

Confirmation Bias in Studies of Nestmate Recognition: A Cautionary Note for Research into the Behaviour of Animals

Ellen van Wilgenburg*, Mark A. Elgar

Department of Zoology, University of Melbourne, Melbourne, Victoria, Australia

Abstract

Confirmation bias is a tendency of people to interpret information in a way that confirms their expectations. A long recognized phenomenon in human psychology, confirmation bias can distort the results of a study and thus reduce its reliability. While confirmation bias can be avoided by conducting studies blind to treatment groups, this practice is not always used. Surprisingly, this is true of research in animal behaviour, and the extent to which confirmation bias influences research outcomes in this field is rarely investigated. Here we conducted a meta-analysis, using studies on nestmate recognition in ants, to compare the outcomes of studies that were conducted blind with those that were not. Nestmate recognition studies typically perform intra- and inter colony aggression assays, with the *a priori* expectation that there should be little or no aggression among nestmates. Aggressive interactions between ants can include subtle behaviours such as mandible flaring and recoil, which can be hard to quantify, making these types of assays prone to confirmation bias. Our survey revealed that only 29% of our sample of 79 studies were conducted blind. These studies were more likely to report aggression among nestmates if they were conducted blind (73%) than if they were not (21%). Moreover, we found that the effect size between nestmate and non-nestmate treatment means is significantly lower in experiments conducted blind than those in which colony identity is known (1.38 versus 2.76). We discuss the implications of the impact of confirmation bias for research that attempts to obtain quantitative syntheses of data from different studies.

Citation: van Wilgenburg E, Elgar MA (2013) Confirmation Bias in Studies of Nestmate Recognition: A Cautionary Note for Research into the Behaviour of Animals. PLoS ONE 8(1): e53548. doi:10.1371/journal.pone.0053548

Editor: Deborah M. Gordon, Stanford University, United States of America

Received: December 1, 2011; **Accepted:** December 3, 2012; **Published:** January 23, 2013

Copyright: © 2013 van Wilgenburg, Elgar. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This research was funded by the Australian Research Council. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: ellenvw@gmail.com

Introduction

“...for it is a habit of mankind to entrust to careless hope what they long for, and to use sovereign reason to thrust aside what they do not desire.” From *History of the Peloponnesian War* 431 B.C., Thucydides

Confirmation bias, a well-documented phenomenon in psychology, is the tendency of humans to seek out evidence and interpret it in a manner that confirms their existing ideas and hypotheses [1–7]. Confirmation bias is often described as a result of automatic processing, occurring more or less unintentionally but nevertheless potentially distorting the data collected in scientific research. Ideally, scientific researchers avoid confirmation bias by searching for falsifying, as well as confirming evidence [8,9]. However, in reality, scientists often have high stakes for obtaining particular research outcomes [10], and the expectations for an experiment can potentially affect which data are collected and how they are interpreted and reported [1,2]. For example, two-thirds of recording errors in several psychological studies were biased in the direction of the observer’s hypothesis [1]. The extent to which observation bias influences the outcomes of a study will depend on the kind of observations that are being made [1,7,11]. Observa-

tions are more prone to bias if 1) the variable is not clearly defined, 2) the subject under observation is hard to perceive, 3) the observations require subjective assessment, and 4) the observer has an interest in the outcome of the study.

Confirmation bias can be avoided by designing experiments in which the observers are blind to the treatment assignment of their subjects [12–18]. For example, to test whether consumers have a taste preference for one brand of pop over another, the identity of the pop should be concealed because otherwise the subjects tend to prefer the brand with which they are more familiar. Nowadays, blind experiments are commonplace in many scientific disciplines, including pharmacology, market research, psychology, physics and certain branches of biology. Indeed, in some fields of research, blinding of experiments is essential for publication.

Such an experimental tradition appears to be less widely adopted in the field of animal behaviour, where researchers collect observational data that may be subject to systematic error. For example, a survey by Gamboa et al. [18] revealed that only 27% of 33 studies that investigated kin-recognition and were published in the journal *Animal Behaviour* between 1987 and 1989 mentioned blind assays. Studies of animal behaviour may be particularly prone to confirmation bias, especially when a certain degree of interpretation is required— typically when the behaviours are rapid, subtle or similar in appearance to other behaviours. Yet

remarkably few studies have investigated the extent to which confirmation bias influences research outcomes in animal behaviour [19–21]. Almost half a century ago Cordaro and Ison [21] conducted an experiment in which they asked students to observe the behaviour of planaria (non-parasitic Turbellaria flatworms). One group of observers were told the planaria would move and turn frequently, whereas the other group of observers were told their planaria rarely move and turn. In reality, the planaria were randomly allocated to the two groups. The group of students anticipating high-activity animals found that the planaria moved on average 18 times and turned 49 times, while the group of students anticipated low-activity animals reported the planaria moved on average once and turned 10 times. Similar studies by Rosenthal and Fode [19] and Marsch and Hanlon [20] conducted on the behaviour of rats and salamanders respectively, also report that *a priori* expectations can bias behavioural observations. However, it has to be noted that the observers in all three studies were undergraduates, who generally have little or no training in conducting behavioural observations. More experienced researchers may make fewer observational errors and their data may therefore be more reliable.

An alternative approach to addressing the question of whether confirmation bias affects research into animal behaviour is to compare the outcomes of published studies that are conducted blind with those that are not. If behavioural observations are influenced by confirmation bias, then the outcomes of studies that have been conducted blind should have smaller effect sizes than similar experiments that were not conducted blind.

In this study, we explore the evidence for confirmation bias in studies of animal behaviour by focussing on a single research topic—nestmate recognition in ants. Our intention is to use studies of nest-mate recognition as a ‘model system’ to highlight the potential impact of confirmation bias, which is a potential issue for all quantitative research, including animal behaviour. Ants, like other social insects, maintain colony cohesion by recognizing and, if necessary, discriminating against conspecifics that are not members of their colony [22–26]. The mechanisms of nestmate recognition have received considerable attention during the past 25 years [27–29]. Aggression assays are useful tools when trying to test hypotheses regarding the influence of context, environment and heritability on nestmate recognition. Experimenters have used a variety of methods testing inter-colony aggression ranging from one-on-one assays to group encounters, which often yield similar results [30]. However, like any other observational method of data acquisition, aggression assays have their limitations and great care must be taken to avoid false positive and negative results when designing nestmate recognition studies [31]. Several authors stress the importance of conducting these types of assays blind to the treatment [18,31]. Nestmate recognition experiments typically involve intra- and inter colony aggression assays with the *a priori* expectation that there should be little or no aggression among nestmates. Since little or no aggression is expected among nestmates, we expect aggression to be less frequently reported in trials involving nestmates that are not conducted blind, compared with those conducted blind – that is, the experimenter has no knowledge of whether the ants involved in the assay comprise nestmates only, or a mixture of nestmates and non-nestmates.

Studies of nestmate recognition are particularly suitable to investigate confirmation bias for a variety of reasons. First, many studies use similar experimental designs, allowing relatively straightforward comparisons across studies. Second, aggressive behaviour in ants can include subtle behaviours such as mandible flaring and recoil that are hard to quantify, making the assays potentially prone to confirmation bias. Using a meta-analysis, we

specifically ask 1) what proportion of studies of nestmate recognition are conducted blind, and 2) do the outcomes of blind studies differ from those of non-blind studies?

Methods

We searched for papers on nestmate recognition in ants using *ISI Web of Science* (Thomson Reuters) search engine, with the search terms “nestmate recognition” or “nest mate recognition”. We conducted the last search in July 2011. To be included in our analyses, studies must have conducted a nestmate recognition experiment on ants that included both a nestmate (control) and non-nestmate aggression assay. Aggression assays had to involve either live, chilled or dead ants. To determine whether a study was conducted blind or not (e.g. whether the observers of the assays were aware of the colony identity of the workers) we carefully read through the method section of each paper. We deemed a study as blind only if this was explicitly stated, and categorised the remaining studies as non-blind. It is possible that some of the studies we deemed non-blind were, in fact, conducted blind. However, the alternative of contacting the authors of all papers to ascertain whether their study was conducted blind or not introduces several sources of bias that we could not control. For example, authors that did not explicitly state their study was conducted blind may be less likely to recall whether the study was done blind or more likely to remember incorrectly.

We treated different experiments included in the same publication, different studies by the same author, and different studies on the same species as independent because leaving them out may lead to greater loss of information and distortion of the results than those caused by their potential non-independence [32]. The studies included in our sample are listed in Table 1. We do not include details of the experimental methods for each study because we see no value in drawing attention to the methods of individual studies. The types of aggression assay and the methods of scoring vary between studies. For example, assays may involve one-on-one encounters in a petri-dish [33], or placing ants into a nest [34] or foraging trail [35]. Aggression may then be scored as simply the presence or absence of aggression [36] or it may be scored on a scale based on specific behaviours thought to represent increasing aggression [35]. A meta-analysis requires a certain level of homogeneity among studies, limiting the number of studies that can be included in the analysis. Since our dataset is so varied, a single type of analysis would necessarily exclude many studies. We therefore chose to analyse our data using two different methods that differ in their selection criteria, resulting in two mostly overlapping, but nonetheless different samples. First, we tested whether there was any difference in the frequency with which aggression was reported in nestmate trials in blind versus non-blind studies. Second, we compared the nestmate vs non-nestmate effect size between blind and non-blind experiments.

Frequency of aggression reported in nestmate trials

We compared the frequency with which aggression is reported in blind *versus* non-blind studies by searching for papers that specifically mention the presence or absence of aggression in nestmate trials. To be included in this analysis studies had to either present their data as presence/absence of aggression, or explicitly mention that aggression was or was not found in the nestmate trials.

Differences in effect sizes of aggressive behavior

Studies included in our analysis of effect sizes had to report descriptive statistics (means, standard deviation or standard error

Table 1. Evidence for aggression among nestmates in studies of nestmate recognition in ants.

Taxa	Aggression among nestmates?	d	Var (d)	Reference
Paraponerinae				
<i>Paraponera clavata</i>	Yes			[46]
Ponerinae				
<i>Odontomachus Bauri</i>	No			[47]
<i>Pachycondyla inversa</i>		0.840	0.076	[48]
<i>Pachycondyla subversa</i>		1.160	0.147	[48]
<i>Pachycondyla villosa</i>		0.762	0.075	[48]
Myrmeciinae				
<i>Myrmecia nigriceps</i>	Yes			[40]
Pseudomyrmecinae				
<i>Pseudomyrmex ferruginea</i>	No			[49]
<i>Pseudomyrmex pallidus</i>		1.248	0.154	[50]
Dolichoderinae				
<i>Iridomyrmex purpureus</i>		3.629	0.157	[35]
<i>Iridomyrmex purpureus</i>	No			[51]
<i>Iridomyrmex purpureus</i>	Yes			[52]
<i>Iridomyrmex purpureus</i>	Yes	0.955	0.045	[53]
<i>Linepithema humile</i>	No			[54]
<i>Linepithema humile</i>				[55]
<i>Linepithema humile</i>		9.678	1.321	[56]
<i>Linepithema humile</i>		1.954	0.006	[57]
<i>Linepithema humile</i>	No	0.225	0.027	[58]
Ectatomminae				
<i>Ectatomma ruidum</i>	No			[59]
<i>Ectatomma ruidum</i>	Yes			[60]
<i>Ectatomma tuberculatum</i>				[61]
Formicinae				
<i>Anoplolepis gracilipes</i>	No			[62]
<i>Camponotus aethiops</i>		1.525	0.047	[63]
<i>Camponotus aethiops</i>		1.059	0.095	[64]
<i>Camponotus aethiops</i>	No	3.069	0.109	[65]
<i>Camponotus atriceps</i>		0.939	0.185	[66]
<i>Camponotus cruentatus</i>				[67]
<i>Camponotus fellah</i>	No	1.512	0.205	[68]
<i>Camponotus fellah</i>	No			[69]
<i>Camponotus fellah</i>	No			[70]
<i>Camponotus floridanus</i>	No			[71]
<i>Camponotus floridanus</i>	No			[72]
<i>Camponotus japonicus</i>	Yes	1.384	0.113	[73]
<i>Camponotus ruffemur (black)</i>	Yes			[25]
<i>Camponotus ruffemur (red)</i>	Yes			[25]
<i>Camponotus rufipes</i>	No	3.484	0.420	[74]
<i>Camponotus vagus</i>	No			[75]
<i>Camponotus yamaokai</i>	No			[76]
<i>Cataglyphis cursor</i>	No			[77]
<i>Cataglyphis cursor</i>	No	2.280	0.114	[78]
<i>Cataglyphis iberica</i>	No			[79]
<i>Cataglyphis niger</i>		4.343	0.480	[80]
<i>Formica exsecta</i>	Yes	0.139	0.083	[81]

Table 1. Cont.

Taxa	Aggression among nestmates?	d	Var (d)	Reference
<i>Formica exsecta</i>	No			[82]
<i>Formica exsecta</i>	No			[83]
<i>Formica japonica</i>	No			[84]
<i>Formica montana</i>	Yes			[85]
<i>Formica pratensis</i>	No			[86]
<i>Formica pratensis</i>	Yes	1.213	0.064	[87]
<i>Formica rufibarbis</i>		1.309	0.130	[88]
<i>Formica selysi</i>	Yes			[89]
<i>Lasius neglectus</i>	No			[90]
<i>Oecophylla smaragdina</i>				[91]
<i>Plagiolepis pygmaea</i>	No	7.295	0.064	[92]
Myrmicinae				
<i>Acromyrmex lobicornis</i>	Yes			[93]
<i>Acromyrmex subterraneus</i>	No			[94]
<i>Aphaenogaster senilis</i>		1.676	0.025	[95]
<i>Aphaenogaster senilis</i>	No			[96]
<i>Apterostigma collare</i>	No			[97]
<i>Cataulacus mckeyi</i>	No			[98]
<i>Leptothorax ambiguus</i>	No			[99]
<i>Leptothorax longispinosus</i>	Yes			[100]
<i>Leptothorax longispinosus</i>		4.951	0.033	[101]
<i>Leptothorax retractus</i>	No			[102]
<i>Leptothorax sp B</i>	Yes	1.661	0.168	[102]
<i>Monomorium pharaonis</i>				[103]
<i>Myrmica rubra</i>		0.506	0.145	[104]
<i>Myrmica rubra</i>	Yes	1.694	0.165	[105]
<i>Pheidole megacephala</i>	No			[106]
<i>S. invicta, S. richteri hybrids</i>	No			[107]
<i>Solenopsis invicta</i>	Yes	3.938	0.235	[108]
<i>Solenopsis invicta</i>	No			[107]
<i>Solenopsis invicta</i>		4.971	0.090	[109]
<i>Solenopsis invicta</i>	No			[110]
<i>Solenopsis invicta</i>		3.064	0.084	[111]
<i>Solenopsis richteri</i>	No			[107]
<i>Temnothorax crassispinus</i>	Yes	0.370	0.020	[112]
<i>Temnothorax nylanderii</i>	Yes	0.183	0.020	[112]
<i>Temnothorax unifasciatus</i>	Yes	-0.174	0.020	[112]
<i>Tetramorium bicarinatum</i>	No			[113]

The table gives the effect sizes measured as Hedge's d and the variance of Hedge's d (Var(d)). (A full table, including assignment of experimental protocol, is available on request to the authors.)

doi:10.1371/journal.pone.0053548.t001

and sample size) of a measure of aggression for both nestmate and non-nestmate trials. We retrieved this information either directly from the text or estimated it from the figures, and converted standard errors to standard deviations.

Statistical methods

We used Fisher's exact test (with each experiment as an independent value) to reveal whether aggression was more frequently reported in nestmate trials that were conducted non-blind than blind. We compared the effect size of blind and non-

blind studies using MetaWin 2.0 [37]. We measured the effect using Hedges d [38], which provides a standardized mean difference between nestmate and non-nestmate mean value of the aggression assay. If a study included several different non-nestmate treatments (for example non-nestmates of colonies from different distances) we calculated the average level of aggression and standard deviations over these treatments.

We compared the effect sizes across studies using mixed effect models that allow for fixed differences between groups of studies (in our case blind versus non-blind), and assume that differences

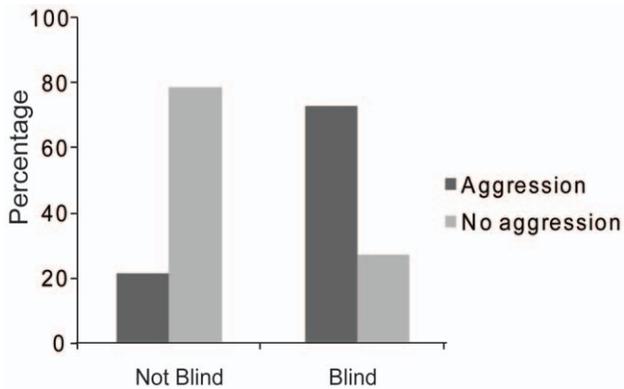


Figure 1. Percentage of non-blind and blind studies reporting aggression in control trials.

doi:10.1371/journal.pone.0053548.g001

among studies within a class are due to both sampling error and random variation [37,39]. Although the statistical power of mixed models is lower than that of fixed-effects models, the assumptions of mixed models are much more likely to be met in most meta-analyses in ecology [37,38]. We report the mean response ratios and their 95% confidence limits. For comparisons between blind and non-blind studies we examined the between-group heterogeneity using a Chi-square test, Q_B [38].

Potential sources of bias

We assessed potential bias both within individual studies included in the meta-analysis and across the meta-analysis as a whole, considering studies excluded for any reason including failure to publish non-significant results. We examined individual studies for any potential sources of bias and did not find any experimental design limitations other than the absence of blinding in the non-blind studies, so there is no evidence for bias within studies that would affect our conclusions. At the level of the whole meta-analysis, publication bias could influence our results if statistically non-significant results are under-reported. This would only affect our conclusions if blind studies were less likely to find significant results than non-blind studies and a greater proportion of blind studies remained unpublished for this reason. While there

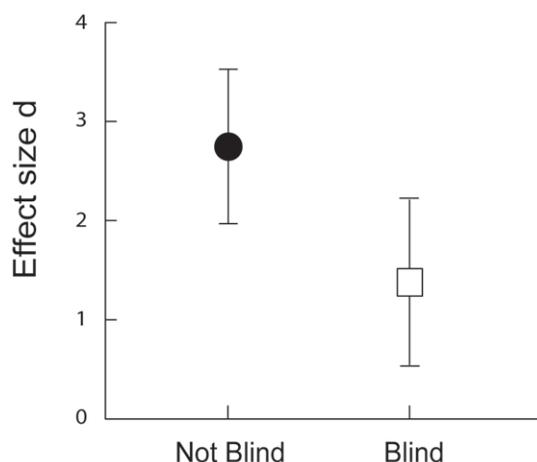


Figure 2. Mean effect size (d) and 95% CI of non-blind and blind studies.

doi:10.1371/journal.pone.0053548.g002

are tests for publication bias [37], we were not able to test for statistically significant differences in publication bias between blind and non-blind studies, particularly since such tests would require much larger sample sizes than are available with the current published literature. So while it seems unlikely that a difference in publication bias would influence our main conclusions, it is possible and should be considered when interpreting our results.

Results

We obtained published reports of 156 nestmate recognition experiments in ants. 79 of these studies involved live, chilled or dead ants and included nestmate controls. In 29% (23 of 79) of these studies, the experiments were conducted blind with respect to colony identity. Six of the studies that were not conducted blind according to colony identity were nonetheless blinded for different treatments among non-nestmate aggression assays (for example genetic distance). We could infer whether aggression was reported for the nestmate controls in 57 of the 79 experiments. We found that blind experiments were significantly more likely to report aggression in the controls than those not conducted blind (11 out of 15, or 73% versus 9 out of 42 or 21%, $P < 0.001$, Figure 1). Thirty-three experiments fulfilled our inclusion criteria for the meta-analysis (see Methods and Table 1) and 15 (45%) of these were conducted blind. Blind experiments yielded a significantly lower treatment effect than non-blind experiments (Blind: $0.46 \leq 1.38 \leq 2.29$; Non-Blind: $1.92 \leq 2.76 \leq 3.83$; $Q_B = 5.61$, $P = 0.018$, Figure 2).

Discussion

Our meta-analysis provides evidence of confirmation bias in studies of nestmate recognition in ants. Experimental aggression assays that investigate nestmate recognition in ants can be conducted blind with respect to the origin of ants. Aggression among nestmates was three times more likely to be reported in blinded than non-blinded experiments. Furthermore, the effect size – the differences between the level of aggression among nestmates and that among non-nestmates – in non-blind experiments was twice that of blind experiments. Remarkably, less than a third of the studies in our sample were conducted blind. While it is possible that some of the studies in our sample were incorrectly labelled non-blind, such incorrect allocations act against the observed pattern that blinded experimental designs are typically reported. Further, the minimum number of incorrectly assigned studies required to render our initial analysis non-significant would be 7, or 12% of the studies included in our sample, which seems an unrealistically high error rate.

There may be several explanations for the magnitude of confirmation bias in the non-blind studies. First, ants are relatively small, fast moving, and their aggressive behaviour may be particularly hard to quantify. While nestmates usually behave amicably, handling of worker ants may occasionally elicit aggression among nestmates and a rapid bite to the leg or a mandible flare can be easily overlooked if such behaviour is not expected. Moreover, aggressive behaviours in ants may appear similar to other non-aggressive behaviours. For example, ants may flare their mandibles as a threat to intruders, but may also open their mandibles in order to solicit trophallaxis. The more frequently the behaviour requires interpretation, the more likely the data become prejudiced. The dramatic effects of confirmation bias revealed in our analysis highlight the impact of automatic, unintentional processing, even when the experiments are typically utilised to address broader questions, and the magnitude of the difference in the behaviour of nestmates and non-nestmates is

often of little consequence. Indeed, reporting an absence of aggression in both nestmate and non-nestmate trials is more remarkable [40].

Our analysis raises the question of whether the degree of confirmation bias revealed in this study reflects research in animal behaviour more generally. While confirmation bias is an issue for almost all kinds of quantitative research, there is likely to be considerable variation across (and within disciplines) in the degree to which it is controlled. Several factors may influence this variation. Most obvious is the degree of prior expectations, which may derive from a compelling theoretical framework and/or empirical evidence – both of which are true for studies of nestmate recognition in ants. On that basis, we might expect similar levels of confirmation bias in, for example, studies of winner and loser effects [41,42] or those that investigate the relationship between predator vigilance and group size in vertebrates [43]. Second, the accuracy of observations may be important: as already noted, behavioural observations on ant aggression may be prone to bias because ants are relatively small and fast moving, and so aggression may be difficult to discern accurately. Finally, observation biases may be more prevalent in studies of animals that have humanlike behaviours [44]. If so, the level of confirmation bias described here may be at the lower end of the spectrum.

Less than a third of the studies in our sample were conducted blind, a statistic similar to that published over 20 years ago for this kind of research [18]. This is surprising, since confirmation bias is widely documented, and textbooks on scientific methods and experimental design encourage blind experimentation [12–13]. While the nature of some experiments or sampling observations in animal behaviour would make it technically impossible to conduct them blind, there may be other explanations why blinding is so rare. Some researchers may choose to conduct open trials in the belief that the behaviour in question is easy to classify and therefore not prone to bias. Such a view is most likely mistaken, as confirmation bias occurs more or less unintentionally and scientists generally do not distort data intentionally [45]. Observations of the behaviour of animals are often thought to be less subjective than, for example, the qualitative observations in human psychology or market research. Our data suggest that, again, this view may be mistaken – in the absence of data such as that provided here, researchers may underestimate the extent to which

confirmation bias can influence the outcomes of a study. Unfortunately, being informed about confirmation bias may not solve the problem entirely: around 75% of studies in special education research, in which the role of confirmation bias had been extensively investigated, made no precaution against it [11]. Finally, open trials may be preferred simply because of the additional costs of conducting experiments blind. These costs may not be trivial because blinding typically requires a second person to label the treatments. It is likely that the most effective way of encouraging researchers to conduct experiments blind is if journals set a benchmark for experimental design. If there is a trade-off between the chance of error and productivity, it may be otherwise unrealistic to expect researchers to utilise methodological standards beyond what is expected by scientific research journals.

The results for most studies that were not conducted blind are likely robust because the treatment effect size in both blind and non-blind studies is much greater than the difference in the effect sizes. Nevertheless, the prevalence of confirmation bias in studies that are not conducted blind has significant implications for synthetic research that relies on published data, such as comparative or meta-analyses. For example, an inter-specific analysis of the variation in the level of aggression expressed toward non-nestmates may yield a distorted pattern if the experimental methodology is linked to particular taxonomic groups. As a precaution, such studies may be advised to include only blind studies in these types of research. We hope that our analysis will stimulate renewed interest in designing experiments in a way that bias is minimized and set a methodological benchmark for research in animal behaviour more generally.

Supporting Information

Checklist S1 PRISMA 2009 Checklist.
(DOC)

Flow Diagram S1 PRISMA 2009 Flow Diagram.
(DOC)

Author Contributions

Conceived and designed the experiments: EvW MAE. Performed the experiments: EvW. Analyzed the data: EvW. Contributed reagents/materials/analysis tools: EvW MAE. Wrote the paper: EvW MAE.

References

- Rosenthal R, Rosnow RL (1969) Artifacts in behavioral research: Academic Press.
- Rosenthal R (1976) Experimenter effects in behavioural research. New York: Irvington Publishers, Inc.
- Nickerson (1998) Confirmation bias: a ubiquitous phenomenon in many guises. *Rev gen psychol* 2: 175–220.
- MacCoun RJ (1998) Biases in the interpretation and use of research results. *Ann Rev Psychol* 49: 259–287.
- Kapchuk TJ (2003) Effect of interpretive bias on research evidence. *Brit Med J* 326: 1453–1455.
- Foster GG, Ysseldyke JE, Reese JH (1975) I wouldn't have seen it if I hadn't believed it. *Except Children* 41: 469–473.
- Balgh DF, Balgh MH (1983) On the psychology of watching birds - the problem of observer-expectancy bias. *Auk* 100: 755–757.
- Popper KR (2002) The logic of scientific discovery: Routledge.
- Popper KR (2002) Conjectures and refutations: the growth of scientific knowledge: Routledge.
- Fanelli D (2010) Do Pressures to Publish Increase Scientists' Bias? An Empirical Support from US States Data. *PLoS ONE* 5.
- Salvia JA, Meisel CJ (1980) Observer bias - A methodological consideration in special-education research. *J Spec Educ* 14: 261–270.
- Glass DJ (2007) Experimental design for biologists: Cold Spring Harbor Laboratory Press.
- Tillberg CV, Breed MD, Hinners SJ (2007) Field and Laboratory Exercises in Animal Behavior. London: Elsevier.
- Krauth J (2000) Experimental design: a handbook and dictionary for medical and behavioral research: Elsevier.
- Sani F, Todman J, Todman JB (2006) Experimental design and statistics for psychology: a first course: Blackwell Pub.
- Bernard C, Greene HC, Henderson LJ (1957) An Introduction to the Study of Experimental Medicine: Dover Publications.
- Romesburg HC (2009) Best Research Practices: Lulu.com.
- Gamboa GJ, Reeve HK, Holmes WG (1991) Conceptual issues and methodology in kin recognition research, A critical discussion. *Ethology* 88: 109–127.
- Rosenthal R, Fode KL (1963) The effect of experimenter bias on the performance of the albino-rat. *Behav Sc* 8: 183–189.
- Marsh DM, Hanlon TJ (2007) Seeing what we want to see: Confirmation bias in animal behavior research. *Ethology* 113: 1089–1098.
- Cordaro L, Ison JR (1963) Psychology of the scientist. 10. Observer bias in classical-conditioning of the planarian. *Psychol Rep* 13: 787–789.
- van Wilgenburg E, Ryan D, Morrison P, Marriott PJ, Elgar MA (2006) Nest- and colony-mate recognition in polydomous colonies of meat ants (*Iridomyrmex purpureus*). *Naturwissenschaften* 93: 309–314.
- van Zweden JSv, Dreier S, d'Ettorre P (2009) Disentangling environmental and heritable nestmate recognition cues in a carpenter ant. *J Insect Physiol* 55: 158–163.
- Foitzik S, Sturm H, Pusch K, D'Ettorre P, Heinze J (2007) Nestmate recognition and intraspecific chemical and genetic variation in Temnothorax ants. *Anim Behav* 73: 999–1007.

25. Menzel F, Schmitt T, Blüthgen N (2009) Intraspecific nestmate recognition in two parabolic ant species: acquired recognition cues and low inter-colony discrimination. *Insect Soc* 56: 251–260.
26. Sunamura E, Hatsumi S, Karino S, Nishisue K, Terayama M, et al. (2009) Four mutually incompatible Argentine ant supercolonies in Japan: inferring invasion history of introduced Argentine ants from their social structure. *Biol Invasions* 11: 2329–2339.
27. van Zweden JS, d'Ettorre P (2010) Nestmate recognition in social insects and the role of hydrocarbons. In: Blomquist GJ, Bagnères A-G, editors. *Insect Hydrocarbons: Biology, Biochemistry, and Chemical Ecology*. Cambridge: Cambridge University Press.
28. Vander Meer RK, Morel L (1998) Nestmate recognition in ants. In: Vander Meer RK, Breed M, Winston M, Espelie KE, editors. *Pheromone communication in social insects*. Boulder, CO: 368 p.: Westview Press. pp. 79–103.
29. d'Ettorre P, Lenoir A (2009) Nestmate and kin recognition. In: Lach L, Parr C, Abbott K, editors. *Ant Ecology*. Oxford: Oxford University Press.
30. Roulston TH, Buczkowski G, Silverman J (2003) Nestmate discrimination in ants: effect of bioassay on aggressive behavior. *Insect Soc* 50: 151–159.
31. Breed MD (2003) Nestmate Recognition Assays as a Tool for Population and Ecological Studies in Eusocial Insects: A Review. *J Kansas Entomol Soc* 76: 539–550.
32. Gurevitch J, Morrow LL, Wallace A, Walsh JS (1992) A Meta-analysis of competition in field experiments. *Am Nat* 140: 539–572.
33. Thomas ML, Payne-Makrisa CM, Suarez AV, Holway DA (2006) When supercolonies collide: territorial aggression in an invasive and uniclonal social insect. *Mol ecol* 15: 4303–4315.
34. Vasquez GM, Silverman J (2008) Intraspecific aggression and colony fusion in the Argentine ant. *Anim Behav* 75: 583–593.
35. Thomas ML, Parry LJ, Allan RA, Elgar MA (1999) Geographic affinity, cuticular hydrocarbons and colony recognition in the Australian meat ant *Iridomyrmex purpureus*. *Naturwissenschaften* 86: 87–92.
36. van Wilgenburg E, Clemencet J, Tsutsui ND (2009) Experience influences aggressive behaviour in the Argentine ant. *Biol letters*.
37. Rosenberg MS, Adams DC, Gurevitch J (1999) MetaWin: Statistical Software for Meta-analysis. 2 ed. Sunderland, Massachusetts: Sinauer associates.
38. Gurevitch J, Hedges LV (2001) Meta-analysis: combining the results of independent experiments. In: Scheiner SM, Gurevitch J, editors. *Design and analysis of ecological experiments*. New York: Oxford University Press.
39. Raudenbush SW (1994) Analyzing effect sizes: Random effects models. In: Cooper H, Hedges LV, editors. *The Handbook of Research Synthesis* New York: Russell Sage Foundation. pp. 302–332.
40. van Wilgenburg E, Dang S, Forti AL, Koumoundouros TJ, Ly A, et al. (2007) An absence of aggression between non-nestmates in the bull ant *Myrmecia nigriceps*. *Naturwissenschaften* 94: 787–790.
41. Hsu YY, Wolf LL (1999) The winner and loser effect: integrating multiple experiences. *Anim Behav* 57: 903–910.
42. Hsu YY, Wolf LL (2001) The winner and loser effect: What fighting behaviours are influenced? *Anim Behav* 61: 777–786.
43. Elgar MA (1989) Predator vigilance and group-size in mammals and birds - A critical-review of the empirical-evidence. *Biol Rev* 64: 13–33.
44. Zuk M (2003) Sexual selections: what we can and can't learn about sex from animals: University of California Press.
45. Hergovich A, Schott R, Burger C (2010) Biased Evaluation of Abstracts Depending on Topic and Conclusion: Further Evidence of a Confirmation Bias Within Scientific Psychology. *Curr Psychol* 29: 188–209.
46. Breed MD, Stiller TM, Fewell JH, Harrison JM (1991) Intercolonial interactions and nestmate discrimination in the giant tropical ant, *Paraponera clavata*. *Biotropica* 23: 301–306.
47. Jaffe K, Marcuse M (1983) Nestmate recognition and territorial behaviour in the ant *Odontomachus bauri* Emery (Formicidae: Ponerinae). *Insect Soc* 30: 466–481.
48. Lucas C, Pho DB, Fresneau D, Jallon JM (2005) Role of cuticular hydrocarbons in the chemical recognition between ant species in the *Pachycondyla villosa* species complex. *J Insect Physiol* 51: 1148–1157.
49. Mintzer A (1982) Nestmate recognition and incompatibility between colonies of the acacia-ant *Pseudomyrmex ferruginea* [sic]. *Behav Ecol Sociobiol* 10: 165–168.
50. Starks PT, Watson RE, Matthew MJ, Dipaola CP (1998) The effect of queen number on nestmate discrimination in the facultatively polygynous ant *Pseudomyrmex pallidus* (Hymenoptera: Formicidae). *Ethology* 104: 573–584.
51. van Wilgenburg E (2007) The influence of relatedness, neighbourhood and overall distance on colony mate recognition in a polydomous ant. *Ethology* 113: 1185–1191.
52. van Wilgenburg E, van Lieshout E, Elgar MA (2005) Conflict resolution strategies in meat ants (*Iridomyrmex purpureus*): ritualised displays versus lethal fighting. *Behaviour* 142: 701–716.
53. van Wilgenburg E, Ryan D, Morrison P, Marriott PJ, Elgar MA (2006) Nest- and colony-mate recognition in polydomous colonies of meat ants (*Iridomyrmex purpureus*). *Naturwissenschaften* 93: 309–314.
54. Caldera EJ, Holway DA (2004) Evidence that queens do not influence nestmate recognition in Argentine ants. *Insect Soc* 51: 109–112.
55. Bjorkman-Chiswell BT, van Wilgenburg E, Thomas ML, Swearer SE, Elgar MA (2008) Absence of aggression but not nestmate recognition in an Australian population of the Argentine ant *Linepithema humile*. *Insect Soc* 55: 207–212.
56. Sunamura E, Hatsumi S, Karino S, Nishisue K, Terayama M, et al. (2009) Four mutually incompatible Argentine ant supercolonies in Japan: inferring invasion history of introduced Argentine ants from their social structure. *Biol Invasions* 11: 2329–2339.
57. Thomas ML, Payne-Makrisa CM, Suarez AV, Tsutsui ND, Holway DA (2006) When supercolonies collide: territorial aggression in an invasive and uniclonal social insect. *Mol Ecol* 15: 4303–4315.
58. Vásquez GM, Silverman J (2008) Queen acceptance and the complexity of nestmate discrimination in the Argentine ant. *Behav Ecol Sociobiol* 62: 537–548.
59. Breed MD, Abel P, Bleuze TJ, Denton SE (1990) Thievery, home ranges, and nestmate recognition in *Ectatomma ruidum*. *Oecologia* 84: 117–121.
60. Breed MD, Snyder LE, Lynn TL, Morhart JA (1992) Acquired chemical camouflage in a tropical ant. *Anim Behav* 44: 519–523.
61. Zinck L, Hora RR, Châline N, Jaisson P (2008) Low intraspecific aggression level in the polydomous and facultative polygynous ant *Ectatomma tuberculatum*. *Entomol Exp Appl* 126: 211–216.
62. Drescher J, Blüthgen N, Feldhaar H (2007) Population structure and intraspecific aggression in the invasive ant species *Anoplolepis gracilipes* in Malaysian Borneo. *Mol Ecol* 16: 1453–1465.
63. Bos N, Guerrieri FJ, d'Ettorre P (2010) Significance of chemical recognition cues is context dependent in ants. *Anim Behav* 80: 839–844.
64. Strocymeyt N, Guerrieri FJ, van Zweden JS, d'Ettorre P (2010) Rapid Decision-Making with Side-Specific Perceptual Discrimination in Ants. *PLoS ONE* 5.
65. van Zweden JS, Dreier S, d'Ettorre P (2009) Disentangling environmental and heritable nestmate recognition cues in a carpenter ant. *J Insect Physiol* 55: 158–163.
66. Vicente Hernandez J, Rodriguez D, Tarano Z, Drijfhout FP, Sainz-Borgo C (2010) Are Postpharyngeal or Mandibular Gland Secretions the Source for Nestmate Recognition in *Camponotus atriceps* (Hymenoptera: Formicidae)? *Sociobiol* 56: 775–793.
67. Boulay R, Cerdá X, Simon T, Roldan M, Hefetz A (2007) Intraspecific competition in the ant *Camponotus cruentatus*: should we expect the 'dear enemy' effect? *Anim Behav* 74: 985–993.
68. Boulay R, Lenoir A (2001) Social isolation of mature workers affects nestmate recognition in the ant *Camponotus fellah*. *Behav Process* 55: 67–73.
69. Katzav-Gozansky T, Boulay R, Vander Meer R, Hefetz A (2004) In-nest environment modulates nestmate recognition in the ant *Camponotus fellah*. *Naturwissenschaften* 91: 186–190.
70. Katzav-Gozansky T, Boulay R, Ionescu-Hirsh A, Hefetz A (2008) Nest volatiles as modulators of nestmate recognition in the ant *Camponotus fellah*. *J Insect Physiol* 54: 378–385.
71. Moore D, Liebig J (2010) Mixed messages: fertility signaling interferes with nestmate recognition in the monogynous ant *Camponotus floridanus*. *Behav Ecol Sociobiol* 64: 1011–1018.
72. Morel L, Vander Meer RK, Lavine BK (1988) Ontogeny of nestmate recognition cues in the red carpenter ant (*Camponotus floridanus*)—behavioral and chemical evidence for the role of age and social experience. *Behav Ecol Sociobiol* 22: 175–183.
73. Liu Z, Yamane S, Wang Q, Yamamoto H (1998) Nestmate recognition and temporal modulation in the patterns of cuticular hydrocarbons in natural colonies of Japanese carpenter ant *Camponotus japonicus* Mayr (Hymenoptera: Formicidae). *J Ethol* 16: 57–65.
74. Jaffe K, Sanchez C (1984) On the nestmate-recognition system and territorial marking behaviour in the ant *Camponotus rufipes*. *Insect Soc* 31: 302–315.
75. Bonavita-Cougourdan A, Clément JL, Lange C (1987) Nestmate recognition: the role of cuticular hydrocarbons in the ant *Camponotus vagus* Scop. *J Entomol Sci* 22: 1–10.
76. Satoh T, Hirota T (2005) Factors affecting internet variation in the aggressiveness of a polygynous ant, *Camponotus yamaokai*. *Entomol Sci* 8: 277–281.
77. Nowbahari M, Lenoir A (1989) Age related changes in aggression in ant *Cataglyphis cursor* (Hymenoptera, Formicidae): Influence on intercolonial relationships. *Behav Process* 18: 173–181.
78. Nowbahari E, Lenoir A, Clément JL, Lange C, Bagnères AG, et al. (1990) Individual, geographical and experimental variation of cuticular hydrocarbons of the ant *Cataglyphis cursor* (Hymenoptera: Formicidae): their use in nest and subspecies recognition. *Biochem Syst Ecol* 18: 63–74.
79. Dahbi A, Lenoir A (1998) Nest separation and the dynamics of the Gestalt odor in the polydomous ant *Cataglyphis iberica* (Hymenoptera, Formicidae). *Behav Ecol Sociobiol* 42: 349–355.
80. Lahav S, Soroker V, Hefetz A, Vander Meer RK (1999) Direct behavioral evidence for hydrocarbons as ant recognition discriminators. *Naturwissenschaften* 86: 246–249.
81. Katzerke A, Neumann P, Pirk CWW, Bliss P, Moritz RFA (2006) Seasonal nestmate recognition in the ant *Formica exsecta*. *Behav Ecol Sociobiol* 61: 143–150.
82. Martin SJ, Viitkainen E, Helanterä H, Drijfhout FP (2008) Chemical basis of nest-mate discrimination in the ant *Formica exsecta*. *Proc Roy Soc B* 275: 1271–1278.

83. Martin SJ, Helantera H, Kiss K, Lee YR, Drijfhout FP (2009) Polygyny reduces rather than increases nestmate discrimination cue diversity in *Formica exsecta* ants. *Insect Soc* 56: 375–383.
84. Akino T, Yamamura K, Wakamura S, Yamaoka R (2004) Direct behavioral evidence for hydrocarbons as nestmate recognition cues in *Formica japonica* (Hymenoptera: Formicidae). *Appl Entomol Zool* 39: 381–387.
85. Henderson G, Andersen JF, Phillips JK, Jeanne RL (1990) Intra-nest aggression and identification of possible nestmate discrimination pheromones in polygynous ant *Formica montana*. *J Chem Ecol* 16: 2217–2228.
86. Beye M, Neumann P, Chapuisat M, Pamilo P, Moritz RFA (1998) Nestmate recognition and the genetic relatedness of nests in the ant *Formica pratensis*. *Behav Ecol Sociobiol* 43: 67–72.
87. Pirk CWW, Neumann P, Moritz RFA, Pamilo P (2001) Intra-nest relatedness and nestmate recognition in the meadow ant *Formica pratensis* (R.). *Behav Ecol Sociobiol* 49: 366–374.
88. van Zweden JS, Brask JB, Christensen JH, Boomsma JJ, Linksvayer TA, et al. (2010) Blending of heritable recognition cues among ant nestmates creates distinct colony gestalt odours but prevents within-colony nepotism. *J Evol Biol* 23: 1498–1508.
89. Rosset H, Schwander T, Chapuisat M (2007) Nestmate recognition and levels of aggression are not altered by changes in genetic diversity in a socially polymorphic ant. *Anim Behav* 74: 951–956.
90. Ugelvig LV, Drijfhout FP, Kronauer DJC, Boomsma JJ, Pedersen JS, et al. (2008) The introduction history of invasive garden ants in Europe: integrating genetic, chemical and behavioural approaches. *BMC Biol* 6:11: 20 p.
91. Newey PS, Robson SKA, Crozier RH (2008) Near-infrared spectroscopy as a tool in behavioural ecology: a case study of the weaver ant, *Oecophylla smaragdina*. *Anim Behav* 76: 1727–1733.
92. Thurin N, Aron S (2008) Seasonal nestmate recognition in the polydomous ant *Plagiolepis pygmaea*. *Anim Behav* 75: 1023–1030.
93. Dimarco RD, Farji-Brener AG, Premoli AC (2010) Dear enemy phenomenon in the leaf-cutting ant *Acromyrmex lobicornis*: behavioral and genetic evidence. *Behav Ecol* 21: 304–310.
94. Richard EJ, Hefetz A, Christides JP, Errard C (2004) Food influence on colonial recognition and chemical signature between nestmates in the fungus-growing ant *Acromyrmex subterraneus subterraneus*. *Chemoecology* 14: 9–16.
95. Ichinose K, Cerda X, Jean-Philippe C, Lenoir A (2005) Detecting nestmate recognition patterns in the fission-performing ant *Aphaenogaster senilis*: A comparison of different indices. *J Insect Behav* 18: 633–650.
96. Lenoir A, Cuisset D, Hefetz A (2001) Effects of social isolation on hydrocarbon pattern and nestmate recognition in the ant *Aphaenogaster senilis* (Hymenoptera, Formicidae). *Insect Soc* 48: 101–109.
97. Singer TL, Espelie KE (1998) Nest and nestmate recognition by a fungus-growing ant, *Apterostigma collare* Emery (Hymenoptera: Formicidae). *Ethology* 104: 929–939.
98. Debout G, Provost E, Renucci M, Tirard A, Schatz B, et al. (2003) Colony structure in a plant-ant: behavioural, chemical and genetic study of polydomy in *Catantolus mckeyi* (Myrmicinae). *Oecologia* 137: 195–204.
99. Stuart RJ (1991) Nestmate recognition in lepto thoracine ants: testing for effects of queen number, colony size and species of intruder. *Anim Behav* 42: 277–284.
100. Stuart RJ (1992) Nestmate recognition and the ontogeny of acceptability in the ant, *Leptothorax curvispinosus*. *Behav Ecol Sociobiol* 30: 403–408.
101. Stuart RJ, Herbers JM (2000) Nestmate recognition in ants with complex colonies: Within- and between-population variation. *Behav Ecol* 11: 676–685.
102. Stuart RJ (1993) Differences in aggression among sympatric, facultatively polygynous *Leptothorax* ant species. *Anim Behav* 46: 809–812.
103. Schmidt AM, d'Ettorre P, Pedersen JS (2010) Low levels of nestmate discrimination despite high genetic differentiation in the invasive pharaoh ant. *Front Zool* 7:20: 14 p.
104. Furst M, Durey M, Nash DR (2011) Testing the adjustable threshold model for intruder recognition on *Myrmica* ants in the context of a social parasite. *Proc Roy Soc B* Online early.
105. Garnas JR, Drummond FA, Groden E (2007) Intercolony aggression within and among local populations of the invasive ant, *Myrmica rubra* (Hymenoptera: Formicidae), in coastal Maine. *Environ Entomol* 36: 105–113.
106. Fournier D, De Biseau JC, Aron S (2009) Genetics, behaviour and chemical recognition of the invading ant *Pheidole megacephala*. *Mol Ecol* 18: 186–199.
107. Fadamiro HY, He X, Chen L (2009) Aggression in imported fire ants: an explanation for shifts in their spatial distributions in Southern United States? *Ecol Entomol* 34: 427–436.
108. Balas MT, Adams ES (1996) Nestmate discrimination and competition in incipient colonies of fire ants. *Anim Behav* 51: 49–59.
109. Morel L, Vander Meer RK, Lofgren CS (1990) Comparison of nestmate recognition between monogynous and polygynous populations of *Solenopsis invicta* (Hymenoptera: Formicidae). *Ann Entomol Soc Am* 83: 642–647.
110. Obin MS (1986) Nestmate recognition cues in laboratory and field colonies of *Solenopsis invicta* Buren (Hymenoptera: Formicidae): Effect of environment and the role of cuticular hydrocarbons. *J Chem Ecol* 12: 1965–1975.
111. Obin MS, Morel L, Vander Meer RK (1993) Unexpected, well-developed nestmate recognition in laboratory colonies of polygynous imported fire ants (Hymenoptera: Formicidae). *J Insect Behav* 6: 579–589.
112. Foitzik S, Sturm H, Pusch K, D'Ettorre P, Heinze J (2007) Nestmate recognition and intraspecific chemical and genetic variation in *Temnothorax* ants. *Anim Behav* 73: 999–1007.
113. Astruc C, Malosse C, Errard C (2001) Lack of intraspecific aggression in the ant *Tetramorium bicarinatum*: a chemical hypothesis. *J Chem Ecol* 27: 1229–1248.