

Development of a controlled-release anti-parkinsonian nanodelivery system using levodopa as the active agent

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Abstract: A new layered organic–inorganic nanocomposite material with an anti-parkinsonian active compound, L-3-(3,4-dihydroxyphenyl) alanine (levodopa), intercalated into the inorganic interlayers of a Zn/Al-layered double hydroxide (LDH) was synthesized using a direct coprecipitation method. The resulting nanocomposite was composed of the organic moiety, levodopa, sandwiched between Zn/Al-LDH inorganic interlayers. The basal spacing of the resulting nanocomposite was 10.9 Å. The estimated loading of levodopa in the nanocomposite was approximately 16% (w/w). A Fourier transform infrared study showed that the absorption bands of the nanocomposite were characteristic of both levodopa and Zn/Al-LDH, which further confirmed intercalation, and that the intercalated organic moiety in the nanocomposite was more thermally stable than free levodopa. The resulting nanocomposite showed sustained-release properties, so can be used in a controlled-release formulation. Cytotoxicity analysis using an MTT assay also showed increased cell viability of 3T3 cells exposed to the newly synthesized nanocomposite compared with those exposed to pure levodopa after 72 hours of exposure.

Keywords: levodopa, layered double hydroxide, coprecipitation, sustained release

Introduction

Levodopa has remained the gold standard symptomatic replacement therapy for Parkinsonism, a progressive neurodegenerative disease, over the past 40 years. However, long-term treatment is often complicated by dyskinesia, a form of motor dysfunction, which is attributed to the discontinuous and intermittent delivery of levodopa to the brain, resulting in nonphysiological pulsatile stimulation of striatal dopamine receptors.^{1,2} Dyskinesia is observed in 40% of patients with Parkinson disease after 5 years (the honeymoon period) of levodopa use and in over 80% of patients after 10 years of using levodopa.²

Tailored nanoparticles are good drug delivery agents because of their sustained-release and controlled-release properties. It is widely believed that reducing the pulsatile stimulation of dopaminergic neurons will reduce the risk of levodopa-induced dyskinesia.²

Use of nanotechnology in the medical science field and in nanomedicine has reached an advanced stage, where much research has been conducted on the synthesis of nanoparticles that can be used as delivery tools for anticancer agents, central nervous system tumor agents,^{3,4} antibiotics,⁵ and nonsteroidal anti-inflammatory drugs⁶ in gene delivery and imaging studies.^{7,8}

Layered double hydroxides (LDHs), also known as hydrotalcite-like compounds or anionic clays, are a broad class of inorganic lamellar compounds with

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a high capacity for anion intercalation. The chemical composition of LDHs is expressed by the general formula $[M_{1-x}^{2+}M_x^{3+}(\text{OH})_2]^{x+}(\text{A}^{n-})_{x/n} \cdot m\text{H}_2\text{O}$, where M^{2+} and M^{3+} are divalent and trivalent metal cations, respectively. These hydroxides are derived from brucite $[\text{Mg}(\text{OH})_2]$, and they can be found in nature or be synthesized in a laboratory via coprecipitation method. The interlayer region contains various amounts of water, which is hydrogen-bonded to the hydroxide layers and/or to the interlayer anions.⁹ A characteristic feature of LDH is weak bonding between the interlayer anions and hydroxide sheets that enables exchange of anions.¹⁰

LDHs are less toxic than other conventional drug carriers and are generally biocompatible, making them an acceptable alternative for drug delivery.¹¹ Anionic drugs, such as levodopa, which replace the interlayer anions lying between the two positive metal hydroxide sheets, enable a controllable ion-exchange mechanism. pH dependency and the ability for controlled-release are other advantages of using LDHs as drug delivery systems. Further, the positively charged outer layer of the nanohybrid delivery system, which can attract a negatively charged cell membrane, enables easy penetration of LDH into cells. This type of research has not as yet focused on neurodegenerative diseases. Hence, the aim of this study was to intercalate levodopa into LDH for the treatment of Parkinson's disease.

Materials and methods

Materials

L-3-(3,4-dihydroxyphenyl) alanine (levodopa, 99% purity) was purchased from Sigma-Aldrich (St Louis, MO, USA). Other chemicals, including $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ and $\text{Al}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$, were of analytical grade and used without further characterization. Deionized water was used throughout the experiment, unless stated otherwise.

Synthesis

Zn/Al-LDH (the nanolayer) was synthesized in nitrate form by adding a solution of sodium hydroxide (1 molar) dropwise to a solution of zinc and aluminum nitrate, with a molar ratio of 2:1, in deionized water under a nitrogen atmosphere while vigorously stirring until a pH of 7.0 was reached. The mixture was aged in an oil bath for 18 hours at 70°C. The white precipitate obtained was centrifuged, washed three times with deionized water, and dried in an oven overnight at 70°C. The resulting Zn/Al-LDH was used in a cytotoxicity study and compared with its nanocomposite and levodopa.

Levodopa-Zn/Al-LDH (the nanocomposite) was synthesized using a direct coprecipitation method. In brief, a solution of levodopa (0.08 molar) was added to a $\text{Zn}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$ and $\text{Al}(\text{NO}_3)_3 \cdot 9\text{H}_2\text{O}$ solutions, at a ratio of 2:1, under constant stirring in the presence of a sustained nitrogen supply at room temperature, and the pH was adjusted to 7.0 using 1.0 molar NaOH. The experiment was protected from direct sunlight exposure because of the sensitivity of levodopa to light. The mixture was aged at 70°C in an oil bath for 18 hours and then centrifuged, filtered, washed with deionized water three times, and dried in an oven overnight.

Controlled-release study

Drug release profiles were determined at room temperature using phosphate-buffered saline at pH 4.8 and 7.4. Approximately 300 mg of the nanocomposite was added to 500 mL of the medium. The accumulated amount of levodopa released into the solution was measured at preset time intervals and at $\lambda_{\text{max}} = 280$ nm using an ultraviolet-visible spectrophotometer (Lambda 35, Perkin-Elmer, Boston, MA, USA).

Cell culture

3T3 cells (fibroblasts) were purchased from the American Type Culture Collection ((ATCC), Manassas, VA, USA), cultured in RPMI 1640 medium supplemented with 10% fetal bovine serum, L-glutamine 15 mmol/L, penicillin 100 U/mL, and streptomycin 100 µg/mL, and were then incubated in 5% CO_2 at 37°C. Cells at 80%–90% confluence were used for seeding and treatment.

Seeding and treatment

For the cytotoxicity study, cells were seeded in a 96-well plate at a density of 0.5×10^5 cells/mL (100 µL/well) and allowed 24 hours to attach before treatment. A stock solution containing 10 mg/mL of the nanocomposite, the nanohybrid, and levodopa in phosphate-buffered saline was made, diluted in RPMI 1640 complete medium to the desired concentration, and subjected to sonication before being added to the cells. Cell treatment was conducted using a series of dilutions ranging from 150 µg/mL to 0 µg/mL (the control), and readings were taken 72 hours after treatment using a microplate reader (EL 800x, Bio-Tek Instruments Inc, Winooski, VT, USA) after adding MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide). This assay is dependent upon reduction of the tetrazolium salt by mitochondrial dehydrogenase in viable cells, which forms a blue formazan product (Mosmann 1983).¹² In brief, 20 µL of MTT solution (5 mg/mL in phosphate-buffered saline) was added to each

well, and the plates were kept in an incubator in 5% CO₂ at 37°C for 3 hours and centrifuged at 1000 rpm for 5 minutes. The supernatant was discarded, and 100 μL of dimethyl sulfoxide was added per well. After keeping this mixture in the dark at room temperature for 30 minutes, absorbance was measured at a wavelength of 570 nm. Experiments were conducted in triplicate, and the cytotoxicity was calculated as:

$$\text{Cell viability (\%)} = [\text{Average}] \text{ test} / [\text{Average}] \text{ control} \times 100.$$

Characterization

Powder X-ray diffraction patterns were recorded in the 2°–60° range on a diffractometer (XRD-6000, Shimadzu, Tokyo, Japan) using CuK_α radiation ($\lambda = 1.5418 \text{ \AA}$) at 30 kV and 30 mA, with a dwell time of 4 degrees per minute. The Fourier transform infrared spectra of the materials were recorded at 400–4000 cm⁻¹ using a Thermo Nicolet Nexus FTIR (model Smart Orbit) (International Equipment Trading Ltd. Vernon Hills, IL, USA). The chemical compositions of the samples were analyzed for zinc and aluminum ion content using inductively coupled plasma atomic emission spectrometry (Optima 2000DV, Perkin-Elmer) under standard conditions. An elemental analyzer was used for C, H, N, and S analyses (CHNS-932, LECO Corporation, Saint Joseph, MI, USA). Thermogravimetric and differential thermogravimetric analyses (Mettler Toledo, Columbus, OH, USA) were carried out at a heating rate of 10°C per minute from 20° to 1000°C under a nitrogen atmosphere (N₂ flow rate of 50 mL per minute). A field emission scanning electron microscope (Nova™ NanoSEM 230, FEI Company, Hillsboro, OR, USA) was used to determine the surface morphology of the samples.

Results and discussion

Powder X-ray diffraction

The X-ray diffraction patterns of levodopa intercalated into Zn/Al-LDH and of free levodopa are shown in Figure 1A and B, respectively. As shown in Figure 1A, the (003), (006), and (009) reflections were found at 2θ values of approximately 8.06°, 16.04°, and 24.52°, respectively. As reported in the literature, the (003) reflection for the precursor Zn/Al-NO₃ LDH appears at 9.98°, indicating a d spacing of 8.90 Å.¹³ The shifting of this reflection toward low 2θ (2θ = 8.06°, corresponding to a d spacing of 10.9 Å) for the nanocomposite indicates intercalation of levodopa into the interlayer galleries of the LDH. The characteristic reflections of levodopa, shown in Figure 1B, were absent from the X-ray diffraction pattern of the nanocomposite, suggesting that levodopa was intercalated into the interlayer

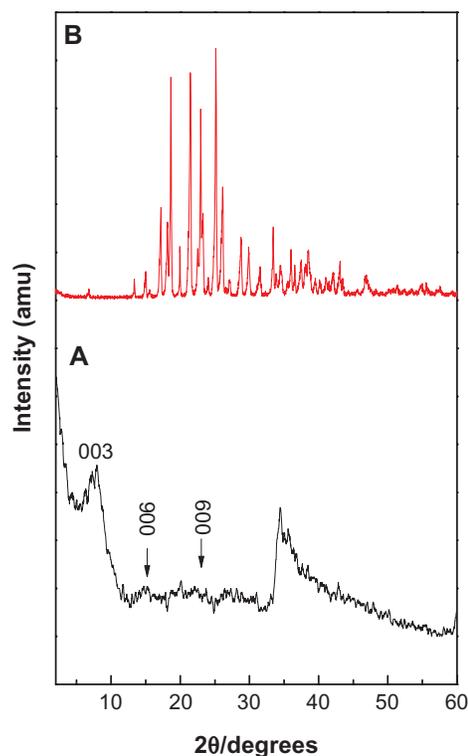


Figure 1 Powder X-ray diffraction patterns prepared using the coprecipitation method for nanocomposite (A) and levodopa (B).

galleries rather than absorbed at the surface. The broader (003) reflection peak in Figure 1A might have been caused by the formation of a secondary phase due to intercalation of the counter ion (NO₃⁻).

Spatial orientation of intercalated levodopa

Figure 2A shows the three-dimensional molecular size of levodopa, which was obtained using ChemOffice 2008 software (Cambridge, MA, USA). The long and short axes (x axis and y axis, respectively) and molecular thickness (z axis) of levodopa were calculated, giving values of 10.4 Å, 7.1 Å, and 6.2 Å, respectively. The X-ray diffraction pattern shows a basal spacing (d) of 10.9 Å for the nanocomposite. The thickness of the Zn/Al-NO₃ LDH layer was 4.8 Å.¹⁴ Therefore, the gallery height of LDH after the intercalation processes was 6.1 Å (10.9–4.8 Å). This gallery height of the nanocomposite, 6.1 Å, is similar to the thickness axis of levodopa (6.2 Å). This result indicates that the levodopa anions are accommodated as an alternate monolayer (see Figure 2B).

Infrared spectroscopy

The Fourier transform infrared spectra for pristine levodopa and levodopa intercalated into the Zn/Al interlayer are shown in Figure 3. The spectrum for levodopa (Figure 3A) shows many intense, sharp absorption peaks that arise from the

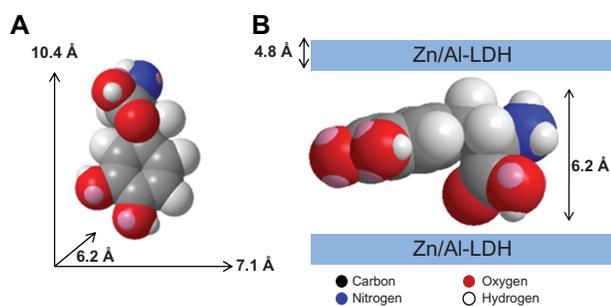


Figure 2 Three-dimensional structure of levodopa (A) and molecular structure model of levodopa intercalated between interlayers of Zn/Al-layered double hydroxide (B).

Abbreviation: LDH, layered double hydroxide.

different functional groups present in the molecules, ie, the primary amine, carboxylic acid, benzene ring, and hydroxyl groups.

The carboxylic group for levodopa shows three absorption peaks: 1245 cm^{-1} for the C–O stretching, 1739 cm^{-1} for the C=O bond, and a broad band at 1400–1440 cm^{-1} for the O–H bending vibration. The bands at 3353, 3189, and 3035 cm^{-1} can be attributed to the $\nu(\text{N–H})$, $\nu(\text{O–H})$, and $\nu(\text{Ar–H})$ vibrations, respectively. The primary amine shows absorption peaks at 1651 cm^{-1} and 1563 cm^{-1} that can be attributed to N–H bending, while the peaks between 1064 cm^{-1} and 1200 cm^{-1} are due to C–N stretching, and the peak at approximately 800 cm^{-1} indicates N–H (oop) bending. The band at 1496 cm^{-1} can be attributed to the C=C bond in the benzene ring.

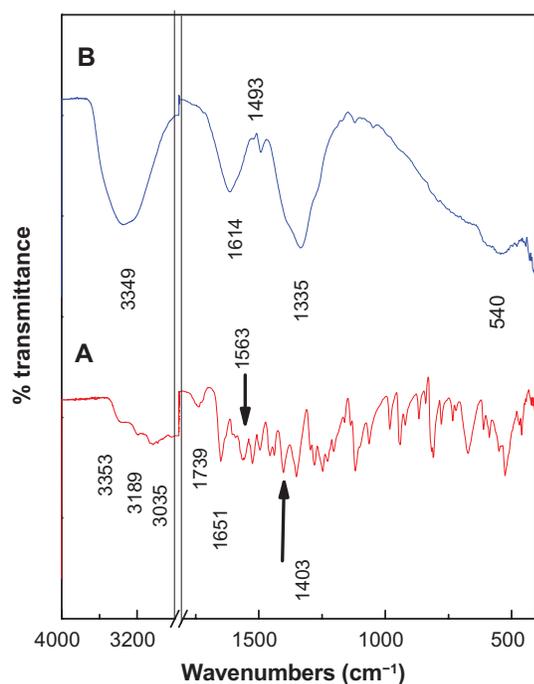


Figure 3 Fourier transform infrared spectra of free levodopa (A) and levodopa-Zn/Al-layered double hydroxide nanocomposite (B).

The Fourier transform infrared spectrum of the levodopa intercalated into Zn/Al-LDH is shown in Figure 3B. The spectrum of the nanocomposite shows the characteristic bands of pure levodopa, which indicate that the levodopa anions were intercalated into the interlayer galleries of LDH. Some of the bands are slightly shifted. For example, the position of the band for the C=C bond shifted from 1496 cm^{-1} for free levodopa to 1493 cm^{-1} for the nanocomposite, and the band at 1118 cm^{-1} for the C–N bond shifted to 1120 cm^{-1} ; these shifts are due to the interaction between the levodopa anion and the interlayers as a result of the intercalation process. The appearance of a new broad peak at 1614 cm^{-1} for the nanocomposite can be assigned to the asymmetric stretching vibration of the COO^- group.¹⁴ The band at 1335 cm^{-1} is due to the nitrate anion, which may not be completely removed from the interlayers during the intercalation process.¹⁵

Elemental analysis

Elemental (CHNS) and inductively coupled plasma analyses were conducted to determine the organic and inorganic compositions of the nanocomposite. As expected, the levodopa-nanocomposite contained both organic and inorganic constituents. This result indicates that the levodopa was incorporated into the Zn/Al-LDH inorganic interlayers.

The percentage loading of levodopa in the nanocomposite is 16.00%, as shown in Table 1. The C/N ratio for free levodopa is 8.09, and this value is higher than the C/N ratio of the nanocomposite, possibly indicating that the nitrate anion was not completely removed from the interlayers during the intercalation process. This result was confirmed by the presence of a very broad peak at low 2θ in the X-ray diffraction pattern. From the elemental chemical analysis and thermogravimetric studies, the empirical formula for the nanocomposite can be proposed as: $[\text{Zn}_{0.64}\text{Al}_{0.36}(\text{OH})_2](\text{levodopa})_{0.09}(\text{NO}_3)_{0.27} \cdot 3.03\text{H}_2\text{O}$.

Thermal analysis

The thermal behavior of the compounds before and after intercalation of levodopa into Zn/Al-LDH was examined using thermogravimetric and derivative thermogravimetric analysis. The thermograms of levodopa and the nanocomposite are shown in Figure 4. The free levodopa (Figure 4A) thermal decomposition exhibited two weight loss events above 270°C. The first sharp event occurred at 288°C with a weight loss of 42.0%, which was due to the decomposition of levodopa. The second weight loss at 381°C–666°C can be attributed to the combustion of levodopa.

After the intercalation process, the thermal decomposition characteristics of the resulting product were different from

Table 1 Elemental chemical compositional for free levodopa and its nanocomposite

Sample	Zn% ^a	Al% ^a	C% ^b	N% ^b	C/N	Drug% ^b	Zn ²⁺ /Al ³⁺	X
Levodopa	–	–	55.69	6.88	8.09	–	–	–
Nanocomposite	16.44	3.85	8.56	4.62	1.85	16.00	1.80	0.36

Notes: ^aCalculated from inductively coupled plasma data; ^bcalculated from CHNS elemental data.

Abbreviations: Zn, zinc; Al, aluminium; C, carbon; N, nitrogen; X, aluminium mole fraction = mole of Al/(mole of Al + mole of Zn).

those of the precursor. The thermal decomposition of the nanocomposite was characterized by two weight loss events, one at 35°C–162°C with a weight loss of 31.0%, due to the removal of the external surface-adsorbed and interlayer water molecules, and a second event at 162°C–960°C with a weight loss of 30.0%. This second weight loss was due to dehydroxylation of the layers and decomposition of levodopa.

Surface properties

In Figure 5, the surface morphologies of the Zn/Al-nanolayer (A and B) and levodopa-nanocomposite (C) are shown. The micrographs were obtained using field emission scanning electron microscopy at (A) 8000×, (B) 15,000×, and (C) 30,000× magnifications. Zn/Al-LDH and the nanocomposite show nonuniform, irregular agglomerates with compact and nonporous plate-like structures, which are commonly observed for LDH and its nanocomposites.

Release behavior of levodopa from the Zn/Al-levodopa nanocomposite

Typical release behavior profiles of levodopa from its nanocomposite at different pHs are shown in Figure 6, indicating that the release rate of levodopa from the nanocomposite is pH-dependent. The release rate at pH 7.4 is remarkably lower than that at pH 4.8. The percentage release of levodopa from the nanocomposite reaches approximately 74% within 2400 minutes when exposed to a pH of 4.8. When the pH is changed to 7.4, the release rate of levodopa from the

nanocomposite is obviously lower. The time for 76% of the drug to be released from the nanocomposite is approximately 8600 minutes. This lower release rate of levodopa from the nanocomposite at pH 7.4 indicates that the levodopa-Zn/Al nanocomposite is indeed a potential drug delivery system. The difference in the release rates at pH 4.8 and pH 7.4 may be due to the difference in the release mechanism of levodopa from the nanocomposite.¹⁶ At an acidic/lower pH of 4.8, Zn/Al-LDH begins to dissolve. At pH 7.4, the Zn/Al-LDH is more stable because of electrostatic interaction between the levodopa anions and the positively charged Zn/Al-LDH layers. Thus, the release mechanism at pH 4.8 should occur through both dissolution of the LDH layers and ion exchange, while the mechanism at pH 7.4 should occur via ion exchange with the ions in the buffer solution.¹⁶

Release kinetics of levodopa from the Zn/Al-levodopa nanocomposite

To investigate the exact mechanism of levodopa release from the Zn/Al-LDH interlayer, the release profile data were fitted using mathematical models that describe various kinetics. The pseudo-first order kinetics mathematical model equation $\ln(q_e - q_t) = \ln q_e - k_1 t$ (q_e and q_t are the equilibrium release amount and the release amount at time t , respectively),¹⁷ the pseudo-second order equation $t/q_t = 1/k_2 q_e^2 + t/q_e$ ¹⁸ and the parabolic diffusion equation $(1 - M_t/M_0)/t = kt^{-0.5} + b$ (M_0 and M_t are the drug content that remained in LDH at the release times of 0 and t , respectively)¹⁹ were used.

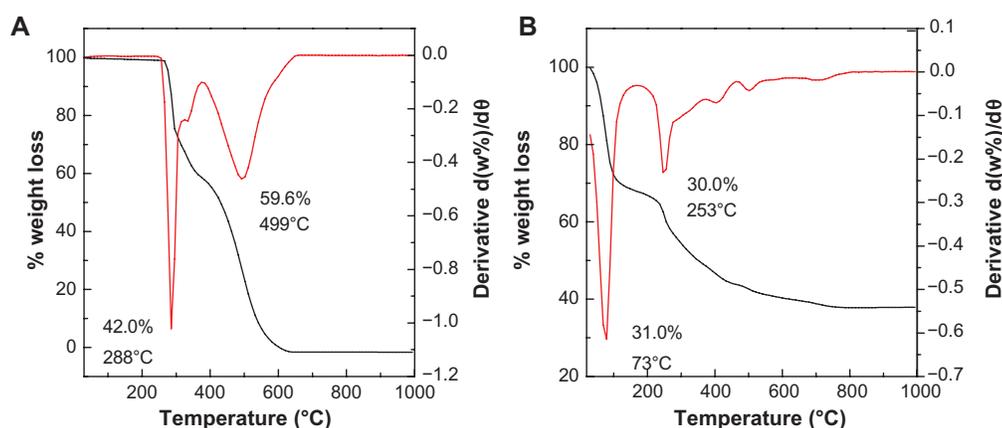


Figure 4 Thermogravimetric and differential thermogravimetric thermograms of levodopa (A) and levodopa-nanocomposite (B).

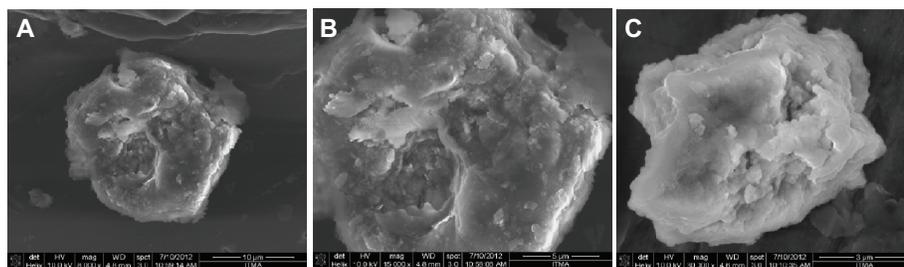


Figure 5 Field emission scanning electron micrographs of Zn/Al-layered double hydroxide (A and B), and of the Zn/Al-layered double hydroxide-levodopa nanocomposite (C).

The fitting results of the drug release profiles at pH 7.4 and 4.8 based on the three different kinetic models are shown in Figure 7. The correlation coefficient (R^2) is tabulated in Table 2. From Table 2, it can be seen that the release of levodopa from Zn/Al-LDH follows the pseudo-second order equation very well for both pH values, with satisfactory coefficients of 0.9988 (pH 7.4) and 0.9952 (pH 4.8). The rate constant for the pseudo-second order equation is 2.85×10^{-5} mg per minute and 4.29×10^{-5} mg per minute at pH 7.4 and pH 4.8, respectively (Table 2). The kinetic results obtained during this work are very similar to those from a kinetic study of the release of camptothecin from Mg/Al-LDH¹⁶ and are also similar to those for perindopril erbumine intercalated into Zn/Al-LDH using the ion-exchange and coprecipitation methods.²⁰

Cytotoxicity analysis of the nanocomposite, nanohybrid, and levodopa

Compared with the control, cell viability was found to decrease with increasing concentrations of levodopa, the nanolayer, and the nanocomposite. The 3T3 cell viability was reduced by approximately 40% when the concentration of levodopa was more than 50 $\mu\text{g/mL}$. For the nanolayer

alone, cell viability was reduced by more than 40% when the concentration of the hybrid exceeded 100 $\mu\text{g/mL}$. We observed that the levodopa nanocomposite reached a higher cell viability compared with the carrier nanolayer or with the drug alone in a dose-dependent manner after 72 hours of exposure, as shown in Figure 8.

In vitro, cytotoxicity testing is an essential step in analyzing the health hazards of synthesized nanomaterials in terms of the cellular response to a toxicant for a biomedical application. Therefore, in the present study, we investigated the cytotoxic effect of the synthesized nanocomposite, nanohybrid, and pristine levodopa doses using a biological model with 3T3 fibroblast cells and an MTT assay.

It was found that exposure to the nanocomposite at a dose level of 5–150 $\mu\text{g/mL}$ caused dose-dependent cytotoxicity in 3T3 cells, as shown in Figure 8, but the same dose of pristine levodopa but more cytotoxicity after 72 hours of exposure compared with the nanocomposite. Findings by other researchers have demonstrated a similar pattern of toxicity in primary rat hepatocytes after exposure to allergic acid and its nanocomposite, ie, the allergic acid caused more toxicity compared with its nanocomposite using the same dose range after 6 hours of exposure.²¹

Our results also demonstrated more toxicity in 3T3 cells following exposure to the nanolayer compared with exposure to the nanocomposite at the same doses after 72 hours. In previously reported studies, a decrease in the viability of human umbilical vein endothelial cells was observed following exposure to an LDH nanolayer compared with that following exposure to its nanocomposite, which was functionalized by poly(sulfobetaine).²² Meanwhile, in another report, Chang liver cells exposed to an LDH nanolayer also exhibited a slight decrease in viability compared with those exposed to its nanocomposite containing cetirizine.²³ Our findings are supported by the above-mentioned studies using nanolayers and their corresponding nanocomposites. The results also indicate that our nanocomposite is likely to be a biocompatible product because it shows less cytotoxicity than pristine

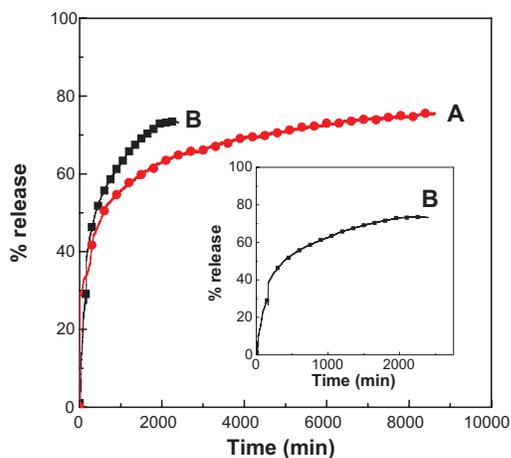


Figure 6 Release profiles of levodopa from the nanocomposite at pH 7.4 (A) and pH 4.8 (B).

Note: Inset shows the release profiles of levodopa from the nanocomposite at pH 4.8 from 0 to 2000 minutes.

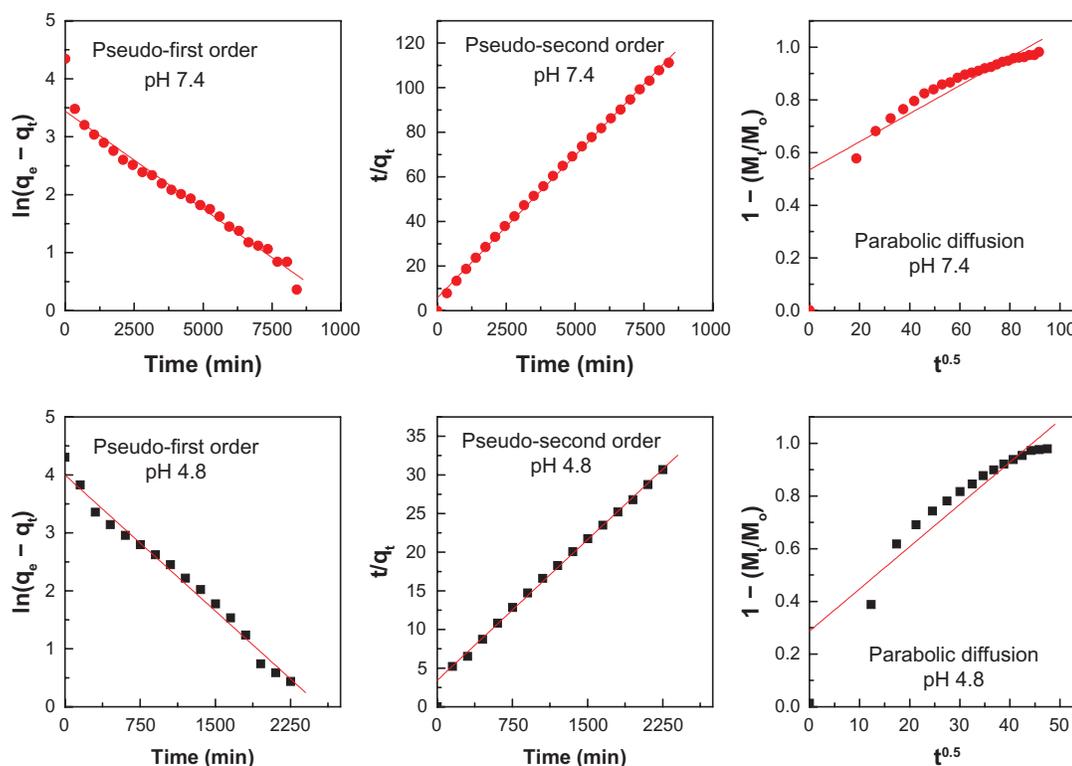


Figure 7 Fits of the levodopa release data for the nanocomposite to the pseudo-first and pseudo-second order kinetics models as well as to the parabolic diffusion model at pH 7.4 and 4.8.

Table 2 Correlation coefficients (R^2), and rate constants (k) obtained by fitting the levodopa release data for the nanocomposite in solutions at pH 4.8 and 7.4

pH	Saturation release (%)	R^2			Pseudo-second order
		Pseudo-first order	Pseudo-second order	Parabolic diffusion model	Rate constant k (mg/min) $\times 10^{-5}$
7.4	76%	0.9839	0.9988	0.8682	2.85
4.8	74%	0.9845	0.9952	0.8838	4.29

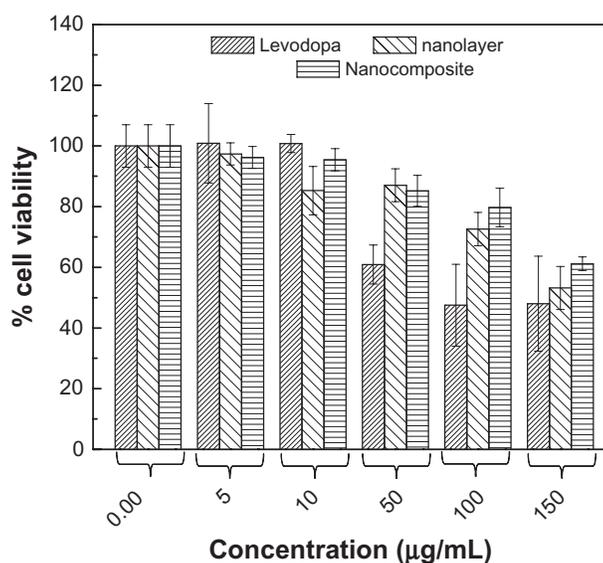


Figure 8 In vitro cytotoxicity study of 3T3 cells after 72 hours of exposure to free levodopa, the nanolayer, and the levodopa nanocomposite.
Notes: Using the graph as well as regression analysis, the IC_{50} (concentration of the drug required to induce 50% cell mortality) for the nanocomposite, nanolayer, and levodopa was $202 \pm 3.55 \mu\text{g/mL}$, $161 \pm 6.42 \mu\text{g/mL}$, and $73 \pm 6.46 \mu\text{g/mL}$, respectively ($P < 0.005$ for all three, showing statistical significance compared with the control).

levodopa, which is very important for the potential use of the nanocomposite in an in vivo study.

Conclusion

Levodopa-Zn/Al-LDH nanocomposites were synthesized using the coprecipitation method with a basal spacing of 10.9 Å and 16.0% (w/w) loading of levodopa. The intercalation of levodopa into the interlayer spacing of LDH is shown by Fourier transform infrared spectra. The intercalated guest molecules of levodopa were found to be arranged in a monolayer manner between the inorganic interlayers. Release of levodopa from the nanocomposite was found to be governed by pseudo-second order kinetics, and the release time of levodopa from the nanocomposite at pH 7.4 was longer than that at pH 4.8. The cytotoxicity study also demonstrated a decrease in the toxicity potential of levodopa in a normal cell line following its intercalation into Zn/Al-LDH. Hence, successful intercalation of levodopa may improve drug delivery systems for the treatment of parkinsonism as well as decrease its potential toxicity to cells.

Acknowledgment

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Disclosure

The authors report no conflicts of interest in this work.

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