

Binding, Conformational Transition and Dimerization of Amyloid- β Peptide on GM1-Containing Ternary Membrane: Insights from Molecular Dynamics Simulation

Moutusi Manna^{1,2}, Chaitali Mukhopadhyay^{1*}

¹ Department of Chemistry, University of Calcutta, Kolkata, India, ² Department of Physics, Tampere University of Technology, Tampere, Finland

Abstract

Interactions of amyloid- β (A β) with neuronal membrane are associated with the progression of Alzheimer's disease (AD). Ganglioside GM1 has been shown to promote the structural conversion of A β and increase the rate of peptide aggregation; but the exact nature of interaction driving these processes remains to be explored. In this work, we have carried out atomistic-scale computer simulations (totaling 2.65 μ s) to investigate the behavior of A β monomer and dimers in GM1-containing raft-like membrane. The oligosaccharide head-group of GM1 was observed to act as scaffold for A β -binding through sugar-specific interactions. Starting from the initial helical peptide conformation, a β -hairpin motif was formed at the C-terminus of the GM1-bound A β -monomer; that didn't appear in absence of GM1 (both in fluid POPC and liquid-ordered cholesterol/POPC bilayers and also in aqueous medium) within the simulation time span. For A β -dimers, the β -structure was further enhanced by peptide-peptide interactions, which might influence the propensity of A β to aggregate into higher-ordered structures. The salt-bridges and inter-peptide hydrogen bonds were found to account for dimer stability. We observed spontaneous formation of intra-peptide D²³-K²⁸ salt-bridge and a turn at V²⁴GSN²⁷ region - long been accepted as characteristic structural-motifs for amyloid self-assembly. Altogether, our results provide atomistic details of A β -GM1 and A β -A β interactions and demonstrate their importance in the early-stages of GM1-mediated A β -oligomerisation on membrane surface.

Citation: Manna M, Mukhopadhyay C (2013) Binding, Conformational Transition and Dimerization of Amyloid- β Peptide on GM1-Containing Ternary Membrane: Insights from Molecular Dynamics Simulation. PLoS ONE 8(8): e71308. doi:10.1371/journal.pone.0071308

Editor: Jie Zheng, University of Akron, United States of America

Received: March 29, 2013; **Accepted:** June 27, 2013; **Published:** August 9, 2013

Copyright: © 2013 Manna, Mukhopadhyay. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the research fellowship to MM (09/028/0690)/2007-EMR-I) by the Council of Scientific and Industrial Research (CSIR), India and also by financial support to CM (SR/S1/PC-60/2009) from the Department of Science and Technology (DST), India. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: chaitalicu@yahoo.com

Introduction

Alzheimer's disease (AD) is a progressive cognitive decline that pathologically characterized by the presence of senile plaques within the gray matter of brain. The neuritic plaques contain filamentous aggregates of amyloid- β (A β) peptides [1], proteolytically cleaved from transmembrane amyloid precursor protein. A β is 39–43 residue-long peptide and the C-terminal properties of A β critically determines its aggregation potential [2]. While A β _{1–40} composes approximately 90% of total secreted A β , the highly amyloidogenic 42-residue form A β _{1–42} is the principal component of the diffuse deposits [1]. Amyloid fibrils were long suspected as proximate disease agents, but recent compelling evidences suggest that small soluble oligomers of A β are the major neurotoxic components [3,4]. A details structural characterization of early aggregation intermediates is thus essential for understanding the origin of AD and to develop therapeutic strategies. Although much is known about the molecular structure of A β -fibrils [5,6], which universally exhibit a characteristic cross β -structure; the transient and disordered nature of low-ordered oligomers has made it difficult to pin down their structural properties and the dynamics of assembly pathway [7,8]. The limited solubility and structural

heterogeneity of the early A β -aggregates stand in the way of crystallization and limits experimental outcome, that average over different morphologies. Though A β has garnered much scientific attention, many fundamental aspects, such as how and where these soluble peptides are transformed into toxic oligomers and the cellular mechanism of toxicity - still remain elusive.

It has been proposed that the interaction of A β with neuronal membrane is integral to its toxicity and progression of AD [9,10]. Over last few decades a great deal of efforts has been made to explore the behavior of A β within different lipid environments. Studies showed that A β could intercalate into the membrane and eventually cause severe membrane damage [11,12]. Subsequent nonspecific membrane leakage or specific ionic transport through channels could perturb ion homeostasis and might be directly related to A β cytotoxicity [11–14]. While there have been other reports revealing the significant catalytic role of membrane in A β fibrillogenesis [10,15]. Association of peptides on membrane surface increases the local peptide concentration and reduces their self-assembly dimension from three to two, thereby speed up the aggregation rate than would occur in solution [10,15]. The compositional/lateral heterogeneity of biological membrane ren-

ders the study of A β -membrane interactions extremely complicated and gives rise to controversial mechanisms of A β induced toxicity in neuronal cells.

Lipid rafts, the cholesterol and sphingolipids enriched highly ordered membrane microdomains, are potential modulators of A β production, aggregation and toxicity [9]. Gangliosides are important components of lipid rafts and are known to play key role in the formation of amyloid fibrils by A β [9]. A recent study showed that the presence of ganglioside enhances both the early pore formation and the fibril-dependent membrane fragmentation process [11]. These sialic acid containing glycosphingolipids are abundant in plasma membrane of neurons and are involved in numerous neurobiological events [16]. Yanagisawa et al. had identified monosialoganglioside GM1-bound A β from AD brain and suggested the involvement of GM1 not only in aggregate formation but also in neurotoxic events [17]. Thereafter, a series of spectroscopic studies had revealed direct binding interactions between GM1 and A β [18–21]. GM1 was shown to facilitate the conformational transition of A β and accelerate amyloid fibril formation [20–23] and also these fibrils exhibited greater cytotoxicity than fibrils formed in solution [24]. Further, the formation of A β -GM1 complex is facilitated in raft-like environments that are enriched in cholesterol [10,20]. However, despite rigorous experimental focus, the exact nature of GM1-A β interactions driving these processes, thus the underlying molecular-mechanism is not yet determined and the aggregation states of A β during these investigations are also unclear. Simulation is one of the most viable techniques to study complex biological systems and being widely used now a day [25–26]. A large number of simulation studies had investigated the structural properties of A β dimers/trimers in aqueous solution [8,27–31], but there are very limited computational attempts to focus on how they behave in presence lipid membranes [32–34], especially within their native raft-like environment. Some recent computational studies had investigated the interactions of pre-formed A β protofilaments or preassembled fibril-like A β oligomers with one-component lipid bilayer [35,36] or with self-assembled monolayers [37]; where these models were based on NMR-derived structures of A β -fibril with significant β -sheet content.

In the present work, we have performed MD simulation (totaling 2.65 μ s) to investigate the effects of GM1 on the accumulation, conformational transition and subsequent dimerization of full length A β _{1–42} - which are the crucial early events during oligomerisation of A β on membrane surface. The goal of the present work is twofold. First we investigated the interactions of monomeric A β with GM1-containing raft-like membrane, composed of GM1/Chol/POPC. For comparison, two control simulations of A β monomer were performed in absence of GM1: (i) in liquid-ordered (L_o) Chol/POPC (containing 25 mol% Chol) bilayer and (ii) in liquid-disordered (L_d) POPC membrane. Another reference system of A β _{1–42} monomer in water was also simulated. We observed that the carbohydrate headgroup of GM1 acted as binding sites for A β and induced a β -hairpin structure at the C-terminus of the peptide. The α -helix to β -strand conformational transition is considered as the key step in the amyloidogenic oligomerisation process, as amyloid peptides must acquire β -sheet conformation to aggregate and polymerize [12]. To further examine whether the GM1-induced structure can form stable A β -oligomers, we had studied the structure and association pattern of three different dimeric arrangements of A β at the interface of GM1/Chol/POPC bilayer. Dimer formation is the first step in aggregation and has shown to be adversely toxic to neurons [4]. The results of the present simulations are compared with the available experimental data where possible.

Methods

System Setup

Lipid bilayers. Three model membranes with different lipid compositions (Table 1) were used in this study. These bilayers with varying fluidity (Table S1 and Text S1) can represent different regions of neuronal membrane and each was equilibrated for 150 ns. (i) *Single-component POPC bilayer:* The bilayer composed of unsaturated POPC lipids (*sn*-1 chain 16:0 and *sn*-2 chain 18:1) represented the fluid-phase membrane at room temperature (as phase transition temperature of POPC is 268 K) [25]. (ii) *Chol/POPC bilayer:* With addition of 25 mol% of Chol, the bilayer became more densely packed, with significant increase in bilayer rigidity and ordering [38]. (iii) *GM1/Chol/POPC bilayer:* Gangliosides are abundant in neuronal membrane and constitute 5–10% of lipids on the outer leaflet of cell membrane [39]. GM1 is an anionic glycosphingolipid containing a large penta-saccharide (four neutral sugar groups and one sialic acid) head group (Figure S1). Initially, coordinates for the ternary lipid-mixture: GM1/Chol/POPC were taken from a previously published work [40], where we had reported the formation of highly ordered GM1-Chol lateral assemblies. For the current study, the initial ternary system was enlarged (total 224 lipids) and equilibrated for further 150 ns (Table 1). In accordance with the experimental findings, here GM1 was present only in the extracellular leaflet, while we had maintained an equal concentration (25 mol%) of Chol in both layers. Our ternary bilayer closely mimicked the composition of lipid-rafts and properties of this bilayer (including very high lipid-tail order and bilayer mechanical strength, Table S1 and Text S1) were supportive of its liquid-ordered nature.

A β -monomer model. A β peptide [D¹AEFRHDS-GY¹⁰EVHHQKLVFF²⁰AEDVGSNK-GA³⁰IIGLMVGGV-V⁴⁰IA] is comprised of hydrophilic N-terminus (residues: 1–28) and hydrophobic C-terminus (residues: 29–40). The starting coordinates of full-length A β _{1–42} were taken from PDB entry: 1Z0Q [41]. This aqueous solution structure of A β _{1–42} derived by solution NMR consisted of two helical domains (residues: 10–22 and 28–32) connected by a turn (residues: 25–26) (Figure S2a). To construct the A β -monomer-membrane systems, peptide was then placed parallel on the top of equilibrated bilayer surface, approximately 35 Å away from the average phosphate plane of the contact monolayer (Figure S2b). A β -membrane systems were energy minimized, followed by simulation with position restraint on the peptide heavy atom and then subjected to 250 ns long unrestraint production run (Table 1). In addition, A β -monomer was also simulated for 250 ns in aqueous solution (Table 1).

Initial dimer structure. The major problem associated with the structural characterization of amyloid aggregates is their heterogeneity. Amyloid fibrils can adopt distinct morphologies differing in shape, size and association pattern (parallel and antiparallel β -sheet alignment) of A β units [5,6]. Compared to fibrils/protofibrils, small oligomers of A β are less ordered and their structures are highly heterogeneous, including significant β -sheet, α -helix and unstructured sections [8,27–29]. Despite ambiguities, emerging findings from earlier studies conjectured that – the hydrophobic C-terminus of A β peptide tends to adopt β -structure and form the central hydrophobic core, while the hydrophilic N-terminus is more solvent exposed [5,6]. Though varieties of biophysical techniques provide valuable information about A β oligomers, their high-resolution structures are still not available. Previous MD simulations had revealed that the small oligomers, like dimers, trimers etc., are capable of adopting diverse variety of intra-/inter-molecular parallel and antiparallel stacking configu-

Table 1. The systems under MD simulation study.

Systems	Number of each components							Simulation time (ns)
	A β	POPC	Chol	GM1	Water	Na ⁺	Cl ⁻	
POPC bilayer	–	128	–	–	6014	16	16	150
Chol/POPC bilayer	–	116	38	–	7170	19	19	150
GM1/Chol/POPC bilayer	–	161	56	7	8790	30	23	150
A β_{42} /Water	1	–	–	–	6929	22	19	250
A β_{42} /POPC	1	128	–	–	6740	21	18	250
A β_{42} /Chol/POPC	1	116	38	–	8757	26	23	250
A β_{42} /GM1/Chol/POPC	1	161	56	7	9555	35	25	250
Dimer1/GM1/Chol/POPC	2	161	56	7	10402	40	27	400
Dimer2/GM1/Chol/POPC	2	161	56	7	10312	40	27	400
Dimer3/GM1/Chol/POPC	2	161	56	7	10704	41	28	400

doi:10.1371/journal.pone.0071308.t001

rations, with possible lateral and frontal packing of peptide monomers [8,27–32,42].

Based on the previous findings, A β_{42} -dimers with three different packing configurations were modeled in the present work (Figure S3). Our final structure of A β_{42} monomer bound to GM1/Chol/POPC bilayer served as starting configuration for A β_{42} units in dimers. Initially, each A β was separated by minimum center of mass distance of 1.6 nm. For first two cases, peptides were placed side-by-side, where their C-terminal β -hairpin segments were kept in adjacent positions, with antiparallel (dimer 1, Figure S3a) and parallel (dimer 2, Figure S3c) orientations. While in another dimer, peptides were stacked on the top of each other (dimer 3, Figure S3e). Dimers were then placed above (\sim 2.2 nm) the surface of GM1/Chol/POPC bilayer (Figure S3b,d,f), energy minimized to remove clashes within the system, equilibrated with position restraint on the peptide and finally followed by 400 ns unrestrained production run for each of the dimer-membrane systems (Table 1).

Parameters and Simulation Protocols

All simulations were performed using GROMACS 3.3.1 package [43,44] and the standard united-atom GROMOS87 force field [45]. The ffmx force field was used to represent peptide in conjunction with Berger lipids [46]. This set of parameters was shown to perform well in many membrane simulations [38,47–48]. The structure and topology of cholesterol molecule was taken from H \ddot{o} ltje et al. [49], with the atomic charge modifications by Olsen et al. [50]. Parameters for GM1 were taken from Segal et al. [51]. The Ryckaert-Bellemans potential was used to model the torsional angles of lipid hydrocarbon tails [52]. The SPC model was used for water molecules [53]. To maintain the electro-neutrality of the system Na⁺ counter ions were added and additional NaCl was added to achieve the physiological salt concentration. All MD simulations were carried out under the isobaric–isothermal (NPT) ensemble with imposed 3D periodic boundary conditions. A time step of 2 fs was used for integrating the equation of motion. The Berendsen algorithm [54] was employed to keep temperature (at 300 K, using coupling constant $\tau_T = 0.1$ ps) and pressure (semiisotropically at 1 bar, using coupling constant $\tau_P = 1.0$ ps) constant. Lennard-Jones interactions were cutoff at a distance of 1.2 nm. For long-range electrostatics we had employed the particle mesh Ewald (PME) method [55] with a real-space cutoff of 1.2 nm. LINCS algorithm

[56] was applied to constrain all bonds. VMD [57] and Pymol [58] were used for visualization.

Results and Discussion

A β -monomer Systems

Distribution of A β in bilayer environment. Figure 1 represents the final snapshots of membrane-bound A β in different bilayer environments. In GM1 containing membrane, A β resided at the oligosaccharide head group of GM1 (Figure 1a and Figure 2a). The voluminous carbohydrate moiety of GM1 prevented deeper insertion of the peptide to the bilayer core. The hydrophobic C-terminus of A β approached the membrane-water interface, while the rest of the peptide remained partly exposed to the aqueous environment (Figure 2b). Our results correlated well with ample experimental findings suggesting accumulation of A β in the carbohydrate region of ganglioside containing model membranes [19,59]. In a previous NMR spectroscopic study, the observed change in NMR signals from the sugar moiety indicated perturbations in exactly the region where A β was located [59]. For our simulation of GM1-depleted Chol/POPC bilayer, the interfacial accumulation of A β was also observed (Figure 1b and Figure 2c); where the central part (Lys¹⁶–Lys²⁸) of the peptide acted as glue to assist its surface association (Figure 2d). In contrary, the peptide got deeply inserted into the liquid-disordered POPC membrane (Figure 1c and Figure 2e). Many residues of the N-terminal segment of A β , including several charged/polar residues, penetrated well below (up to 5 Å) the average phosphate plane, with aromatic Phe¹⁹ and Phe²⁰ residues favorably immersed into membrane hydrophobic interior (Figure 2f). Such orientation might attribute to A β -induced membrane perturbation [60]. Thus the positioning of A β_{1-42} in membranes was found to be governed by the lipid-composition and consequently its strength of interaction with membrane was observed to vary in the order: POPC>Chol/POPC>GM1/Chol/POPC (Figure S4).

Lipid-peptide interactions: Involvement of ganglioside-GM1. To identify specific lipid-A β interactions, the hydrogen bond (H-bond) participation of the peptide with the surrounding lipid molecules (Figure 3a and Figure S5) were calculated. An H-bond was defined by an acceptor-hydrogen distance less than 2.8 Å and a donor-hydrogen-acceptor angle greater than 120° [38]. In GM1-containing membranes both GM1 and POPC formed direct H-bonds with A β (Figure 3a). The basic His¹³, polar

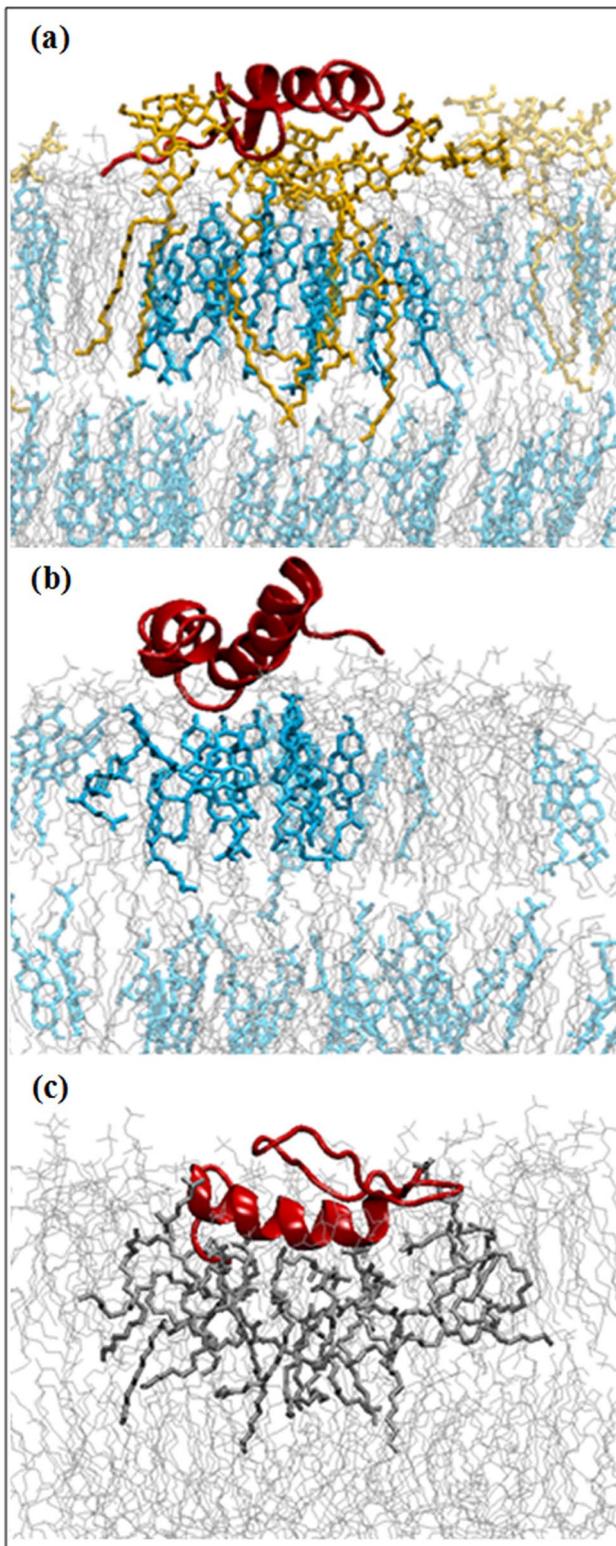


Figure 1. Final snapshots of membrane-bound A β ₁₋₄₂ monomer in different lipid bilayers. A β in (a) GM1/Chol/POPC, (b) Chol/POPC, and (c) POPC bilayers. Here, peptide was presented as red cartoon. Phospholipids, cholesterol and GM1 were shown in grey, cyan and orange, respectively. The lipids those were adjacent to the peptide made highlighted. Water molecules and ions were not shown for clarity. doi:10.1371/journal.pone.0071308.g001

Ser⁸ and C-terminal hydrophobic residues, like, Leu³⁴, Val⁴⁰, Ala⁴² of the peptide were H-bonded with GM1 sugar groups (Figure 3a and Table S2). While residues like, Asn²⁷, Lys²⁸, Val³⁹, those penetrated deeper into the membrane, formed H-bonds with polar head-groups of POPC (Figure 3a). Recently, His¹³/His¹⁴ has been identified as key residues for binding A β to GM1 [61,62]. The penta-saccharide headgroup of GM1 (Figure S1) can provide numerous sites for the hydrogen bonding to the peptide. Further scrutiny for the involvement of different sugar moieties showed that Neu5Ac (i.e. sialic acid) has the major contribution for binding A β to the membrane, while GalNAc also significantly participate in H-bonding (Figure 3b). Our findings were consistent with previous experiments that reported an important role of sialic acid in ganglioside-specificity of A β . [20]. In addition, we were also able to capture CH- π stacking interaction between Phe²⁰ and the terminal galactose ring (Figure 3c). The pyranosyl ring of galactose has two chemically distinct faces: one apolar face with hydrocarbyl (CH) groups and other polar face with -OH groups. The apolar side of galactose acted as complimentary surface for stacking interactions with aromatic residue (Figure 3c). The current findings were in line with earlier reports showing that the glycosphingolipid-binding domain should contain basic and aromatic residues to mediate specific glycosphingolipid-protein interactions [61].

In GM1-containing membrane, no direct interaction between Chol and A β was observed (Figure 3a). The hydroxyl (-OH) group of Chol was found to form several polar interactions with GM1 oligosaccharide head group as well as -NH and -OH of sphingosine and restricts the conformation of glycan moiety of GM1 to be significantly tilted towards membrane plane (Figure 3d). Fantini et al. had showed that such Chol-induced glycolipid conformation is crucial for optimum recognition of A β [63]. In absence of GM1 (in Chol/POPC bilayer), Chol directly interacted with A β and formed hydrogen bonds mainly with the basic Lys¹⁶ and Lys²⁸ residues of peptide (Figure S5). The charged residues may acted as hook to initiate the early binding of A β to the bilayer and found to be crucial for A β -membrane interaction [64–66].

Effects on membrane morphology. Binding of A β exerted reciprocal effects on membrane biophysical properties, which were depicted by (i) the in-plane distribution of $\Delta Z_i^P = Z_i^P - \langle Z^P \rangle$, where Z_i^P was Z-coordinates of the phosphorus atom of i^{th} POPC molecule in the contact monolayer and $\langle Z^P \rangle$ was the average Z-value of this surface (Figure 4a–c) [47] and (ii) the molecular order parameter, S_{mol} , of POPC acyl tails (Figure 4d–f). In POPC bilayer the deeper penetration of peptide strongly perturbed the local bilayer structure (Figure 4c,f). Lipids adjacent to the peptide become highly disordered, as depicted from the drastic drop in the order parameter values (Figure 4f). Simultaneously we observed formation of grooves or dents over a broad region underneath the peptide, with some portion of the bilayer even depressed up to 6–8 Å (Figure 4c). Such membrane-destabilizing effects of A β often can damage bilayer integrity and are known to be precursor to the formation of pore structures and ion channels. With the incorporation of 25 mol % Chol, the bilayer became more ordered and densely packed (Table S1); that prevented the deeper insertion of peptide (Figure 1b and Figure 2c) and protected membrane from A β -induced perturbation (Figure 4b,e). Since in Chol/POPC bilayer, A β was found to bind at the Chol-rich region (Figure 1b), two opposing effects were operating at the peptide contact area: (1) A β -induced membrane destabilization and (2) Chol-induced bilayer ordering; here the latter predominated over the other. Moreover the greater increase in the ordering of palmitoyl tail was probably due to the better packing of Chol

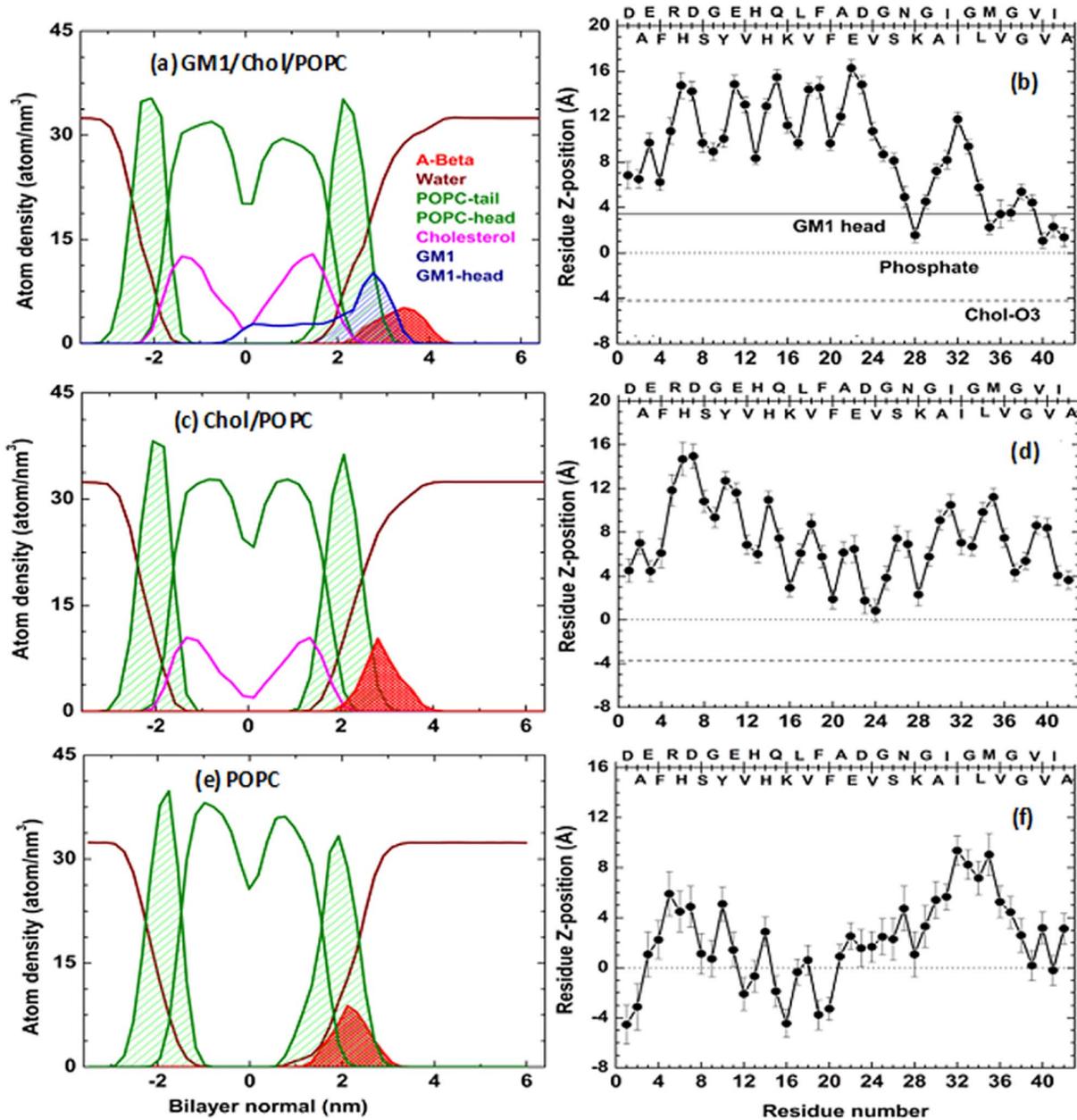


Figure 2. Location and orientation of A β -monomer at membrane-interfaces. (a,c,e) Atom density profiles (last 100 ns average) of peptide, lipids and water were plotted along Z-axis. For POPC (green) and GM1 (blue) the shaded area under the curve represented their head-group regions. (b,d,f) The average (last 100 ns) distance of the center of mass of peptide residues from bilayer interfaces. The horizontal lines represent the average planes of GM1 penta-saccharide head (solid line), Chol -OH oxygen (dashed line) and phosphorus atom of POPC (dotted line at Z=0 position). doi:10.1371/journal.pone.0071308.g002

around the saturated tail rather than the unsaturated one (Figure 4e). With addition of GM1 the rigidity of the ternary GM1/Chol/POPC bilayer was enhanced further (Table S1, Figure 4d). Here A β was found to bind at the interface of four GM1 molecules (Figure 4a) and remained partially solvent exposed. Such distribution of A β on membrane surface might increase the possibility of peptide-peptide interactions leading to aggregation. The interior of GM1-containing membrane remained almost unperturbed on A β invasion (Figure 4a,d). Thus, lipid-composition/bilayer fluidity can modulate the mode of A β -membrane interactions, responsible for the diverse mechanisms of A β -induced toxicity in neural cells.

Peptide conformation. The structural transition of A β is a central step in amyloidogenic oligomerisation process. To investigate the impact of lipid-composition on peptide secondary structure content, we have plotted the secondary structure profile of A β at various bilayer interfaces (Figure 5a,b,c), calculated on the basis of DSSP (defined secondary structure of protein) program [67]. In GM1-containing membrane, the N-terminal part almost retained its overall helicity (Figure 5c). While we observed the formation of an anti-parallel β -sheet near peptide C-terminus (Figure 5c,f) that remained immersed into the membrane-water interface, surrounded by GM1 sugar moieties (Figure S6). The β -sheet segment was formed by two β -strands composed of residues

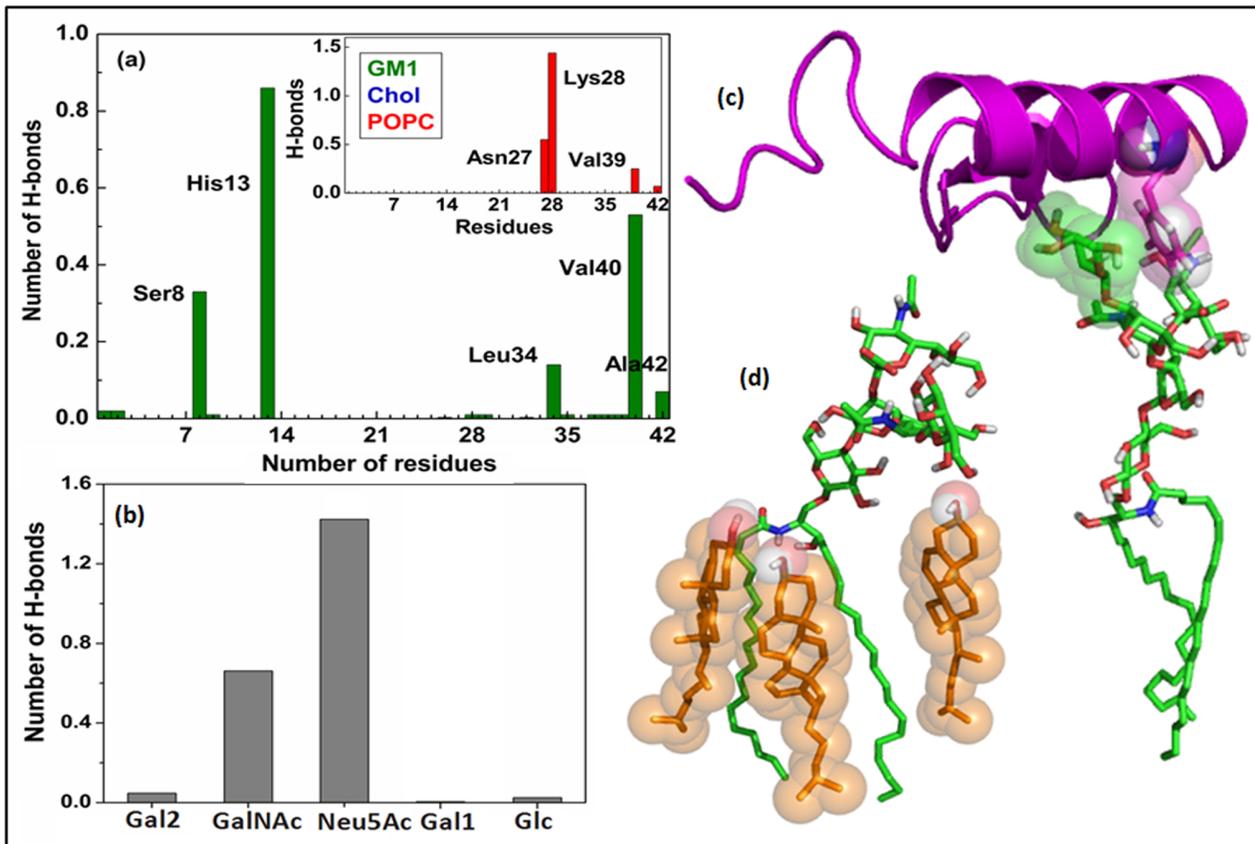


Figure 3. Interactions of A β with GM1-containing ternary bilayer. (a) Number of H-bonds formed by A β with GM1 (in green) and POPC (in red) in the ternary GM1/Chol/POPC bilayer. (b) Involvement of different sugar-residues in A β -GM1 interaction (the terminal galactose was termed as Gal2, whereas internal galactose was as Gal1). (c) Snapshot represents the CH- π stacking interaction between Phe²⁰ and the pyranosyl ring of terminal galactose. Here A β was shown in purple cartoon and GM1 as green sticks. (d) Figure showed how cholesterol (in orange) constrained GM1 head-group conformation.

doi:10.1371/journal.pone.0071308.g003

Ala³⁰-Ile³¹-Ile³² and residues Val³⁶-Gly³⁷, linked by a turn [residues: Gly³³-Leu³⁴-Met³⁵] (Figure 5c,f). The inter-strand H-bonds (backbone): between -NH of Val³⁶ with -C=O of Ile³² and also between -C=O of Val³⁶ with -NH of Ile³² (Figure 5g), stabilized the β -hairpin-like structure and allowed it to evolve over last 150 ns of simulation. In contrary to the GM1-containing membrane, in POPC (Figure 5a,d) and Chol/POPC bilayers (Figure 5b,e) we did not observe appearance of any β -strand component even after 250 ns of simulations. No significant β -structure was also formed in aqueous solution (Figure S7). It has been reported earlier that GM1 amplifies the α -helix to β -sheet conformational transition of A β and facilitates the formation of toxic β -sheet-rich fibrils [20,22]. Such conformational change of A β was not detected in presence of ganglioside-free vesicles composed of various phospholipids and sphingomyelin (as characterized by CD-spectroscopy) [21,68]. In presence of anionic lipid, a recent study had reported formation of small β -hairpin in A β [66]. The helix/ β -sheet mixed conformations of A β was previously predicted as possible intermediates for A β -oligomerisation [69]. However, the experimental studies were unable to distinguish the monomeric and different oligomeric states of A β . Owing to the high aggregation propensity of A β , small oligomers are hard to detect experimentally. Thus, to further investigate the role of GM1 in the early-stages of A β -aggregation, it was imperative to investigate whether the GM1-induced structure of

monomeric A β ₄₂ can trigger ordered oligomer formation with increased inter-peptide interactions.

A β ₄₂-dimers on GM1-containing Ternary Membrane

According to the nucleated polymerization model [70], the assembly of monomeric A β into oligomeric structure is crucial for amyloid-fibril formation. Here we have investigated the structure, association pattern and stabilizing forces of three different dimers on the surface of GM1-containing membrane. Our results showed that all dimers were stable over 400 ns simulation time scale and exhibited no tendency to get dissociated. All of these structures were well equilibrated, as can be seen by the leveling off of RMSD after 50 ns (Figure S8). These structures were stabilized by inter-peptide contact and hydrogen-bonding interactions and the initial separation distance between two A β was dropped down to <1.3 nm (Table S3). Energetically, Dimer1 exhibited strongest peptide-peptide interaction and was the most stable among all (Figure S9, Table S3 and Text S2). The results of Dimer1 were presented in the main text, while that of other two dimers were given in the supporting information. Moreover all three dimers were favorably accumulated on membrane surface (Figure S10, Table S4 and Text S3).

Secondary structure of A β ₄₂ dimers. To characterize the structural changes associated with the first step of assembly from monomeric to dimeric states, the secondary structure contents of

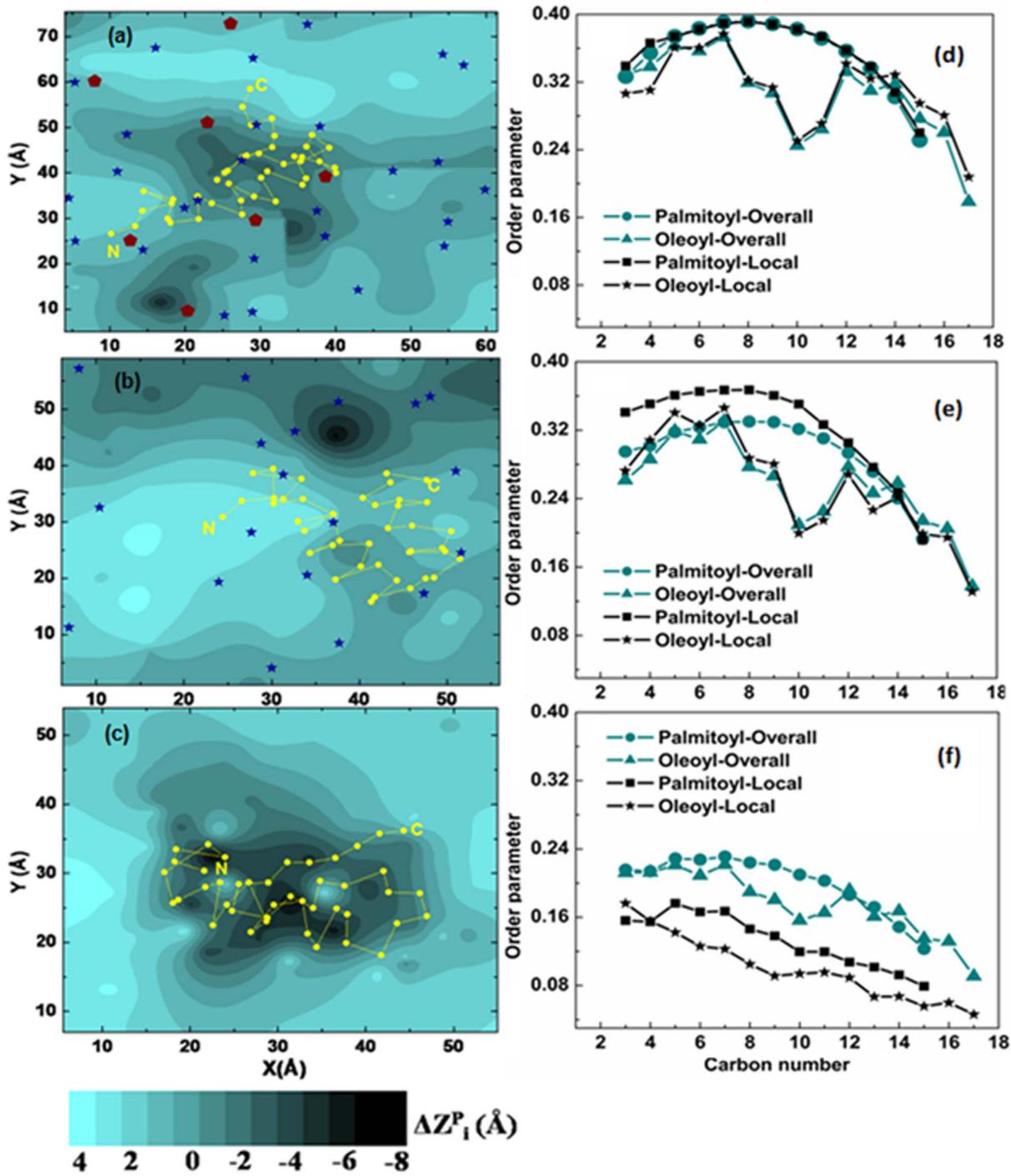


Figure 4. Effects of A β binding on bilayer properties. (a–c) The surface distribution of the change in bilayer thickness (defined in terms of Z coordinates of phosphorus atoms of individual POPC lipids as a function of their average in-plane positions) around A β in (a) GM1/Chol/POPC (b) Chol/POPC and (c) POPC bilayers. The average (X–Y) positions of peptide C $_{\alpha}$ atoms, Chol –OH oxygen and oxygen atom of glycosidic linkage of GM1 connecting ceramide tail with sugar head group, were shown in yellow (circle), blue (stars) and maroon (pentagons), respectively. (d–f) Order parameters of POPC lipid tails in (d) GM1/Chol/POPC (e) Chol/POPC and (f) POPC bilayers (last 100 ns average). To assess the effect of peptide, proximal lipids [i.e. local lipids, which with a lipid non-hydrogen atom within 10 Å of a peptide non-hydrogen atom] were considered separately. doi:10.1371/journal.pone.0071308.g004

dimers were plotted as function of time (Figure 6a, Figure S11a,c). We observed that significant amount β -structure persists for all dimers, along with turn and helical contents (Figure 6 and Figure S11). As depicted in Figure 6, dimerization appears to involve an increase in β -structure. For Dimer1, the β -content reached \sim 20%

during last 100 ns simulation (Figure 6a) and was in nice agreement with previous estimates: \sim 15–30% from Circular Dichroism (CD) and NMR spectroscopy [18,71] and \sim 15–26% from simulations [8,27]. All dimers were found to contain β -sheets at C-terminus formed between different sets of residues and with

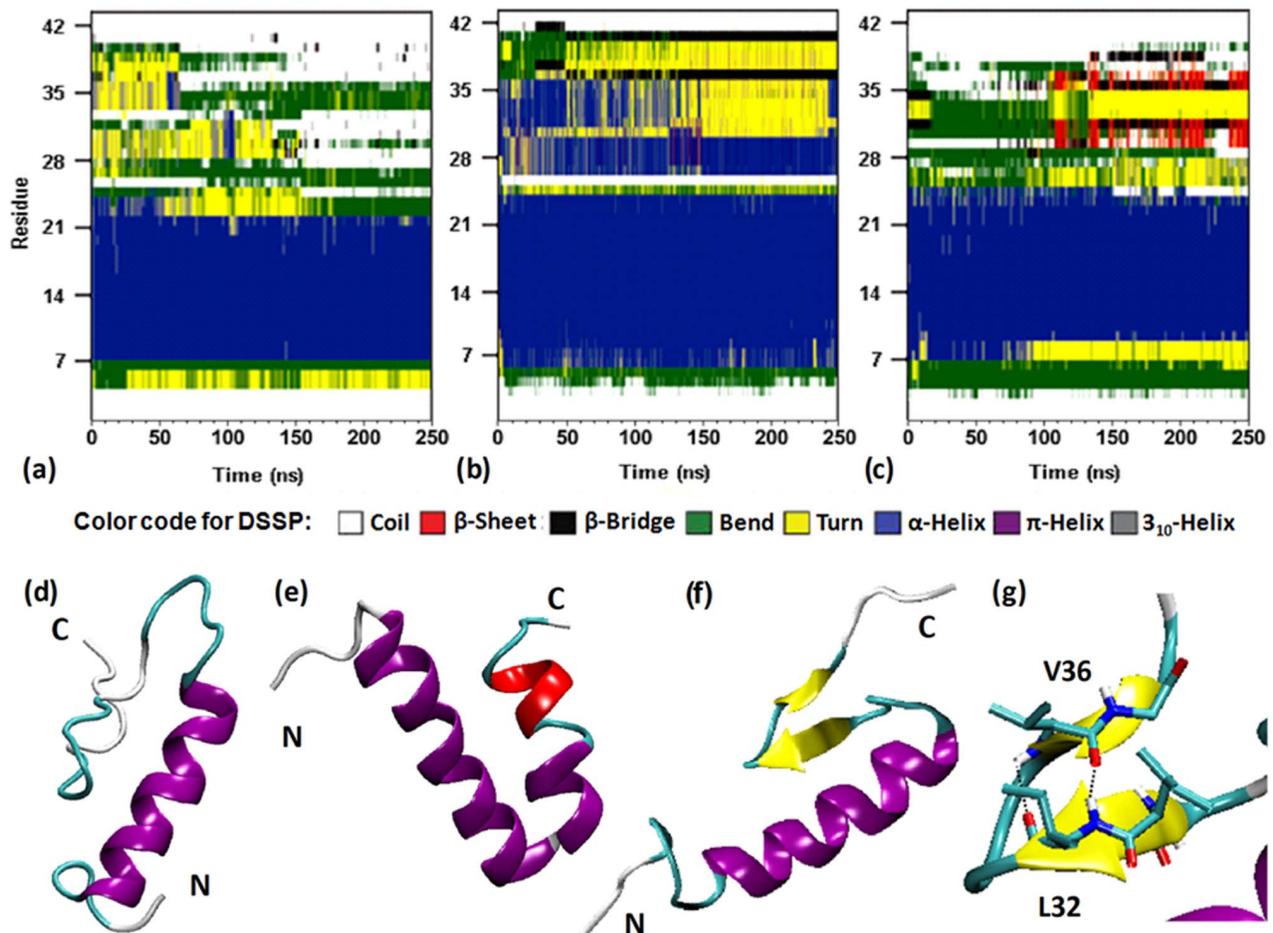


Figure 5. Membrane-induced structural conversion of A β -monomer. The secondary structure profiles of A β in (a) POPC, (b) Chol/POPC, and (c) GM1/Chol/POPC membranes. The final snapshots of A β on (d) POPC, (e) Chol/POPC, and (f) GM1/Chol/POPC bilayer surfaces. (g) H-bonds stabilize the β -hairpin structure. The default color code for DSSP plots (a-c)- random coil: white, β -sheet: red, β -bridge: black, bend: green, turn: yellow, α -helix: blue, π -helix: purple, 3_{10} -helix: gray. In snapshots (d-g), the secondary structures of A β were colored based on the default representation in VMD (α -helix: purple, 3_{10} -helix: blue, π -helix: red, extended- β : yellow, bridge- β : tan, turn: cyan, coil: white). doi:10.1371/journal.pone.0071308.g005

different interlocking patterns (Figure 6 and Figure S11). The average secondary structure probabilities of each residues of dimer1 were plotted in Figure 6b; where we observed the formation of well-reserved β -strands connected by short loops at peptides' C-terminus. Interestingly, the occurrence of turn at V²⁴-N²⁷ region (Figure 6b) was in nice agreement with solid-state NMR fibril studies and complimentary MD simulations [6,72]. The D²³-K²⁸ salt-bridge stabilizes the turn conformation (Figure 7a) (discussed in details later). The structure with intact D²³-K²⁸ salt-bridge and conserved V²⁴-G²⁵-N²⁷ turn has long been identified as aggregation-prone structure [72,73]. As depicted in Figure 6, β -strands in dimer were up to 5–7 residues long, hence were more extended than isolated monomers. Some β -strands were tending to be curved (Figure 6), perhaps lacking enough interactions to stabilize the flat extended sheets observed in mature fibrils. In a recent study the formation of a short segment of parallel intermolecular β -sheet was reported during assembly of three A β peptides in a mixed bilayer [33]. The small oligomers of A β do not have β -sheet structure characteristics of fibril, rather are composed of several shorter and loosely aggregated stands, which might polymerize in later stages during fibrillization [5].

Stabilizing forces. Characterization of driving forces for amyloid aggregation, in terms of key intra- and inter-peptide interactions, is an outstanding problem. According to the principal of amyloid self-assembly: maximizing hydrophobic and favorable electrostatic interactions enhanced fibril stability [73]. Salt bridges can play crucial role in oligomerisation and in stabilizing fibril-like structures. In the present work, we had calculated the formation time percentage of possible salt-bridges during the dimer trajectories and had listed in Table S5. A salt-bridge was considered when the distance between an amide N and a carboxyl O is less than 4.5 Å [38]. Interestingly, we observed spontaneous formation of intra-molecular D²³-K²⁸ salt-bridge in Dimer1 (Figure 7a,b) over 290 ns to 400 ns simulation time span. The D²³-K²⁸ salt-bridge was also found to be present in NMR-derived structures of A β -fibrils [5] and its importance has been emphasized in recent studies [73,74]. In addition, we found that intra-molecular E²²-K²⁸ salt-bridge (in Dimer1) as well as intermolecular salt-bridges between: K²⁸-E¹¹ and K²⁸-E³ in Dimer1; R⁵-E³ and R⁵-D⁷ in Dimer2 and R⁵-D¹, K¹⁶-D⁷ and K²⁸-E²² in Dimer3 were also populated (Table S5). Both intra- and inter-peptide charge-pairs have significant contributions in dimer stability.

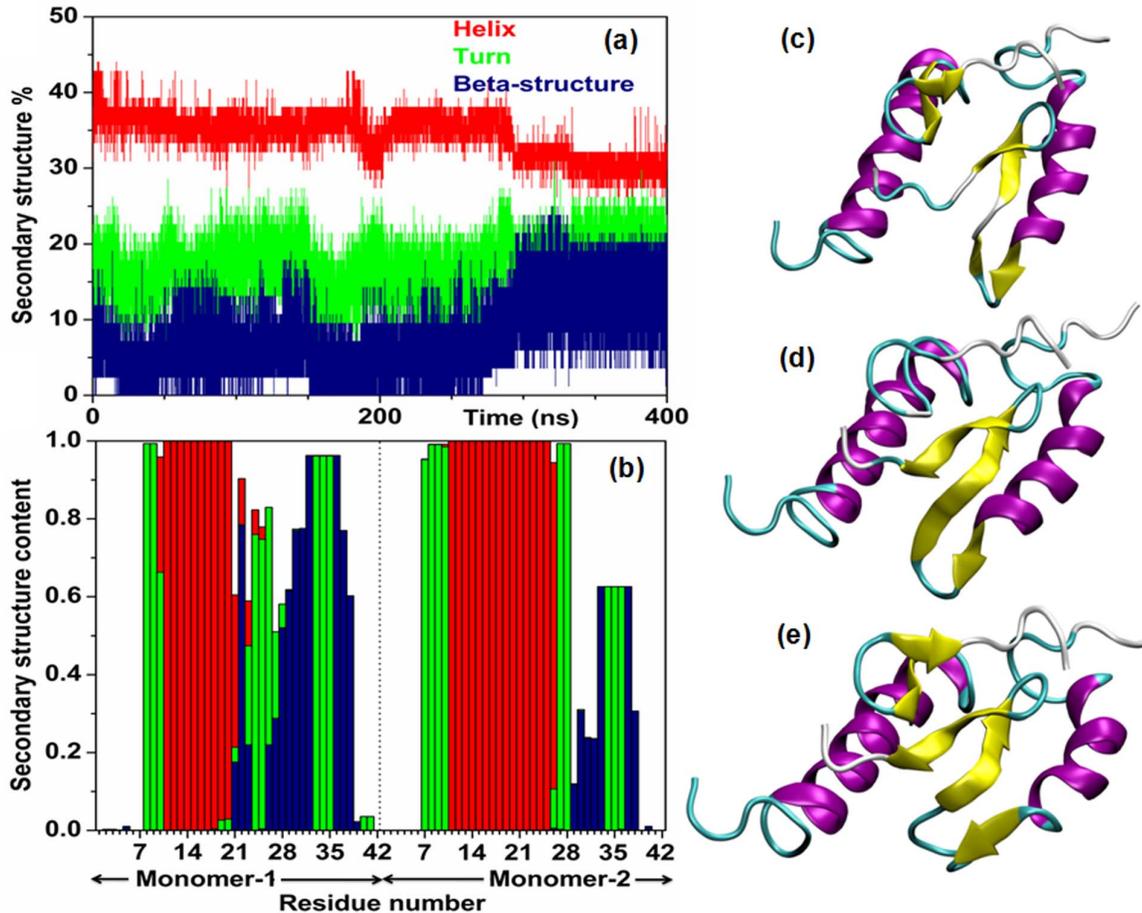


Figure 6. Secondary structures of A β -dimer. (a) Time profile of secondary structural contents and (b) average (last 125 ns) secondary structural content per residues of Dimer1 (calculated from DSSP); plotted with helix (sum of α -, 3_{10} - and π - helices) in red, turn in green and β -structure (sum of extended β -strands and isolated β -bridges) in blue. Snapshots of Dimer1 at (c) 200 ns, (d) 275 ns and (e) 400 ns. In snapshots, the secondary structures of A β were colored based on the default representation in VMD (α -helix: purple, 3_{10} -helix: blue, π -helix: red, extended- β : yellow, bridge- β : tan, turn: cyan, coil: white).

doi:10.1371/journal.pone.0071308.g006

The interface of dimer is characterized by electrostatic interactions. Figure 7c, Figure S12 and Figure S13 depict the inter-peptide hydrogen bonds, which helped two A β -monomers to stick together within the dimers. For Dimer1, the predominant H-bonds formed between: H¹³(1)-A⁴²(2) (numbers in brackets indicate peptide number within dimers, i.e., 1 for peptide-1 and 2 for peptide-2), A⁴²(1)-H¹³(2), N²⁷(1)-Y¹⁰(2) and L³⁴(1)-I⁴¹(2) (Figure 7c). The charged/polar residues of dimers, like, D^{1,7}, E^{3,11,22}, H^{6,13,14}, Y¹⁰, S²⁶, N²⁷ and K²⁸, exhibited high H-bonding propensity (Figure 7c, Figure S12 and Figure S13) and their side-chains were mainly involved in the bonding network (Table S6, Table S7 and Table S8). Additionally, we observed important participation of many hydrophobic residues (like, A^{2,21,30,42}, I^{31,41}, L³⁴, G³⁸ and V³⁹) in H-bond (main-chain) formation (Figure 7c, Figure S12, Figure S13 and Table S6–S8). Among them the C-terminus residues I⁴¹ and A⁴² are well known for their crucial role in the stability and toxicity of A β -fibrils [2]. Detailed knowledge about specific interactions may facilitate designing of possible inhibitors of A β -aggregation and developing therapeutic strategies against AD.

The hydrophobic, hydrophilic and total solvent-accessible surface area (SASA) for monomer and all dimers were also measured (Table S9). The significant drop in SASA values (by

16.85 nm² for Dimer1, 14.93 nm² for Dimer2 and 15.39 nm² for Dimer3) upon monomer to dimer conversion indicated favorable peptide-association. Together, all these factors conferred stability to the dimers and might be important for protofibril/fibril formation in later stages.

Conclusions

The behavior of A β -peptide within the cell membrane is integral to the manifestation of Alzheimer's disease. Membrane composition defines the A β -lipid interactions and has significant implications in the context of age- and disease-related evolution of brain lipid expression and A β -deposition in AD [61]. We found that A β buried itself into the liquid-disordered POPC membrane and significantly perturbed membrane structure as a consequence of strong peptide-lipid interactions. Inclusion of cholesterol enhanced bilayer rigidity and protected membrane from A β -induced disruption. With addition of GM1, bilayer became more densely packed. The oligosaccharide head group of GM1 acted as privileged sites for A β binding- aided by hydrogen-bonding and CH- π stacking interactions with nearby sugar moieties. Such distribution of A β at the membrane-water interface may promote further peptide-peptide interactions characteristic of aggregation. Notably, the GM1-containing membrane exhibited a significant

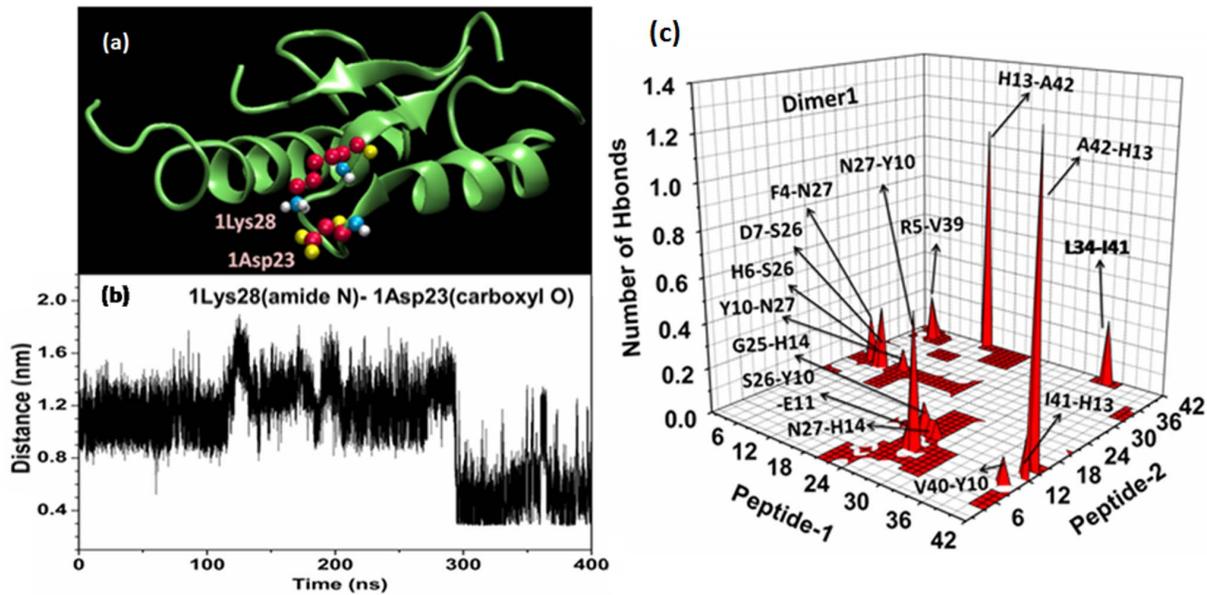


Figure 7. Factors accounting for dimer stability. (a) The snapshot showed the presence of intra-molecular Lys²⁸-Asp²³ salt-bridge in monomer-1 of Dimer1. Here the corresponding Lys²⁸ and Asp²³ were colored according to atom type: C in red, O in yellow, N in blue and H in white. (b) The time dependence of minimum distance between amide N of Lys²⁸ and carboxyl O of Asp²³ in monomer-1 of Dimer1, as highlighted in Figure 7a. (c) 3D-plot showing the inter-peptide hydrogen-bonding interactions within Dimer1 (last 200 ns average). The notation used here for leveling H-bond between a pair of residues was XA-YB, where X was the amino acid residue of peptide-1 with its corresponding residue number A and Y was the residue of peptide-2 with its corresponding residue number B. doi:10.1371/journal.pone.0071308.g007

control over the process of β -strand formation in A β . For GM1-bound A β , we observed spontaneous formation of β -hairpin at the C-termini (residues: Ala³⁰-Gly³⁷) of the peptide. As the toxic A β -oligomers and mature fibrils are rich in β -strand content, the formation of β -structure has long been considered to be central to the aggregation cascade.

For our simulations of A β -dimers on GM1-containing membrane, the overall β -strand content was found to enhance considerably upon A β -A β interaction. Amyloid formation is a slow process (needs several hours to days) [11,24] and still far beyond the reach of current atomistic simulation studies. However, it is worth noting that we had started with a helical peptide structure (PDB entry: 1Z0Q) without any β -strand content, while ended up with dimers having considerable β -structure, e.g., \sim 20% β -content in Dimer1- in nice agreement with previous experiments. Our results showed that dimerization was favored by several inter- and intra-peptide salt-bridges and peptide-peptide hydrogen bonds. These forces can influence the propensity of A β to aggregate into higher-ordered structure. In Dimer1, we observed spontaneous formation of key structural elements, like, intra-peptide D²³-K²⁸ salt-bridge and turn at V²⁴-GSN²⁷ region, which nucleate and stabilize the β -hairpin like structure and might act as template for larger oligomer growth. Altogether, the present work provides molecular level insight into the effects of ganglioside-GM1 on A β -membrane association, peptide's conformational transition and aggregation- the three most vital early stages of membrane-induced A β -fibrillogenesis.

Supporting Information

Figure S1 Structure of ganglioside GM1. GM1 [Gal β (1-3) GalNAc β (1-4) [Neu5Ac α (2-3)] Gal β (1-4) Glc β 1-Ceramide] contains an oligosaccharide head group composed of: glucose (Glc), internal galactose (termed as Gal1), *N*-acetylneur-

aminic acid (Neu5Ac) or sialic acid, *N*-acetylgalactosamine (GalNAc), and terminal galactose (termed as Gal2). (TIF)

Figure S2 Description of initial structures for A β -monomer systems. (a) Starting structure of A β ₁₋₄₂ (PDB entry: 1Z0Q) and (b) its initial position with respect to bilayer surface. The dashed line represented the average phosphate plane of the upper bilayer leaflet. (TIF)

Figure S3 Initial structures of three different dimers. In (a) Dimer1 and (c) Dimer2: peptides were placed side-by-side, where their C-terminal β -hairpin segments were in adjacent positions, with antiparallel and parallel orientations, respectively. In (e) Dimer3: peptides were stacked on the top of each other. The right column showed the initial snapshots of dimers: (b) Dimer1, (d) Dimer2 and (f) Dimer3, placed on the top of GM1/Chol/POPC bilayer. The image rendering was done with VMD. Here peptides were shown in cartoon and their secondary structures were colored based on the default representation in VMD (α -helix: purple, 3_{10} -helix: blue, π -helix: red, extended- β : yellow, bridge- β : tan, turn: cyan, coil: white). Phospholipids were shown as thin gray lines, cholesterol as green sticks and GM1 as orange van der Waals spheres. (TIF)

Figure S4 Lipid-peptide interaction energies in three different A β -monomer-membrane systems. The average values of last 100 ns data were presented here. (TIF)

Figure S5 Number of A β -monomer-lipid H-bonds. H-bonds in (a) Chol/POPC and (b) POPC bilayers. (TIF)

Figure S6 $\text{A}\beta$ -monomer bound at the interface of four GM1 molecules.

(TIF)

Figure S7 Secondary structure of $\text{A}\beta$ -monomer in aqueous solution. (a) The secondary structure profile of $\text{A}\beta$ in aqueous solution based on DSSP calculation. (b) The final snapshot of $\text{A}\beta$ at 250 ns, prepared by VMD. The default color code of DSSP plot (a): random coil: white, β -sheet: red, β -bridge: black, bend: green, turn: yellow, α -helix: blue, π -helix: purple, 3_{10} -helix: gray. In snapshot (b) the secondary structures were colored based on the default representation in VMD (α -helix: purple, 3_{10} -helix: blue, π -helix: red, extended- β : yellow, bridge- β : tan, turn: cyan, coil: white).

(TIF)

Figure S8 Root mean square deviations (RMSD) plotted against time for all three dimers.

(TIF)

Figure S9 $\text{A}\beta$ - $\text{A}\beta$ interaction energies in dimers (last 200 ns average).

(TIF)

Figure S10 Preferential location of $\text{A}\beta$ -dimers on GM1-containing membrane surface. Time dependence of the distance between the center of mass of $\text{A}\beta$ -dimers and the average plane of phosphorus atoms in the contact monolayer.

(TIF)

Figure S11 Secondary structures of $\text{A}\beta$ -dimers. Time profile of secondary structural contents of (a) Dimer2 and (c) Dimer3 and their snapshots: (b) Dimer2 and (d) Dimer3 near the end of simulation. In snapshots the secondary structures of $\text{A}\beta$ were colored based on the default representation in VMD (α -helix: purple, 3_{10} -helix: blue, π -helix: red, extended- β : yellow, bridge- β : tan, turn: cyan, coil: white).

(TIF)

Figure S12 Inter-peptide H-bonds in Dimer2. 3D-plot showing the inter-peptide hydrogen-bonding interactions within Dimer2 (last 200 ns average). The notation used here for leveling H-bond between a pair of residues was XA-YB, where X was the amino acid residue of peptide-1 with its corresponding residue number A and Y was the residue of peptide-2 with its corresponding residue number B.

(TIF)

Figure S13 Inter-peptide H-bonds in Dimer3. 3D-plot showing the inter-peptide hydrogen-bonding interactions within Dimer3 (last 200 ns average). The notation used was same as Figure S12.

(TIF)

Table S1 Properties of equilibrated bilayers.

(DOC)

Table S2 Details of crucial $\text{A}\beta$ -monomer-GM1 hydrogen bonding interactions.

(DOC)

Table S3 Properties of dimers (averaged over last 200 ns trajectory) on GM1-containing membrane.

(DOC)

Table S4 Interaction of dimers with membrane surface (averaged over last 200 ns trajectory).

(DOC)

Table S5 The formation time percentage (last 200 ns average) of intra- and inter-molecular salt-bridges in $\text{A}\beta$ -dimers. Listed were those, which had more than 10% existence.

(DOC)

Table S6 Details of inter-peptide hydrogen-bonding interactions within Dimer1. Listed were those which have H-bond ≥ 0.1 .

(DOC)

Table S7 Details of inter-peptide hydrogen-bonding interactions within Dimer2. Listed were those which have H-bond ≥ 0.1 .

(DOC)

Table S8 Details of inter-peptide hydrogen-bonding interactions within Dimer3. Listed were those which have H-bond ≥ 0.1 .

(DOC)

Table S9 Solvent-accessible surface area (SASA) for monomer (M) and dimers (D).

(DOC)

Text S1 Description of additional data for Table S1.

(DOC)

Text S2 Description of additional data for Table S3 and Figure S9.

(DOC)

Text S3 Description of additional data for Table S4 and Figure S10.

(DOC)

Acknowledgments

We would like to thank the Department of Chemistry, University of Calcutta, Kolkata, India, and the RCAMOS of the Indian Association for the Cultivation of Science, Kolkata, India, for the computational facilities.

Author Contributions

Conceived and designed the experiments: MM CM. Performed the experiments: MM. Analyzed the data: MM. Contributed reagents/materials/analysis tools: MM CM. Wrote the paper: MM CM.

References

- Iwatsubo T, Odaka A, Suzuki N, Mizusawa H, Nukina N, et al. (1994) Visualization of $\text{A}\beta_{42,43}$ and $\text{A}\beta_{40}$ in senile plaques with end-specific $\text{A}\beta$ monoclonals: evidence that an initially deposited species is $\text{A}\beta_{42(43)}$. *Neuron* 13: 45–53.
- Jarrett JT, Berger EP, Lansbury PT Jr (1993) The carboxy terminus of the β amyloid protein is critical for the seeding of amyloid formation: implications for the pathogenesis of Alzheimer's disease. *Biochemistry* 32: 4693–4697.
- Kirkitadze MD, Bitan G, Teplow DB (2002) Paradigm shifts in Alzheimer's disease and other neurodegenerative disorders: the emerging role of oligomeric assemblies. *J Neurosci Res* 69: 567–577.
- Walsh DM, Klyubin I, Fadeeva JV, Cullen WK, Anwyl R, et al. (2002) Naturally secreted oligomers of amyloid β protein potently inhibit hippocampal long-term potentiation in vivo. *Nature* 416: 535–539.
- Ahmed M, Davis J, Aucoin D, Sato T, Ahuja S, et al. (2010) Structural conversion of neurotoxic amyloid- β_{1-42} oligomers to fibrils. *Nat Struct Mol Biol* 17: 561–567.
- Luhers T, Ritter C, Adrian M, Riek-Loher D, Bohrmann B, et al. (2005) 3D structure of Alzheimer's amyloid- $\beta(1-42)$ fibrils. *Proc Natl Acad Sci U S A* 102: 17342–17347.
- Laganowsky A, Liu C, Sawaya MR, Whitelegge JP, Park J, et al. (2012) Atomic view of a toxic amyloid small oligomer. *Science* 335: 1228–1231.
- Zhu X, Bora RP, Barman A, Singh R, Prabhakar R (2012) Dimerization of the full-length Alzheimer amyloid β -peptide ($\text{A}\beta_{42}$) in explicit aqueous solution: a molecular dynamics study. *J Phys Chem B* 116: 4405–4416.
- Rushworth JV, Hooper NM (2011) Lipid rafts: linking Alzheimer's amyloid- β production, aggregation, and toxicity at neuronal membranes.

- International J Alzheimer's Disease, volume 2011, Article ID 603052, 14 pages, doi:10.4061/2011/603052.
- Matsuzaki K (2007) Physicochemical interactions of amyloid β -peptide with lipid bilayers. *Biochim Biophys Acta* 1768: 1935–1942.
 - Sciacca MF, Kotler SA, Brender JR, Chen J, Lee DK, et al. (2012) Two-step mechanism of membrane disruption by A β through membrane fragmentation and pore formation. *Biophys J* 103: 702–710.
 - Kagan BL, Thundimadathil J (2010) Amyloid peptide pores and the beta sheet conformation. *Adv Exp Med Biol* 677: 150–167.
 - Jang H, Zheng J, Nussinov R (2007) Models of β -Amyloid ion channels in the membrane suggest that channel formation in the bilayer is a dynamic process. *Biophys J* 93: 1938–1949.
 - Chang Z, Luo Y, Zhang Y, Wei G (2011) Interactions of A β _{25–35} β -barrel-like oligomers with anionic lipid bilayer and resulting membrane leakage: an all-atom molecular dynamics study. *J Phys Chem B* 115: 1165–1174.
 - Nikolic A, Baud S, Rauscher S, Pomès R (2011) Molecular mechanism of β -sheet self-organization at water-hydrophobic interfaces. *Proteins* 79: 1–22.
 - Nagai Y (1995) Functional role of gangliosides in bio-signaling. *Behav Brain Res* 66: 99–104.
 - Yanagisawa K, Odaka A, Suzuki N, Ihara Y (1995) GM1 ganglioside-bound amyloid β -protein (A β): a possible form of preamyloid in Alzheimer's disease. *Nat Med* 1: 1062–1066.
 - Yagi-Utsumi M, Matsuo K, Yanagisawa K, Gekko K, Kato K (2011) Spectroscopic characterization of inter-molecular interaction of amyloid β promoted on GM1 micelles. *Int J Alzheimer's Disease* 2011, 8 pages (doi:10.4061/2011/925073).
 - Mikhalyov I, Olofsson A, Gröbner G, Johansson LBA (2010) Designed fluorescent probes reveals interactions between amyloid- β (1–40) peptides and GM1 ganglioside in micelles and lipids vesicles. *Biophys J* 99: 1510–1519.
 - Kakio A, Nishimoto S, Yanagisawa K, Kozutsumi Y, Matsuzaki K (2002) Interactions of amyloid β -protein with various gangliosides in raft-like membranes: importance of GM1 ganglioside-bound form as an endogenous seed for Alzheimer amyloid. *Biochemistry* 41: 7385–7390.
 - Matsuzaki K, Horikiri C (1999) Interactions of amyloid β -Peptide (1–40) with ganglioside-containing membranes. *Biochemistry* 38: 4137–4142.
 - Chi EY, Frey SL, Lee KYC (2007) Ganglioside G_{M1}-mediated amyloid-beta fibrillogenesis and membrane disruption. *Biochemistry* 46: 1913–1924.
 - Lemkul JA, Bevan DR (2011) Lipid composition influences the release of Alzheimer's amyloid β -peptide from membranes. *Protein Sci* 20: 1530–1545.
 - Fukunaga S, Ueno H, Yamaguchi T, Yano Y, Hoshino M (2012) GM1 cluster mediates formation of toxic A β fibrils by providing hydrophobic environments. *Biochemistry* 51: 8125–8131.
 - Manna M, Mukhopadhyay C (2009) Cause and effect of melittin-induced pore formation: a computational approach. *Langmuir* 25: 12235–12242.
 - Niemelä PS, Ollila S, Hyvönen MT, Karttunen M, Vattulainen I (2007) Assessing the nature of lipid raft membranes. *PLoS Comput Biol* 3: e34. doi:10.1371/journal.pcbi.0030034.
 - Wei G, Jewett AI, Shea JE (2011) Structural diversity of dimers of Alzheimer amyloid- β (25–35) peptide and polymorphism of resulting fibrils. *Phys Chem Chem Phys* 12: 3622–3629.
 - Jang S, Shin S (2006) Amyloid β -peptide oligomerization in silico: dimers and trimers. *J Phys Chem B Lett* 110: 1955–1958.
 - Kittner M, Knecht V (2010) Disordered versus fibril-like amyloid β (25–35) dimers in water: structure and thermodynamics. *J Phys Chem B* 114: 15288–15295.
 - Barz B, Urbanc B (2012) Dimer formation enhances structural differences between amyloid β -protein (1–40) and (1–42): an explicit-solvent molecular dynamics study. *PLoS One* 7: e34345. Doi:10.1371/journal.pone.0034345.
 - Baftizadeh F, Biarnes X, Pietrucci F, Affinito F, Laio A (2012) Multidimensional view of amyloid fibril nucleation in atomistic details. *J Am Chem Soc* 134: 3886–3894.
 - Davis CH, Berkowitz ML (2010) A molecular dynamics study of the early stages of amyloid- β (1–42) oligomerization: the role of lipid membranes. *Proteins* 78: 2533–2545.
 - Zhao LN, Chiu S-W, Benoit J, Chew LY, Mu Y (2011) Amyloid β peptides aggregation in a mixed membrane bilayer: a molecular dynamics study. *J Phys Chem B* 115: 12247–12256.
 - Strudel B, Lee JW, Whittleston CS, Wales DJ (2010) Transmembrane structures for Alzheimer's A β _{1–42} oligomers. *J Am Chem Soc* 132: 13300–13312.
 - Tofoleanu F, Buchete N-V (2012) Molecular interactions of Alzheimer's A β protofibrils with lipid membranes. *J Mol Biol* 421: 572–586.
 - Jang H, Connelly L, Arce FT, Ramachandran S, Kagan BL, et al. (2013) Mechanisms for the insertion of toxic, fibril-like β -amyloid oligomers into the membrane. *J Chem Theory Comput* 9: 822–833.
 - Wang Q, Zhao C, Zhao J, Wang J, Yang J-C, et al. (2010) Comparative molecular dynamics study of A β adsorption on the self-assembled monolayers. *Langmuir* 26: 3308–3316.
 - Manna M, Mukhopadhyay C (2011) Cholesterol driven alteration of the conformation and dynamics of phospholamban in model membranes. *Phys Chem Chem Phys* 13: 20188–20198.
 - Alberts B, Johnson A, Lewis J, Raff M, Roberts K, et al. (2002) *Molecular biology of the cell*, 4th ed., Garland Science, Taylor & Francis Group, New York, NY.
 - Mondal S, Mukhopadhyay C (2008) Molecular level investigation of organization in ternary lipid bilayer: a computational approach. *Langmuir* 24: 10298–10305.
 - Tomaselli S, Esposito V, Vangone P, van Nuland NAJ, Bonvin AMJJ, et al. (2006) The α -to- β conformational transition of Alzheimer's A β (1–42) peptide in aqueous media is reversible: a step by step conformational analysis suggests the location of beta conformation seeding. *Chem Bio Chem* 7: 257–267.
 - Ma B, Nussinov R (2002) Stabilities and conformations of Alzheimer's β -amyloid peptide oligomers (A β _{16–22}, A β _{16–35}, and A β _{10–35}): sequence effects. *Proc Natl Acad Sci U S A* 99: 14126–14131.
 - Lindahl E, Hess B, van der Spoel D (2001) GROMACS 3.0: a package for molecular simulation and trajectory analysis. *J Mol Model* 7: 306–317.
 - Berendsen HJC, van der Spoel D, van Drunen R (1995) GROMACS: A message-passing parallel molecular dynamics implementation. *Comput Phys Commun* 91: 43–56.
 - Van Gunsteren WF, Berendsen HJC (1987) *Gromos-87 manual*; Biomos BV: Nijenborgh 4, 9747 AG Groningen, The Netherlands.
 - Berger O, Edholm O, Jähnig F (1997) Molecular dynamics simulations of a fluid bilayer of dipalmitoylphosphatidylcholine at full hydration, constant pressure, and constant temperature. *Biophys J* 72: 2002–2013.
 - Manna M, Mukhopadhyay C (2011) Molecular dynamics simulations of the interactions of kinin peptides with an anionic POPG bilayer. *Langmuir* 27: 3713–3722.
 - Polyansky AA, Volynsky PE, Nolde DE, Arseniev AS, Efremov RG (2005) Role of lipid charge in organization of water/lipid bilayer interface: insights via computer simulations. *J Phys Chem B* 109: 15052–15059.
 - Höltje M, Förster T, Brandt B, Engels T, von Rybinski W, et al. (2001) Molecular dynamics simulations of stratum corneum lipid models: fatty acids and cholesterol. *Biochim Biophys Acta* 1511: 156–167.
 - Olsen BN, Schlesinger PH, Baker NA (2009) Perturbations of membrane structure by cholesterol and cholesterol derivatives are determined by sterol orientation. *J Am Chem Soc* 131: 4854–4865.
 - Jedlovsky P, Segal M, Vallauri R (2009) GM1 ganglioside embedded in a hydrated DOPC membrane: a molecular dynamics simulation study. *J Phys Chem B* 113: 4876–4886.
 - Ryckaert JP, Bellemans A (1975) Molecular dynamics of liquid n-butane near its boiling point. *Chem Phys Lett* 30: 123–125.
 - Berendsen HJC, Postma JPM, van Gunsteren WF, Hermans J (1981) Interaction models for water in relation to protein hydration. *Intermolecular Forces*, B. Pullman, Ed., Reidel, Dordrecht, The Netherlands 331–342.
 - Berendsen HJC, Postma JPM, van Gunsteren WF, DiNola A, Haak JR (1984) Molecular dynamics with coupling to an external bath. *J Chem Phys* 81: 3684–3690.
 - Essmann U, Perera L, Berkowitz ML, Darden T, Lee H, et al. (1995) A smooth particle mesh Ewald method. *J Chem Phys* 103: 8577–8593.
 - Hess B, Bekker H, Berendsen HJC, Fraaije JGE (1997) LINCS: a linear constraint solver for molecular simulations. *J Comput Chem* 18: 1463–1472.
 - Humphrey W, Dalke A, Schulten K (1996) VMD: visual molecular dynamics. *J Mol Graphics* 14: 33–38.
 - DeLano WL (2002) *The PyMOL molecular graphics system*. DeLano Scientific, San Carlos, CA.
 - Yagi-Utsumi M, Kameda T, Yamaguchi Y, Kato K (2010) NMR characterization of the interactions between lyso-GM1 aqueous micelles. *FEBS Letts* 584: 831–836.
 - Lemkul JA, Bevan DR (2009) Perturbation of membranes by the amyloid β -peptide - a molecular dynamics study. *FEBS J* 276: 3060–3075.
 - Fantini J, Yahi N (2010) Molecular insights into amyloid regulation by membrane cholesterol and sphingolipids: common mechanisms in neurodegenerative diseases. *Expert Rev Mol Medicine* 12: e27.
 - Williamson MP, Suzuki Y, Bourne NT, Asakura T (2006) Binding of amyloid β -peptide to ganglioside micelles is dependent on histidine-13. *Biochem J* 397: 483–490.
 - Yahi N, Aulas A, Fantini J (2010) How cholesterol constrains glycolipid conformation for optimal recognition of Alzheimer's β amyloid peptide (A β _{1–40}). *PLoS One* 5: e9079.
 - Yu X, Zheng J (2012) Cholesterol promotes the interaction of Alzheimer β -amyloid monomer with lipid bilayer. *J Mol Biol* 421: 561–571.
 - Qiu L, Buic C, Reay A, Vaughn MW, Cheng KH (2011) Molecular dynamics simulations reveal the protective role of cholesterol in β -amyloid protein-induced membrane disruptions in neuronal membrane mimics. *J Phys Chem B* 115: 9795–9812.
 - Davis CH, Berkowitz ML (2009) Structure of the amyloid- β (1–42) monomer absorbed to model phospholipid bilayers: a molecular dynamics study. *J Phys Chem B* 113: 14480–14486.
 - Kabsch W, Sander C (1983) Dictionary of protein secondary structure: pattern recognition of hydrogen-bonded and geometrical features. *Biopolymers* 22: 2577–2637.
 - Choo-Smith L-P, Surewicz WK (1997) The interaction between Alzheimer amyloid β (1–40) peptide and ganglioside G_{M1}-containing membranes. *FEBS Letts* 402: 95–98.
 - Kirkpatrick MD, Condrón MM, Teplow DB (2001) Identification and characterization of key kinetic intermediates in amyloid β -protein fibrillogenesis. *J Mol Biol* 312: 1103–1119.

70. Xue WF, Homans SW, Radford SE (2008) Systematic analysis of nucleation-dependent polymerization reveals new insights into the mechanism of amyloid self-assembly. *Proc Natl Acad Sci U S A* 105: 8926–8931.
71. Walsh DM, Hartley DM, Kusumoto Y, Fezoui Y, Condron MM, et al. (1999) Amyloid β -protein fibrillogenesis. Structure and biological activity of protofibrillar intermediates. *J Biol Chem* 274: 25945–25952.
72. Tarus B, Straub JE, Thirumalai D (2008) Structures and free-energy landscape of the wild type and mutants of the A β _{21–30} peptide are determined by an interplay between intrapeptide electrostatic and hydrophobic interactions. *J Mol Biol* 379: 815–829.
73. Thirumalai D, Reddy G, Straub JE (2012) Role of water in protein aggregation and amyloid polymerization. *Acc Chem Res* 45: 83–92.
74. Sciarretta KL, Gordon DJ, Petkova AT, Tycko R, Meredith SC (2005) A β 40-Lactam (D23/K28) models a conformation highly favorable for nucleation of amyloid. *Biochemistry* 44: 6003–6014.