

Brief Communication



1,3-Dibenzyl-5-Fluorouracil Prevents Ovariectomy-Induced Bone Loss by Suppressing Osteoclast Differentiation

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ABSTRACT


Osteoclasts (OCs) are clinically important cells that resorb bone matrix. Accelerated bone destruction by OCs is closely linked to the development of metabolic bone diseases. In this study, we screened novel chemical inhibitors targeting OC differentiation to identify drug candidates for metabolic bone diseases. We identified that 1,3-dibenzyl-5-fluorouracil, also named OCI-101, is a novel inhibitor of osteoclastogenesis. The formation of multinucleated OCs is reduced by treatment with OCI-101 in a dose-dependent manner. OCI-101 inhibited the expression of OC markers via downregulation of receptor activator of NF- κ B ligand and M-CSF signaling pathways. Finally, we showed that OCI-101 prevents ovariectomy-induced bone loss by suppressing OC differentiation in mice. Hence, these results demonstrated that OCI-101 is a good drug candidate for treating metabolic bone diseases.

Keywords: Osteoclasts; Osteoclastogenesis; 1,3-Dibenzyl-5-fluorouracil; Ovariectomy; Osteoporosis

INTRODUCTION

Bone is continuously maintained by the process of bone remodeling through the balanced actions of bone-resorbing osteoclasts (OCs) and bone-forming osteoblasts (1-3). OCs, the only cells with bone-resorbing activity, play clinically important roles in metabolic bone diseases, such as postmenopausal osteoporosis and osteoarthritis, which are characterized by accelerated bone destruction by OCs (2,4). Therefore, inhibition of excessive OC bone-resorbing activity and OC differentiation play a key role in the treatment of metabolic bone diseases (2,3).

Mature bone-resorbing OCs are multinucleated giant cells derived from cell fusion of monocyte/macrophage lineage precursors by a process of cellular differentiation called OC differentiation or osteoclastogenesis (2,3). Osteoclastogenesis is mainly regulated by 2 cytokines, M-CSF and RANK ligand (RANKL), which are produced by osteoblasts and activated immune cells (2,3). RANK-RANKL signaling is a key process in OC differentiation

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Conflict of Interest

The authors declare no potential conflicts of interest.

Abbreviations

BMD, bone mineral density; BMM, bone marrow macrophage; BMOC, bone marrow-derived osteoclast; IKK, inhibitor of κ B kinase; KCB, Korea Chemical Bank; micro-CT, micro-computed tomography; OC, osteoclast; OVX, ovariectomy; RANKL, receptor activator of NF- κ B ligand; TRAF, tumor necrosis factor receptor-associated factor; TRAP, tartrate-resistant acid phosphatase.

Author Contributions

Conceptualization: Yu J,¹ Jeon H, Hwang JM; Data curation: Yu J,¹ Jeon H, Hwang JM, Kim T; Formal analysis: Lee ZW, Rho J; Funding acquisition: Rho J; Investigation: Yu J,¹ Jeon H, Hwang JM; Methodology: Park HW, Yu J,² Kim T; Project administration: Rho J, Lee ZW; Resources: Lee ZW; Supervision: Rho J; Writing - original draft: Jeon H, Yu J; Writing - review & editing: Rho J.
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and bone-resorbing function. Upon RANKL stimulation, RANK recruits tumor necrosis factor receptor-associated factors (TRAFs) to activate downstream signaling cascades of adaptors/kinases, such as I κ B kinases (IKKs), Vav3, c-Src and MAPKs, including p38, JNK and ERK (3,5). M-CSF also activates p38, JNK, ERK and Akt signaling that regulate OC proliferation and survival (2,3). The final consequence of RANKL and M-CSF signaling cascades is the activation of osteoclastogenic transcription factors, such as NF- κ B, AP-1, cAMP-response element-binding protein and NFATc1, which induce the expression of OC markers, such as tartrate-resistant acid phosphatase (TRAP), OC-associated receptor, dendritic cell-specific transmembrane protein, B-lymphocyte induced maturation protein 1 and cathepsin K (3,6-8).

Osteoporosis is most common in postmenopausal women (9). Estrogen has a protective effect on bones by inhibiting bone loss (10). In postmenopausal osteoporotic patients, estrogen deficiency can lead to excessive bone resorption due to increases in both OC differentiation and OC activity, which causes rapid bone loss (9). Thus, estrogen or estrogen signal-related therapies targeting OC differentiation/activity, including estrogen replacement and selective estrogen receptor modulators, have been regarded as treatments for postmenopausal osteoporosis (11). In long-term therapy, however, these therapeutic options are limited by adverse effects, including an increased risk of uterine or breast cancer and cardiovascular events (9,12). Bisphosphonates (a classic anti-resorptive agent) and denosumab (a human monoclonal antibody to RANKL) are currently the most widely used therapies for the treatment of postmenopausal osteoporosis (13). Bisphosphonates, such as alendronate, risedronate and zoledronate, are considered the first-line treatment for postmenopausal osteoporosis (13,14). Bisphosphonates bind to calcium in bone and inhibit bone resorption by reducing OC activity or by inducing OC apoptosis, leading to reduced fracture risk (14-16). Denosumab blocks the RANK-RANKL interaction, thereby inhibiting both OC differentiation and OC activity and enhancing bone strength (17). Denosumab is also considered both a first-line agent and an alternative to bisphosphonates in the treatment of postmenopausal osteoporosis (18). In long-term therapy, bisphosphonate treatment results in a plateau in bone mineral density (BMD) response after 3-5 years, while denosumab treatment leads to continuously increasing BMD for up to 10 years of treatment (19-21). Denosumab treatment increases BMD and reduces cortical porosity at all measured skeletal sites and decreases expression of biochemical markers of bone turnover (18,22). Therefore, denosumab administration targeting RANK-RANKL signaling can be considered the preferred choice for the treatment of postmenopausal osteoporosis (18). Both bisphosphonates and denosumab are generally considered to be safe, but prolonged medication is also associated with adverse effects, such as jaw osteonecrosis, hypocalcemia, gastrointestinal symptoms and atypical femoral fractures (18,23,24). Thus, it is necessary to find new and more effective drug candidates for the management of postmenopausal osteoporosis and inflammatory bone-related diseases.

In this study, we discovered a novel chemical inhibitor named OCI-101, 1,3-dibenzyl-5-fluorouracil, targeting OC differentiation. We demonstrate that OCI-101 acts as an inhibitor of osteoclastogenesis. The formation of multinucleated OCs is dose-dependently inhibited by OCI-101 treatment. Finally, we show that OCI-101 prevents ovariectomy (OVX)-induced bone loss by suppressing OC differentiation in mice.

MATERIALS AND METHODS

Reagents, antibodies and mice

1,3-Dibenzyl-5-fluorouracil was purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). A chemical library with a total of 16,380 unique chemical compounds was obtained from the Korea Chemical Bank (KCB). Recombinant human soluble RANKL and human M-CSF were prepared as previously described (25,26). Antibodies were purchased from Cell Signaling Technology (Beverly, MA, USA). Mice were purchased from Daehan Biolink Co. (Umsung, Korea), and the animal study was approved (approval No. CNU-00326 and CNU-00114) by the Animal Experiment Ethics Committee of Chungnam National University.

OC differentiation, analysis and cytotoxicity assay

Osteoclastogenesis was performed as described previously (25,26). Briefly, bone marrow cells collected from the tibias and femurs of 6-week-old C57BL/6J male mice were cultured at 1×10^5 cells/well in 96-well plates in the presence of 50 ng/ml M-CSF and 200 ng/ml RANKL for 4 days. Bone marrow-derived OCs (BMOCs) were further analyzed by TRAP staining and solution assays. For the TRAP solution assay, BMOCs were fixed and incubated with the *p*-nitrophenyl phosphate substrate for 30 min. The supernatant was then mixed with 1 N NaOH, and TRAP activities were analyzed by measuring the absorbance at 405 nm in each well using a microplate reader (Bio-Rad, Hercules, CA, USA). For the TRAP staining assay, the cells were stained with naphthol AS phosphate and fast red violet for 20–30 min at room temperature. TRAP-positive multinucleated BMOCs (TRAP⁺ MNCs) with more than 3 nuclei were considered multinucleated OCs. For the cytotoxicity assay, bone marrow cells (1×10^5 cells/well in 96-well plates) were differentiated into bone marrow-derived macrophages (BMMs) with 50 ng/ml M-CSF alone in the presence or absence of OCI-101 (25–100 μ M) for 4 days. Real-time PCR analysis and pit formation assay were performed as previously described (27,28). The cytotoxicity of OCI-101 in BMMs was analyzed using the Cell Counting Kit-8 (Dojindo Lab., Kumamoto, Japan) according to the manufacturer's recommendations. The analyses of RANKL-induced signaling using immunoblotting were performed as described previously (25,26). BMMs were preincubated with 100 μ M OCI-101 for 12 h before RANKL or M-CSF stimulation. To induce RANKL or M-CSF signaling, BMMs precultured for 2 h without RANKL or M-CSF. The cells were stimulated with RANKL (200 ng/ml) or M-CSF (50 ng/ml) for the indicated times, and the cell lysates were subjected to immunoblot analysis with antibodies.

Bone analysis by micro-computed tomography (micro-CT) and histological analysis

Bone analysis was performed as previously described (28,29). Briefly, C57BL/6J female mice (8-week-old, $n=10-11$ per group) were randomly divided into 4 groups: sham operation (Sham), OVX with a vehicle, ovariectomy with a low dose of OCI-101 (OVX + OCI-101 [1 mg/kg]) and ovariectomy with a high dose of OCI-101 (OVX + OCI-101 [10 mg/kg]). OVX was performed by surgical removal of ovaries, and sham surgery was performed by identifying both ovaries. A week after surgery, the mice were injected intraperitoneally with OCI-101 (1–10 mg/kg) or vehicle each day for 3 wk. For micro-CT analysis, femurs were fixed with 10% formalin, and trabecular morphometry of distal femurs was performed using micro-CT (SkyScan 1076, Bruker micro-CT; Bruker, Kontich, Belgium). For histological analysis, the femurs were decalcified in 15% EDTA solution at 4°C for 3 wk and embedded in paraffin. Paraffin sections were stained with TRAP and hematoxylin, and the TRAP⁺ OCs were counted by visualization under a microscope.

Statistical analysis.

The data represent the mean \pm SD (at least $n=3$ per group). Statistical analyses were performed using 2-tailed Student's t-test or 1- or 2-way ANOVA. The p-values <0.05 were considered statistically significant.

RESULTS

OCI-101 inhibits OC differentiation

Inhibitors of osteoclastogenesis were screened by OC differentiation assays using a KCB chemical library containing a total of 16,380 unique chemical compounds at a chemical concentration of 5 μ M. We identified several inhibitory candidates, named the OCI series (data not shown). Among them, we discovered a novel chemical inhibitor named OCI-101, or 1,3-dibenzyl-5-fluorouracil, that targeted OC differentiation (**Fig. 1A**). First, we examined the cytotoxicity of OCI-101 by a cell proliferation assay with BMMs. BMMs were stimulated for 4 days with M-CSF alone (50 ng/ml) in the presence of OCI-101 compounds (0–100 μ M). We did not observe OCI-101 cytotoxicity at any of the tested concentrations in BMMs after 4 days of culture (**Fig. 1B**). To test the inhibitory effect of OCI-101 on OC differentiation, BMMs were differentiated into BMOCs by stimulation with M-CSF (50 ng/ml) and RANKL (200 ng/ml) in the presence or absence of OCI-101. After 4 days of culture, the cells were fixed, and the fixed cells were analyzed by TRAP solution assay and TRAP staining to determine the number of differentiated multinucleated OCs (TRAP⁺ MNC). OCI-101-treated BMMs showed a marked dose-dependent reduction in the differentiation of multinucleated OCs when compared to that of the mock-treated control (**Fig. 1C**). Furthermore, the number of TRAP⁺ MNCs and TRAP activity were dose-dependently decreased by OCI-101 treatment (**Fig. 1D**). Similarly, the resorption area was reduced significantly by OCI-101 (**Fig. 1E**). Consistent with these results, we observed that the expression of OC markers was inhibited by OCI-101 (**Fig. 1F**). These results suggest that OCI-101 is an inhibitor targeting OC differentiation.

OCI-101 inhibits RANKL and M-CSF signaling pathway

We next analyzed the effect of OCI-101 on the RANKL- and M-CSF-induced signaling pathway. To induce RANKL signaling, BMMs precultured for 2 h without RANKL treatment were stimulated with 200 ng/ml RANKL at the indicated times. We then examined the activation of NF- κ B and MAPKs by RANKL stimulation in OCI-101-treated BMMs. Upon RANKL stimulation, the phosphorylation of I κ B α , p65 and ERK was inhibited by OCI-101 treatment, but not JNK phosphorylation, compared to the mock-treated control. In contrast, the phosphorylation of p38 was slightly enhanced by OCI-101 treatment (**Fig. 2A**). Furthermore, we observed that the phosphorylation of JNK, Akt and ERK following M-CSF stimulation was inhibited by OCI-101 treatment, but not I κ B α , p65 and p38 phosphorylation (**Fig. 2B**). These results indicate that OCI-101 negatively regulates osteoclastogenesis via modulating multiple RANKL- and M-CSF-induced signaling pathways.

OCI-101 inhibits OVX-induced OC formation in mice

To examine the effects of OCI-101 on OC formation *in vivo*, C57BL/6 female mice were subjected to OVX or sham surgery. OVX mice were injected intraperitoneally with OCI-101 or vehicle each day for 3 wk. The femurs were collected, fixed and analyzed by TRAP staining and hematoxylin counterstaining. Finally, we compared bone histological morphology and OC formation in the trabecular region of femurs. OVX mice showed significantly reduced trabecular bones compared to those of sham control mice (**Fig. 3A**). After treatment with OCI-101 in OVX mice,

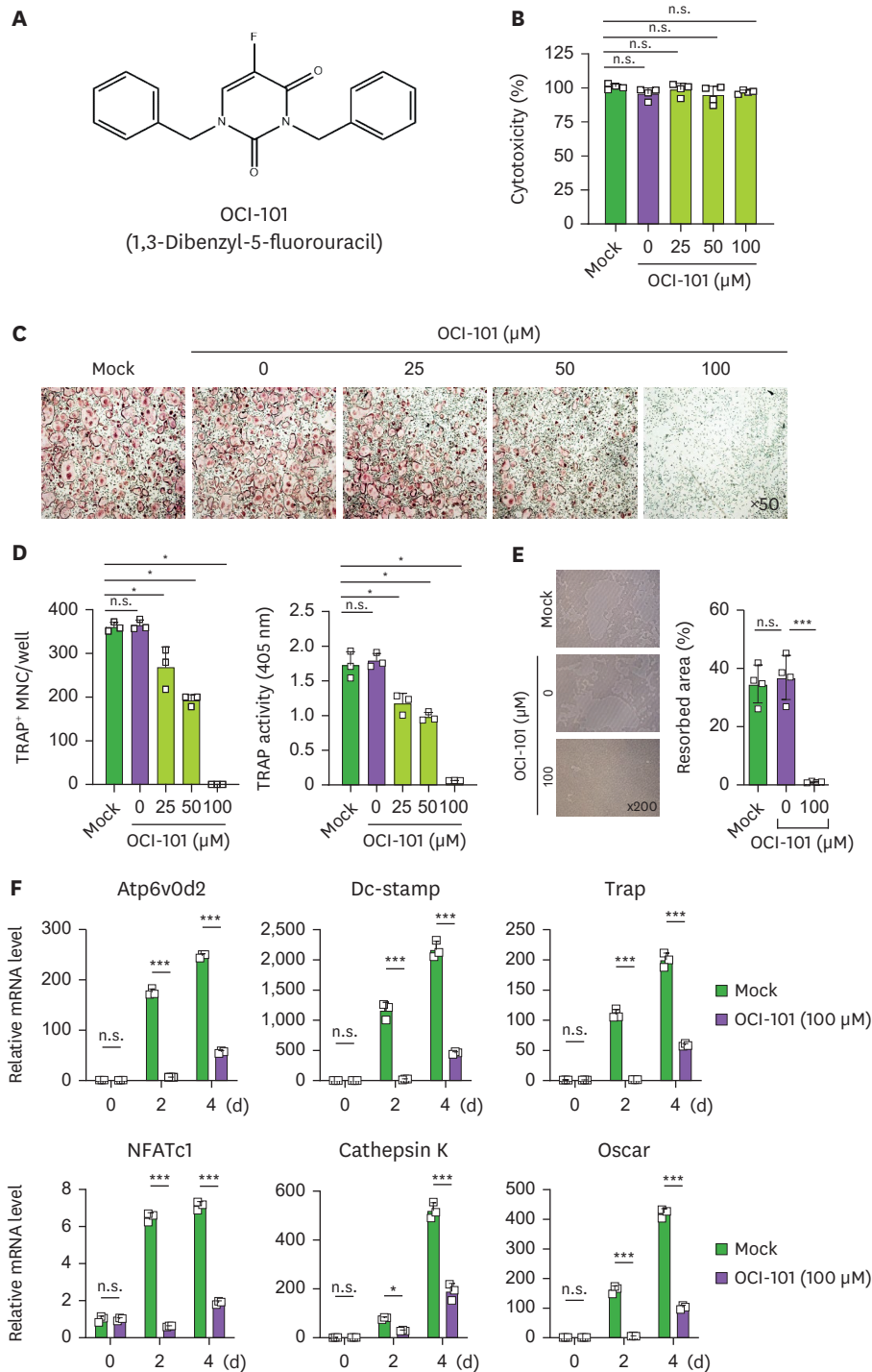


Figure 1. OCI-101 inhibits OC differentiation. (A) Chemical structures of OCI-101. (B) Cytotoxicity tests for OCI-101 in BMMs. Bone marrow cells were cultured with the indicated doses of OCI-101 in the presence of M-CSF (50 ng/ml) for 4 days. Cytotoxicity was measured by a Cell Counting Kit-8 assay kit. (C) The inhibition of BMOC differentiation by OCI-101. BMOCs were differentiated with the indicated doses of OCI-101 in the presence of M-CSF (50 ng/ml) and RANKL (200 ng/ml) for 4 days. BMOCs were analyzed by TRAP staining and photographed (original magnification, $\times 50$). (D) Summary of the inhibitory effects of OCI-101 in the BMOC formation assay. The number of TRAP⁺ MNCs (≥ 3 nuclei) was counted (left panel). TRAP solution assays were performed (right panel). (E) The inhibition of resorption pit formation by OCI-101. Resorption pits were visualized (left panel, original magnification, $\times 200$) and analyzed by using Image J software (right panel). (F) The inhibition of OC marker expression by OCI-101. Total RNA was prepared from BMOCs and subjected to real-time PCR analysis for OC marker expression. The data were normalized to β -actin.

n.s., not significant.

* $p < 0.05$; *** $p < 0.001$.

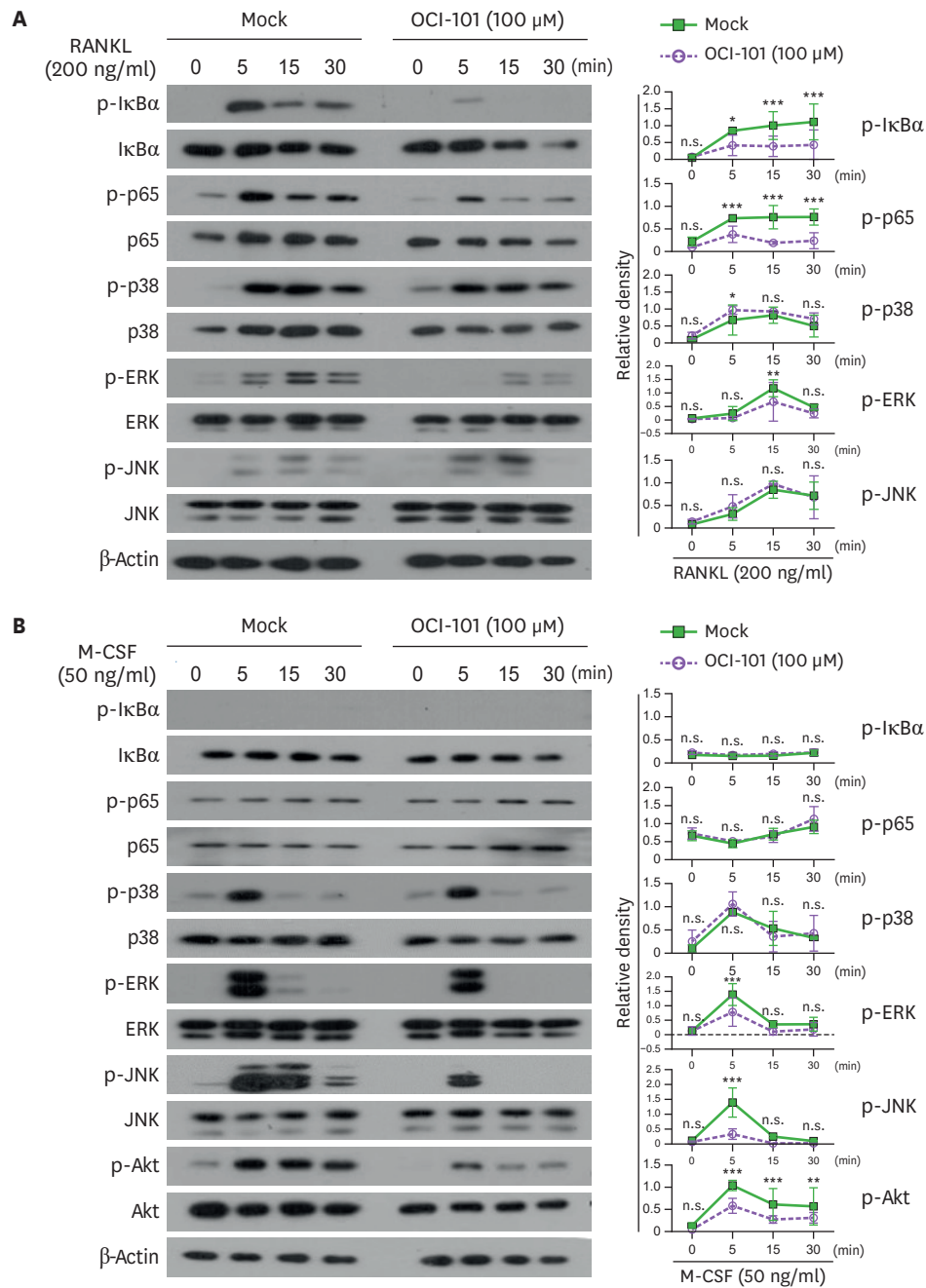


Figure 2. OCI-101 inhibits RANKL- and M-CSF-induced signaling. (A) The effect of OCI-101 on RANKL-induced signaling pathway. The cultured BMMs were preincubated with 100 μM OCI-101 for 12 h, and the cells were stimulated with RANKL (200 ng/ml) for the indicated times. The cells were then analyzed by immunoblotting with antibodies recognizing phosphorylated and total IκBα, p65, p38, ERK and JNK (left panel). β-Actin was used as the loading control. The relative level of phosphorylated proteins was calculated after normalization to total protein input (right panel). (B) The effect of OCI-101 on M-CSF-induced signaling pathway. n.s., not significant. *p<0.05; **p<0.01; ***p<0.001.

we observed that the extent of bone loss in the trabecular bones of the femur in OVX mice was not significantly altered by 1 mg/kg OCI-101 treatment, while OVX-induced bone loss in the trabecular bones was rescued by 10 mg/kg OCI-101 treatment (Fig. 3A). Consistent with the results, the number of TRAP⁺ OCs in the trabecular region of femurs was decreased (26.9%

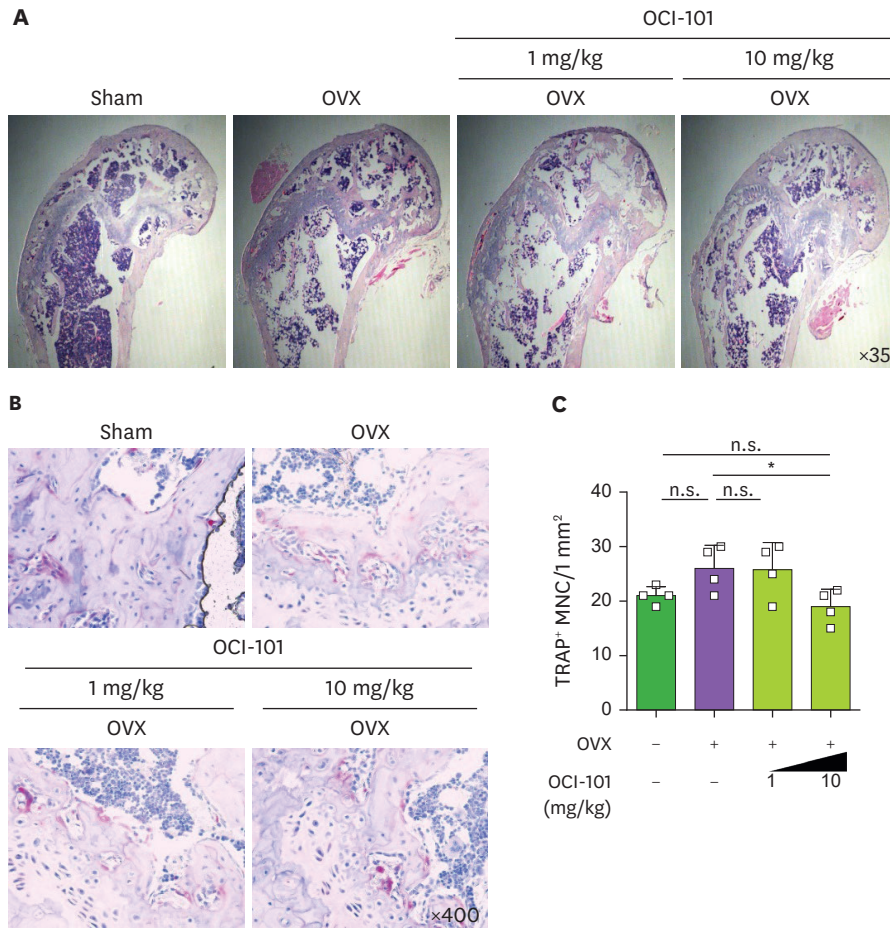


Figure 3. Morphological and histological analyses of trabecular bones in OCI-101-treated OVX mice. (A) Rescued phenotype of bone histological morphology by OCI-101 treatment in OVX mice. Femurs of mice (8-week-old, n=10–11/group) were fixed with 10% formalin, decalcified and embedded in paraffin. The paraffin sections were stained with TRAP and hematoxylin (original magnification, $\times 35$). Sham, sham operation. OVX, ovariectomy. (B) The number of TRAP⁺ OCs in the distal femurs of OCI-101-treated OVX mice was decreased. Paraffin sections were stained with TRAP and hematoxylin (original magnification, $\times 200$). (C) The number of TRAP⁺ OCs in the distal femur was counted. n.s., not significant. * $p < 0.05$.

reduction, $p < 0.05$) in OCI-101 (10 mg/kg)-treated OVX mice compared to that of OVX control mice (Fig. 3B and C). Hence, these results indicate that OCI-101 acts as an inhibitor in OVX-induced bone loss by reducing OC formation *in vivo*.

OCI-101 prevents OVX-induced bone loss in mice

We next analyzed bone microarchitecture changes in OCI-101-treated OVX mice by micro-CT. Bone parameters and 3-dimensional images of the trabecular region of the femurs were measured and quantified using micro-CT (Fig. 4A). Based on the imaging analysis, the extent of bone loss in the trabecular bones of the femur in OVX mice was not significantly altered by 1 mg/kg OCI-101 treatment (Fig. 4A). However, we observed that the increased bone loss of the trabecular bones in OVX mice was clearly inhibited by 10 mg/kg OCI-101 treatment (Fig. 4A). Consistent with the results, we observed the rescued phenotypes in the analysis of bone parameters after 10 mg/kg OCI-101 treatment (Fig. 4B). The BMD of the femurs was significantly increased (12.8% higher [$p < 0.001$]) with high-dose OCI-101 treatment (10 mg/kg) in OVX mice compared to that in the femurs of OVX control mice (Fig. 4B, first panel). The trabecular bone volume fraction was 34.8% higher ($p < 0.01$) in OVX mice treated with a

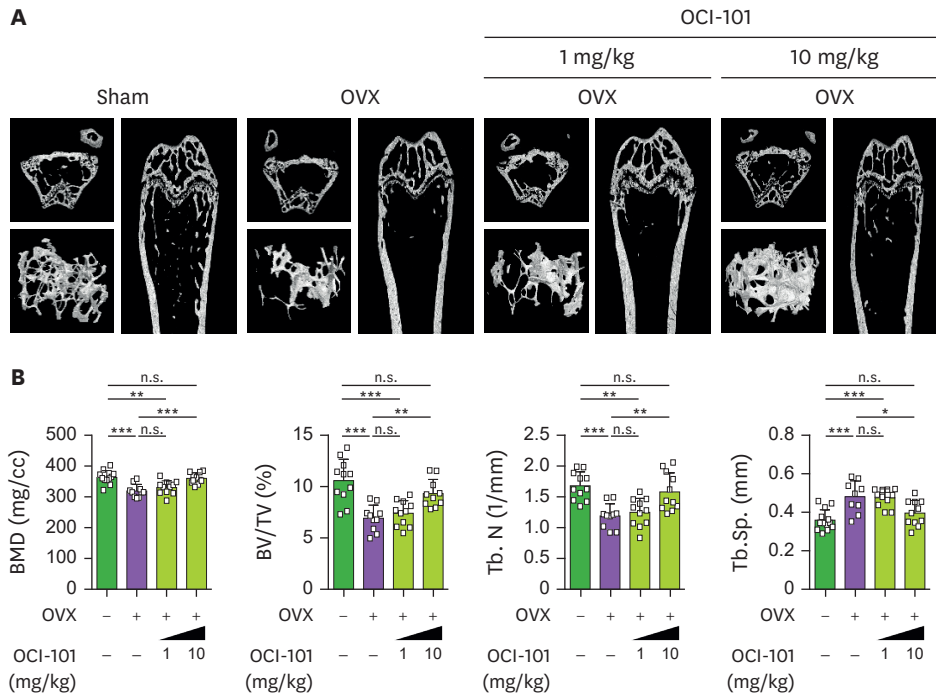


Figure 4. Micro-CT analyses of trabecular bones in OCI-101-treated OVX mice. (A) Three-dimensional microstructural analysis of the distal femur in OCI-101-treated OVX mice. Femurs of mice (8-week-old, n=10–11/group) were fixed with 10% formalin and analyzed by micro-CT. Micro-CT images of the femur are shown. (B) Three-dimensional trabecular structural parameters in distal femurs from OCI-101-treated OVX mice. The following parameters were analyzed: BMD, BV/TV, Tb. N and Tb.Sp. n.s., not significant; BV/TV, trabecular bone volume fraction, Tb. N, trabecular number; Tb.Sp, trabecular spacing. *p<0.05; **p<0.01; ***p<0.001.

high concentration (10 mg/kg) of OCI-101 than in OVX control mice (**Fig. 4B**, second panel). Furthermore, the Tb. N was 32.9% higher (p<0.01) in OCI-101 (10 mg/kg)-treated OVX mice than in OVX control mice, while trabecular spacing was 17.8% lower (p<0.05) in OCI-101 (10 mg/kg)-treated OVX mice than in OVX control mice (**Fig. 4B**, third and fourth panels). Taken together, these results suggest that OCI-101 has protective effects on the OVX-induced osteoporotic-like bone phenotype *in vivo*.

DISCUSSION

OCs are terminally differentiated and specialized multinucleated cells exclusively responsible for physiological and pathological bone destruction (2,3). Mature OCs are formed by cell fusion of TRAP⁺ mononuclear OC precursors during osteoclastogenesis (2,3). Therefore, targeting osteoclastogenesis is considered a good therapeutic strategy for management of metabolic bone diseases, including postmenopausal osteoporosis and osteoarthritis, which are characterized by accelerated bone destruction by OCs. In our current study, we identified a novel inhibitor, OCI-101, targeting OC differentiation. Our findings reveal that 1) OCI-101 is an inhibitor of osteoclastogenesis, 2) OCI-101 inhibits multiple RANKL- and M-CSF-induced signaling, and 3) OCI-101 has protective effects against OVX-induced osteoporotic-like bone loss.

RANK-RANKL signaling is a critical osteoclastogenic pathway. The stimulation of RANK by RANKL recruits TRAFs, such as TRAF2, 5, and 6, to the cytoplasmic tail of trimeric RANK, thereby finally activating the transcription factors AP-1 and NF-κB (2,3). Among the TRAFs,

TRAF6 plays a pivotal role in the activation of downstream signaling cascades, including MAPK and NF- κ B pathways during RANKL-induced osteoclastogenesis (30). RANKL-induced NF- κ B activation can trigger the induction of RANKL target gene expression (31,32). Particularly NFATc1, a master regulator of osteoclastogenesis, is one of the key target genes of NF- κ B in RANKL-induced osteoclastogenesis (33). Thus, NF- κ B activation is crucial for RANKL-induced osteoclastogenesis. In our current study, we showed that OCI-101 inhibits RANKL-induced OC differentiation via downregulation of NF- κ B pathways (Figs. 1 and 2A). The phosphorylation of I κ B α and p65 in the NF- κ B pathway was significantly reduced by OCI-101 treatment (Fig. 2A). The phosphorylation of I κ B α by the IKK complex can lead to ubiquitin-mediated I κ B α degradation, resulting in activation and nuclear translocation of NF- κ B (34). The phosphorylation of p65 induces a conformational change that promotes its stability and interaction with cofactors of transcriptional machinery, thereby increasing NF- κ B transcriptional activity (35,36). Thus, this process may explain that TRAF6-mediated IKK signaling pathways are inhibited by OCI-101 in RANKL-induced osteoclastogenesis.

The significance of RANKL-induced MAPK pathway activation is also well documented in osteoclastogenesis (2,3). In particular, the phosphorylation of p38, JNK and ERK in OC precursors by RANKL-induced TRAF6 signal transduction pathways is crucial for OC differentiation (37,38). The phosphorylation of p38 activates microphthalmia-associated transcription factors and NFATc1, which are responsible for the induction of osteoclastogenic genes, such as TRAP and cathepsin K (39,40). The phosphorylation of JNK is involved in the activation of the transcription factor c-Jun in RANKL-induced osteoclastogenesis (41,42). RANKL-induced ERK activation induces c-Fos phosphorylation and matrix metalloproteinase 9 expression (43,44). In our current study, we showed that the phosphorylation of ERK, but not that of p38 or JNK, by RANKL stimulation was reduced by OCI-101 treatment (Fig. 2A). Thus, it is possible that TRAF6 itself or the TRAF6 upstream signaling pathway is not the direct target of OCI-101. Hence, we presume that OCI-101 plays functionally distinct roles in blocking ERK activation of RANKL-RANK-TRAF6-mediated signaling pathways. RANKL-induced ERK activation is crucial for osteoclastogenesis (45,46). ERK1 deficiency reduces OC differentiation, activation and survival (47). Furthermore, it has been reported that M-CSF signaling is also linked to ERK activation in osteoclastogenesis (44). M-CSF-induced ERK activation leads to OC proliferation and survival (48,49). Collectively, ERK activation is mediated by both RANKL- and M-CSF-induced signaling in osteoclastogenesis. Based on our observations showing the reduction of ERK phosphorylation by OCI-101 (Fig. 2), we hypothesized that the inhibitory effects of OCI-101 might be linked to both RANKL- and M-CSF-induced ERK activation in osteoclastogenesis. However, a direct target of OCI-101 for the ERK signaling pathway was not identified in this study; thus, further studies are required to elucidate how OCI-101 modulates the RANKL-induced ERK activation pathway in osteoclastogenesis.

In conclusion, we show that OCI-101 is a novel inhibitor of osteoclastogenesis. OCI-101 inhibits OC differentiation by blocking multiple RANKL- and M-CSF-induced signaling pathways. Finally, we show that OCI-101 prevents OVX-induced osteoporotic-like bone loss by suppressing OC differentiation in mice. Hence, these results show that OCI-101 is a good drug candidate to treat postmenopausal osteoporosis and inflammatory bone-related diseases.

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