

Physiological concentrations of dietary polyphenols regulate vascular endothelial cell expression of genes important in cardiovascular health

Sonja K. Nicholson, Gregory A. Tucker and John M. Brameld*

Division of Nutritional Sciences, School of Biosciences, University of Nottingham, Sutton Bonington Campus, Loughborough, Leics LE12 5RD, UK

(Received 12 August 2009 – Revised 2 November 2009 – Accepted 17 November 2009 – First published online 21 December 2009)

Previous cell culture-based studies have shown potential health beneficial effects on gene expression of dietary polyphenols, including those found in red wine and green tea. However, these studies have tended to use higher concentrations (2–100 μM) than those observed in blood (0.1–1 μM) after consuming polyphenol-rich foods or beverages. The present study investigated effects of physiological concentrations of different classes of dietary polyphenol on the expression of genes important in cardiovascular health (endothelial NO synthase (eNOS), endothelin-1 (ET-1) and vascular endothelial growth factor (VEGF)) by cultured vascular endothelial cells (human umbilical vein endothelial cells) in the absence or presence of H_2O_2 . Resveratrol and quercetin (0.1–1 μM) increased eNOS and VEGF mRNA expression particularly in the absence of H_2O_2 (50 μM) and decreased H_2O_2 -induced ET-1 mRNA expression ($P < 0.001$ for polyphenol \times H_2O_2 interactions). Similarly, resveratrol and quercetin decreased endothelin secretion into the media, blocking the stimulatory effect of 50 μM - H_2O_2 ($P < 0.001$ for polyphenol \times H_2O_2 interaction). Of the nine other polyphenols tested, only epigallocatechin gallate had similar effects on both the eNOS and ET-1 mRNA expression, but to a lesser extent than resveratrol at an equimolar concentration (0.1 μM). The observed effects on gene expression would be expected to result in vasodilation and thereby reduced blood pressure. Since only three of the eleven polyphenols tested had biological activity, it is unclear whether particular structures are important or whether the effects might relate to the relatively high antioxidant capacities of the three active polyphenols.

Dietary polyphenols: Endothelial cells: Endothelial NO synthase: Endothelin-1: Gene expression

Epidemiological studies⁽¹⁾ have shown an association between the intake of plant foods, particularly fruit and vegetables, and a decrease in CHD and other CVD. Similarly, increased consumption of green tea is associated with a reduced risk of CVD⁽²⁾, while rat studies indicate that consumption of green tea reduces the increased blood pressure induced by fructose feeding⁽³⁾. These beneficial effects of plant-derived foods and beverages on cardiovascular health are suggested to be due to the presence of polyphenols. Previous studies have shown that polyphenols may well be beneficial to cardiovascular health⁽⁴⁾, particularly those present in red wines (e.g. resveratrol and quercetin), which are thought to account for the ‘French paradox’.

The mechanism of action for the beneficial effects of polyphenols on CVD may be due to their antioxidant activity. However, many polyphenols are metabolised in the body, and these metabolites have much lower antioxidant capacities than their parent compounds, suggesting that antioxidant activity may not be their only mechanism of action⁽⁵⁾. Polyphenols thought to provide health benefits include resveratrol, quercetin, epicatechin gallate (ECG) and epigallocatechin gallate (EGCG), which belong to different classes of polyphenol and may therefore have differing effects dependent upon their chemical structures. Tea and polyphenols present in

teas (particularly ECG and EGCG) have previously been reported to have anti-carcinogenic, anti-proliferative and anti-angiogenic properties⁽⁶⁾, as well as having effects on gene expression. These anti-angiogenic properties may relate to local expression of vascular endothelial growth factor (VEGF), a potent stimulator of angiogenesis. Red wine polyphenols have been shown to reduce expression of the vasoconstrictor, endothelin-1 (ET-1) in cultured vascular cells^(7–9) and to increase expression of endothelial NO synthase (eNOS), the enzyme responsible for the production of the vasodilator NO^(10,11). However, cell culture-based studies investigating these effects have tended to use supraphysiological concentrations of individual polyphenols. Maximum concentrations of polyphenols in the blood after consumption of polyphenol-rich foods or beverages tend to be about 0.1–1 μM ⁽¹²⁾, whereas previous *in vitro* studies have tended to treat cells with much higher concentrations (2–100 μM).

Stress is known to elevate the expression of certain genes in several cell types including human umbilical vein endothelial cells (HUVEC) and smooth muscle cells⁽⁹⁾. Different methods to induce stress or to elevate gene expression include stretching cells cultured on a flexible membrane (cyclic strain)⁽⁹⁾ or treatment with H_2O_2 ⁽⁹⁾ or angiotensin II⁽⁸⁾. Previous studies have shown that supraphysiological concentrations

Abbreviations: ECG, epicatechin gallate; EGCG, epigallocatechin gallate; eNOS, endothelial NO synthase; ET-1, endothelin-1; HUVEC, human umbilical vein endothelial cells; VEGF, vascular endothelial growth factor.

* **Corresponding author:** Dr John M. Brameld, fax +44 115 9516122, email john.brameld@nottingham.ac.uk

(10–100 μM) of resveratrol blocked the stimulatory effects⁽⁹⁾ of H₂O₂, cyclic strain⁽⁹⁾ and angiotensin II treatment⁽⁸⁾ on ET-1 mRNA expression.

The present study investigated the effects of physiological concentrations of different classes of dietary polyphenol on the expression of genes important in cardiovascular health (eNOS, ET-1 and VEGF) in cultured vascular endothelial cells (HUVEC) under stress-free and oxidative stress conditions (i.e. with or without H₂O₂).

Materials and methods

Materials and chemicals

Unless stated otherwise, all chemicals (including polyphenols) were obtained from Sigma-Aldrich (St Louis, MO, USA). The endothelin ELISA kit was obtained from Biomedica (Vienna, Austria) via Oxford Biosystems (Oxford, UK).

Preparation of individual polyphenols

A 100 μM stock solution of each polyphenol was prepared in PBS and diluted further with PBS before being added to culture media.

Cell culture

HUVEC were purchased from Clonetics (Cambrex, Inc., Walkersville, MD, USA) as a frozen stock (passage 3), defrosted and cultured in endothelial basal medium 2 growth media plus bullet kit (Cambrex, Inc.) containing 2% fetal calf serum, antibiotics and growth factor supplements at 37°C and 5% CO₂. The cells were passaged three times to provide a pool of cells to be used for experiments. Experiments were performed in six-well plates (Corning, Inc., Corning, NY, USA) when the cells were 80–85% confluent, at which stage the endothelial basal medium 2 growth media (2% foetal calf serum, with growth factors) was replaced with endothelial basal medium 2 control media (0.8% foetal calf serum, no growth factors) and incubated for 24 h (37°C, 5% CO₂). Treatments were prepared in endothelial basal medium 2 containing 0.8% fetal calf serum and antibiotics but no other supplements. This consisted of 70% media and 30% polyphenol and/or H₂O₂ in PBS, with PBS alone used as control. Final concentrations of individual polyphenols in the culture media ranged from 0.001 to 1.0 μM, while concentrations of H₂O₂ ranged from 6.25 to 50 μM. After 24 h incubation with treatments, culture media were stored at –40°C for measurement of endothelin secretion using an ELISA kit and total RNA was extracted from the cells using TRIzol (Invitrogen, Carlsbad, CA, USA), both according to the manufacturer’s instructions.

Quantitative real-time PCR

Total RNA was DNase-treated before quantifying RNA yields using a Nanodrop spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA) and checking RNA integrity by running on 1% agarose gels. Complementary DNA was synthesised using Moloney murine leukaemia virus reverse transcriptase and buffer (Promega, Fitchburg, WI, USA). Gene- and species-specific primers and probes for real-time PCR (Table 1) were all purchased from Applied Biosystems

Table 1. Real-time PCR primers and probes

Gene (product code)	GenBank accession number	Amplicon length (bp)	Exon boundary	Probe sequence 5'–3'	Probe concentration (μM)	Primer concentrations (μM)	Assay concentration
GAPDH (Hs00266705_g1)	NM_002046.3	74	2–3	GGACTCATGACCACAGTCCATGCCA	5	18	20X
eNOS (Hs00167166_m1)	NM_000603.3	80	14–15	GAATGGAGAGAGCTTTGCAGCTGCC	5	18	20X
VEGF (Hs00173626_m1)	NM_001025366.1	77	1–2	TCCACCATGCCAAGTGGTCCCAGGC	5	18	20X
ET-1 (Hs00174961_m1)	NM_001955.2	62	2–3	ACACTCCCCGAGACGTTGTTCCGTA	5	18	20X

GAPDH, glyceraldehyde 3-phosphate dehydrogenase; eNOS, endothelial NO synthase; VEGF, vascular endothelial growth factor; ET-1, endothelin-1.

(Foster City, CA, USA). Real-time PCR was carried out on triplicate complementary DNA samples from control or treated HUVEC following the standard curve method on the Roche Lightcycler 480 diagnostic system using 384-well plates and TaqMan Universal PCR master mix (Applied Biosystems). As there were no significant treatment effects on glyceraldehyde 3-phosphate dehydrogenase mRNA (Tables 2–6), glyceraldehyde 3-phosphate dehydrogenase was selected as the housekeeping gene to allow quantification of the expression of eNOS, ET-1 and VEGF.

Statistical analyses

Relative mRNA expression data are expressed as means, with the standard error of the differences of the means shown in tables or the standard deviations shown in figures. All the data were analysed by one- or two-way ANOVA using Genstat version 9.1 (2007). $P < 0.05$ was considered as statistically significant. A *post hoc* Dunnett's test was used to identify significant differences between the control and treatment groups, but only when there were no significant interactions.

Results

There were no effects ($P > 0.1$) of any of the treatments on the expression of glyceraldehyde 3-phosphate dehydrogenase (Tables 2–6), so glyceraldehyde 3-phosphate dehydrogenase was used as the control gene to normalise the expression of eNOS, ET-1 and VEGF mRNA.

Effects of hydrogen peroxide on human umbilical vein endothelial cells gene expression

The dose-dependent effects of H_2O_2 (6.25–50 μM) on the expression of eNOS, ET-1 and VEGF mRNA were determined. Table 2 shows a dose-dependent increase in the expression of all the three genes with H_2O_2 treatment with maximum increases of about 220% (relative to controls) observed with 50 μM - H_2O_2 ($P < 0.001$ for all).

Dose–response effects of resveratrol and quercetin on human umbilical vein endothelial cells gene expression

The dose-dependent effects of resveratrol and quercetin (0.001–1.0 μM) on the expression of eNOS, ET-1 and VEGF mRNA were investigated in the absence or presence

of H_2O_2 (50 μM). Both the polyphenols had similar effects, but resveratrol was more potent than quercetin at equimolar concentrations (Tables 3 and 4).

As before, addition of 50 μM - H_2O_2 increased the eNOS, ET-1 and VEGF mRNA expression (Tables 3 and 4), but the effects of the polyphenols were gene specific. Resveratrol induced dose-dependent increases in the expression of eNOS and VEGF mRNA in the absence of H_2O_2 (Table 2), but the two treatments were not additive ($P < 0.001$ for $H_2O_2 \times$ resveratrol interactions). In contrast, resveratrol (at 0.1 and 1 μM) reduced the ET-1 mRNA expression in the absence of H_2O_2 (Table 3) and reduced or blocked the effect of 50 μM - H_2O_2 ($P < 0.001$ for both $H_2O_2 \times$ resveratrol interaction).

Similarly, quercetin induced dose-dependent increases in the expression of eNOS and VEGF mRNA in the absence of H_2O_2 (Table 4), but there was no additive effect with H_2O_2 on eNOS mRNA expression, while higher concentrations of quercetin reduced VEGF expression induced by 50 μM - H_2O_2 ($P < 0.001$ for both $H_2O_2 \times$ quercetin interactions). Like resveratrol, quercetin (at 0.1 and 1 μM) reduced ET-1 mRNA expression compared with controls (Table 4) and reduced or blocked the effect of 50 μM - H_2O_2 ($P < 0.001$ for both $H_2O_2 \times$ quercetin interaction).

Effects of resveratrol, quercetin and hydrogen peroxide on endothelin secretion

Since both quercetin and resveratrol (at 0.1 μM) reduced basal- and H_2O_2 (50 μM)-induced expression of ET-1 mRNA, their effects on the secretion of endothelin by HUVEC were examined. The effects were very similar to the effects on mRNA expression: in the absence of H_2O_2 , both quercetin and resveratrol (0.1 μM) reduced the secretion of endothelin relative to controls (Fig. 1). In the absence of quercetin and resveratrol, H_2O_2 (50 μM) significantly increased the secretion of endothelin, but this was blocked by the addition of either resveratrol or quercetin (0.1 μM), with resveratrol being slightly more potent ($P < 0.001$, $H_2O_2 \times$ polyphenol interaction).

Effects of other dietary polyphenols on human umbilical vein endothelial cells gene expression

In order to investigate structure–function relationships in the observed effects, a variety of polyphenols (caffeic acid, genistein, phloretin, phloridzin, gallic acid, epicatechin,

Table 2. Dose-dependent effects of hydrogen peroxide on human umbilical vein endothelial cells gene expression (Mean values with their standard error of the differences)

H_2O_2 concentration (μM)	0	6.25	12.5	25	50	SED	<i>P</i> value
GAPDH	3.60	3.60	3.62	3.60	3.62	0.02	0.562
eNOS*	0.61	0.77†	0.99†	1.12†	1.35†	0.01	<0.001
VEGF*	0.62	0.76†	0.92†	1.08†	1.36†	0.04	<0.001
ET-1*	0.60	0.73†	0.96†	1.05†	1.30†	0.02	<0.001

GAPDH, glyceraldehyde 3-phosphate dehydrogenase; eNOS, endothelial NO synthase; VEGF, vascular endothelial growth factor; ET-1, endothelin-1.

* Gene expression normalised to GAPDH.

† Mean values were significantly different from those of the control group ($P < 0.05$, *post hoc* Dunnett's test).

Table 3. Dose-dependent effects of resveratrol (R) in the absence or presence of hydrogen peroxide (50 μM) on human umbilical vein endothelial cells gene expression

(Mean values with their standard error of the differences)

R concentration (μM)	H ₂ O ₂ concentration (μM)														P value	
	0							50								
	0	0.001	0.005	0.01	0.05	0.1	1.0	0	0.001	0.005	0.01	0.05	0.1	1.0		SED
GAPDH	3.66	3.62	3.70	3.69	3.67	3.63	3.65	3.68	3.69	3.67	3.70	3.67	3.65	3.66	0.03	0.362
eNOS*	0.64	0.73	0.82	0.93	1.08	1.19	1.22	1.22	1.24	1.27	1.32	1.38	1.39	1.49	0.02	<0.001
VEGF*	0.63	0.68	0.73	0.80	0.88	0.91	0.97	1.21	1.20	1.22	1.26	1.29	1.33	1.36	0.04	<0.001
ET-1*	0.66	0.84	0.76	0.69	0.59	0.45	0.43	1.39	1.48	1.41	1.31	1.29	0.74	0.68	0.03	<0.001

GAPDH, glyceraldehyde 3-phosphate dehydrogenase; eNOS, endothelial NO synthase; VEGF, vascular endothelial growth factor; ET-1, endothelin-1.

* Gene expression normalised to GAPDH.

ECG, epigallocatechin and EGCG) were compared with resveratrol (all at 0.1 μM) in the absence or presence of 50 μM -H₂O₂ (Tables 5 and 6). These studies were carried out in two batches (each table represents a batch) but were performed at exactly the same time, therefore allowing direct comparisons. Although previous studies had shown a significant effect of resveratrol and H₂O₂ on VEGF expression, the effects of the different polyphenols on VEGF expression were not examined here. As in previous studies, significant interactions ($P < 0.001$) were observed between polyphenol and H₂O₂ treatments on both the eNOS and ET-1 mRNA expression. As before, H₂O₂ treatment (50 μM) increased eNOS and ET-1 mRNA expression relative to control treatment (Tables 5 and 6) and resveratrol (0.1 μM) increased eNOS mRNA expression in the absence of H₂O₂, but there was no additive effect of resveratrol and H₂O₂ (Table 5). Similar to resveratrol, gallic acid and EGCG (Table 6) had stimulatory effects on eNOS, with both appearing to be additive to the effect of H₂O₂. The effects of the catechins on eNOS mRNA appeared to increase in the order epicatechin < ECG < epigallocatechin < EGCG (Table 6), which might indicate a structure–function relationship, although only epigallocatechin and EGCG appeared to have significant effects. In contrast, genistein, phloretin and phloridzin (Table 5) all reduced eNOS expression in the presence and absence of H₂O₂ (50 μM), opposite to the effect of resveratrol. Both resveratrol (Table 5) and EGCG (Table 6)

reduced ET-1 expression compared with controls and blocked the stimulatory effect of H₂O₂ on ET-1 mRNA. None of the other polyphenols studied appeared to have significant effects on ET-1 expression, although there was some variability in expression.

Discussion

H₂O₂ alone was found to significantly increase the expression of eNOS, VEGF and ET-1 mRNA in a dose-dependent manner (6.25–50 μM) after 24 h treatment. Previous studies have shown similar effects, with 25 μM -H₂O₂ increasing ET-1 expression in HUVEC⁽⁹⁾ and 100 μM -H₂O₂ increasing eNOS expression in bovine aortic endothelial cells⁽¹³⁾.

Quercetin and resveratrol were found to significantly increase the expression of eNOS in a dose-dependent manner after 24 h. This agrees with previous studies using supraphysiological concentrations of resveratrol, where 33 μM and 10–100 μM up-regulated eNOS mRNA expression in HUVEC and HUVEC-derived EA.hy 926 cells, respectively⁽¹⁰⁾. In the present study, the combination of 50 μM -H₂O₂ and quercetin or resveratrol (0.001–1.0 μM) increased eNOS expression, but the two effects were not completely additive, suggesting that the effects are either via similar mechanisms or have simply reached a maximum level. Although eNOS protein or activity levels were not measured, a recent study⁽¹⁴⁾ showed that both resveratrol and quercetin

Table 4. Dose-dependent effects of quercetin (Q) in the absence or presence of hydrogen peroxide (50 μM) on human umbilical vein endothelial cells gene expression

(Mean values with their standard error of the differences)

Q concentration (μM)	H ₂ O ₂ concentration (μM)														P value	
	0							50								
	0	0.001	0.005	0.01	0.05	0.1	1.0	0	0.001	0.005	0.01	0.05	0.1	1.0		SED
GAPDH	3.66	3.64	3.63	3.64	3.66	3.64	3.68	3.65	3.64	3.65	3.68	3.68	3.66	3.67	0.02	0.780
eNOS*	0.67	0.74	0.78	0.81	0.86	0.93	0.98	1.27	1.24	1.30	1.31	1.33	1.36	1.39	0.02	<0.001
VEGF*	0.62	0.64	0.76	0.81	0.87	0.90	0.93	1.21	1.25	1.23	1.22	1.16	1.14	1.11	0.03	<0.001
ET-1*	0.62	0.87	0.72	0.64	0.62	0.57	0.52	1.46	1.56	1.54	1.52	1.45	0.94	0.88	0.04	<0.001

GAPDH, glyceraldehyde 3-phosphate dehydrogenase; eNOS, endothelial NO synthase; VEGF, vascular endothelial growth factor; ET-1, endothelin-1.

* Gene expression normalised to GAPDH.

Table 5. Effects of polyphenols (Poly) with differing structures (all 0.1 μM) in the absence or presence of hydrogen peroxide (50 μM) on human umbilical vein endothelial cells gene expression

(Mean values with their standard error of the differences)

Poly (all 0.1 μM)	H ₂ O ₂ concentration (μM)												P value	
	0						50							
	Cont	R	C	G	Pt	Pd	Cont	R	C	G	Pt	Pd		SED
GAPDH	3.47	3.52	3.45	3.44	3.51	3.55	3.49	3.48	3.49	3.46	3.46	3.50	0.03	0.209
eNOS*	0.64	1.23	0.75	0.32	0.49	0.30	1.25	1.31	0.96	0.98	0.75	0.99	0.04	<0.001
ET-1*	0.59	0.47	0.70	0.50	0.63	0.69	1.19	0.60	1.00	1.06	0.92	1.04	0.04	<0.001

Cont, control; R, resveratrol; C, caffeic acid; G, genistein; Pt, phloretin; Pd, phloridzin; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; eNOS, endothelial NO synthase; ET-1, endothelin-1.

* Gene expression normalised to GAPDH.

(at 100 μM) increased NO synthesis by HUVEC-derived EA.hy 926 cells, indicating that the effects on mRNA expression described here are likely to be matched at the protein level.

In contrast, treatment of HUVEC with either quercetin or resveratrol (at 0.1 or 1.0 μM) resulted in decreased expression of ET-1 mRNA, particularly in the presence of 50 μM -H₂O₂. These findings are consistent with previous studies in HUVEC showing that resveratrol (100 μM) inhibited the expression of ET-1 mRNA induced by H₂O₂ (25 μM) or cyclic strain⁽⁹⁾. Similarly, resveratrol (at 1–100 μM) reduced ET-1 mRNA expression and gene promoter activity induced by angiotensin II in rat aorta smooth muscle cells⁽⁸⁾. Therefore, resveratrol's ability to block stress-induced ET-1 mRNA expression is not specific to treatment with H₂O₂, but importantly the present studies demonstrate the effects at physiologically relevant concentrations.

As well as increasing ET-1 mRNA expression, H₂O₂ (50 μM) also significantly increased secretion of endothelin protein into the media. A similar effect on endothelin protein levels was observed in human aortic vascular smooth muscle cells treated with 200 μM -H₂O₂ for 8 h⁽¹⁵⁾. Quercetin and resveratrol (0.1 μM) significantly reduced the secretion of endothelin and blocked the stimulatory effects of H₂O₂ (50 μM); again in agreement with Ruef *et al.*⁽¹⁵⁾, who showed that supraphysiological concentrations of resveratrol and quercetin (100 and 50 μM , respectively) blocked the effect of H₂O₂ (200 μM) on ET-1 protein content in human aortic vascular smooth muscle cells.

Studies on the effects of equimolar concentrations of resveratrol and nine other dietary polyphenols showed that resveratrol, gallic acid and EGCG all increased eNOS mRNA expression in the presence and absence of H₂O₂ (50 μM), but only resveratrol and EGCG decreased ET-1 expression in the absence and presence of H₂O₂ (50 μM). In contrast, genistein, phloridzin and phloretin reduced the expression of eNOS, particularly in the absence of H₂O₂ (50 μM) but had little or no effect on ET-1 mRNA expression. The stimulatory effects of gallic acid on eNOS mRNA expression agree with a previous study⁽¹¹⁾ showing that 1 μM -gallic acid increased eNOS expression in HUVEC-derived EA.hy 926 cells, although another study⁽¹⁴⁾ showed no effect of gallic acid on NO synthesis by EA.hy 926 cells. Interestingly, Appeldoorn *et al.*⁽¹⁴⁾ also demonstrated the stimulatory effects of resveratrol, quercetin, EGCG and ECG (all at 100 μM) on NO synthesis, with no effects of epicatechin, epigallocatechin, phloretin, caffeic acid and a number of other polyphenols. The stimulatory effects of resveratrol, quercetin and EGCG were matched by increases in eNOS mRNA expression⁽¹⁴⁾, whereas the effect of ECG on eNOS mRNA expression was not statistically significant, agreeing with the data described here using lower concentrations of individual polyphenols. Although the observed inhibitory effects of phloretin on eNOS mRNA expression contrast with its lack of effect on NO synthesis⁽¹⁴⁾, inhibitory effects on NO synthesis would be difficult to measure in the culture system used, since the basal level of NO synthesis was already low⁽¹⁴⁾. The inhibitory effects of genistein on eNOS mRNA expression in HUVEC described here are in contrast to

Table 6. Effects of individual polyphenols (Poly; all 0.1 μM) in the absence or presence of hydrogen peroxide (50 μM) on human umbilical vein endothelial cells gene expression

(Mean values with their standard error of the differences)

Poly (all 0.1 μM)	H ₂ O ₂ concentration (μM)												P value	
	0						50							
	Cont	Ga	EC	ECG	EGC	EGCG	Cont	Ga	EC	ECG	EGC	EGCG		SED
GAPDH	3.49	3.49	3.47	3.51	3.49	3.48	3.49	3.48	3.50	3.46	3.50	3.51	0.02	0.217
eNOS*	0.67	0.97	0.55	0.66	0.75	1.06	1.20	1.36	1.19	1.23	1.28	1.33	0.02	<0.001
ET-1*	0.62	0.60	0.88	0.77	0.84	0.43	1.20	1.09	1.22	1.03	1.11	0.96	0.02	<0.001

Cont, control; Ga, gallic acid; EC, epicatechin; ECG, epicatechin gallate; EGC, epigallocatechin; EGCG, epigallocatechin gallate; GAPDH, glyceraldehyde 3-phosphate dehydrogenase; eNOS, endothelial NO synthase; ET-1, endothelin-1.

* Gene expression normalised to GAPDH.

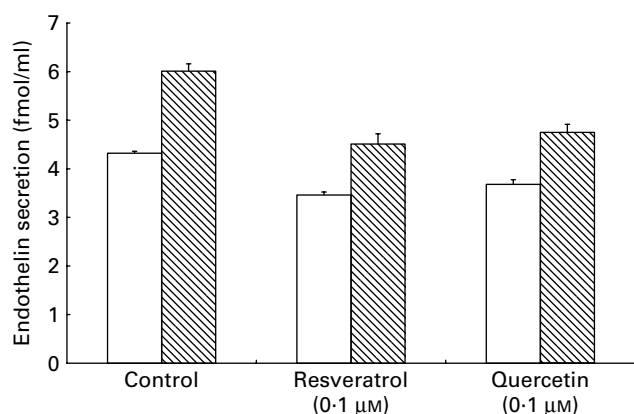


Fig. 1. Effects of resveratrol and quercetin (0.1 μM) in the presence or absence of hydrogen peroxide (50 μM) on secretion of endothelin by cultured human umbilical vein endothelial cells. Means and standard deviations (n 3) are shown. \square , Minus hydrogen peroxide (50 μM); ▨ , plus hydrogen peroxide (50 μM).

a previous study⁽¹⁶⁾, where genistein increased the expression of eNOS mRNA and protein (at 1–10 and 0.1–10 μM , respectively) in human aortic endothelial cells, possibly through activation of eNOS transcription.

Recently, Appeldoorn *et al.*⁽¹⁴⁾ demonstrated low recovery (<50%) of several polyphenols, including quercetin, from the culture medium after 24 h incubation, suggesting that some polyphenols may be unstable or being metabolised in the culture system. Using HPLC, we observed similar losses of quercetin in cell culture medium (without cells) during incubation under the same conditions used for gene expression studies (pH 7.4, 37°C, 100% humidity) with an apparent half-life of approximately 6 h. Hence, it is unclear whether the observed effects are due to quercetin *per se* or a breakdown product, but the data are still physiologically relevant since the conditions used for cell culture would match those encountered when quercetin is absorbed from the gut into the blood stream.

In conclusion, resveratrol and quercetin increased eNOS mRNA expression and decreased ET-1 mRNA expression in HUVEC in a dose-dependent manner in the absence and presence of H_2O_2 . Similar to their effects on ET-1 mRNA expression, resveratrol and quercetin decreased endothelin secretion, again in the absence and presence of H_2O_2 (50 μM). Of the nine other polyphenols tested, only EGCG had similar effects to resveratrol and quercetin on both eNOS and ET-1 mRNA expression but to a lesser extent than resveratrol at an equimolar concentration (0.1 μM). Whether these effects are dependent upon structure and/or antioxidant capacities is not clear, although the three active polyphenols (resveratrol, quercetin and EGCG) had the highest antioxidant capacities as measured by ferric reducing antioxidant power assay (data not shown). The observed effects on gene expression would be expected to result in a potent vasodilation and thereby reduced blood pressure. Indeed, quercetin is suggested to reduce diastolic blood pressure in hypertensive human subjects⁽¹⁷⁾; although closer inspection indicates that the statistical significance is due to the quercetin group having a higher baseline blood pressure but reaching a similar endpoint blood pressure to controls⁽¹⁷⁾. Further *in vivo* studies investigating the effects of individual polyphenols on blood pressure in human subjects or animals are therefore required.

Acknowledgements

S. K. N. was funded by a Biotechnology and Biological Sciences Research Council (BBSRC) committee PhD studentship. J. M. B., G. A. T. and S. K. N. planned the work; S. K. N. carried out the laboratory analyses; S. K. N. and J. M. B. statistically analysed the data; J. M. B., S. K. N. and G. A. T. wrote the manuscript. The authors have no conflict of interest to declare.

References

1. Brat P, George S, Bellamy A, *et al.* (2006) Daily polyphenol intake in France from fruit and vegetables. *J Nutr* **136**, 2368–2373.
2. Grassi D, Aggio A, Onori L, *et al.* (2008) Tea, flavonoids and nitric oxide-mediated vascular reactivity. *J Nutr* **138**, 1554S–1560S.
3. Wu LY, Juan CC, Hwang LS, *et al.* (2004) Green tea supplementation ameliorates insulin resistance and increases glucose transporter IV content in a fructose-fed rat model. *Eur J Nutr* **43**, 116–124.
4. Nicholson SK, Tucker GA & Brameld JM (2008) Effects of dietary polyphenols on gene expression in human vascular endothelial cells. *Proc Nutr Soc* **67**, 42–47.
5. Scalbert A, Manach C, Morand C, *et al.* (2005) Dietary polyphenols and the prevention of diseases. *Crit Rev Food Sci Nutr* **45**, 287–306.
6. Tang FY, Chiang EPI & Shih CJ (2007) Green tea catechin inhibits ephrin-A1-mediated cell migration and angiogenesis of human umbilical vein endothelial cells. *J Nutr Biochem* **18**, 391–399.
7. Corder R, Douthwaite JA, Lees DM, *et al.* (2001) Endothelin-1 synthesis reduced by red wine. *Nature* **414**, 863–864.
8. Chao HH, Juan SH, Liu JC, *et al.* (2005) Resveratrol inhibits angiotensin II-induced endothelin-1 gene expression and subsequent proliferation in rat aortic smooth muscle cells. *Eur J Pharmacol* **515**, 1–9.
9. Liu JC, Chen JJ, Chan P, *et al.* (2003) Inhibition of cyclic strain-induced endothelin-1 gene expression by resveratrol. *Hypertension* **42**, 1198–1205.
10. Wallerath T, Deckert G, Ternes T, *et al.* (2002) Resveratrol, a polyphenolic phytoalexin present in red wine, enhances expression and activity of endothelial nitric oxide synthase. *Circulation* **106**, 1652–1658.
11. Wallerath T, Li H, Godtel-Ambrust U, *et al.* (2005) A blend of polyphenolic compounds explains the stimulatory effect of red wine on human endothelial NO synthase. *Nitric Oxide* **12**, 97–104.
12. Scalbert A & Williamson G (2000) Dietary intake and bioavailability of polyphenols. *J Nutr* **130**, 2073S–2085S.
13. Drummond GR, Cai H, Davis ME, *et al.* (2000) Transcriptional and posttranscriptional regulation of endothelial nitric oxide synthase expression by hydrogen peroxide. *Circ Res* **86**, 347–354.
14. Appeldoorn MM, Venema DP, Peters THF, *et al.* (2009) Some phenolic compounds increase the nitric oxide level in endothelial cells *in vitro*. *J Agric Food Chem* **57**, 7693–7699.
15. Ruef J, Moser M, Kubler W, *et al.* (2001) Induction of endothelin-1 expression by oxidative stress in vascular smooth muscle cells. *Cardiovasc Pathol* **10**, 311–315.
16. Si H & Liu D (2008) Genistein, a soy phytoestrogen, upregulates the expression of human endothelial nitric oxide synthase and lowers blood pressure in spontaneously hypertensive rats. *J Nutr* **138**, 297–304.
17. Edwards RL, Lyon T, Litwin SE, *et al.* (2007) Quercetin reduces blood pressure in hypertensive subjects. *J Nutr* **137**, 2405–2411.