

# The Oxidative Stress in Cataract Patients

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## ABSTRACT

**Background:** The recent studies on cataract formation focus on the primary role of the systemic oxidative stress which is generated outside the lens. Our research was directed to assess the oxidative stress by measuring the lipid peroxidation products in the form of the Thiobarbituric Acid Reactive Substances (TBARS) and the antioxidant enzyme levels in the blood. The antioxidant therapy may have a role to play in delaying the onset and the progression of age related cataracts.

**Material and Method:** This was a case control study. It comprised of 100 age matched subjects (50 with cataracts and 50 controls) with their ages ranging from 45- 75 years. Oxidative stresses such as the Thiobarbituric Acid Reactive Substances (TBARS) and the antioxidant enzymes, Superoxide Dismutase

(SOD) and Glutathione Peroxidase( GPX ) were investigated in all the patients and the controls.

**Results:** Significantly increased levels of serum lipid peroxide in the form of Malondialdehyde (MDA ) ( $p < 0.001$ ) were observed in the cataract patients as compared to the controls. Significantly decreased blood levels of SOD and GPX were observed in all the patients.

**Conclusion:** In the present study, it was concluded that oxidative stress plays an important role in the onset and the progression of cataracts. The pro-oxidant i.e. serum malondialdehyde (MDA) levels were increased in the cataract patients. The blood levels of the enzymatic anti-oxidants, SOD and GPX were decreased. The plasma TBARS can be used as biomarkers of the degeneration in the lens.

**Key Words:** Cataract age related, Plasma TBARS, Oxidative stress, Superoxide dismutase, Glutathione peroxidase, Lipid peroxidation(LPO)

## INTRODUCTION

Cataract is a complete or a partial opacification of sufficient severity on or in the human lens or in the capsule, which impairs the vision. It is one of the leading causes of reversible blindness in the world today. The pathophysiology behind the age related cataracts is complex and it has yet to be fully understood. It is believed that oxidation is a very early or initial event in the overall process in the sequence of events which lead to cataracts [1,2].

J.J Harding proposed a variety of factors which were implicated in the maturity onset of cataractogenesis: a low antioxidant defence capacity, high lipid peroxidation, an augmented non enzymatic glycosylation, a reduced chaperone function of the alpha crystallins and an increased permeability of the lens membrane [3,4].

The lipid peroxidation represents the oxidative tissue damage which is caused by hydrogen peroxide, the superoxide anions and the hydroxyl radicals, which results in the structural alteration of the membrane, with the release of the cell and the organelle contents and the loss of the essential fatty acids, with the formation of cytosolic aldehyde and peroxide products. Malondialdehyde (MDA) is the major end product of the free radical reaction on the membrane fatty acids [5].

SOD is an enzymatic antioxidant which provides defence, that acts by quenching  $O_2$  and converting it into  $H_2O_2$ . This protects the cell membrane from the damage which is caused by the Reactive Oxygen Species ( ROS). But the decreased SOD levels

may lead to increased lipid peroxidation, resulting in cellular rigidity and deformability [6]. An altered activity of SOD in the cataract patients has been revealed recently [3].

Glutathione Peroxidase (GPX) scavenges the highly reactive lipid hydroperoxide in the aqueous phase of the cell membrane. During aging, the lens loses its antioxidant potencies such as may be seen with the decrease of glutathione or the expression of the antioxidant enzymes [7].

Researchers have shown that the oxidative stress which is caused due to the accumulation of free radicals plays a role in the pathogenesis of cataracts and that this process can be prevented or ameliorated by antioxidants. Studies on the antioxidant status of the lens and the blood in cataract patients have been extensively reported. However, very few studies have been conducted on the Indian patients with cataracts. In the developing countries like India, cataracts evolve earlier in life and they are 3 times more prevalent than those in the developed countries [8]. The present study invites attention to the possible role of the Reactive Oxygen Species (ROS) in the progression of cataracts after estimating the levels of the antioxidant enzymes, SOD and GPX in the blood. The lipid peroxidation product, Malondialdehyde (MDA) was also estimated.

## MATERIALS AND METHODS

Fifty patients with cataracts, who were in the age range of 45-75 years, who presented to the Outpatients Department of Ram Lal

Eye hospital which is attached to Govt. Medical College, Amritsar, India, were included in the study. Before the start of the study, the approval of the institutional ethical committee was obtained.

A group of 50 normal healthy individuals who were age matched, who were from the same population, served as the controls. Clinical and biochemical research was carried out on all the subjects. During the selection of the subjects from both the groups, it was made sure that they were without a previous medical history of any chronic disease or metabolic disorder. A diagnosis of cataract was established by an ophthalmologist after doing a complete ocular examination. The patients who took any antioxidant drugs were excluded from the study.

After obtaining a written informed consent, venous blood was collected from the subjects under aseptic conditions by venipuncture by using a 10ml sterile disposable syringe and a needle. The serum was separated by centrifugation at 3000rpm for 10 minutes at room temperature. 3ml of whole blood was collected in a heparinized vial for the estimation of the GPX levels in the whole blood. The samples were stored at 4°C before analysis and all the samples were analyzed on the same day of the collection.

The serum lipid peroxide levels were measured by precipitating the lipoproteins with trichloroacetic acid and boiling them with thiobarbituric acid, which reacted with malondialdehyde to form a pink colour, as per the 'Kei satoh' method [9]. The resulting chromogen was extracted with n-butyl alcohol and the absorbance of the organic phase was determined at the wavelength of 530nm. The determined values were expressed in terms of malondialdehyde in nmol/ml.

Serum Superoxide Dismutase (SOD) was analyzed by applying the method of Marklund and Marklund (1974) which was modified by Nandi and Chatterjee in 1988 [10]. Glutathione peroxidase was estimated by the method of Paglia and Valentine [11] by using Ransel-Randox reagent kits which were manufactured by Randox Labs. Ltd. (Crumlin UK).

All the results were expressed as mean  $\pm$  SD. The statistical analysis was done by using the Student's t' test. The P values which were <0.001 was considered as highly significant.

## RESULTS

The present study was conducted on 50 cataract patients who were aged 45-75 years. 50 age matched healthy individuals served as the controls. The estimated mean serum MDA levels in the cataract patients and the controls were  $5.43 \pm 1.69$  nmol/ml and  $2.42 \pm 0.46$  nmol/ml. The serum lipid peroxide concentration in the form of MDA was significantly higher in the cataract patients ( $p < 0.001$ ) as compared to that in the controls [Table/Fig-1]. [Table/Fig-2] shows that the mean serum concentration of superoxide dismutase in the cataract patients and in the control group was  $2.75 \pm 0.40$  units/ml and  $4.25 \pm 1.20$  units/ml respectively. The level of the antioxidant enzyme, SOD was significantly decreased in the cataract patients as compared to that in the controls.

Subjects	No. of cases(n)	Range (nmol/ml)	Mean	$\pm$ SD	S.E
Controls	50	1.5-2.8	2.42	0.46	0.07
Patients	50	2.3-7.5	5.43	1.69	0.24

**[Table/Fig-1]:** Comparison of Serum MDA Levels in Controls and Patients Under Study

Subjects	No. of cases	Range units/ml	Mean	$\pm$ SD	S.E
Controls	50	2.8-6.66	4.25	1.20	0.17
Patients	50	2.0-3.3	2.75	0.40	0.06

**[Table/Fig-2]:** Comparison of Serum Sod Levels in Controls and Cataract Patients Under Study

Subjects	No. of cases	Range units/gm Hb	Mean	$\pm$ SD	S.E
Control	50	42.05-93.46	70.29	10.53	1.49
Patients	50	12.93-45.83	26.23	11.90	1.68

**[Table/Fig-3]:** Comparison of Blood Glutathione Peroxidase Levels In Controls and Patients Under Study

The mean glutathione peroxide level in the blood of the cataract patients was  $26.23 \pm 11.90$ . The blood level of GPX in the controls was  $70.29 \pm 10.53$ . The blood levels of GPX were decreased in the cataract patients as compared to those in the control group. The decreases were statistically significant [Table/Fig-3].

## DISCUSSION

Cataract is the leading cause of blindness, accounting for 50% of the blindness cases worldwide. Although significant progress has been made towards identifying the risk factors for cataract, there is no proven primary prevention or medical treatment for it. The surgical removal of the cataract remains the only therapy.

The ocular lens which is continually exposed to light and ambient oxygen, is at a high risk of photooxidative damage which results in a cataract. The oxygen free radicals appear to impair not only the lens crystallins which aggregate and precipitate, forming opacities, but also the proteolytic enzymes whose function is to eliminate the damaged proteins. Apart from an enzymatic defence system which consists of superoxide dismutase, catalase and glutathione peroxidase against the excited oxygen species, the lens contains the antioxidants vitamins C and E and presumably beta-carotene as another line of defence [11].

The pathophysiology behind the age related cataracts is complex and it has yet to be fully understood. It is believed that oxidation is a very early or initiating event in the overall process in the sequence of events which lead to the formation of cataracts [12-14]. Oxidative stress may result from an imbalance between the production of the reactive oxygen species and the cellular antioxidant defence mechanisms. In the cells of the eyes, the reactive oxygen species may initiate a surge of toxic biochemical reactions such as peroxidation of the membrane lipids and extensive damage of the proteins, which cause intracellular protein aggregation and precipitation [15].

Lipid peroxidation represents the oxidative tissue damage which is caused by hydrogen peroxide, the superoxide anions and the hydroxyl radicals, resulting in a structural alteration of the membrane, with the release of the cell and the organelle contents and the loss of cytosolic aldehyde and peroxide products. Malenaldehyde is a major end product of the free radical reaction on the membrane fatty acids.

In our study, an increase in the MDA level [Table/Fig-1] was seen, which indicated an increase in the oxidative stress or a decrease in the antioxidant defence mechanism. In the cases of the development of age related cataracts, LPO may also be the real cause

of destruction of the plasma membrane of the lenticular fibres and the subsequent oligomerization of the crystalline lens [16]. The lipid peroxidation may be linked to the premature development of senile cataracts [17]. Therefore, it can be stated that LPO is one of the possible causes of the cataract progression.

SOD is an enzymatic antioxidant while provides the first line of defence that acts by quenching  $O_2^-$  and converting it into  $H_2O_2$ . There may be two reasons for the lowering of the SOD levels:

1) As more and more ROS like  $O_2^-$  are produced, SOD will be used up in the process when it converts  $O_2^-$  to  $H_2O_2$ .

2)  $H_2O_2$  also causes inhibition of the SOD activity. There are several classes of SOD that differ in their metal binding ability, their distribution in the different cell compartments and in their sensitivity to various reagents.

Among these, the Cu and the Zn superoxide dismutase  $SOD_1$  is widely distributed and it comprises 90% of the total SOD. This ubiquitous enzyme, which requires Cu and Zn for its activity has a great physiological significance and a therapeutic potential.

SOD removes  $O_2^-$  by catalyzing a dismutation reaction which involves the oxidation of one  $O_2^-$  to oxygen and the reduction of another  $O_2^-$  to hydrogen peroxide.



The discovery of SOD led to the realization that the  $O_2^-$  which was formed in vivo in the living organisms and SOD removes (Halliwell, 1991).

Chinese researchers found the lower activities of several RBC antioxidant enzymes (SOD and catalase) and a significantly decreased erythrocyte GPX level in the subjects with the senile lens changes [18]. The POLA study showed a strong association of the high levels of the erythrocyte SOD with an increased risk of nuclear cataracts [19]. In our study, a significant decrease in the serum SOD level ( $P < 0.001$ ) was observed in the cataract patients as compared to that in the controls [Table/Fig-2].

The GSH antioxidant system is the body's powerhouse for diffusing and disposing the radicals that threaten the cell and tissue and cause organ damage, thus slowing the approach of age [20]. One of the several possibilities for the occurrence of a lower GSH conc. in the blood is an increased GSH consumption for the removal of peroxides and xenobiotics [21].

GSH is the obvious compound which defends the lens against oxidative insults, being directly involved in reducing the disulfides, being a pivotal cofactor in the detoxication of  $H_2O_2$  and acting as a free radical quencher.

The present study also indicated an age related decrease in the glutathione peroxidase activity [Table/Fig-3].

Our results confirmed some previous findings that had correlated with human cataracts [22]. It has been suggested that a decrease in the antioxidant status of the erythrocytes may increase the oxidative damage in the tissues, which includes the oxidative modification of the lens proteins which are observed in cataracts. However, in contrast to these data, the increased blood levels of the antioxidant enzymes have been reported to be associated with cataracts [22-24].

## CONCLUSION

The oxidative stress of the lens had a direct influence on the solubility of the lens proteins, which led to an increase in the opacity of the lens. The antioxidant enzyme activity levels reflected the changes which took place in the development of senile cataracts. At present, the only remedy is surgical removal of the cataractous lens and substituting it with a lens which is made of synthetic polymers. Therefore, there is a search for an intervention which will help in delaying the onset and in the slowing down of the aetiology of the cataracts. The assays of these enzymes and the plasma TBARS can be used as the biomarkers of the degeneration in the lens.

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## REFERENCES

- [1] Cekic S, Zlatanovic G, Cvetkovic T, Petrovic B. The oxidative stress in cataractogenesis. *Bosian Journal Of Basic Medical Sciences*. 2010; 10 (3): 265-69.
- [2] Mohan M, Sperduto RD, Augra SK, Milton RC, Mathur RL, Underwood BA et al. An Indo-US case control study on age related cataracts. *Arch Ophthalmol*. 1989; 107: 670- 76.
- [3] Virgolici B, Stoian I, Muscurel C, Maracine M, Popescu L, Moraru C, et al. The systemic redox modifications in senile cataracts. *Rom. J. Intern. Med*. 2009; 47(3): 279-87.
- [4] Harding JJ. The physiology, biochemistry, pathogenesis and the epidemiology of cataracts. *Current Opinion in Ophthalmology*. 1992; 3: 3-12.
- [5] Donma O, Yorulniaz E, Pekel H, Suyugul N. The blood and lens lipid peroxidation and the anti-oxidant status in senile individuals and in senile and diabetic cataractous patients. *Curr. Eye Res*. 2002; 25(1): 9-16.
- [6] Yamanaka N, Fukushima M, Koizami K, Nishida K, Kato T, Ota K. Enhancement of the DNA chain breakage by the bleomycin and biological free radicals producing system. *Oxygen Biomembrane (new York) North Holland*. 1998; 56-69.
- [7] Saadat M, Farvardin, Jahroni M, Saadat H. The null genotype of glutathione-S-transferase M1 is associated with the senile cataract susceptibility in non smoker females. *Biochem Biophys. Res Commun*. 2004; 1287-91.
- [8] Satoh K. The serum lipid peroxide levels in cerebrovascular disorders which were determined by a new colorimetric method. *Clin Chim Acta*. 1978; 90: 37-43.
- [9] Marklund, Marklund. Assaying the superoxide dismutase activity in animal tissues. Modified by Nandi et al. *J Biochem*. 1988; 13(3): 305-15.
- [10] Paglia DE, Valentine WN. Studies on the quantitative and the qualitative characterization of the erythrocyte glutathione peroxidase. *J Lab Clin Med* 1967; 70: 158-69.
- [11] David LL, Shearer TR. The role of proteolysis in the lenses: A review. *Lens Eye Tox Res*. 1989; 6: 725-47.
- [12] Chitkara DK. Cataract formation mechanisms second ed Yanoff M, *Duker JS ed Ophthalmology Mosby*. 2004; 4: 273-79.
- [13] Boulton M, Saxby I.A. Age changes second ed Yanoff M, *Duker JS Ophthalmology Mosby*. 2004; 4: 261-68.
- [14] Taylor A, Jacques P, Chylack LT, Haukinson SF, Khu PM, Rogers et al. The long term intake of vitamins and carotenoids and the odds of early age related cortical and posterior sub capsular lens opacities, 1- 4. *Am J Clin. Nutr*. 2002; 75: 540-49.
- [15] Boscica F, Grattagliano L, Vandermiale G et al. The protein oxidation and the lens opacity in humans. *Invest. Ophthalmol. Vis. Sci*. 2000; 41: 2461-65.
- [16] Babizhayev MA. The accumulation of lipid peroxidation products in human cataracts. *Acta ophthalmol*. 1989; 67: 281-87.
- [17] Virgolici B, Stoian I, Muscurel C, Maracine M, Moraru C, Dinu V. The plasma redox status in age related cataracts. *Romanian Journal of Internal Medicine*. 2009; 47(3): 279-87.

- [18] Xue AN, Cai QY, Wang SQ, Zhou AS, Li Fu P. *Chen Biomed Environ Sc.* 1996; 9 (2-3): 144-48.
- [19] Delcourt C, Cristol JP, Tessier F, Leger CL, Mitchel F, Papoz L. The risk factors for cortical, nuclear and posterior subcapsular cataracts: The POLA study. *Pathologies Oculaires Liees Epidemiol.* 2000 Mar; 151 (5): 497-504.
- [20] Leutner S, Schindowski K, Frolich L, Muller WE. An enhanced ROS generation in the lymphocytes in Alzheimers patients. *Pharmacopsychiatry.* 2005; 38(6): 312-15.
- [21] Meister A. The selective modification of the glutathione metabolism. *Sciences.* 1983; 220(4596): 472-77.
- [22] Nourmohamadi I, Ladan G, Mehdi M, Abbas GJ. Evaluation of the erythrocyte glutathione peroxidase, superoxide dismutase and the total antioxidant levels in cataract patients. *Arch Iranian Med.* 2001; 4: 123-26.
- [23] Delcourt C, Cristol JP, Leger GL, Descamps B, Papoz L. The association of the antioxidant enzymes with cataracts and age related macular degeneration. *Ophthalmology.* 1999; 106: 215-22.
- [24] Delcourt C, Carriere I, Delage M, Descamps B, Cristol JP, Papoz L. The association of cataracts with antioxidant enzymes and other risk factors: the French age related eye diseases (POLA) prospective study. *Ophthalmology.* 2003; 110: 2318-26.

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