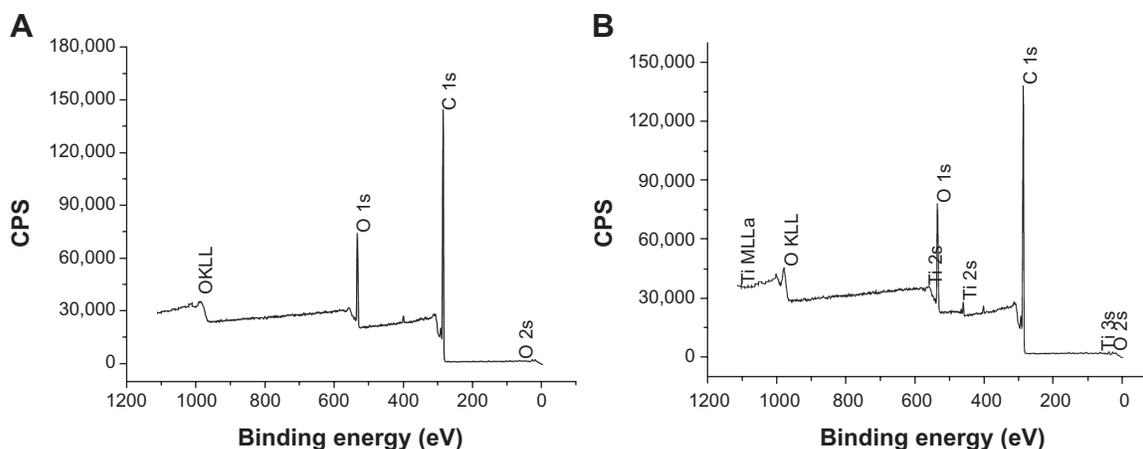


Figure 3 X-ray photoelectron spectroscopy of PEEK **(A)** and n-TiO₂/PEEK composite **(B)**.

Abbreviation: PEEK, poly(ether-ether-ketone).

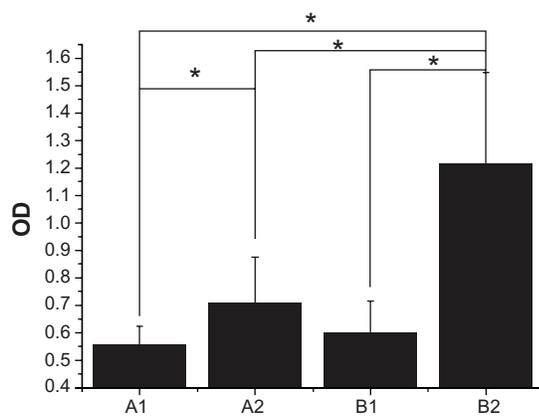


Figure 4 Cell attachment on PEEK and n-TiO₂/PEEK before and after blasted treatment. (A1) Smooth PEEK, (A2) rough PEEK, (B1) smooth n-TiO₂/PEEK, and (B2) rough n-TiO₂/PEEK.
Note: * $P < 0.05$.

Abbreviations: OD, optical density; PEEK, poly(ether-ether-ketone).

Cytotoxicity

The cytotoxicity of the various materials with smooth or rough surfaces was evaluated in terms of cell proliferation and viability at 3, 7, and 14 days. The relative growth rate in the four groups was more than 80% with cytotoxicity grade 1, suggesting a lack of severe cytotoxicity in both the short term (3 days) and long term (7 and 14 days, Figure 5). The smooth n-TiO₂/PEEK group showed significantly higher ($P < 0.05$) cell proliferation and viability in the short term (3 days), compared with rough PEEK. During the short culture period (3 days), the smooth PEEK showed higher cell proliferation and viability ($P < 0.05$) than the rough n-TiO₂/PEEK group.

Cell morphology

In general, cells in the rough groups seemed to spread more efficiently, with numerous pseudopods (Figure 6), while cells

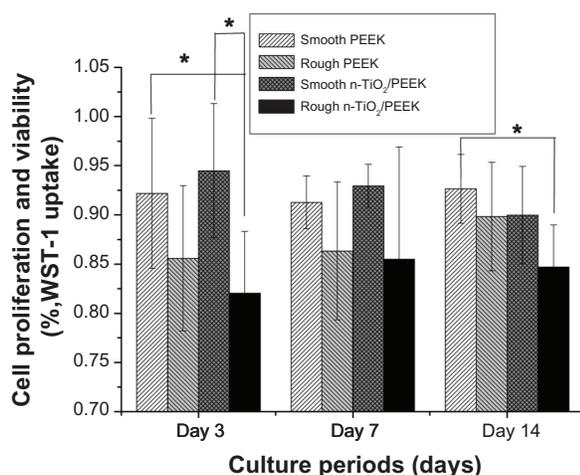


Figure 5 Cytotoxicity of the samples, smooth PEEK, rough PEEK, smooth n-TiO₂/PEEK and rough n-TiO₂/PEEK, after 3, 7, and 14 days as measured by WST-1 assay.
Note: * $P < 0.05$.

Abbreviation: PEEK, poly(ether-ether-ketone).

on the smooth n-TiO₂/PEEK exhibited better spreading than the smooth PEEK groups. Pseudopods seemed to prefer to anchor at the areas where n-TiO₂ was exposed. After long-term culture (7 and 14 days), no significant difference could be found among the groups.

Flow cytometric analysis

The cell cycle of the MG-63 cultures on the samples was further evaluated by measuring the DNA content of nuclei labeled with propidium iodide, and different cytodieresis phases (G0G1: pre-DNA synthesis of resting, S: DNA synthesis phase, G2M: post-DNA synthesis and mitosis) were analyzed. Figure 7 shows the total percentage of osteoblast MG-63 cells in the S and G2M phases for the four groups after the various culture periods (3, 7, and 14 days). For the short-term culture period (3 days), smooth n-TiO₂/PEEK showed a larger percentage of cells in the S and G2M phases compared with the rough group ($P < 0.05$), with no significant difference found in the other groups. For the long-term culture periods (7 and 14 days), although there was no significant difference found between rough PEEK and rough n-TiO₂/PEEK, these two groups showed a larger percentage than the other groups ($P < 0.05$).

Biocompatibility in vivo

Microcomputed tomography

Microcomputed tomography makes it possible to calculate not only the new bone bonding to the implants in terms of percent of bone volume/tissue volume but also to reconstruct three-dimensional images. As shown in Figure 8, the bone volume/tissue volume of n-TiO₂/PEEK was about twice as large as that of PEEK ($P < 0.05$). These results could be seen clearly in the three-dimensional reconstructions (Figure 9).

Histologic analysis

Fluorochrome bone marker labels (calcein, calcein blue, and tetracycline) were clearly observed for bone tissue bonding to the cylindrical implants. More bone deposition and remodeling was seen around the n-TiO₂/PEEK implants (Figure 10), suggesting a greater degree of bone regeneration than around the pure PEEK implants.

Discussion

While PEEK has exceptional biomechanical properties, there is a need to improve further its bioactivity for dental and orthopedic applications. The purpose of this study was to evaluate the in vitro and in vivo bioactivity of n-TiO₂/PEEK with different surface morphologies. n-TiO₂/PEEK composites were

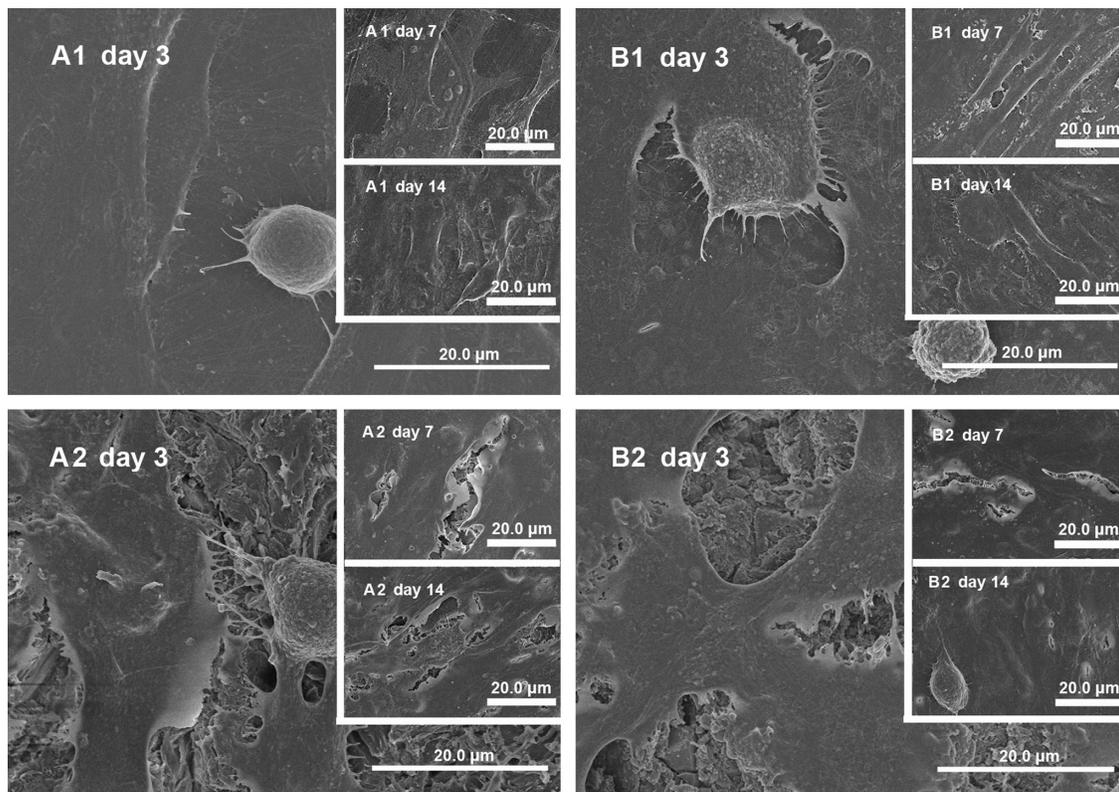


Figure 6 Cell morphology. (A1), (A2), (B1), and (B2) MG-63 cells cultured on disks of smooth PEEK, rough PEEK, smooth n-TiO₂/PEEK and rough n-TiO₂/PEEK for 3 days (A1, A2, B1 and B2), 7 days (A1, A2, B1 and B2) and 14 days (A1, A2, B1 and B2), respectively. **Note:** Bar 20.0 μm.

Abbreviation: PEEK, poly(ether-ether-ketone).

successfully fabricated. They did not cause severe cytotoxicity and there was no significant difference in nuclear DNA content compared with the other two materials. It was found that n-TiO₂ significantly improved osteoblast cell attachment, and cells on n-TiO₂/PEEK showed better spreading than on pure PEEK. These unique properties of n-TiO₂ significantly

enhanced new bone regeneration and bonding to the implants during in vivo testing.

In recent years, n-TiO₂ particles have attracted considerable attention, not only because of their unique physicochemical properties, especially in biomedical applications, but also because of their inflammatory, proliferative, cytotoxic, and

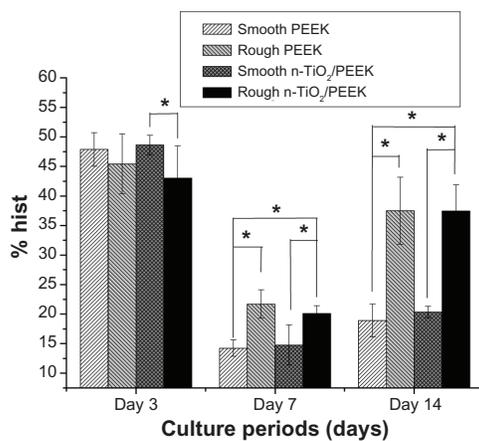


Figure 7 Cell cycle analysis of osteoblasts grown on smooth PEEK, rough PEEK, smooth n-TiO₂/PEEK, and rough n-TiO₂/PEEK for 3, 7, and 14 days.

Note: *P < 0.05.

Abbreviation: PEEK, poly(ether-ether-ketone).

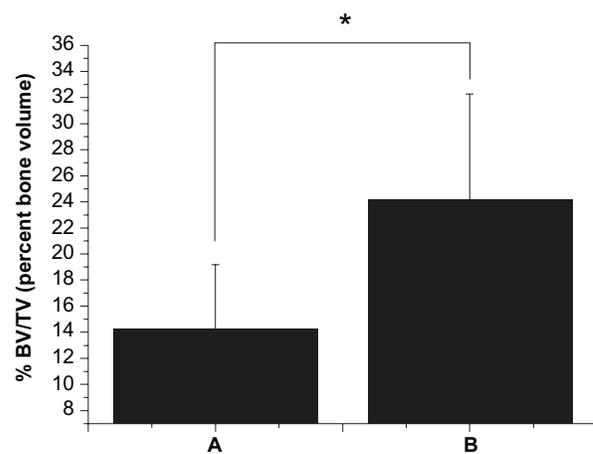


Figure 8 BV/TV of PEEK (A) and n-TiO₂/PEEK (B) was calculated after microcomputed tomography.

Note: *P < 0.05.

Abbreviations: BV/TV, percent bone volume; PEEK, poly(ether-ether-ketone).

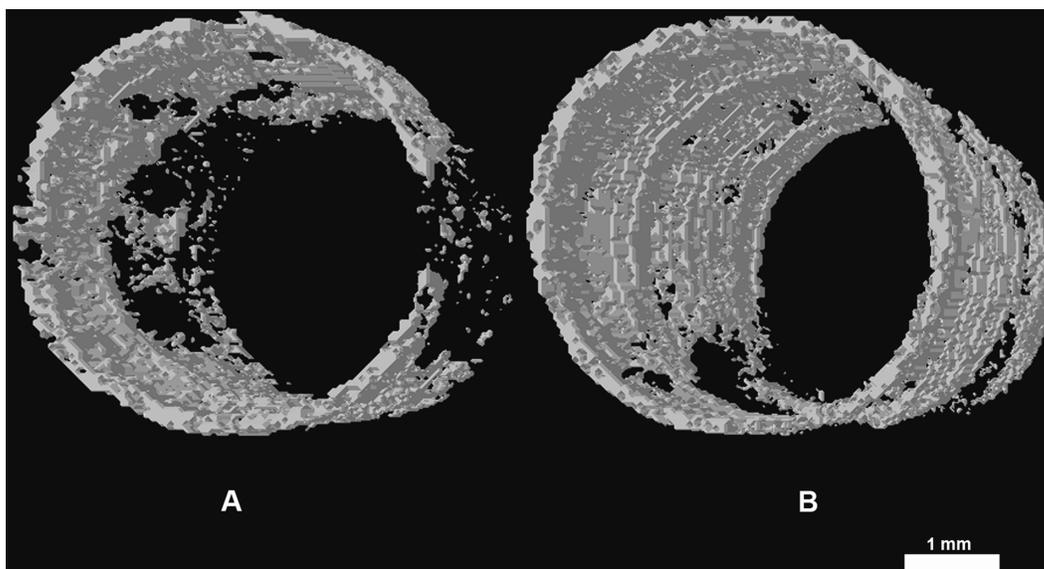


Figure 9 Three-dimensional images showing regeneration of microstructures about 0.5 mm in thickness bonding to PEEK (A) and n-TiO₂/PEEK (B) implants in the marrow. **Abbreviation:** PEEK, poly(ether-ether-ketone).

genotoxic effects. The chemistry and surface morphologic effect of the n-TiO₂ nanostructure also significantly enhanced the proliferation and differentiation of osteoblasts.^{1,21,22} A superthin (300 pm–6.3 nm) TiO₂ coating fabricated by slow sputter deposition of molten TiO₂ nanoparticles is sufficient to enhance biologic properties of microroughened titanium significantly, and at this thickness range it does not alter the previously created microtopography of the titanium surface.¹ On the other hand, some reports have shown that various cultured cell models show cytotoxicity when cultured with different doses of free TiO₂ nanoparticles,^{23–25} and animal models have show an inflammatory response and tumorigenesis when exposed to free TiO₂ nanoparticles.^{23,26,27} Free TiO₂ nanoparticles induce rapid phosphorylation in polymorphonuclear neutrophils, with a great potential to induce apoptosis.²⁴

Considering the aforementioned studies, we believe that the excellent biocompatibility of n-TiO₂ nanocomposites is mainly due to their exceptional bioactivity and the surface morphologic effects of the TiO₂ layer. Cytotoxicity, inflammatory response, or tumorigenesis are due to free TiO₂ nanoparticles which could break into the intracellular environment²³ and tissues from culture media or by administration via nasal administration, oral administration, or instillation.^{26,27} It is important to utilize the chemistry and surface morphologic effect of n-TiO₂ and to limit free n-TiO₂ particles when these particles are used to fabricate biomedical materials, especially orthopedic and dental implant materials.

In this study, a n-TiO₂/PEEK composite was fabricated successfully. n-TiO₂ could be visualized on the surface of n-TiO₂/PEEK composites (Figure 2) which showed appropriate homogeneity throughout (Figure 1). Elements of n-TiO₂ were detected on the composite surface by XPS (Figure 3). Therefore, the n-TiO₂/PEEK composite could also be considered as a superthin n-TiO₂ layer exposed on the PEEK surface in a safe way such that n-TiO₂ was bound into the PEEK matrix, with only some parts of each nanoparticle exposed to the surface. Several kinds of successfully fabricated PEEK nanocomposites have been reported, such as strontium-containing (Sr)-HA/PEEK, nanosized SiO₂/PEEK and nanosized Al₂O₃/PEEK.^{20,28} When reinforced with 0–30 vol% Sr-HA, the bending modulus of Sr-HA/PEEK composites increases with the volume ratio of Sr-HA (25 vol%, 9.6 GPa; 30 vol%, 10.6 GPa), while the bending strength of the composites decreases with addition of Sr-HA (25 vol%, 93.8 MPa; 30 vol%, 89.1 MPa).²⁰ The 25 vol% Sr-HA was chosen to produce the composites for in vitro testing, which was due to the desired mechanical performance. In this study, 40 wt% n-TiO₂ was chosen to fabricate the n-TiO₂/PEEK composite, which was based on an optimization between the bending modulus (3.8 GPa) and bending strength (93 MPa).

In vitro studies showed that n-TiO₂ did not cause severe cytotoxicity or disturb cell cycle progression, but improved the bioactivity of PEEK. Although the rough n-TiO₂/PEEK group showed lower cell proliferation and viability (Figure 5) than the other groups, n-TiO₂ did not cause severe cytotoxicity in the n-TiO₂/PEEK nanocomposite, and the relative

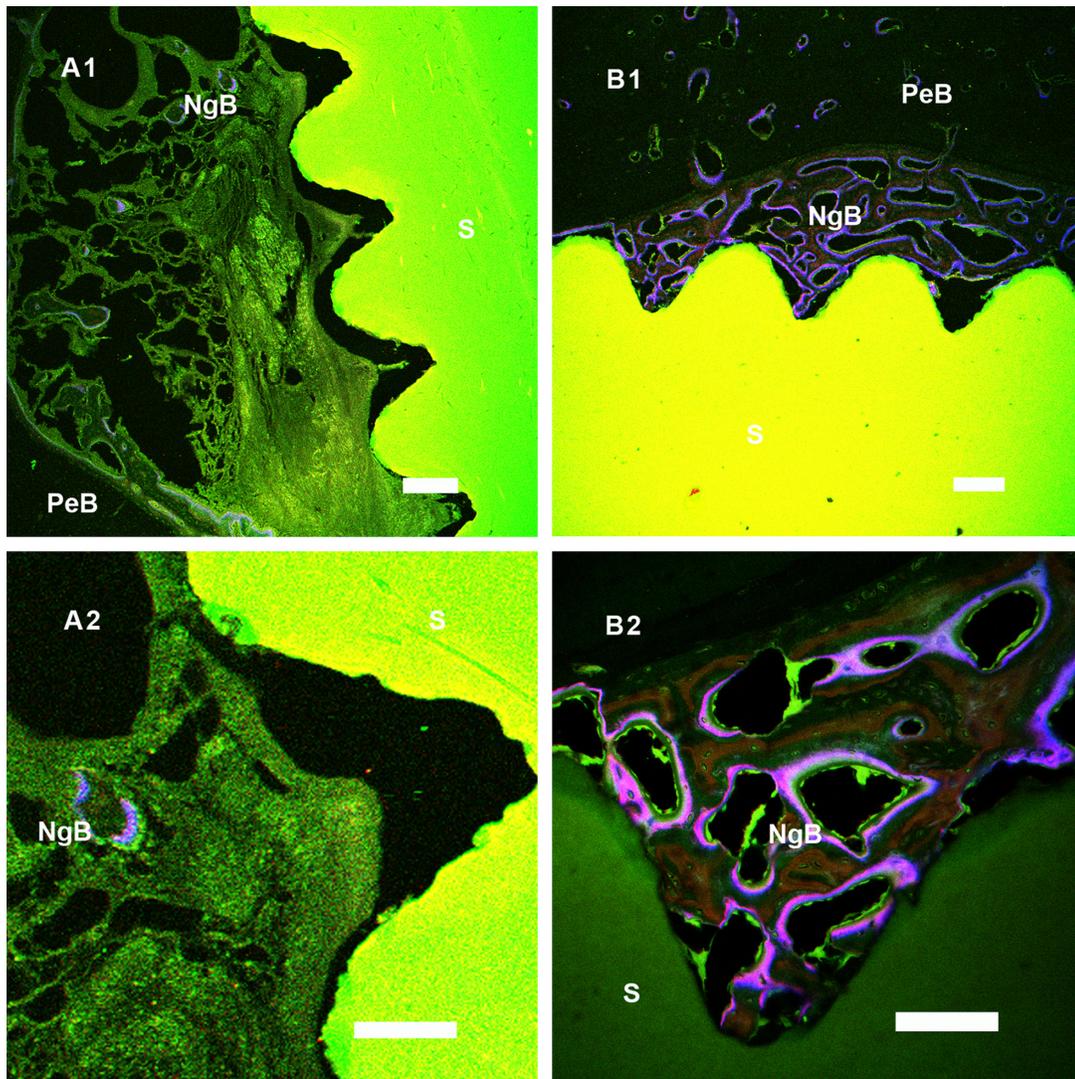


Figure 10 In histological analysis, new bone formation around the PEEK and n-TiO₂/PEEK were detected by bone labelling (calcein, calcein blue, and tetracycline). Group A was PEEK and group B was n-TiO₂/PEEK. Original magnification of (A1) and (B1) was 5× and (A2) and (B2) was 20×.

Notes: A1 and B1 bar, 200 μm; A2 and B2 bar, 100 μm.

Abbreviations: S, sample; NgB, newly grown bone deposition and remodeling zone; PeB, pre-existing bone tissue zone; PEEK, poly(ether-ether-ketone).

growth rate of the n-TiO₂/PEEK composites was more than 80%. Flow cytometric analysis (Figure 7) showed that n-TiO₂ did not disturb cell cycle progression compared with the pure PEEK groups. After long-term culture (for 7 and 14 days), osteoblast cells on both rough PEEK and n-TiO₂/PEEK surfaces contained a significantly larger portion in the S and G2M phases compared with cells on the smooth surface. This suggested that the rough-surfaced PEEK was beneficial to the proliferation of osteoblast cells. Other studies have reported that the rough surface of substrate materials improves proliferation and differentiation of osteoblasts.^{3,21,29,30}

Furthermore, in cell attachment testing (Figure 4), n-TiO₂ significantly improved cell attachment to n-TiO₂/PEEK compared with pure PEEK. This is consistent with the cell

morphology test (Figure 6). Cells on smooth n-TiO₂/PEEK appeared better spread, with numerous pseudopods. The pseudopods of osteoblasts on the smooth n-TiO₂/PEEK disks seemed to anchor exposed n-TiO₂ particles to the surface of the composite and formed a superthin coating on the PEEK surface, indicating that the surface morphologic effect of n-TiO₂ promotes the bioactivity of n-TiO₂/PEEK. A similar effect was observed in other studies using n-TiO₂. An n-TiO₂ superthin coated Ti surface significantly improves osteoblast cell attachment.¹ Ti coating on PEEK also enhances cell attachment, proliferation, and differentiation of osteoblasts.⁷ All of these studies suggest that the chemistry and surface morphologic effect of n-TiO₂ particles plays an important role in the exceptional bioactivity of n-TiO₂.

Our in vitro test results were consistent with those of in vivo testing. Microcomputed tomography is effective and reproducible for monitoring tumor-associated bone destruction,^{31,32} analyzing the three-dimensional structure of scaffolds,³³ measuring subtle differences in bone growth into three-dimensional scaffolds, and calculating new bone deposition around implants.^{34,35} In this study, the percent of bone volume/tissue volume of regenerated bone in the marrow bonding to the implants was calculated, and three-dimensional images were constructed. n-TiO₂/PEEK showed higher bone volume/tissue volume than pure PEEK (Figure 8). Three-dimensional (Figure 9) and histologic images (Figure 10) showed the same results, suggesting that n-TiO₂ enhances the bioactivity of n-TiO₂/PEEK. In terms of the potential risks of free n-TiO₂ particles, the novel bioactivity of n-TiO₂, such as chemistry and surface morphologic effect, is still attractive. It is important to utilize its outstanding bioactive properties and limit free n-TiO₂ when it is used in biomaterials, especially orthopedic and dental implants.

Conclusion

In this study, n-TiO₂/PEEK composites were fabricated and their in vitro and in vivo bioactivity was assessed. In in vitro testing, instead of causing cytotoxicity or disturbing cell cycle progression, n-TiO₂ improved the bioactivity of PEEK. It promoted cell attachment and improved osteoblast cell spreading. In the in vivo studies, enhanced bone regeneration around the implants by n-TiO₂ was indicated by higher bone volume/tissue volume and vivid visualization in three-dimensional and histologic images.

A special effect of n-TiO₂ on the composite surface was also observed. Bioactivity evaluation of the nanocomposites revealed that pseudopods of osteoblasts preferred to anchor at areas where n-TiO₂ presented at the surface. This study also showed that the surface roughness of n-TiO₂/PEEK composites played a prominent role in promoting cell attachment and regeneration of new bones. Our study suggests that n-TiO₂/PEEK composites utilize the attractive bioactivity of n-TiO₂ as well as the outstanding mechanical properties of PEEK, and could be a potential substitute for metal implant material in orthopedic and dental applications.

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Disclosure

The authors report no conflicts of interest in this work.

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