Chlamydia pneumoniae in atheroma: consideration of criteria for causality

Allan Shor, James I Phillips, Gloria Ong, Brenda J Thomas, David Taylor-Robinson

Abstract

Aims—(1) To seek evidence of the existence of *Chlamydia pneumoniae* in a spectrum of atheromatous lesions in different types of arteries from individuals of different ages, using a polymerase chain reaction (PCR) assay supported by electron microscopy and immunocytochemistry; (2) to use electron microscopy to examine interactions between *C pneumoniae* and the cells present in the arterial tissue; (3) to assess the extent to which the data fulfil the criteria for causality.

Methods—At necropsy examination, 35 arterial specimens were taken from 25 subjects. The grade of atheroma was determined macroscopically and microscopically and the tissues coded and examined by the three techniques.

Results—Of the 35 specimens, 24 had macroscopic or microscopic atheromatous lesions of varying degree. Twenty two of the 35 specimens were examined by electron microscopy, *C pneumoniae*-like bodies being found in 11 (50%); seven specimens were examined by the immunocytochemical method, positive staining being detected in three; and all specimens were examined by the PCR technique, 15 (43%) being PCR positive. Overall, of the 24 specimens with lesions, 17 (71%) were positive by at least one of the three tests, whereas of the 11 specimens without lesions, only one was positive. The positive specimens comprised 10 of 19 aortas, three of six iliac arteries, and one coronary and one pulmonary artery. *C pneumoniae* was detected in four of six specimens in which there were early changes and in a 20 year old subject. Concerning the 25 subjects, of 17 who had atheromatous arteries, 14 (82%) were *C pneumoniae* positive and of the eight who had normal arteries, none was positive.

Conclusions—There is a strong correlation between *C pneumoniae* and arterial atheromatous lesions. The organism may contribute to the disease process by damaging smooth muscle cells.

(*J Clin Pathol* 1998;**51**:812–817)

Keywords: atherosclerosis; *Chlamydia pneumoniae*

School of Pathology, University of the Witwatersrand, South Africa
A Shor

National Centre for Occupational Health, Johannesburg, South Africa
J I Phillips

MRC Sexually Transmitted Diseases Research Group, Imperial College School of Medicine at St Mary's, Paddington, London W2, UK
G Ong
B J Thomas
D Taylor-Robinson

Correspondence to:
Dr James Ian Phillips, National Centre for Occupational Health, PO Box 4788, Johannesburg 2000, South Africa; email: jim@ncoh.pwv.gov.za

Accepted for publication 1 June 1998

Chlamydia pneumoniae was first described by Grayston *et al* in 1986.[1] In common with the other three species of the genus *Chlamydia*, it is a pathogen. It causes upper respiratory tract infections and is estimated to be responsible for up to 10% of community acquired pneumonia.[2]

Seroepidemiological studies in several countries have shown the incidence of IgG antibodies to *C pneumoniae* to be in excess of 50% and higher in men than in women.[2] The interpretation of these studies is that the majority of the population has been exposed to the organism. Epidemiological studies have also shown an association of seropositivity for *C pneumoniae* with coronary heart disease[3–9] and with carotid artery disease.[10] These findings have been reviewed by Jackson and Grayston.[11]

C pneumoniae was first detected by Shor *et al*,[12] using electron microscopy, in atheromatous lesions of the coronary artery obtained at necropsy from South African patients. Subsequent studies from several centres in different parts of the world have shown the presence of *C pneumoniae* in both postmortem and surgically obtained lesions derived from the aorta, carotid, coronary, iliac, femoral, and popliteal arteries.[11–21] Recently, viable *C pneumoniae* organisms were isolated from atherosclerotic coronary[22 23] and carotid[24] arteries.

In view of the enormous morbidity and mortality caused worldwide by atherosclerosis, and the lack of agreement about its aetiology,[25] the association with *C pneumoniae* is potentially of great importance. As *C pneumoniae* is a known human pathogen, the possibility of a contributory or causative role has to be considered. However, elucidating whether *C pneumoniae* initiates, contributes to, or is incidental to atherogenesis is a difficult "chicken or egg" problem. Ong *et al* have suggested that looking for *C pneumoniae* using the polymerase chain reaction (PCR) in normal tissue and in arteries with early and advanced atheromatous lesions would be a sensible approach.[19] Another approach would be to examine the interactions of *C pneumoniae* using the electron microscope to assess cell damage. A further approach would be to apply the criteria which determine whether an association is causal, as proposed by Hill.[26] With these issues in mind, we have sought evidence of the existence of *C pneumoniae* in a spectrum of atheromatous lesions in different types of arteries from individuals of different ages, using a PCR assay, supported by electron microscopy and immunocytochemistry. In addition, electron microscopy was used to examine interactions between *C pneumoniae* and the cells present in the arterial tissue.

Methods

Arterial tissue was obtained at necropsy examination from the aorta, carotid, cerebral, coronary, femoral, iliac, and pulmonary arter-

Table 1   Details of subjects and specimens examined

No	Age	Cause	Artery	Macro	Micro	EM	PCR	IC
1	26	Trauma	Aorta	0	0	−	−	−
2	70	Nat	Aorta	2	2	+	+	+
3	20	Trauma	Aorta	1	1	−	−	−
4a	26	Trauma	Aorta	1	1	−	−	−
4b				0	0	−	−	−
5	86	CCF	Aorta	3	3	+	+	n/r
6a	37	Trauma	Iliac	2	2	+	+	+
6b			Iliac	2	2	n/r	−	n/r
6c			Iliac	2	2	n/r	+	n/r
7a	57	MI	Iliac	2	2	+	−	n/r
7b			Iliac	2	2	n/r	−	n/r
7c			Iliac	2	2	n/r	−	n/r
8a	75	Pneumo	Aorta	2	2	+	−	+
8b			Carotid	0	0	n/r	−	n/r
8c			Carotid	3	3	n/r	+	n/r
9a	73	COPD	Carotid	2	2	+	+	n/r
9b			Carotid	0	0	n/r	+	n/r
9c			Femoral	2	2	n/r	−	n/r
10a	79	Nat	Aorta	2	2	+	+	n/r
10b				2	2	n/r	+	n/r
11	0	Trauma	Aorta	0	0	−	−	n/r
12	25	Trauma	Carotid	0	0	−	−	n/r
13	25	Trauma	Aorta	0	0	−	−	n/r
14	30	Trauma	Aorta	0	0	−	−	n/r
15	28	Trauma	Aorta	1	1	−	−	n/r
16	34	Trauma	Aorta	1	1	+	+	n/r
17	24	Trauma	Cerebral	0	0	−	−	n/r
18	52	Trauma	Coronary	2	2	+	+	n/r
19	40	Trauma	Aorta	1	1	+	+	n/r
20	30	Trauma	Aorta	0	0	−	−	n/r
21	22	Trauma	Aorta	0	0	−	−	n/r
22	?	Nat	Aorta	2	2	n/r	+	n/r
23	?	Nat	Cerebral	3	3	n/r	−	n/r
24	?	?	Pulmonary	2	2	n/r	+	n/r
25	28	Trauma	Aorta	0	1	n/r	+	n/r

?, unknown; +, positive; −, negative; CCF, congestive cardiac failure; COPD, chronic obstructive pulmonary disease; EM, electron microscopy; IC, immunocytochemistry; Macro, macroscopic grade; MI, myocardial infarction; Micro, microscopic grade; Nat, natural causes; n/r = no result; PCR, polymerase chain reaction; Pneumo, pneumonia.

ies. To reduce the risk of chlamydial DNA cross contamination during dissection, usually only one artery was collected from each subject (table 1).

The cause of death in most cases was trauma. The 35 tissue samples were from 25 males with an age range from newborn to 86 years. Collection of tissue was between 12 and 36 hours after death. The 25 subjects were examined consecutively at necropsy by one pathologist.

At collection, tissue was noted to be macroscopically normal or affected by atheroma, the degree being graded as follows: grade 0, no visible lesion; grade 1, fatty streak; grade 2, fibrolipid plaque; grade 3, advanced lesion with fibrosis, calcification, ulceration, or haemorrhage.

Each specimen was given a code number and divided into three, one portion being frozen and stored at −70°C for PCR studies. A second portion was fixed in a solution of 10% phosphate buffered formalin for histology, and a third portion was fixed in a 2.5% phosphate buffered glutaraldehyde solution for electron microscopy. Each tissue sample was given a coded number and examined by a histopathologist, electron microscopist, and staff of the PCR laboratory, none of whom had access to any specimen information other than a coded number.

Tissue collected in buffered formalin was left to fix for 48 hours and processed routinely before being embedded in wax and sectioned at 5 μm. Sections were stained with haematoxylin and eosin and Masson's trichrome technique.

The presence and degree of atheroma of the intima was assessed by light microscopy and graded as follows: grade 0, no lesion; grade 1, smooth muscle cell damage, macrophage and foam cell infiltration; grade 2, central necrotic area with overlying fibrosis; grade 3, dense fibrosis, calcification, ulceration, neovascularisation, or haemorrhage.

For electron microscopy, the arterial tissue was processed by a standard technique,[27] sectioned, and stained with lead citrate and uranyl acetate. Semithin, 1 μm sections were stained with Azure 2 for comparison with sections for light microscopy.[28] A Jeol 1200 EX 2 instrument was used to examine 90 nm sections on 100 mesh copper grids. Component cells and the matrix of the arteries were identified and the presence of organisms with chlamydial morphology was noted.

Additional sections from the wax blocks used to examine the morphology and grade the degree of atheroma were stained using the chlamydia genus-specific CF-2 antibody according to the immunoperoxidase method of Grayston *et al*.[20]

The tissue samples for the PCR assay, identifiable only by code numbers, were sent by courier in a frozen condition from Johannesburg to London, where they were stored immediately in liquid nitrogen. The DNA extractions and PCR assays were performed according to the method of Ong *et al*.[19] When DNA was extracted from the tissues, a negative control—comprising a tube of extraction buffer only—was processed in an identical manner. Each batch of samples in the PCR cycle was accompanied by a negative control, in which water was substituted for the tissue DNA fragments, and a positive control, to which was added sufficient *C pneumoniae* DNA for the detection of 10 organisms. The first round PCR primers used spanned bases 1053 to 1076 and 1518 to 1540 of the major outer membrane protein (MOMP) gene. The second round primers spanned bases 1053 to 1076 and bases 1254 to 1280 of the MOMP gene. Samples which gave a positive result were confirmed by Southern blot analysis, using a 30 base pair oligonucleotide probe based between the DNA sequences of the two second round primers. The 30 base pair probe spanned bases 1106 to 1135 of the MOMP gene. Samples that gave a negative PCR result were retested for any inhibitors to the PCR assay by seeding an aliquot of the sample DNA with 100 fg of *C pneumoniae* DNA and repeating the assay. To minimise aerosol contamination, the DNA extraction from the tissue samples was carried out in a laminar flow cabinet in a different room from that used for the PCR assay. Mixing of the pre-PCR reagents and the addition of the DNA aliquot were carried out in separate cabinets. In the second round PCR, the already amplified first round DNA was added in a further laminar flow cabinet in a different room from that used for mixing of pre-PCR reagents and the addition of sample DNA. All cabinets were swabbed with methanol and irradiated with ultraviolet light for 30 minutes before use.
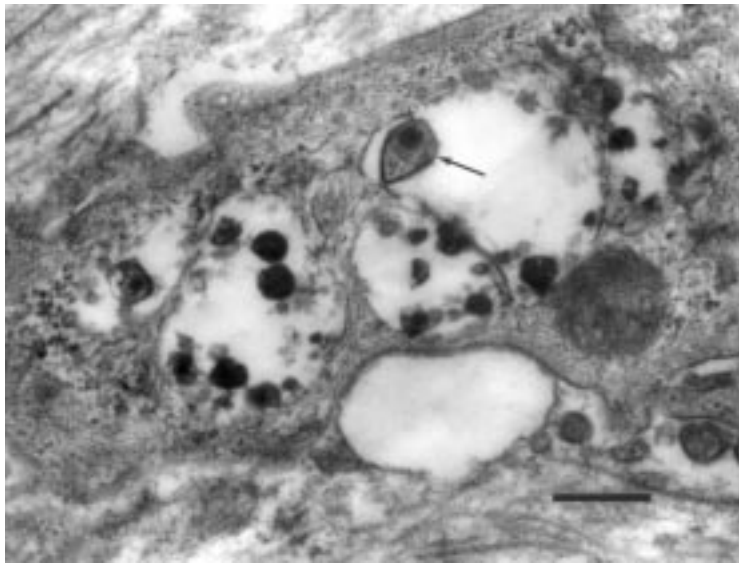
Figure 1   Transmission electron micrograph of an atheromatous arterial wall. Elementary bodies of Chlamydia pneumoniae in vacuoles within a smooth muscle cell. The organisms appear pear shaped in some profiles (arrowed) and contain a central, electron-dense core. The scale bar represents 300 nm.
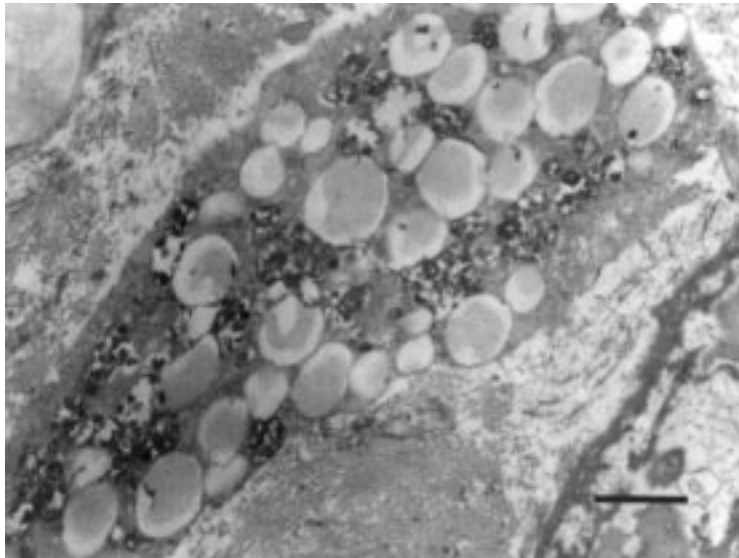

Figure 2   Transmission electron micrograph of a foam cell in an area of atheroma within an artery. The foam cell is identified as a smooth muscle cell full of lipid droplets and vacuoles containing Chlamydia pneumoniae. The scale bar represents 1 µm.

On completion of all the laboratory investigations, the specimen code number and data were revealed to allow analysis of the data.

Results

MACROSCOPIC AND MICROSCOPIC LESIONS

Of the 35 specimens, 23 were found to have macroscopic lesions: five of grade 1, 14 of grade 2, and four of grade 3. Microscopic examination of the specimens showed 24 to have lesions: six of grade 1, 14 of grade 2, and four of grade 3. Thus one macroscopically normal specimen, an aorta from patient No 25, was seen to have a grade 1 microscopic lesion (table 1).

ELECTRON MICROSCOPY

Twenty two specimens were suitable for examination by electron microscopy. Some specimens were not suitable either because of calci-

fication, which rendered the tissue impossible to section, or because of failure to orientate the tissue correctly. Structures similar in morphology and size to those of elementary bodies of *C pneumoniae* were detected in 11 of the specimens. They comprised membrane bound, often pear shaped, 100 to 300 nm particles, containing an electron-dense core measuring up to 100 nm diameter (fig 1) and were identified in smooth muscle cells, in foam cells, and in the extracellular debris of atheromatous lesions. The *C pneumoniae*-like organisms were observed only in areas of tissue damage, and smooth muscle cells containing them were always altered pathologically. The changes observed were vacuolation of the cytoplasm with concurrent reduction in myofilaments and an accumulation of cytoplasmic lipid (fig 2).

IMMUNOCYTOCHEMISTRY

There was consistent background staining in all but seven of the 35 specimens examined, and this staining could not be completely extinguished by routine blocking techniques.[29] As a result, the histopathologist was unable to interpret the immunocytochemically stained sections from 28 specimens. Therefore, only seven specimens were assessed by immunocytochemistry; in these, positive staining—clearly distinguishable from any background staining—was detected in three. The positive staining was observed in smooth muscle cells and foam cells and occasionally in the central necrotic core region.

PCR ASSAY

All the 35 specimens were subjected to analysis by the PCR and 15 were positive for *C pneumoniae*.

CORRELATION OF THE RESULTS OF THE VARIOUS TESTS

Of the 22 specimens examined by electron microscopy, the result of the PCR assay was in agreement in 19. Eight were positive by both tests, 11 were negative by both tests, and three were positive by electron microscopy but negative by the PCR assay. Of the seven specimens examined by immunocytochemistry, the result of electron microscopy was in agreement in six specimens, as was the result of the PCR assay. Overall, of the 35 specimens examined, 18 were positive by at least one of the methods employed. When the 25 subjects rather than specimens are considered, 14 of the 17 who had atheromatous arteries had vessels that were *C pneumoniae* positive, whereas none of the eight who had normal arteries was positive.

TYPE OF VESSEL AND OCCURRENCE OF
C PNEUMONIAE

Seven different arteries were represented. Of 19 aortas examined, 10 were positive for *C pneumoniae* by at least one of the methods employed. Of six iliac arteries examined, three were positive; of five carotid arteries examined, three were positive, as were one coronary and one pulmonary artery. Two cerebral arteries and one femoral artery were negative.

GRADE OF LESION AND OCCURRENCE OF
C PNEUMONIAE

On microscopic examination, one of the macroscopically normal aorta specimens, from patient No 25, was seen to have a grade 1 lesion. *C pneumoniae* was demonstrated in this lesion by the PCR assay. Of the 24 subjects whose specimens had macroscopic or microscopic evidence of a lesion, *C pneumoniae* was detected in 17: in four of the six grade 1 lesions, 11 of the 14 grade 2 lesions, and two of the four grade 3 lesions. Eleven of the specimens had no lesion by macroscopic or microscopic assessment. Of these, only one was positive for *C pneumoniae*, in this case by the PCR assay. This tissue sample was from a normal area between atheromatous lesions in the carotid artery of a 73 year old man (patient No 9b). Unfortunately, owing to failure to orientate this specimen correctly, this area could not be examined by electron microscopy to determine whether there was any cell damage.

AGE OF SUBJECT AND OCCURRENCE OF
C PNEUMONIAE

The age of three of the subjects was not available. Of the remaining 22, 10 were less than 30 years old. Four of these had grade 1 atheromatous lesions, two of which were *C pneumoniae* positive. Of the 12 subjects who were 30 or more years old, 10 had atheromatous lesions, eight of grade 2 or 3, and all 10 were confirmed *C pneumoniae* positive.

Discussion

We are confident that the measures taken to prevent cross contamination of arteries by chlamydial DNA during their collection and processing means that the results obtained by use of the PCR assay are not spurious. Immuno-electronmicroscopy was not used to identify the particles seen, but their structure and size was compatible with the elementary bodies of *C pneumoniae*. Background staining made the interpretation of the results of immunocytochemistry difficult for all but seven samples. Such staining was a problem also encountered by Ong *et al*.[19] Nevertheless, where interpretation was possible, the agreement of the results obtained by immunocytochemistry with those of the PCR assay and electronmicroscopy indicates that the results of immunocytochemistry are not non-specific and caused, as has been suggested,[30] by cross reactivity of the chlamydial antibody with a diseased tissue component. Overall, there was good agreement between the results of the three independent techniques. Based on these, the important observations in this study are as follows: first, the detection of *C pneumoniae* in 71% of arterial atheromatous lesions, in keeping with the results of some previous studies in which the organism has been detected in 60% to 100% of lesions[13 18]; second, the detection, for the first time, of the organism in a lesion of the pulmonary artery (the organism has now been described in this artery and in lesions of the aorta, carotid, coronary, femoral, popliteal,[21] and iliac arteries); third, the occurrence of *C pneumoniae* only in areas of tissue damage as

shown by electron microscopy and immunocytochemistry; and, finally, the finding that minimal lesions are *C pneumoniae* positive as often as severe lesions.

Whether *C pneumoniae* has a causal role in atheroma is the important "chicken or egg" issue. *C pneumoniae* has been isolated by culture from atheromatous arteries[22–24]; however, the organism appears difficult to culture consistently, which makes the fulfilment of Koch's postulates difficult to achieve. However, the question of association or causation has been addressed by Hill,[26] who highlighted nine aspects to consider before deciding on an interpretation of causality. These are concerned with an examination of the association in terms of its strength, consistency, specificity, temporality, plausibility, coherence, analogy, biological gradient, and experimental evidence. We shall consider them in the light of existing data and those provided by this study.

Apart from one report of failure to detect *C pneumoniae* in coronary artherectomy specimens,[31] which has been criticised by Jackson *et al*,[32] there is a strong positive and consistent association between *C pneumoniae* and arterial atheromatous lesions. This was evident in our present study and in reports from several laboratories in different countries based on the use of the PCR assay[12–19 21 33] and culture.[22–24]

In relation to specificity, whereas cytomegalovirus has been detected in both normal and atheromatous arteries,[15] *C pneumoniae* has been found more specifically in atheromatous lesions in this and other studies.[33]

To fulfil the criterion of temporality, the organism must be found in the earliest lesions. It was clear in this study that the existence of *C pneumoniae* in arteries was related to the age of the subjects. Below the age of 30 years, fewer subjects had atheromatous lesions and those that had, had the less severe lesions. Nevertheless, *C pneumoniae* was detected in half of them. Thus *C pneumoniae* has been documented here and elsewhere in fatty streaks[12] which are considered to be the earliest manifestation of atheroma. Indeed, in subject No 25, a 28 year old, a lesion in the aorta was noted only microscopically, but *C pneumoniae* was detected by the PCR technique. Overall, our observations are in keeping with the findings of Kuo *et al*,[15] who demonstrated *C pneumoniae* in the coronary arteries of young adults (15 to 35 years old).

Is a causal association plausible? The answer would seem to be in the affirmative. *C pneumoniae* is a pathogenic organism which has been shown in vitro to be capable of infecting aortic smooth muscle cells, endothelial cells, and macrophages,[34 35] all of which are involved in atherogenesis.[36 37] Furthermore, the results of seroepidemiological studies have shown that infection with *C pneumoniae* is common,[2] occurring at an early age and at intervals throughout life and—like coronary heart disease—more commonly in males.[2]

Would a causal role for *C pneumoniae* be coherent with existing knowledge about atherogenesis? For some time there has been an

accepted response to injury hypothesis to explain atherogenesis, and microorganisms, including viruses, have been proposed as initiators of injury.[36][37] The electronmicroscopic findings suggest that *C pneumoniae* may be contributing to the disease process by damaging smooth muscle cells. Smooth muscle cells containing *C pneumoniae* show vacuolation, loss of myofilaments and an accumulation of lipid. The resultant foam cell is a characteristic feature of atheroma.[37] In addition, an analogy has been drawn between *C pneumoniae* and *C trachomatis*, the latter causing chronic, fibrotic, and necrotic lesions in trachoma and lymphogranuloma venereum.[38] Recent reports of an association between C reactive protein, interleukin 6, and coronary heart disease support the concept of an underlying inflammatory process, such as a chronic infection, playing a role in atherogenesis.[39–41]

There are no data concerning a biological gradient to fulfil Hill's criterion, but preliminary experimental evidence from mouse and rabbit models[42][43] indicate that infection with *C pneumoniae* can produce an atherosclerotic lesion in the aorta.

With regard to experimental evidence, eradication experiments have been suggested.[19][20] Some data have been published on the effects of antichlamydial agents on patients at risk for coronary heart disease, and these initial results appear to show a beneficial effect associated with antibiotic administration.[44][45] In the rabbit model, treatment with an antichlamydial agent prevents the development of atherosclerosis.[43]

In conclusion, we have confirmed a strong positive association between *C pneumoniae* and atheroma. The extent of the association has been expanded to include the pulmonary artery. *C pneumoniae* has been demonstrated in the earliest lesions of atheroma and in subjects as young as 20 years. The observation that infected smooth muscle cells in the arterial wall become vacuolated, lipid filled, and contribute to the foam cell population is consistent with a possible causal role for *C pneumoniae* in the development of atheromatous lesions. Indeed, from the work in this and other studies, Hill's criteria for causation have been fulfilled to a large extent. However, unequivocal evidence is still lacking. If *C pneumoniae* existed only in the older lesions then no case could be made for it as an initiator of atheroma. Demonstration of the organism in subjects as young as 20 years and in the earliest lesions of atheroma is consistent with it having a causal role. Unfortunately, it is still not possible to establish whether *C pneumoniae* predates and initiates an early lesion or infiltrates an early lesion after it has begun to develop. Resolution of this conundrum may lie in the outcome of preventive measures, most notably antibiotic treatment. Initial results in humans[44][45] treated with antichlamydial agents are encouraging. The prevention of atherosclerosis in rabbits infected with *C pneumoniae* and treated with antibiotics[43] strengthens the aetiological link between *C pneumoniae* and atherosclerosis. Data from more clinical trials are needed, along with an examination of the interaction of *C pneumoniae* with arterial tissue.

1 Grayston JT, Kuo C-C, Wang SP, *et al*. A new Chlamydia psittaci strain, TWAR, isolated from acute respiratory tract infections. *N Engl J Med* 1986;**315**:161–8.
2 Grayston JT. Infections caused by Chlamydia pneumoniae strain TWAR. *Clin Infect Dis* 1992;**15**:757–63.
3 Saikku P, Mattila K, Nieminen MS, *et al*. Serological evidence of an association of a novel Chlamydia, TWAR, with chronic coronary heart disease and acute myocardial infarction. *Lancet* 1988;ii:983–6.
4 Leinonen M, Linnanmaki E, Mattila K, *et al*. Circulating immune complexes containing chlamydial lipopolysaccharide in acute myocardial infarction. *Microb Pathog* 1990;**9**:67–73.
5 Saikku P, Leinonen M, Tenkanen L, *et al*. Chronic Chlamydia pneumoniae infection as a risk factor for coronary heart disease in the Helsinki heart study. *Ann Intern Med* 1992;**116**:273–8.
6 Linnanmaki E, Leinonen M, Mattila K, *et al*. Chlamydia pneumoniae specific circulating immune complexes in patients with chronic coronary heart disease. *Circulation* 1993;**87**:1130–4.
7 Thom DH, Wang S-P, Grayston JT, *et al*. Chlamydia pneumoniae strain TWAR antibody and angiographically demonstrated coronary heart disease. *Arterioscler Thromb* 1991;**11**:547–51.
8 Thom DH, Grayston JT, Siscovick DS, *et al*. Association of prior infection with Chlamydia pneumoniae and angiographically demonstrated coronary artery disease. *JAMA* 1992;**268**:68–72.
9 Patel P, Kendall MA, Carrington D, *et al*. Association of Helicobacter pylori and Chlamydia pneumoniae infections with coronary heart disease. *BMJ* 1995;**311**:711–14.
10 Melnick S, Shahar E, Folsom A, *et al*. Past infection by Chlamydia pneumoniae strain TWAR and asymptomatic carotid atherosclerosis. *JAMA* 1993;**95**:499–504.
11 Jackson L, Grayston T. Chlamydia pneumoniae and Mycoplasma pneumoniae infections. *Curr Opin Infect Dis* 1996;**9**:89–93.
12 Shor A, Kuo C-C, Patton DL. Detection of Chlamydia pneumoniae in the coronary artery atheroma plaque. *S Afr Med J* 1992;**82**:158–61.
13 Kuo C-C, Shor A, Campbell LA, *et al*. Demonstration of Chlamydia pneumoniae in atherosclerotic lesions of coronary arteries. *J Infect Dis* 1993;**167**:841–9.
14 Kuo C-C, Gown AM, Benditt EP, *et al*. Detection of Chlamydia pneumoniae in aortic lesions of atherosclerosis by immunocytochemical stain. *Arterioscler Thromb* 1993;**13**:1501–4.
15 Kuo C-C, Grayston JT, Campbell LA, *et al*. Chlamydia pneumoniae (TWAR) in coronary arteries of young adults (15–35 years old). *Proc Natl Acad Sci USA* 1995;**92**:6911–14.
16 Campbell LA, O'Brien ER, Cappuccio AL, *et al*. Detection of Chlamydia pneumoniae (TWAR) in human coronary artherectomy tissue. *J Infect Dis* 1995;**172**:585–8.
17 Ouchi K, Fujii B, Kamamoto Y, *et al*. Detection of Chlamydia pneumoniae in atherosclerotic lesions of coronary arteries and large arteries [abstract]. 35th Interscience Conference on Antimicrobial Agents and Chemotherapy, San Francisco, 17—20 September 1995:294 (abstract K37).
18 Juvonen J, Juvonen T, Laurila A, *et al*. Demonstration of Chlamydia pneumoniae in the walls of abdominal aortic aneurysms [abstract]. Third International Congress on the Macrolides, Azalides and Streptogramins. Lisbon, 24–26 January 1996.
19 Ong G, Thomas BJ, Mansfield AO, *et al*. Detection and widespread distribution of Chlamydia pneumoniae in the vascular system and its possible implications. *J Clin Pathol* 1996;**49**:102–6.
20 Grayston JT, Kuo C-C, Coulson AS, *et al*. Chlamydia pneumoniae (TWAR) in atherosclerosis of the carotid artery. *Circulation* 1995;**92**:3397–400.
21 Kuo C-C, Coulson AS, Campbell LA, *et al*. Detection of Chlamydia pneumoniae in atherosclerotic plaques in the walls of arteries of lower extremities from patients undergoing bypass operation for arterial obstruction. *J Vasc Surg* 1997;**26**:29–31.
22 Ramirez JA, the Chlamydia pneumoniae/Atherosclerosis Study Group. Isolation of Chlamydia pneumoniae from the coronary artery of a patient with coronary atherosclerosis. *Ann Intern Med* 1996;**125**:979–82.
23 Maas M, Krause E, Kruger S, *et al*. Coronary arteries harbour viable Chlamydia pneumoniae. Fourth international symposium on modern concepts in endocarditis and cardiovascular infections [abstract]. Yverdon-Les-Baines, Switzerland, May 1997 (abstract 101).
24 Jackson LA, Campbell LA, Kuo C-C, *et al*. Isolation of Chlamydia pneumoniae from a carotid endarterectomy specimen. *J Infect Dis* 1997;**176**:292–5.
25 Woolf N. *Pathology of atherosclerosis*. London: Butterworth, 1982.
26 Hill AB. The environment and disease: association or causation? *Proc R Soc Med* 1965;**58**:295–300.

27 Phillips JI, Isaacson C, Carman H. Ochronosis in black South Africans who used skin lighteners. *Am J Dermatopathol* 1986;**8**:14–21.
28 Phillips JI. Lack of cilia and squamous metaplasia in upper respiratory tract biopsies from children. *S Afr Med J* 1989;**76**:355–7.
29 Nadji M. Immunoperoxidase techniques. *Am J Dermatopathol* 1986;**8**:32–6.
30 Wissler RW. Significance of Chlamydia pneumoniae (TWAR) in atherosclerotic lesions. *Circulation* 1995;**92**:3376.
31 Weiss SM, Roblin PM, Gaydos CA, *et al.* Failure to detect Chlamydia pneumoniae in coronary atheromas of patients undergoing atherectomy. *J Infect Dis* 1996;**173**:957–62.
32 Jackson LA, Campbell LA, Kuo C-C, *et al.* Detection of Chlamydia pneumoniae in atheroma specimens. *J Infect Dis* 1996;**174**:893–6.
33 Jackson LA, Campbell LA, Schmidt RA, *et al.* Specificity of detection of Chlamydia pneumoniae in cardiovascular atheroma. *Am J Pathol* 1997;**150**:1785–90.
34 Knoebel E, Vijayagopal P, Figueroa JE, *et al.* In vitro infection of smooth muscle cells by Chlamydia pneumoniae. *Infect Immun* 1997;**65**:503–6.
35 Gaydos CA, Summersgill JT, Sahney NN, *et al.* Replication of Chlamydia pneumoniae in human macrophages, endothelial cells and aortic artery smooth muscle cells. *Infect Immun* 1996;**64**:1614–20.
36 Ross R. The pathogenesis of atherosclerosis: a perspective for the 1990's. *Nature* 1993;**362**:801–9.
37 Hajjar DP, Nicholson AC. Atherosclerosis. *Am Scientist* 1995;**83**:460–7.
38 Blanchard T, Bailey R, Holland M, *et al.* Chlamydia pneumoniae and atherosclerosis. *Lancet* 1993;**341**:825.
39 Maseri A, Biasucci LM, Liuzzo G. Inflammation in ischaemic heart disease. *BMJ* 1996;**312**:1049–50.
40 Mendall MA, Patel P, Ballam L, *et al.* C-reactive protein and its relation to cardiovascular risk factors: a population-based cross-sectional study. *BMJ* 1996;**312**:1061–5.
41 Ridker PM, Cushman M, Stampfer MJ, *et al.* Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men. *N Engl J Med* 1997;**336**:973–9.
42 Moazed TC, Kuo C-C, Grayston JT, *et al.* Murine models of Chlamydia pneumoniae infection and atherosclerosis. *J Infect Dis* 1997;**175**:883–90.
43 Muhlestein JB, Anderson JL, Hammond EH, *et al.* Infection with Chlamydia pneumoniae accelerates the development of atherosclerosis and treatment with azithromycin prevents it in a rabbit model. *Circulation* 1998;**97**:633–6.
44 Gupta S, Leathan EW, Carrington D, *et al.* Elevated Chlamydia pneumoniae antibodies, cardiovascular events and azithromycin in male survivors of myocardial infarction. *Circulation* 1997;**96**:404–7.
45 Gurfinkel E, Bozovich G, Daroca A, *et al.* and ROXIS study group. Randomised trial of roxithromycin in non-Q-wave coronary syndromes: Roxis pilot study. *Lancet* 1997;**350**:404–7.

JCP

Chlamydia pneumoniae in atheroma: consideration of criteria for causality.

A Shor, J I Phillips, G Ong, B J Thomas and D Taylor-Robinson

Updated information and services can be found at:

http://jcp.bmj.com/content/51/11/812

These include:

Email alerting service	Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/