

Evaluation of Measles Vaccine Virus as a Vector to Deliver Respiratory Syncytial Virus Fusion Protein or Epstein-Barr Virus Glycoprotein gp350

Hoyin Mok[§], Xing Cheng[§], Qi Xu, James R. Zengel, Bandita Parhy, Jackie Zhao, C. Kathy Wang and Hong Jin^{*}

MedImmune LLC., 319 North Bernardo Ave, Mountain View, California, USA

Abstract: Live attenuated recombinant measles vaccine virus (MV) Edmonston-Zagreb (EZ) strain was evaluated as a viral vector to express the ectodomains of fusion protein of respiratory syncytial virus (RSV F) or glycoprotein 350 of Epstein-Barr virus (EBV gp350) as candidate vaccines for prophylaxis of RSV and EBV. The glycoprotein gene was inserted at the 1st or the 3rd position of the measles virus genome and the recombinant viruses were generated. Insertion of the foreign gene at the 3rd position had a minimal impact on viral replication *in vitro*. RSV F or EBV gp350 protein was secreted from infected cells. In cotton rats, EZ-RSV F and EZ-EBV gp350 induced MV- and insert-specific antibody responses. In addition, both vaccines also induced insert specific interferon gamma (IFN- γ) secreting T cell response. EZ-RSV F protected cotton rats from pulmonary replication of RSV A2 challenge infection. In rhesus macaques, although both EZ-RSV F and EZ-EBV gp350 induced MV specific neutralizing antibody responses, only RSV F specific antibody response was detected. Thus, the immunogenicity of the foreign antigens delivered by measles vaccine virus is dependent on the nature of the insert and the animal models used for vaccine evaluation.

Keywords: Measles vaccine, Edmonston-Zagreb, RSV F, EBV gp350, Cotton rats, Rhesus monkeys.

INTRODUCTION

Live attenuated viruses have been developed and used to help protect humans from various infectious diseases. Live attenuated vaccine viruses simulate natural infection and are highly immunogenic and effective in inducing both humoral and cellular immune responses. Because of their demonstrated safety and effectiveness against infectious diseases, different live attenuated vaccine viruses have been explored as vectors to express heterologous antigens [1]. Measles vaccine (MV) strain has a long history of demonstrated safety in humans. It induces long lasting immunity including antibody and T cell responses [2-4]. Aerosol administration of measles vaccine is also being developed and has been shown to induce similar or better immunogenicity compared to conventional subcutaneous or intramuscular vaccination [5, 6]. This aerosol administration route may offer advantages in helping to protect individuals from mucosal pathogens.

The development of reverse genetics for negative stranded RNA viruses including measles virus has made it possible to explore the utility of the negative stranded RNA viruses to deliver foreign antigens. Measles vaccine virus has been explored by several laboratories to express foreign antigens such as the spike and nucleocapsid proteins of SARS coronavirus, HIV gp160, HPV L1 and hepatitis B surface antigens [7-10]. The vectored MV induces robust antibody and T cell responses against both insert antigens

and MV [7-18], making it an attractive option for vaccination against measles as well as other infectious agents. Although some studies showed that MV could boost measles virus immune response in school aged children who had previously received measles vaccine [19, 20], the pre-existing immunity to measles virus may reduce the immunogenicity of the vectored measles vaccine [21, 22]. Thus, the utility of MV vector in measles seropositive population remains to be determined. Several studies performed in the early 1980s in developing countries revealed that high seroconversion rates were achieved with EZ measles vaccine despite the presence of high titer of maternal measles virus antibody [23]. Therefore, the EZ strain might be immunogenic in populations with histories of measles vaccination or preexisting MV immunity.

In this study, we explored the use of the EZ vaccine strain to deliver viral antigens for prevention of RSV and EBV associated diseases. RSV causes severe lower respiratory tract illness in infants and the elderly. Bronchiolitis from RSV infection in infants is associated with recurrent wheezing and asthma in childhood [24]. No vaccine is available to prevent morbidity and mortality from RSV infection. The surface fusion (F) protein of RSV is a major antigenic protein that induces neutralizing antibodies, CD8⁺ cytotoxic T lymphocytes, and protection against subsequent RSV challenge in mice or cotton rats [25]. High levels of F neutralizing antibodies are sufficient to protect the lower respiratory tract infection [26] and have been used for prophylactic purpose in infants.

EBV is a gamma herpes virus that causes infectious mononucleosis in adolescents [27]. In addition, EBV has been associated with certain malignancies including Burkitt's lymphoma, post-transplantation lymphoprolifera-

*Address correspondence to this author at the MedImmune, 319 North Bernardo Ave, Mountain View, CA 94043, USA; Tel: 650 603 2367; Fax: 650 603 3367; E-mail: jinh@medimmune.com

[§]Both authors contributed to the work equally.

tive disease, lymphoma and nasopharyngeal carcinoma [28]. There is no prophylactic EBV vaccine available. The surface glycoprotein gp350, which binds to human CD21 receptor on B cells to mediate viral entry, has been identified as a protective antigen. The adjuvanted EBV gp350 protein has been shown to be efficacious in a randomized phase II study in preventing infectious mononucleosis [29]. Here, we report the construction of an infectious measles virus by reverse genetics system and the insertion of the F protein of respiratory syncytial virus or gp350 of EBV into MV virus as a separate transcriptional unit. The vectored viruses were then evaluated in cotton rats and rhesus macaques for their immunogenicity.

MATERIALS AND METHODS

Viruses and cells. RSV A2 strain was obtained from ATCC (Manassas, MA). EZ vaccine strain was from Serum Institute of India (Pune, India). Vero and HEP-2 cells were maintained in Dulbecco's modified Eagles's medium (Invitrogen, Carlsbad, CA) supplemented with 2% fetal bovine serum (FBS), 2mM L-glutamine, and 1%

penicillin/streptomycin at 37°C with 5% CO₂.

Construction of Measles vaccine virus rescue system.

Antigenomic cDNA was synthesized by DNA2.0 (Menlo Park, CA) based on the sequence of live attenuated measles virus Edmonston-Zagreb strain (NCBI DNA accession number F266290). The T7 RNA polymerase promoter sequence was added to the 5' end of the antigenomic cDNA and the hepatitis delta virus ribozyme and a T7 RNA polymerase terminator sequences were added to the 3' end of the antigenomic cDNA. Sequence at nucleotides 5502 (Sca I) and 9343 (Mlu I) was modified to facilitate cDNA cloning. The three amino acid residues at positions 3682, 3862 and 10555 (3682_{K→R}, 3862_{S→N}, 10555_{K→R}) that were different from the EZ vaccine strains were corrected by site-directed mutagenesis. The MV antigenomic cDNA was cloned into pUC18 vector and denoted as pEZ (Fig. 1). Supporting plasmids encoding measles N, P and L genes were cloned into pCITE plasmids under the control of T7 promoter as previously described and were designed pN, pP and pL respectively [30].

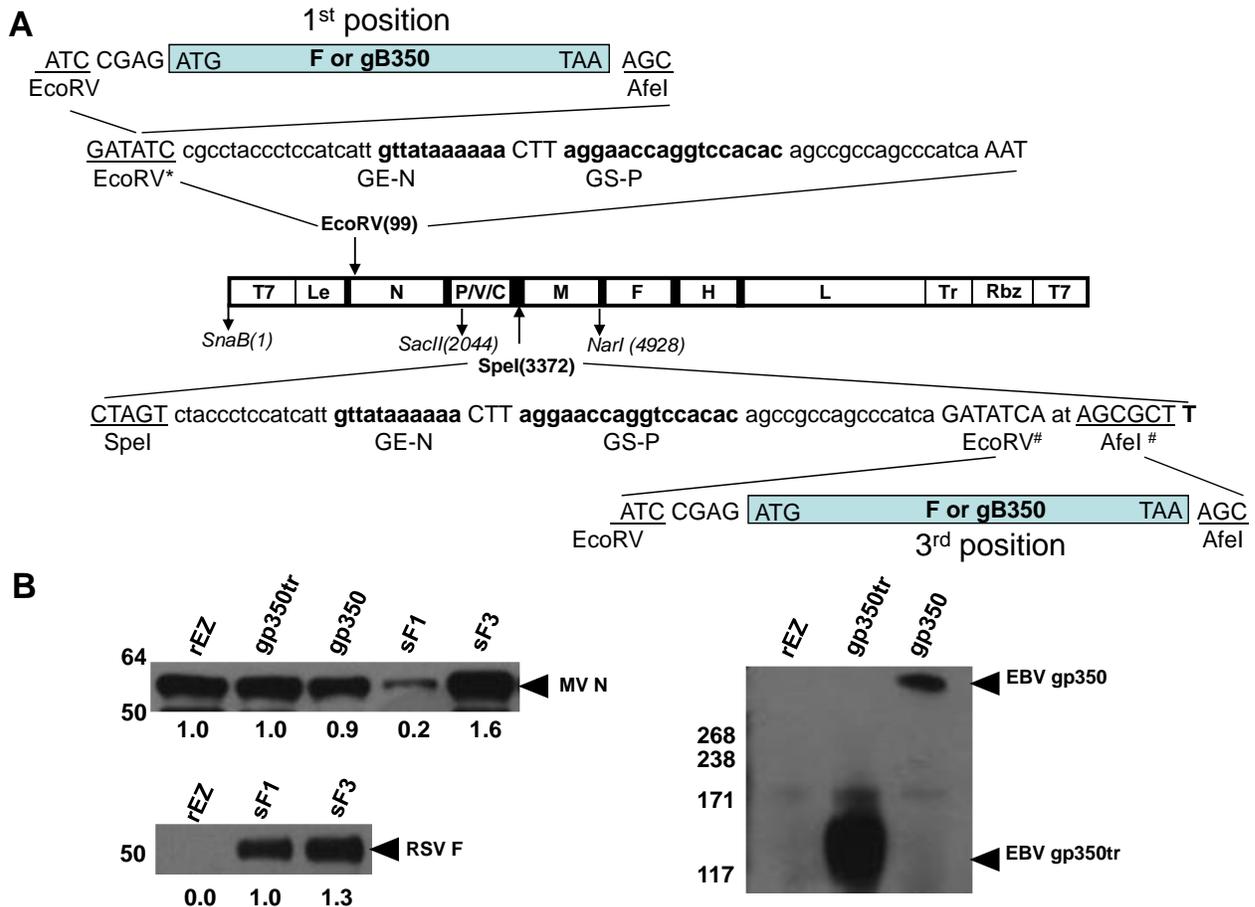


Fig. (1). Construction of measles vaccine vectors encoding ectodomains of RSV F and EBV gp350 and protein expression in virus-infected Vero cells. (A) RSV F ectodomain and EBV gp350 ectodomains were inserted in the 1st and 3rd transcription units of the measles Zagreb strain genome. Insertion was performed in the non-coding region 3' proximally of the MV N gene or between P and M genes of pEZ under the control of additional measles transcriptional units containing gene stop (GE)-gene start (GS) sequences. *RSV F gene (aa. 1-524) was inserted into EcoRV site using blunt ligation. #EBV gp350 full length (aa. 1-960), EBV gp350 truncated (aa. 1-470) or RSV F gene was inserted between EcoRV and Afel site. (B) Vero cells were infected at an MOI of 3.0 with measles viruses encoding soluble RSV F (sF1 or sF3) or soluble EBV gp350 (gp350 or gp350 tr). After 18 h, western blots were used to detect presence of measles virus nucleoprotein in infected Vero cell lysates or RSV F protein, EBV gp350 protein in the supernatant. Black arrowheads indicate the predicted molecular masses of the proteins. Relative intensity of the MV N protein bands was expressed as the intensity of the band relative to the rEZ control.

Construction of measles virus vectored antigenomic cDNA. The ectodomain of RSV F and EBV gp350 ectodomains were inserted into the non-coding region 3' proximal of the N gene or in the junction between the P and M genes of pEZ (Fig. 1). Each inserted gene contained measles virus transcriptional unit consisting of the sequences of N gene stop (GE-N) and P gene start (GS-P). To insert the RSV F gene into the promoter proximal location, a pUC-SnaBI/SacII (pUC-S/S) subclone that contained the T7 promoter and MV sequence of 1 to 2044 with a silent mutation at nt 821 to knockout EcoRV enzyme site was generated. A gene cassette containing the ectodomain of RSV-F (aa 1-524) followed by GE-N and GS-P sequence were inserted into EcoRV site at nt 99. The genomic DNA was in multiples of 6 nucleotides (the rule of six). The SnaB I-Sac II fragment was swiped back to pEZ antigenomic cDNA (Fig. 1A) and the resulting cDNA was denoted as pEZ-RSVsF1.

To insert the RSV F gene and EBV gp350 gene into pEZ between the P and M genes, a pUC-SacII/NarI (pUC-S/N) subclone that contained the MV sequence of 2045 nt to 4924 nt was constructed. A gene cassette containing gene end (GE) of N, gene start (GS) of P and ectodomain of RSV-F (aa 1-524), full length (aa 1-861) or truncated (aa 1-470) ectodomain of gp350 at SpeI site (nt 3372). The Sac II to Nar I fragment was swiped back to pEZ antigenomic cDNA (Fig. 1A) and the resulting cDNA was denoted as pEZ-RSVsF3, pEZ-EBVgp350 and pEZ-EBVgp350tr respectively.

Generation of recombinant measles virus and vectored measles viruses. Rescue of the recombinant viruses from MV antigenomic cDNA was performed as previously described [16, 30, 31]. Briefly, HEP-2 cells were infected with MVA-T7 at a multiplicity of infectivity (moi) of 5 for an hour prior to transfection. A DNA mixture of pEZ (0.8 µg), pN (0.4 µg), pP (0.4 µg) and pL (0.2 µg) were co-transfected into MVA-T7 infected HEP-2 cells using Lipofectamine 2000 (Invitrogen, Carlsbad, CA). The culture medium was replaced with DMEM containing 2% FBS after overnight incubation at 37°C. The transfected cells were incubated for an additional 3 to 4 days and the culture supernatant was used to infect fresh Vero cells to amplify the rescued virus. The rescued viruses were then plaque purified twice in Vero cells. The viruses rescued from pEZ, pEZ-RSVsF1, pEZ-RSVsF3, pEZ-EBVgp350 and pEZ-EBVgp350tr were designated as rEZ, sF1, sF3, gp350 and gp350tr, respectively.

Evaluation of recombinant measles viruses for their replication in cell cultures. Replication kinetics of rEZ, gp350, gp350tr, sF1 or sF3 viruses was compared in Vero cells. Vero cells in six-well plates were infected with each virus at a MOI of 0.01 in triplicates. After a 1 hour adsorption at room temperature, the cells were washed three times with phosphate-buffered saline (PBS) and incubated with 2 mL of complete DMEM at 35°C with 5% CO₂. Aliquots of culture supernatant were collected daily from each well for 7 days and stored at -80°C until virus titration. Viral titers were determined by plaque assay in Vero cells as previously described [32]. Replication kinetic was performed twice in triplicate and each supernatant sample was titered in duplicate.

Western blot. Vero cells were infected at a MOI of 3.0 with rEZ, gp350, gp350tr, sF1 or sF3 for 18 hours at 37°C. Culture supernatant and cells were harvested separately. The cells were scraped and pelleted for 10 sec at 6,000 rpm in microfuge tubes and lysed in lysis buffer (50mM Tris-HCl, 150 mM NaCl, 1% Triton X-100, 0.5% [vol/vol] protease inhibitor cocktail, pH 8.0) (Sigma, St. Louis, MO) for 10 min on ice. The resulting cell lysates were cleared by centrifugation at 13,000 rpm for 5 min.

Proteins from culture supernatant or cell lysates were separated by electrophoresis and transferred onto a PVDF membrane. Viral proteins were detected with anti-measles nucleoprotein (clone 2D7, Santa Cruz Biotechnology), polyclonal rabbit EBV gp350 [33] or monoclonal antibody against RSV F proteins (palivizumab, MedImmune, Gaithersburg, MD) antibodies. Intensities of the bands were quantified by ImageQuant software (GE Healthcare, Piscataway, NJ). Relative intensity was determined by comparing pixel intensity of the band relative to the rEZ control.

Cotton rat studies. Specific-pathogen-free 6- to 8-week-old cotton rats were purchased from Harlan (Indianapolis, IN). The animal study protocol was approved by MedImmune's Institutional Animal Care and Use Committee. Animals were anesthetized with isoflurane by inhalation and vaccinated intramuscularly (i.m.) with 10⁵ PFU of rEZ, gp350tr, gp350, sF1 or sF3 in a 200 µl inoculum volume. The control groups were inoculated intramuscularly (i.m) with PBS, 10⁵ PFU of UV-inactivated sF3, or intranasally (i.n) with 10⁶ PFU of RSV A2 strain. The animals were boosted with the same dose of viruses 4 weeks later.

The animals were observed for clinical signs daily and bled at 14-day intervals to measure antibody responses. Twenty-eight days after the second immunization, the cotton rats from measles virus vectored RSV vaccine groups were challenged with 10⁶ PFU of RSV A2 strain i.n. The lungs were harvested on day 4 post-challenge and viral titers were determined by plaque assay.

Rhesus macaque study. Adult rhesus macaques aged 3-9 year-old were used for the immunogenicity study at Valley Biosystems (West Sacramento, CA). All experimental procedures were approved by the Institutional Animal Use and Care Committee at MedImmune and Valley Biosystems. Measles vaccine immunization history of the rhesus macaques was confirmed by neutralization assay prior to the study. A total of 16 rhesus macaques (8 measles virus sero-positive, 8 naïve) were used in the study. Half of the animals (4 measles virus sero-positive, 4 naïve) received two doses of gp350 intramuscularly using 10⁵ PFU/inoculation and the other half received two doses of sF3 at the same dose intramuscularly. The inoculations were performed at 4 week intervals. Blood was drawn from each animal every 14 days to assess immune responses. Animals were anesthetized with ketamine intramuscularly at each procedure. Clinical symptoms were monitored by veterinarians for the duration of the study.

EBV gp350 and RSV F protein-specific ELISA assay. EBV gp350- or RSV F-specific enzyme-linked immunosorbent assay (ELISA) were performed to determine the levels

of gp350 or F protein-specific antibodies. Briefly, 100 ng of purified recombinant soluble EBV gp350 or RSV F protein expressed from mammalian cells was adsorbed onto Immulon 2B plates overnight in PBS at 4°C. The plate was blocked with 5% BSA in PBS. Serial dilutions of animal sera in PBS/1%BSA were added to the plate and incubated for an hour at room temperature. After washing, HRP-conjugated anti-cotton rat IgG (Immunology Consultants Laboratory, Newberg, OR) or anti-rhesus macaques IgG (Rockland Immunochemicals, Boyertown, PA) were added and incubated for 1 hour. Finally, the plate was developed with tetramethylbenzidine peroxidase substrate (KPL, Rockford, IL) and stopped with HCl. Absorbances of the samples were read at 450 nm and the titer for each sample was calculated by determining the dilution at which the absorbance reached 2 times over the reading of no serum control. The lower limits of detection were 3.3 log₂ and 6.6 log₂ for EBV gp350 ELISA and RSV F ELISA, respectively.

Neutralization assays. Measles virus specific neutralizing antibodies were determined as described previously [34] in 96-well plates using Vero cells. Neutralizing antibody titer is defined as the highest dilution that inhibits 50% CPE. RSV specific neutralizing antibody was determined by the plaque reduction neutralization assay as described previously [35]. The lower limit of detection was 4.3 log₂ and 3.0 log₂ for MV microneutralization and RSV plaque reduction assays, respectively. EBV specific neutralizing antibody titer was determined by the method described by Sashihara *et al.*, [36].

ELISPOT assay. ELISpot assays were performed with spleens from vaccinated and control cotton rats 14 days after the second dose and peripheral blood mononucleocyte cells (PBMC) of rhesus monkeys 28 days after the second dose. Splenocytes were isolated as previously described [37]. Splenocytes were counted using Vi-Cell (Beckman Coulter, Brea, CA). Gamma interferon (IFN-γ) and Interleukin 4-secreting T cells were quantified in an enzyme-linked immunospot (ELISpot) assay using R&D DuoSet ELISA System for cotton rat IFN-γ and IL-4 antibodies (R&D Systems, Minneapolis, MN) as previously described using 10 μg/mL of EBV gp350 or RSV F proteins for stimulation [38]. Spots were scanned and counted by ImmunoSpot spot Analyzer (Cellular Technology, Cleveland, OH). The numbers of IFN-γ or IL-4 secreting cells were expressed as the number cells per 10⁶ cells.

Statistics. GraphPad Prism software was used to analyze the data (GraphPad Software Inc., San Diego, CA). All data were expressed as the mean and standard error. The data were also analyzed by Mann-Whitney rank sum test to compare data distribution between any two experimental groups. Titer comparison between viruses in replication kinetic experiment was performed using the Kruskal-Wallis test, assuming non-parametric distribution.

RESULTS

Cloning and Expression of RSV and EBV Antigens from Measles Virus

The reverse genetics system for measles Edmonston-Zagreb (EZ) vaccine virus was established by transfecting

MV antigenomic plasmid pEZ together with pP, pN and pL into MVA-T7 infected HEp-2 cells to recover infectious EZ virus. To generate rEZ expressing RSV F or EBV gp350, the F ectodomain of RSV and EBV gp350 were cloned into the MV antigenomic cDNA (Fig. 1A). The F was cloned at the two positions of the viral genome: one proximal to the N gene (1st position) and the other between the P and M genes (3rd position). EBV gp350 ectodomain and its N-terminal half of the gp350 (amino acid 1-470, gp350tr) that encoded the receptor binding site to CD21 [39] were also cloned into the EZ genome at the 1st and 3rd positions. Recombinant EZ with RSV F, EBV gp350 and EBV gp350tr inserted at the 1st or 3rd position were generated. However, viruses with gp350 and gp350tr inserted at the 1st position replicated poorly *in vitro* and were not evaluated further. The rescued recombinant viruses expressing RSV F or EBV gp350 or gp350tr were designed as sF1 (F at the 1st position), sF3 (F at the 3rd position), gp350 and gp350tr (both at the 3rd position), respectively.

To confirm that these recombinant measles viruses expressed the desired antigens, the expression of the inserts in infected Vero cells was examined by Western blotting analysis. In addition to MV N protein that was detected in infected cell lysate (Fig. 1B), RSV F and EBV gp350 were detected in the culture supernatant of the infected cells (Fig. 1B), indicating the production of secreted form of RSV F and EBVgp350 proteins from virus infected cells. The observation of lower MV N expression in Vero cells infected with sF1 compared to other viruses may be attributed to the insertion of RSV F proximal to the MV N gene, which moved the MV N gene to the 2nd position of the genome and reduced its mRNA transcript level. The relative expression of RSV F to MV N, however, was shown to be higher for sF1 compared to sF3. This is consistent with the observation that genes proximal to 3' end of the genome are transcribed more abundantly than those downstream and more RSV F was produced from sF1 than sF3 per virion.

Viral replication efficiency of these recombinant viruses was assessed by growth kinetics in Vero cells. Both recombinant EZ (rEZ) and EZ vaccine viruses grew to a peak titer of 7.9 log₁₀ PFU/mL on day 5 (Fig. 3A). rEZ with either RSVF (Fig. 2A) or EBVgp350 (Fig. 2B) inserts had comparable growth kinetics but slightly lower (between 0.1-0.7 log₁₀ lower) than rEZ. The peak titers between the viruses were not statistically different. sF3 had a faster growth kinetics and reached peak titer on day 3 instead of day 5. Therefore, insertion of RSV F or EBV gp350 into the genome of EZ did not significantly reduce viral growth *in vitro*.

Evaluation of Antibody Response in Cotton Rats

Cotton rats had previously been demonstrated to be semi-permissive to measles virus infection [40] and are the best available small animal model for MV infection. In addition, cotton rats are susceptible to RSV infection and have been used in the evaluation of RSV vaccines [41, 42] and anti-RSV antibody investigated for prophylactic application [43]. The ability of rEZ vectored vaccines to induce humoral responses to measles virus and RSV F or EBV gp350 protein were assessed in cotton rats. Cotton rats were immunized

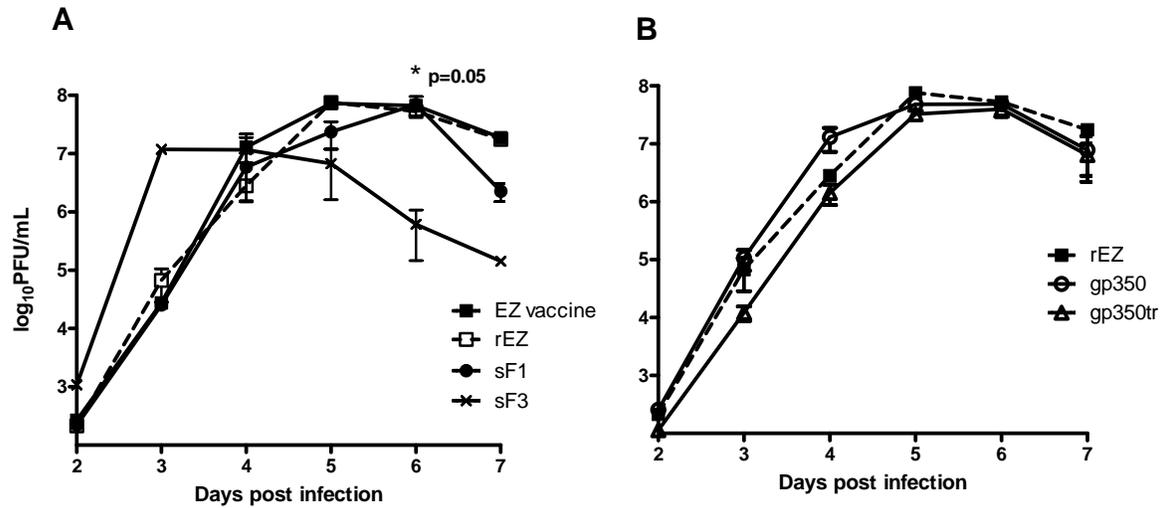


Fig. (2). Growth kinetics of measles vectors encoding soluble RSV F or soluble EBV gp350 was similar to that of recombinant measles constructs. Vero cells were infected at MOI of 0.01 with measles vectors. Cells were incubated at 35°C for 7 days. Cells and supernatant were sampled daily for virus titers using standard plaque assay. * denotes statistically significant difference in titer of sF3 versus sF1, rEZ and EZ using the Kruskal-Wallis test.

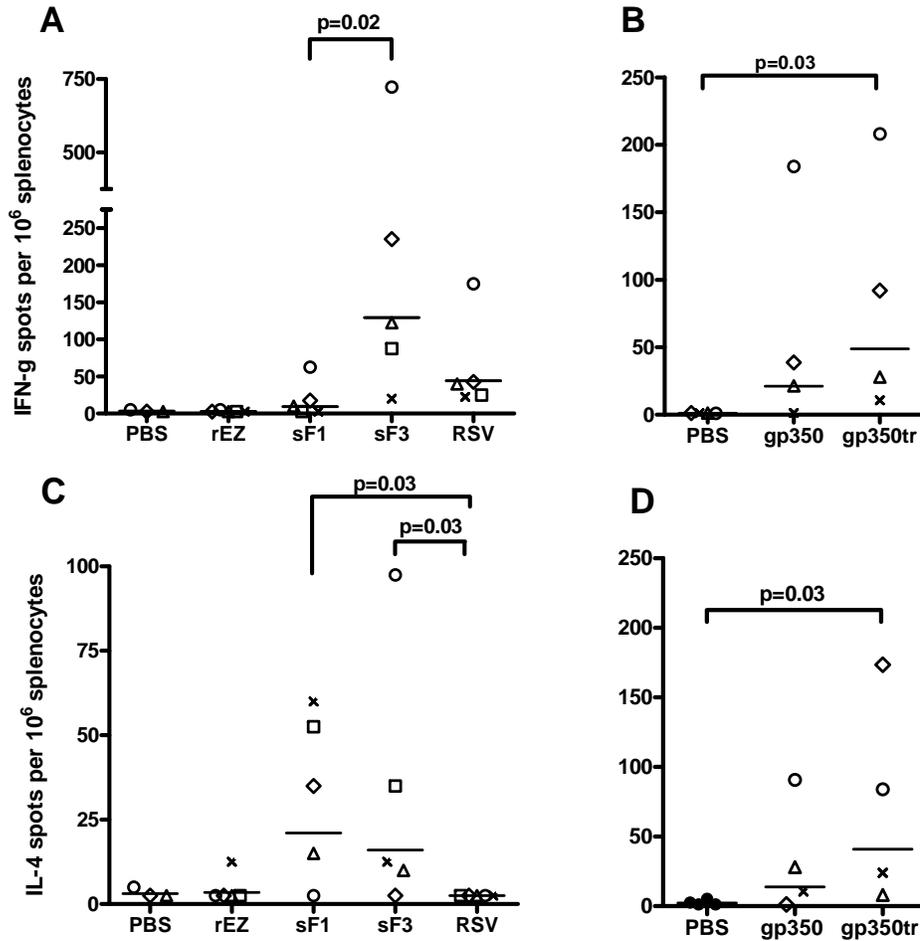


Fig. (3). RSV F-specific and EBV gp350-specific splenocytes were induced in the spleens of cotton rats immunized i.m. with measles vectors. Cotton rats were vaccinated on days 0 and 28 with measles vectors. Splenocytes were harvested 7 days after the second measles vectors vaccination. A total of 3×10^5 cells were stimulated with either RSV F protein (A and C) or EBV gp350 protein (B and D) *in vitro* for 20 h, and the numbers of IFN- γ spot-forming cells (A and B) and IL-4 spot-forming cells (C and D) were quantified by an ELISPOT assay. Spots were counted with an automated counting device and are expressed as numbers of spots per 10^6 cells. Groups are compared using Mann-Whitney rank sum test. Each symbol represents an individual animal.

Table 1. Antibody response of cotton rats to rEZ vectored gp350 and RSV F

Inoculation	MV Neutralizing Ab (Mean log ₂ ± SD Titer) ^{b,c}		EBV-gp350 or RSV-F ELISA Ab (Mean log ₂ ± SD Titers) ^{c,e}		RSV or EBV Neutralizing Ab (Mean log ₂ ± SD Titer) ^{b,c}	
	Dose 1 ^a	Dose 2 ^a	Dose 1	Dose 2	Dose 1	Dose 2
rEZ	8.4 ± 0.4	10.1 ± 0.9	< 3.3	< 3.3	< 3.3	< 3.3
gp350	8.7 ± 0.6	11.0 ± 0.7	3.3 ± 0.1	7.3 ± 0.3	< 3.3	< 3.3
gp350tr	8.2 ± 0.2	10.2 ± 0.8	3.3 ± 0.1	4.3 ± 0.1	< 3.3	< 3.3
rEZ	7.1 ± 0.4	8.7 ± 0.6	< 6.6	< 6.6	< 3.0	< 3.0
sF1	6.7 ± 0.5	8.7 ± 0.9	15.9 ± 2.7	23.7 ± 1.2	5.4 ± 0.7	7.5 ± 1.0
sF3	6.3 ± 0.0	8.7 ± 1.1	15.5 ± 0.9	22.1 ± 0.8	4.2 ± 1.0	8.9 ± 1.1

^aSerum samples were measured 4 weeks post dose 1 and 2 weeks post dose 2.

^bTiter is defined as log₂ reciprocal dilution of serum that inhibited cytopathic effects in 50% of the wells containing measles infected Vero cells.

^cTiter is defined as log₂ of the highest dilution of serum that gave OD₄₅₀ reading 2x of the background.

^dRSV titer is defined as log₂ reciprocal dilution of serum that reduced plaques on HEp-2 cells by 50%.

^eTiter on day 0 for all assays were under the limit of detection.

with two doses of rEZ viruses intramuscularly at 4 weeks interval. Sera were collected 4 weeks post dose 1 and 2 weeks post dose 2. Measles virus neutralizing antibodies (MV Ab) were induced rapidly after one dose administration, reaching titers of 6.3-8.7 log₂. No significant differences in MV Ab titers were observed among the animals receiving sF1, sF3, gp350 or gp350tr compared to rEZ. The 2nd dose increased MV Ab titers by 2-4 folds (Table 1). The control animals vaccinated with UV-inactivated sF3 or RSV A2 virus did not have detectable MV Ab titer (data not shown).

RSV-specific antibody responses were measured by RSV F-specific ELISA and microneutralization assay. sF1 or sF3 induced RSV F-specific IgG antibody in the vaccinated cotton rats (about 15.5-15.9 log₂ post dose 1 and 22.1-23.7 log₂ post dose 2). The RSV F-specific IgG antibody titers were similar between the sF1 and sF3 vaccinated groups. RSV neutralizing antibody titers were also measured against RSV A2 strain. After the first dose, sF1 and sF3 induced RSV neutralizing Ab and titers were boosted to 7.5-log₂ and 8.9-log₂ after the second dose (Table 1). There was no statistical difference in antibody response between sF1 and sF3 vaccinated animals and those intranasally infected with 2 doses of RSV A2 strain (9.4 log₂ ± 1.5, data not shown). Animals vaccinated with UV-inactivated sF3 did not have detectable RSV antibody (data not shown), indicating that virus immunogenicity was dependent on live virus infection.

EBV-specific antibody response was measured by EBV gp350-specific ELISA assay. EBV specific ELISA Ab response was detected after one dose administrations of gp350 or gp350tr; after the 2nd dose the titers reached 7.3 and 4.3 log₂ for gp350 and gp350tr vaccinated animals, respectively (Table 1). Gp350 induced a significantly higher EBV gp350-specific IgG titer than gp350tr. EBV specific neutralizing antibody could not be detected by the EBV neutralization assay.

Evaluation of Cell Mediated Immune Response in Vaccinated Cotton Rats

Cell mediated immune response is important for viral clearance and may play an important role in vaccine mediated protective immune response [44]. RSV F-specific

and EBV gp350-specific T cell responses in splenocytes were examined by IFN-γ and IL-4 ELISpot on day 7 post the 2nd dose. The frequencies of RSV F-specific IFN-γ secreting cells were statistically higher in sF3 vaccinated group (average 130 spots/10⁶ splenocytes) compared to sF1 vaccinated (average 9 spots/10⁶ splenocytes) or RSV A2-infected animals (average 45 spots/10⁶ splenocytes) (Fig. 3A). The RSV F-specific IL-4 secreting cells were low, averaged about 20 spots/10⁶ splenocytes for both sF1 and sF3 groups. Surprisingly, no RSV-specific IL-4 secreting cells detected in RSV A2-infected animals (Fig. 3C). Thus, sF3 and RSV induced a Th1-biased response based on the higher IFN-γ to IL-4 secreting cell ratio.

The frequencies of EBV gp350-specific IFN-γ and IL-4 secreting cells for gp350 (average 23 spots and 15 spots/10⁶ splenocytes for IFN-γ and IL-4 respectively) and gp350tr vaccinated groups (average 49 spots and 41 spots/10⁶ splenocytes for IFN-γ and IL-4 respectively) were not statistically different, but were statistically higher than the PBS control group for the gp350tr group (Fig. 3B, D).

Protection Against RSV A2 Challenge in sF1 and sF3 Vaccinated Cotton Rats

Animal model is not available for assessing EBV vaccine induced protective immune response. Cotton rat is the best available small animal model to evaluate RSV vaccine induced protection against wt RSV challenge infection. To assess if the immune responses induced from sF1 and sF3 could provide protection against RSV A2 challenge, vaccinated cotton rats were challenged with RSV A2 intranasally and virus titers in the lungs were measured. Similar to RSV infected animals, sF1 or sF3 offered complete protection against RSV A2 challenge with viral titers in the lungs that was at least 4.5 log₁₀ lower than the control animals. In contrast, cotton rats vaccinated with 10⁵ PFU of UV-inactivated sF3 had viral titer similar to the PBS control group, indicating that viral replication is necessary for induction of protective immune response (Fig. 4).

Immunogenicity of EZ Vectored Vaccines in Rhesus Macaques

Rhesus macaques are fully permissive to measles virus infection and have been used for evaluation of measles

vaccine and measles virus pathogenesis studies [45-47]. The ability of rEZ vectored RSV F and EBV gp350 to induce measles virus specific or F or gp350 specific immune responses was examined in this more permissive animal model. Since sF3 demonstrated better immunogenicity than sF1 in cotton rats, only sF3 was evaluated in measles virus seropositive (MV+) rhesus macaques that had prior measles vaccination and measles virus seronegative (MV-) rhesus macaques. No abnormal clinical signs were observed and serum chemistries were within the normal range in all monkeys throughout the study (data not shown). sF3 or gp350 vaccinated monkeys induced a similar level of MV neutralizing antibody in both MV- and MV+ monkeys. A second dose of vaccination with sF3 or gp350 increased MV neutralizing Ab titers by about 4-fold in all groups (Table 2). The Ab titer differences between the groups were not statistically significant.

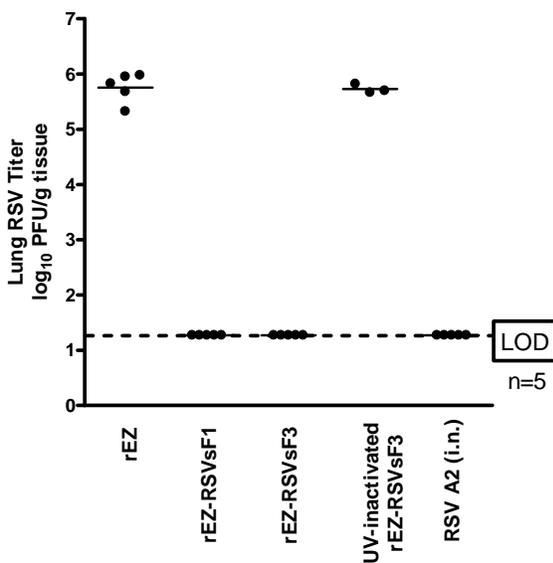


Fig. (4). Reduction of titers of RSV in the lungs of cotton rats vaccinated with measles-vectored RSV F after challenge. Naïve cotton rats were immunized with 10^5 PFU of sF1 or sF3 on day 0 and 28. Cotton rats were then challenged with 10^6 PFU of RSV A2 on day 56. Lungs were harvested on day 60 and RSV titers were determined in the lung homogenates by standard plaque assay. Each experimental group represents 5 individual animals except in UV-inactivated sF3 group (n=3).

Antibody responses to the inserted genes, however, were relatively low. sF3 inoculated animals had RSV F-specific IgG titer higher than the pre-vaccination level (titers were under the detection limit). F specific IgG Ab titers were $9.3 \log_2$ and $8.8 \log_2$ in MV+ and MV- groups after dose 1, respectively. After a second dose, three out of 4 animals in MV+ group had more than 4-fold increase in RSV F-specific IgG Ab titer but none of the animals in MV- group had 4-fold increase in Ab response (Fig. 5A). After two doses of vaccination, RSV neutralizing Ab titers in all groups were very low in both MV+ and MV- groups. Seven out of 8 animals had a detectable level of RSV neutralizing Ab. The RSV Nt Ab titers were not statistically different between the two groups ($5.0 \log_2$ for MV+ versus $4.5 \log_2$ for MV- groups) (Table 2). Ab responses to EBV gp350 in the gp350 vaccinated monkeys were not detected at all time points. Similar to antibody responses, IFN- γ secreting cells in PBMC to measles viral antigens were significantly higher (~10-fold) than the responses to either RSV F or EBV gp350 (Fig. 5B) while PBMC, when stimulated with measles viral antigens, RSV F or EBV gp350, did not secrete any IL-4. Thus, compared to the cotton rat studies, rEZ vectored viruses were poorly immunogenic in the primate model.

DISCUSSION

Reverse genetics has been developed for generating recombinant measles virus vectored vaccines encoding foreign antigens based on the Edmonston-Schwarz and AIK-C vaccine strain [15, 48]. The reverse genetic system described in this study was based on the EZ vaccine strain, which is used predominantly outside the US. The EZ strain might be immunogenic in population that has prior measles vaccine immunization [23]. The ectodomains of the RSV F and EBV gp350 glycoproteins instead of full length proteins were engineered to be expressed by EZ to avoid the incorporation of the foreign surface glycoproteins onto the measles virus envelope. The expression of heterologous membrane proteins on measles virus surface may alter the binding of measles to its natural receptors and thus change the tissue tropism of measles virus. We showed that both RSV F and EBV gp350 was expressed efficiently *in vitro*. However, rEZ vectored vaccines induced insert specific responses in cotton rats but poor responses in rhesus macaques although both cotton rats and rhesus macaques have previously been shown to be permissive to measles virus infection to various degrees [47, 49-51].

Table 2. Antibody Response of Rhesus Macaques to rEZ Vectored gp350 or F

Inoculation	Prior MV Vaccination ^a	MV Neutralizing Ab (mean $\log_2 \pm$ SD Titer)			RSV or EBV ELISA Ab (Mean $\log_2 \pm$ SD Titer)		RSV or EBV Neutralizing Ab (Mean $\log_2 \pm$ SD Titer) ^c	
		Day 0	Dose 1 ^b	Dose 2 ^b	Dose 1	Dose 2	Dose 1	Dose 2
gp350	+	4.3 ± 1.2	5.7 ± 0.5	7.3 ± 0.6	< 3.3	< 3.3	< 3.3	< 3.3
gp350	-	< 3.3	5.4 ± 1.1	7.3 ± 0.1	< 3.3	< 3.3	< 3.3	< 3.3
sF3	+	4.2 ± 0.7	5.2 ± 1.5	7.5 ± 1.2	9.3 ± 1.9	11.8 ± 1.0	3.3 ± 0.5	5.0 ± 0.8
sF3	-	< 3.3	5.9 ± 1.1	7.9 ± 1.1	8.8 ± 1.8	9.5 ± 2.1	3.3 ± 0.5	4.5 ± 1.7

^aPreviously vaccinated with Measles vaccine.

^bSerum samples were measured 4 weeks post dose 1 and 2 weeks post dose 2.

^cRSV antibody titer is defined as \log_2 reciprocal dilution of serum that reduced plaques on HEp-2 cells by 50%; neutralization assay was performed with 10% guinea pig complement.

Titers of prevaccinated animals were under the limit of detection in all the assays.

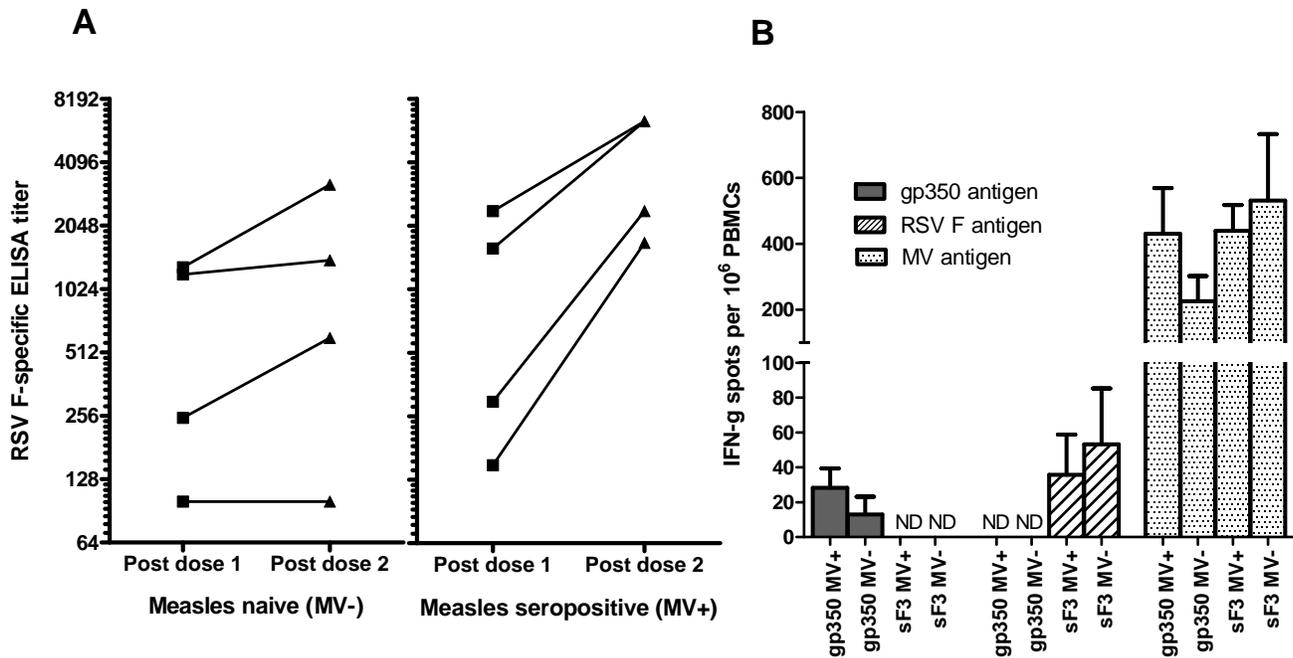


Fig. (5). RSV F specific antibody response and cellular response in rhesus macaques vaccinated with measles vectors. Measles seronegative and sero-positive rhesus macaques were immunized with 10^5 PFU of sF3 or gp350 on day 0 and 28. (A) RSV F titers were measured on day 28 (post dose 1) and 42 (post dose 2) using ELISA. (B) PBMCs were isolated from whole blood on day 56. The numbers of IFN- γ spot forming cells were quantified with an ELISpot assay when PBMCs were stimulated with measles viral antigens, RSV F or EBV gp350 antigens *in vitro*. Spots were counted with an automated counting device and are expressed as numbers of spots per 10^6 cells \pm SEM. ND = not determined.

One of the important determinants for inducing a robust immune response *in vivo* is the amount of the expressed antigen level and antigen presentation. We have evaluated the effect of the gene insertion location and antigen expression level because transcription of nonsegmented negative strand RNA viruses is obligatorily sequential [52]. The genes located at proximal to the 3' promoter are produced more abundantly than the ones distal to the promoter [53]. Indeed, when RSV F was introduced 3' proximally to the N gene, it produced 10 times more secreted RSV F protein than when inserted between the P and M junction (data not shown). In comparison, insertion of the gp350 of EBV at the 1st position of EZ significantly affected virus replication. Insertion of the EBV gp350 gene between the P and M genes had less effect on virus growth, similar to that shown for Hepatitis B surface antigen (HBsAg) (14). The gene insertion position affected immunogenicity of EZ vectored vaccines in cotton rats. rEZ encoding RSV F ectodomain induced antibody response against measles and RSV F. rEZ-sF3 induced MV specific antibody response comparable to rEZ and RSV specific Ab response comparable to those induced by intranasal RSV infection in cotton rats; it also induced cell mediated response as measured by IFN- γ ELISpot assay. Surprisingly, sF1 was inferior to sF3 in its ability to induce both RSV neutralizing antibody and RSV F-specific IFN- γ secreting T cell responses, suggesting that sF1 may be more attenuated than sF3 *in vivo*. Immune response is dependent on viral infectivity and the nature of the inserts. rEZ vectored EBV gp350 and gp350tr induced lower gp350 specific IgG Ab

response and undetectable neutralizing antibody response in cotton rats.

The cotton rats immunized with MV expressing RSV F were also challenged with wt RSV. After challenge, the number of IFN- γ secreting cells was higher than IL-4 secreting cells, which was consistent with a Th1-biased response (data not shown). RSV enhanced disease has been associated with FI-RSV vaccine [54, 55]. A chimeric parainfluenza virus vectored RSV F [35] did not cause enhanced disease in hamsters and has been proven to be safe in phase I clinical trial in seronegative children [56]. Recently, a report from another group has shown that measles virus vectored RSV F did not cause unusual pathology in cotton rats after RSV challenge, thus, the likelihood of the MV vector F to cause enhanced disease is very low [15].

Since cotton rats are partially permissive to MV, MV vectored vaccine was further evaluated in rhesus macaques. Rhesus macaque is fully permissive to measles virus infection and the infected monkeys develop symptoms similar to those experienced by humans, such as skin rash, fever and conjunctivitis [47]. This monkey model was used in the neurovirulence testing of MV vaccine [57]. In addition, since rhesus macaques could be infected with human measles virus, the breeding colony is commonly vaccinated with MV. This allowed us to examine the effect of prior vaccination on vectored measles and mimic our target populations that either have maternal antibodies or have measles vaccination history. rEZ and rEZ expressing gp350 and sF3 induced a good level of antibody response to

measles virus irrespective of their sero-status to MV. The antibody response was slightly lower in magnitude than what were observed in the cotton rats as the dose per kg used in the rhesus was ~ 90 times lower than in cotton rats. The antibody response to the F and gp350 inserts was much lower in rhesus macaques. A previous report showed that two doses of MV-HIV gp140 at 5×10^6 TCID₅₀ (25 times higher dose than our study) induced a robust Ab response towards HIV gp140 in monkeys [10]. However, three doses of 10^4 TCID₅₀ (1/15 our dose) of MV vectored HBV surface antigen induced a very low response in monkeys [14]. Thus, the low humoral responses to the vectored inserts in rhesus macaques may be due to a lower dose used in this study.

Measles vaccine virus has been shown to bind signaling lymphocyte activation molecule (SLAM; also known as CD150) on activated immune cells and CD46 [58], which is ubiquitously expressed on human cells. It is thought that virus infects the respiratory tract through SLAM on dendritic cells. In cotton rats, although human CD46 and CD150 homologs have not been reported [59], measles virus could replicate in the lungs of these animals after intranasal infection [40]. We have evaluated different routes of administration (both intranasal and intramuscular) of MV vectored RSV F in cotton rats and found that both inoculation routes offered protection against RSV challenge after immunization (data not shown). Rhesus macaques possess both CD46 and CD150 receptor homologs [60] and are susceptible to measles virus infection and develop diseases similar to humans. The difference in receptors between the two animal species may be able to explain why insertion of the insert into EZ strain have further attenuated viruses and reduced immunogenicity of the virus in rhesus macaques but not in cotton rats.

We also examined replication of EZ vectored RSV F and EBV gp350 *in vivo* by using real time qRT-PCR since differential in growth kinetics of recombinant measles virus may explain the difference in antibody responses. However, we were not able to detect measles genomes in PBMC isolated from whole blood 24 hours post vaccination. This may be attributed to the low copies of measles virus circulating in the blood or the time point used to measure measles kinetics *in vivo* was not optimal. In conjunction, we have examined replication of EZ vectors in PBMC of rhesus macaques *ex vivo* to determine if insertion of the foreign gene affected replication of these viruses in PBMC. PHA-activated PBMC cells were infected with rMV at MOI of 1.0 and growth kinetics were determined for 7 days. The peak titer of the recombinant viruses containing RSV F or EBV gp350 were only 0.5 log₁₀ lower than rEZ (data not shown), indicating the vectored vaccine could infect rhesus cells. We also investigated genetic stability of the insert by serially passaging the recombinant viruses in Vero cells 10 times and did not find the loss of the inserted gene expression as reported previously [61, 62].

Other negative stranded RNA viruses including Newcastle disease virus (NDV), vesicular stomatitis virus, Sendai virus and parainfluenza viruses [35, 63-65] had been evaluated as vectors to deliver foreign antigens and were shown to elicit both humoral and cellular responses in non-human primates and to confer protection to virus challenge in some cases [63, 66-69]. The vectored vaccines were

normally delivered to non-human primates by either intranasal or intratracheal inoculation route. We found intranasal and intramuscular vaccination of EZ induced similar antibody response in cotton rats. It is possible that intranasal immunization might be more immunogenic than intramuscular vaccination in monkeys.

Based on our study, MV vector is probably not suitable for delivery of the EBV gp350 antigen. Immunogenicity of adjuvanted gp350 protein vaccine has been demonstrated in young adults [29], adjuvanted gp350 might be the right approach for EBV vaccine development. The amount of gp350 delivered by MV is likely not sufficient to induce gp350 specific immune response. The F protein of RSV delivered by MV is more immunogenic than EBV gp350, however, additional evaluation of immunogenicity of MV-RSV F must be conducted before proceeding to testing this vaccine in humans.

ACKNOWLEDGEMENTS

We thank Rosemary Broom for obtaining EZ vaccine strain, MedImmune's Animal Care Facility for conducting the cotton rat studies, Jeff Roberts and Phil Allen at Valley Biosystems for conducting the monkey study, Jennifer Woo, Jason Marshall, Stacie Lambert, Darren Heeke, Lorrie Gemmell for conducting T cells assays, Chin-Fen Yang for sequencing support, Dr. Gary van Nest for reviewing the manuscript.

CONFLICT OF INTEREST

None declared.

REFERENCES

- Liniger M, Zuniga A, Naim HY. Use of viral vectors for the development of vaccines. *Expert Rev Vaccines* 2007; 6(2): 255-66.
- Amanna IJ, Carlson NE, Slifka MK. Duration of humoral immunity to common viral and vaccine antigens. *N Engl J Med* 2007; 357(19): 1903-15.
- Gans HA, Yasukawa LL, Alderson A, et al. T cell immunity to measles viral proteins in infants and adults after measles immunization. *Viral Immunol* 2004; 17(2): 298-307.
- Ovsyannikova IG, Dhiman N, Jacobson RM, Vierkant RA, Poland GA. Frequency of measles virus-specific CD4+ and CD8+ T cells in subjects seronegative or highly seropositive for measles vaccine. *Clin Diagn Lab Immunol* 2003; 10(3): 411-6.
- Hiremath GS, Omer SB. A meta-analysis of studies comparing the respiratory route with the subcutaneous route of measles vaccine administration. *Hum Vaccin* 2005; 1(1): 30-6.
- Low N, Kraemer S, Schneider M, Restrepo AM. Immunogenicity and safety of aerosolized measles vaccine: systematic review and meta-analysis. *Vaccine* 2008; 26(3): 383-98.
- Cantarella G, Liniger M, Zuniga A, et al. Recombinant measles virus-HPV vaccine candidates for prevention of cervical carcinoma. *Vaccine* 2009; 27(25-26): 3385-90.
- del Valle JR, Devaux P, Hodge G, Wegner NJ, McChesney MB, Cattaneo R. A vectored measles virus induces hepatitis B surface antigen antibodies while protecting macaques against measles virus challenge. *J Virol* 2007; 81(19): 10597-605.
- Liniger M, Zuniga A, Tamin A, et al. Induction of neutralising antibodies and cellular immune responses against SARS coronavirus by recombinant measles viruses. *Vaccine* 2008; 26(17): 2164-74.
- Lorin C, Mollet L, Delebecque F, et al. A single injection of recombinant measles virus vaccines expressing human immunodeficiency virus (HIV) type 1 clade B envelope glycoproteins induces neutralizing antibodies and cellular immune responses to HIV. *J Virol* 2004; 78(1): 146-57.
- Despres P, Combredet C, Frenkiel MP, Lorin C, Brahic M, Tangy F. Live measles vaccine expressing the secreted form of the West

- Nile virus envelope glycoprotein protects against West Nile virus encephalitis. *J Infect Dis* 2005; 191(2): 207-14.
- [12] Iankov ID, Haralambieva IH, Galanis E. Immunogenicity of attenuated measles virus engineered to express *Helicobacter pylori* neutrophil-activating protein. *Vaccine* ; 29(8): 1710-20.
- [13] Lorin C, Delebecque F, Labrousse V, *et al.* A recombinant live attenuated measles vaccine vector primes effective HLA-A0201-restricted cytotoxic T lymphocytes and broadly neutralizing antibodies against HIV-1 conserved epitopes. *Vaccine* 2005; 23(36): 4463-72.
- [14] Reyes-del Valle J, Hodge G, McChesney MB, Cattaneo R. Protective anti-hepatitis B virus responses in rhesus monkeys primed with a vectored measles virus and boosted with a single dose of hepatitis B surface antigen. *J Virol* 2009; 83(17): 9013-7.
- [15] Sawada A, Komase K, Nakayama T. AIK-C measles vaccine expressing fusion protein of respiratory syncytial virus induces protective antibodies in cotton rats. *Vaccine* 2010; 29(7): 1481-90.
- [16] Zuniga A, Wang Z, Liniger M, *et al.* Attenuated measles virus as a vaccine vector. *Vaccine* 2007; 25(16): 2974-83.
- [17] Wang Z, Hangartner L, Cornu TI, *et al.* Recombinant measles viruses expressing heterologous antigens of mumps and simian immunodeficiency viruses. *Vaccine* 2001; 19(17-19): 2329-36.
- [18] Brandler S, Lucas-Hourani M, Moris A, *et al.* Pediatric measles vaccine expressing a dengue antigen induces durable serotype-specific neutralizing antibodies to dengue virus. *PLoS Negl Trop Dis* 2007; 1(3): e96.
- [19] Rager-Zisman B, Bazarsky E, Skibin A, *et al.* The effect of measles-mumps-rubella (MMR) immunization on the immune responses of previously immunized primary school children. *Vaccine* 2003; 21(19-20): 2580-8.
- [20] Sepulveda-Amor J, Valdespino-Gomez JL, Garcia-Garcia Mde L, *et al.* A randomized trial demonstrating successful boosting responses following simultaneous aerosols of measles and rubella (MR) vaccines in school age children. *Vaccine* 2002; 20(21-22): 2790-5.
- [21] Wong-Chew RM, Beeler JA, Audet S, Santos JI. Cellular and humoral immune responses to measles in immune adults re-immunized with measles vaccine. *J Med Virol* 2003; 70(2): 276-80.
- [22] Albrecht P, Ennis FA, Saltzman EJ, Krugman S. Persistence of maternal antibody in infants beyond 12 months: mechanism of measles vaccine failure. *J Pediatr* 1977; 91(5): 715-8.
- [23] Tidjani O, Grunitsky B, Guerin N, *et al.* Serological effects of Edmonston-Zagreb, Schwarz, and AIK-C measles vaccine strains given at ages 4-5 or 8-10 months. *Lancet* 1989; 2(8676): 1357-60.
- [24] McIntosh K, Chanock RM. Respiratory syncytial virus. Second Edition ed. New York: Raven Press, Ltd., 1990.
- [25] Olmsted RA, Elango N, Prince GA, *et al.* Expression of the F glycoprotein of respiratory syncytial virus by a recombinant vaccinia virus: comparison of the individual contributions of the F and G glycoproteins to host immunity. *Proc Natl Acad Sci U S A* 1986; 83(19): 7462-6.
- [26] Connors M, Collins PL, Firestone CY, Murphy BR. Respiratory syncytial virus (RSV) F, G, M2 (22K), and N proteins each induce resistance to RSV challenge, but resistance induced by M2 and N proteins is relatively short-lived. *J Virol* 1991; 65(3): 1634-7.
- [27] Fleisher G, Henle W, Henle G, Lennette ET, Biggar RJ. Primary infection with Epstein-Barr virus in infants in the United States: clinical and serologic observations. *J Infect Dis* 1979; 139(5): 553-8.
- [28] Lopes V, Young LS, Murray PG. Epstein-Barr virus-associated cancers: aetiology and treatment. *Herpes* 2003; 10(3): 78-82.
- [29] Sokal EM, Hoppenbrouwers K, Vandermeulen C, *et al.* Recombinant gp350 vaccine for infectious mononucleosis: a phase 2, randomized, double-blind, placebo-controlled trial to evaluate the safety, immunogenicity, and efficacy of an Epstein-Barr virus vaccine in healthy young adults. *J Infect Dis* 2007 196(12): 1749-53.
- [30] Schneider H, Spielhofer P, Kaelin K, *et al.* Rescue of measles virus using a replication-deficient vaccinia-T7 vector. *J Virol Methods* 1997; 64(1): 57-64.
- [31] Radecke F, Spielhofer P, Schneider H, *et al.* Rescue of measles viruses from cloned DNA. *EMBO J* 1995; 14(23): 5773-84.
- [32] Mann GF, Allison LM, Copeland JA, Agostini CF, Zuckerman AJ. A simplified plaque assay system for measles virus. *J Biol Stand* 1980; 8(3): 219-25.
- [33] Jackman WT, Mann KA, Hoffmann HJ, Spaete RR. Expression of Epstein-Barr virus gp350 as a single chain glycoprotein for an EBV subunit vaccine. *Vaccine* 1999; 17(7-8): 660-8.
- [34] Christe KL, McChesney MB, Spinner A, *et al.* Comparative efficacy of a canine distemper-measles and a standard measles vaccine for immunization of rhesus macaques (*Macaca mulatta*). *Comp Med* 2002; 52(5): 467-72.
- [35] Tang RS, Schickli JH, MacPhail M, *et al.* Effects of human metapneumovirus and respiratory syncytial virus antigen insertion in two 3' proximal genome positions of bovine/human parainfluenza virus type 3 on virus replication and immunogenicity. *J Virol* 2003; 77(20): 10819-28.
- [36] Sashihara J, Burbelo PD, Savoldo B, Pierson TC, Cohen JI. Human antibody titers to Epstein-Barr Virus (EBV) gp350 correlate with neutralization of infectivity better than antibody titers to EBV gp42 using a rapid flow cytometry-based EBV neutralization assay. *Virology* 2009; 391(2): 249-56.
- [37] Mok H, Lee S, Utley TJ, *et al.* Venezuelan equine encephalitis virus replicon particles encoding respiratory syncytial virus surface glycoproteins induce protective mucosal responses in mice and cotton rats. *J Virol* 2007; 81(24): 13710-22.
- [38] Zhan X, Slobod KS, Krishnamurthy S, *et al.* Sendai virus recombinant vaccine expressing hPIV-3 HN or F elicits protective immunity and combines with a second recombinant to prevent hPIV-1, hPIV-3 and RSV infections. *Vaccine* 2008; 26(27-28): 3480-8.
- [39] Tanner J, Whang Y, Sample J, Sears A, Kieff E. Soluble gp350/220 and deletion mutant glycoproteins block Epstein-Barr virus adsorption to lymphocytes. *J Virol* 1988; 62(12): 4452-64.
- [40] Wyde PR, Ambrose MW, Voss TG, Meyer HL, Gilbert BE. Measles virus replication in lungs of hispid cotton rats after intranasal inoculation. *Proc Soc Exp Biol Med* 1992; 201(1): 80-7.
- [41] Jin H, Zhou H, Cheng X, Tang R, Munoz M, Nguyen N. Recombinant respiratory syncytial viruses with deletions in the NS1, NS2, SH, and M2-2 genes are attenuated *in vitro* and *in vivo*. *Virology* 2000; 273(1): 210-8.
- [42] Murphy BR, Sotnikov AV, Lawrence LA, Banks SM, Prince GA. Enhanced pulmonary histopathology is observed in cotton rats immunized with formalin-inactivated respiratory syncytial virus (RSV) or purified F glycoprotein and challenged with RSV 3-6 months after immunization. *Vaccine* 1990; 8(5): 497-502.
- [43] Wu H, Pfarr DS, Johnson S, *et al.* Development of motavizumab, an ultra-potent antibody for the prevention of respiratory syncytial virus infection in the upper and lower respiratory tract. *J Mol Biol* 2007; 368(3): 652-65.
- [44] Munoz JL, McCarthy CA, Clark ME, Hall CB. Respiratory syncytial virus infection in C57BL/6 mice: clearance of virus from the lungs with virus-specific cytotoxic T cells. *J Virol* 1991; 65(8): 4494-7.
- [45] de Vries RD, Lemon K, Ludlow M, *et al.* *In vivo* tropism of attenuated and pathogenic measles virus expressing green fluorescent protein in macaques. *J Virol*; 84(9): 4714-24.
- [46] McChesney MB, Miller CJ, Rota PA, *et al.* Experimental measles. I. Pathogenesis in the normal and the immunized host. *Virology* 1997; 233(1): 74-84.
- [47] Zhu YD, Heath J, Collins J, *et al.* Experimental measles. II. Infection and immunity in the rhesus macaque. *Virology* 1997; 233(1): 85-92.
- [48] Combredet C, Labrousse V, Mollet L, *et al.* A molecularly cloned Schwarz strain of measles virus vaccine induces strong immune responses in macaques and transgenic mice. *J Virol* 2003; 77(21): 11546-54.
- [49] Niewiesk S, Prince G. Diversifying animal models: the use of hispid cotton rats (*Sigmodon hispidus*) in infectious diseases. *Lab Anim* 2002; 36(4): 357-72.
- [50] de Swart RL, Ludlow M, de Witte L, *et al.* Predominant infection of CD150+ lymphocytes and dendritic cells during measles virus infection of macaques. *PLoS Pathog* 2007; 3(11): e178.
- [51] El Mubarak HS, Yuksel S, van Amerongen G, *et al.* Infection of cynomolgus macaques (*Macaca fascicularis*) and rhesus macaques (*Macaca mulatta*) with different wild-type measles viruses. *J Gen Virol* 2007; 88(Pt 7): 2028-34.
- [52] Abraham G, Banerjee AK. Sequential transcription of the genes of vesicular stomatitis virus. *Proc Natl Acad Sci U S A* 1976; 73(5): 1504-8.

- [53] Ray J, Fujinami RS. Characterization of *in vitro* transcription and transcriptional products of measles virus. *J Virol* 1987 Nov;61(11):3381-7.
- [54] Kapikian AZ, Mitchell RH, Chanock RM, Shvedoff RA, Stewart CE. An epidemiologic study of altered clinical reactivity to respiratory syncytial (RS) virus infection in children previously vaccinated with an inactivated RS virus vaccine. *Am J Epidemiol* 1969; 89(4): 405-21.
- [55] Kim HW, Canchola JG, Brandt CD, *et al.* Respiratory syncytial virus disease in infants despite prior administration of antigenic inactivated vaccine. *Am J Epidemiol* 1969; 89(4): 422-34.
- [56] Bernstein DI, Malkin E, Abughali N, Falloon J, Yi T, Dubovsky F. Phase I Study of the Safety and Immunogenicity of a Live, Attenuated Respiratory Syncytial Virus and Parainfluenza Virus Type 3 Vaccine in Seronegative Children. *Pediatr Infect Dis J* 2011 Sep 15.
- [57] Sharova OK, Rozina EE, Shteinberg LS, Gordienko NM, Kolyanova IS. Morphological characteristics of the pathological process in the central nervous system of monkeys infected with variants of measles virus strain L-16. *Acta Virol* 1979; 23(5): 393-7.
- [58] Dorig RE, Marciel A, Chopra A, Richardson CD. The human CD46 molecule is a receptor for measles virus (Edmonston strain). *Cell* 1993; 75(2): 295-305.
- [59] Pfeuffer J, Puschel K, Meulen V, Schneider-Schaulies J, Niewiesk S. Extent of measles virus spread and immune suppression differentiates between wild-type and vaccine strains in the cotton rat model (*Sigmodon hispidus*). *J Virol* 2003; 77(1): 150-8.
- [60] Dorig RE, Marciel A, Richardson CD. CD46, a primate-specific receptor for measles virus. *Trends Microbiol* 1994; 2(9): 312-8.
- [61] Binder JJ, Hoffman MA, Palmenberg AC. Genetic stability of attenuated mengovirus vectors with duplicate primary cleavage sequences. *Virology* 2003; 312(2): 481-94.
- [62] Mueller S, Wimmer E. Expression of foreign proteins by poliovirus polyprotein fusion: analysis of genetic stability reveals rapid deletions and formation of cardioviruslike open reading frames. *J Virol* 1998; 72(1): 20-31.
- [63] Bukreyev A, Huang Z, Yang L, *et al.* Recombinant newcastle disease virus expressing a foreign viral antigen is attenuated and highly immunogenic in primates. *J Virol* 2005; 79(21): 13275-84.
- [64] Hurwitz JL, Soike KF, Sangster MY, *et al.* Intranasal Sendai virus vaccine protects African green monkeys from infection with human parainfluenza virus-type one. *Vaccine* 1997; 15(5): 533-40.
- [65] Perez M, Clemente R, Robison CS, *et al.* Generation and characterization of a recombinant vesicular stomatitis virus expressing the glycoprotein of Bornavirus. *J Virol* 2007; 81(11): 5527-36.
- [66] Kato M, Igarashi H, Takeda A, *et al.* Induction of Gag-specific T-cell responses by therapeutic immunization with a Gag-expressing Sendai virus vector in macaques chronically infected with simian-human immunodeficiency virus. *Vaccine* 2005; 23(24): 3166-73.
- [67] Matano T, Kobayashi M, Igarashi H, *et al.* Cytotoxic T lymphocyte-based control of simian immunodeficiency virus replication in a preclinical AIDS vaccine trial. *J Exp Med* 2004; 199(12): 1709-18.
- [68] Rose NF, Marx PA, Luckay A, *et al.* An effective AIDS vaccine based on live attenuated vesicular stomatitis virus recombinants. *Cell* 2001; 106(5): 539-49.
- [69] Tang RS, MacPhail M, Schickli JH, *et al.* Parainfluenza virus type 3 expressing the native or soluble fusion (F) Protein of Respiratory Syncytial Virus (RSV) confers protection from RSV infection in African green monkeys. *J Virol* 2004; 78(20): 11198-207.

Received: December 16, 2011

Revised: January 3, 2012

Accepted: January 16, 2012

© Mok *et al.*; Licensee Bentham Open.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.