

Feeding microparticle protein diet combined with *Lactobacillus* sp. on existence of intestinal bacteria and growth of broiler chickens

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ABSTRAK

Penelitian bertujuan untuk mengevaluasi pengaruh protein mikropartikel dari bungkil kedelai dan tepung ikan yang dikombinasikan dengan *Lactobacillus* sp. terhadap fisiologis saluran pencernaan dan produktivitas ayam broiler. Materi penelitian adalah 193 ekor broiler, dengan bobot badan awal $481,0 \pm 67,2$ g, diberi ransum perlakuan pada hari ke 21 sampai umur 42 hari. Penelitian disusun dalam rancangan acak lengkap (RAL) dengan 8 perlakuan dan 4 ulangan (masing-masing 6 ekor). Perlakuan meliputi : bungkil kedelai dan tepung ikan (tanpa dibuat mikropartikel) 21% (T0), bungkil kedelai dan tepung ikan (tanpa dibuat mikropartikel) 18% (T1), protein mikropartikel 21% (T2), protein mikropartikel 18% (T3), T0 + *Lactobacillus* sp. 1,2 mL (T4), T1 + *Lactobacillus* sp. 1,2 mL (T5), T2 + *Lactobacillus* sp. 1,2 mL (T6), T3 + *Lactobacillus* sp. 1,2 mL (T7). Parameter yang diamati adalah total bakteri asam laktat (BAL), populasi *Coliform*, pH di ileum, laju digesta, pertambahan bobot badan harian (PBBH) dan bobot karkas. Data diolah dengan analisis ragam dan jika terdapat pengaruh nyata ($P < 0,05$) dilanjutkan dengan uji Duncan. Hasil penelitian menunjukkan bahwa ransum menggunakan protein mikropartikel yang ditambah *Lactobacillus* sp. 1,2 mL terjadi peningkatan populasi BAL dan PBBH, namun populasi *Coliform* menurun serta memperlambat laju digesta, tanpa berpengaruh terhadap bobot karkas. Kesimpulan penelitian adalah pemberian ransum protein mikropartikel terendah (18%) dengan penambahan *Lactobacillus* sp. 1,2 mL (T7) dapat memperbaiki kondisi fisiologis saluran pencernaan berdasarkan populasi BAL dan *Coliform* serta meningkatkan pertambahan bobot badan harian dengan bobot karkas yang sama.

Kata kunci : broiler, *Lactobacillus* sp., mikroba usus, protein mikropartikel

ABSTRACT

The objective of the study was to evaluate feeding effect of microparticle protein derived from soybean meal and fish meal combined with *Lactobacillus* sp. at 1.2 mL on physiological condition of digestive tract and productivity of broiler. Total of 192 broiler, with initial body weight was 481.0 ± 67.2 g, given dietary treatment starting on day 21 until day 42. The experiment was assigned in a completely randomized design with 8 treatments and 4 replications. Dietary treatment were 21% intact protein (T0), 18% intact protein (T1), 21% microparticle protein (T2), 18% microparticle protein (T3), T0+*Lactobacillus* sp.1.2 mL (T4), T1+*Lactobacillus* sp.1.2 mL (T5), T2+*Lactobacillus* sp.1.2 mL (T6), T3+*Lactobacillus* sp.1.2 mL (T7). Parameters measured were total ileal lactic acid bacteria (LAB), *Coliform* and pH, rate of passage, daily body weight gain (DWG) and also carcass weight. Data were subjected to analysis of variance and followed by Duncan multiple range test ($P < 0.05$). Experimental results showed that microparticle protein diet added with *Lactobacillus* sp. ($P < 0.05$) increased total LAB and DWG, but decreased *Coliform* population, and slowed down the rate of passage. However, carcass

weight was not significantly affected by treatments. Feeding microparticle protein (18%) with addition of *Lactobacillus* sp. (1.2 mL) can improve to be better condition of digestive tract based on higher LAB and lower *Coliform* populations, and increase daily body weight gain with the same carcass weight.

Keywords: broiler; intestinal microbe, Lactobacillus sp., microparticle protein

INTRODUCTION

Broiler is a meat type chicken that systematically selected to have high growth rate, therefore, it can achieve high meat production for market in a short raising period. For this purpose, it needs to prepare and serve diet containing high protein to fulfill the bird requirement based on the physiological period and rearing purpose. It is well known that when the diet composed of high source of protein the cost per production unit would be expensive. An effort can be implemented to reduce feed cost that is by lowering the portion of using protein source ingredients. However, the lack of protein supply brings about the consequence of protein deficiency for growing bird. On the other hand, protein supply in term of quantity and quality is the main limiting factor on chicken performance. Costs reduction by lowering dietary protein is a possible way, but it should be anticipated the possibility of low protein intake which can retard growth. Feeding low-protein diet should be accompanied by the efforts of increasing nutrients digestibility, especially proteins, in order to maintain the adequacy of protein supply so that it would not exert a negative effect on productivity. Processing feed component, especially protein source ingredient such as fish meal and soybean meal, to become microparticle protein can ensure high protein digestibility and absorption. Smaller particle sizes can be associated with larger surface area per volume of diet, thus allowing higher digestibility due to the intensive contact with digestive enzyme in poultry (Mingbin *et al.*, 2015)

There were very limited information concerning the study of feeding microparticle protein, but research about particle size of feed ingredients in particular were available elsewhere. Therefore, the present study was conducted to evaluate feeding effect of microparticle diet in broiler chicken. Feeding microparticle protein diet is suggested to be efficient when combined with natural additive such as probiotic *Lactobacillus* sp. Dietary inclusion of non-antibiotic additives, such as probiotics, have been increasingly important to be applied for poultry productivity

improvement. Probiotics can be used as an alternative to antibiotic growth factor because it can produce clean poultry product and consumer health friendly. Feed additives in the form of probiotic *Lactobacillus* sp. in general provide a positive impact through the decreased intestinal pH and the increase in population of beneficial bacteria such as lactic acid bacteria (LAB) and lowered total *Coliform* (Roberfroid *et al.*, 2010).

The development of intestinal microbes, primarily beneficial bacteria, can be affected by diets and nutritional supply, and finally it can promote growth of the animal. Probiotic inclusion into the low protein diet can be applied to be a foundation of developing improved diet in relation to nutrient absorption as well as to prevent poultry from gastrointestinal tract diseases caused by pathogenic bacteria (Pan and Yu, 2014). Research concerning the feeding combination of *Lactobacillus* sp. and microparticle protein diet has not been much done previously. Smaller particle size of proteins allows more effective absorption, due to an intensive work of digested enzyme supported by the healthy gastrointestinal tract. Therefore, research concerning feeding effect of microparticle protein diet added with *Lactobacillus* sp. as probiotic was conducted to evaluate their effect on dynamics of intestinal microbe and growth performance in broiler chicken.

MATERIALS AND METHODS

Experimental Animal and Diet

The experimental animals used in the present study were 192 birds of day old broiler chicken of Lohman strain with initial body weight of 481.0 ± 67.2 g. Microparticle preparation was performed based on the slightly modified method of Jambrak *et al.* (2014) using virgin coconut oil as particle stabilisator. Birds were maintained from 1 day to 20 days in a brooder cage given commercial diet, and at 21 days old the chickens were weighed and selected for uniform body weight prior to be moved randomly into the battery cage. The experimental diets (Table 1) were given from day 21 to 42 days old. *Lactobacillus* sp. was mixed

Table 1. Composition and Nutritional Content of Experimental Diet

Ingredient	Experimental Diet			
	Intact Protein 21 %	Intact Protein 18 %	Microparticle Protein 21 %	Microparticle Protein 18 %
	----- (%) -----			
Yellow corn	48	50.5	48	50.2
Rice bran	14	20	14	20
Intact soybean meal	27	21	–	–
Microparticle soybean meal	–	–	27	21
Intact fish meal	10	7.5	–	–
Microparticle fish meal	–	–	10	7.5
CaCO ₃	0.5	0.5	0.5	0.5
Vitamin and mineral	0.5	0.5	0.5	0.5
Total	100	100	100	100
Nutrient Composition *(%)				
Metabolizable energy **(kcal/kg)	2978.41	2948.32	2978.41	2948.32
Crude protein	21.29	18.12	21.29	18.12
Ether extract	2.81	2.57	2.81	2.57
Crude fiber	4.27	4.77	4.27	4.77
Calcium	1.03	0.88	1.03	0.88
Phosphorus	0.65	0.61	0.65	0.61
Methionine***	0.45	0.39	0.45	0.39
Lysin***	1.37	1.12	1.37	1.12
Arginine***	1.51	1.28	1.51	1.28

*Chemical analysis at the Laboratory of Nutrition and Feed Science, Faculty of Animal and Agricultural Sciences, Diponegoro University (2016); **Calculated value based on the formula of Carpenter and Clegg (1965); ***Calculated value based on Table of National Research Council (1994)

with a small portion of diet based on the treatment level and given in the morning at 05.00 am daily in order to ensure the feed containing *Lactobacillus* sp. can be completely consumed. Other portion of feed without *Lactobacillus* sp. were provided thereafter to fulfill the amount of daily requirement.

Experimental Design

The present experiment was assigned in a completely randomized design with 8 treatments and 4 replications (6 birds each). Dietary treatment were as follows : 21% intact protein diet (T0), 18% intact protein diet (T1), 21%

microparticle protein diet (T2), 18% microparticle protein diet (T3), T0+*Lactobacillus* sp. at 1.2 ml (T4), T1+*Lactobacillus* sp. at 1.2 ml (T5), T2+*Lactobacillus* sp. at 1.2 ml (T6), T3+*Lactobacillus* sp. at 1.2 ml (T7). One ml was equal to 10⁸ cfu.

Research Parameters

Parameters measured were total ileal LAB, *Coliform* and pH, rate of passage, daily body weight gain (DWG) and also carcass weight. Total LAB and *Coliform* were measured according to total plate count (TPC) based on the method of Fardiaz (1993). Potential hydrogen (pH)

measurement using pH meter. Rate of passage was determined using ferric oxide (Fe_2O_3) at 0, 03% as an indicator to make sure that the time of excreta dropping can be exactly observed. Daily body weight gain was calculated based on the difference between the final body weight and the initial weight divided by the duration of observation. Carcass weight obtained after chickens were slaughtered and cleaned up from the feathers, legs, head and viscera.

Statistical Analysis

Data were subjected to analysis of variance and followed by Duncan multiple range test at 5% probability when the treatment indicated significant effect.

RESULTS AND DISCUSSION

The results showed that *Lactobacillus* sp. inclusion at 1.2 ml into microparticle protein diet significantly ($P < 0,05$) increased ileal lactic acid bacteria (LAB) population (Table 2) and daily body weight gain (Table 3), but decreased total ileal *Coliform*, and slowed down the rate of passage (Table 2). However, carcass meat was not significantly affected by treatments (Table 3). The ileal lactic acid bacteria (LAB) in T4 to T7

treatments showed the highest value compared to other treatments (T0 to T3) due to the addition of *Lactobacillus* sp. at 1.2 ml. *Lactobacillus* sp. is known to have ability to ferment soybean oligosaccharide derived from soybean meal microparticle which produced lactic acid as well as short chain fatty acid (SCFA). Descriptive evaluation indicated the higher SCFA values in T7, namely 55.62, 29.61 and 16.24 mmol/L compared to T0 was 17.88, 10.63 and 10.03 mmol/L for acetate, propionate, and butirate, respectively, (Table 2). These metabolite products brought about the decrease in ileal pH due to inclusion of *Lactobacillus* sp. (T4, T5, T6, and T7 was 6.05, 6.00, 5.45 and 5.75, respectively) (Table 2) irrespective of dietary protein, and finally increased LAB population. Harimurti *et al.* (2007) showed that *Lactobacillus* was able to survive at the low gastrointestinal tract pH of chicken, therefore this condition implicated to the increasing population of endogenous lactic acid bacteria. On the other hand, decreased total pathogenic bacteria, in this case *Coliform*. Addition of *Lactobacillus* sp. (1.2 ml) was assumed to have a more dominant effect when combined with microparticle protein diet. However, when diet with both intact protein and microparticles without *Lactobacillus* sp. were fed

Table 2. Intestinal Bacteria Population, pH and Rate of Passage in Broiler Chicken Fed Microparticle Protein Added with *Lactobacillus* sp.

Variables	Treatments							
	T0	T1	T2	T3	T4	T5	T6	T7
Total ileal lactic acid bacteria (LAB), 10^8 cfu/g	1.75 ^b	2.37 ^b	2.55 ^b	2.50 ^b	3.82 ^a	3.72 ^a	3.87 ^a	4.55 ^a
Total ileal <i>Coliform</i> , 10^6 cfu/g	5.9 ^a	4.3 ^b	2.7 ^c	2.8 ^c	2.4 ^{cd}	2.2 ^{cd}	1.8 ^{cd}	0.8 ^d
Ileal pH	6.72 ^a	6.70 ^a	6.52 ^{ab}	6.45 ^{ab}	6.05 ^{abc}	6.00 ^{bc}	5.45 ^c	5.75 ^c
Rate of passage, min	180.1 ^b	188.2 ^b	240.2 ^a	245.9 ^a	245.0 ^a	240.0 ^a	267.9 ^a	284.9 ^a
Short chain fatty acid (SCFA), mmol/L								
Asetat	17.88	20.78	21.97	33.90	33.70	33.15	52.73	55.62
Propionat	10.63	12.37	12.08	19.69	20.53	23.53	25.69	29.61
Butirat	10.03	16.20	17.80	17.59	12.60	17.21	16.29	16.24

^{a-d}Mean values in the same raw followed by different superscript indicate significant difference ($P < 0.05$)

Table 3. Productivity in Broiler Chicken Fed Microparticle Protein Added with *Lactobacillus* sp.

Variables	Treatments							
	T0	T1	T2	T3	T4	T5	T6	T7
Daily body weight gain, g/bird/day	27.4 ^b	27.9 ^b	28.2 ^b	30.9 ^{ab}	30.6 ^{ab}	31.8 ^{ab}	31.09 ^{ab}	34.0 ^a
Carcass weight, g	751.0	756.3	710.3	705.3	745.8	735.8	732.8	759.8

^{a-b}Mean values in the same raw followed by different superscript indicate significant difference (P<0.05)

did not affect the population of LAB. The results of the present study were consistent with the previous finding (Krismiyanto *et al.*, 2014) that feeding normal level of intact protein diet added with prebiotic inulin showed the increase in LAB population. Similarly, Faradilla (2015) reported that a combination of inulin from dahlia tuber at 1.2% and *Lactobacillus* sp. at 1.2 ml (10^8 cfu/ml) produced the highest LAB amount compared to other treatments combination. The improved bacterial balance modulates the better intestinal health and it can be associated with the higher body resistance and productive performance.

Lactic acid bacteria (LAB) is also produce other metabolite such as bacteriocins that could be able to inhibit pathogenic bacterial growth. *Lactobacillus* sp. at 1.2 ml could compete with pathogenic bacteria such as *Coliform* in the ileum (5.9×10^6 cfu/g in T0 vs. 0.8×10^6 cfu/g in T7, Table 2), so that *Coliform* population substantially decreased (Table 2). Chichlowski *et al.* (2007) reported that *Lactobacilli* can prevent attachment of pathogenic bacteria such as *E.coli* and *Salmonella* on the intestinal lumen. The mechanism of probiotics work in altering the physical micro environment of the intestinal tract and inhibit pathogenic bacteria growth can be explained via two ways. First, probiotic organism competes with pathogens for nutrient, thus preventing them from acquiring energy for growth and function of gastrointestinal. Second, as it has been discussed previously that probiotics produce a variety of organic acid such as SCFA and lactic acid as an apart of their metabolic activity. Organic acids produced by the prebiotic bacteria could decrease pH of the gut and inhibit the survival of pathogenic bacteria such as *E. coli* and *Salmonella* (Haryati, 2011).

In relation to SCFA production which is attributable to fermentative activity of

Lactobacillus sp. addition is accomplished by the increase in intestinal LAB population. One mechanism with regard to the important interference of oligosaccharides derived from soybean meal called as soybean oligosaccharide may provide protection against harmful microorganism by substituting the soluble receptor analogues. In case of the present study, specific oligosaccharide in the form of soybean oligosaccharide can be obtained from the soybean meal treated with ultrasonic bath. Ultrasonic bath treatment did not only change intact protein to be microparticle but also lowered particle size of soybean oligosachharide causing more available either to deceive pathogenic bacteria through receptor substitution or to be function as “food source” for *Lactobacillus* sp. Reducing harmful bacteria to be vulnerable by inhibiting the formation of fimbriae and other membrane-based adenosines can be an effective control of digestive tract and microbial colonization in the mucosal tissue. The specificity of mucosal attachment provides potential control of gastrointestinal through the function of specific oligosaccharide structure (Ouwehand and Vaughan, 2006). Decreasing pH and increasing population of LAB due to the fermentation effect of soybean oligosaccharide provided an impact on medium viscosity of digesta and thus slowed down the rate of passage (180.1 min in T0 vs. 284.9 min in T7, Table 2). Potential hydrogen (pH) can be associated with the rate of passage (Hetland *et al.*, 2004). Further effect is that the rate of passage is closely related to nutrient digestibility because the slower rate of passage the higher nutrient digestibility. This condition was also supported by feeding effect of microparticle protein which can assumed that smaller diet particle can be more effectively attached by digested enzyme.

The increased lactic acid bacteria (LAB) and the decreased *Coliform* populations were also

supported by the slower rate of passage lead to healthier intestinal of the chicken and better growth of the villi. The better villi growth as an indication of intestinal health improvement was attributable to bacteriocin produced by lactic acid bacteria to suppress the growth of pathogenic bacteria at the competitive state in the small intestine (competitive exclusion) (Dicks and Bones, 2010). As a comparison, Adil *et al.* (2010) and Jamilah *et al.* (2014) reported that organic acids, such as citric acid, had a positive effect on the development and growth of intestinal villi, which leads to the increased nutrient absorption. The higher intestinal villi, the more nutrients can be absorbed due to the wider surface of digestive area. This phenomenon was consistent with those previously discussed that the decreased pathogenic bacteria population brought about healthier gastrointestinal and provided a positive impact on intestinal villi growth which facilitated the more nutrients absorption to accelerate body weight (Faradilla, 2015). The mechanism as described above is the important factor to increase protein supply for body protein deposition which leads to the increase in body weight. Suthama *et al.* (2010) indicated that meat protein deposition or meat protein mass was an indicator of quality broiler growth, depending on the differences in protein synthesis compared to protein degradation. Increased protein intake or supply is also supported by the slower rate of passage because the longer digesta spent in the intestine is assumed to be higher nutrient digestibility, especially protein.

The results of this study suggested that the increase in protein digestibility which supported the higher meat protein mass was closely related to daily weight gain. A comparative finding (Fajrih *et al.*, 2014) revealed that crossbred local chickens fed normal level of intact protein diet with added prebiotic inulin of dahlia tuber at 1.2% increased health status since heterophyl/lymphocyte ratio decreased and bursa fabricius weight increased, and improved body weight gain. It can, therefore, be assumed that effectiveness of natural feed additive supplementation, such as probiotic bacteria as well as prebiotic substances, can modulate the balance of intestinal microbes, depending on the birds strain, dietary protein level and condition. Clear phenomenon had been proved in crossbred Indonesian local chickens given reduced protein diet (18% vs. 20%) with the inclusion of *Lactobacillus sp.* at 1,2 mL improved productive

characteristics and performance. The increased meat protein mass and breast meat color (Abdurrahman *et al.*, 2016a), and the decreased fat mass and cholesterol of meat (Abdurrahman *et al.*, 2016b) were the parameters determining the quality of productive performances due to probiotic supplementing effect. Therefore, a positive effect of *Lactobacillus sp.* inclusion into the diet composed of microparticle protein, although with reduced protein content, was indicated in the present study.

CONCLUSION

Inclusion of *Lactobacillus sp.* at 1.2 mL into the lower (18%) microparticle protein diet can increase the population of lactic acid bacteria and daily weight gain, but decrease total ileal *Coliform* with slower rate of passage in broiler chickens.

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