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Rat Dlx5 is expressed in the subventricular zone and promotes neuronal differentiation

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Abstract

The molecular mechanisms and potential clinical applications of neural precursor cells have recently been the subject of intensive study. Dlx5, a homeobox transcription factor related to the distal-less gene in *Drosophila*, was shown to play an important role during forebrain development. The subventricular zone (SVZ) in the adult brain harbors the largest abundance of neural precursors. The anterior SVZ (SVZa) contains the most representative neural precursors in the SVZ. Further research is necessary to elucidate how Dlx5-related genes regulate the differentiation of SVZa neural precursors. Here, we employed immunohistochemistry and molecular biology techniques to study the expression of Dlx5 and related homeobox genes Er81 and Islet1 in neonatal rat brain and *in vitro* cultured SVZa neural precursors. Our results show that Dlx5 and Er81 are also highly expressed in the SVZa, rostral migratory stream, and olfactory bulb. Islet1 is only expressed in the striatum. In cultured SVZa neural precursors, Dlx5 mRNA expression gradually decreased with subsequent cell passages and was completely lost by passage four. We also transfected a Dlx5 recombinant plasmid and found that Dlx5 overexpression promoted neuronal differentiation of *in vitro* cultured SVZa neural precursors. Taken together, our data suggest that Dlx5 plays an important role during neuronal differentiation.

Key words: Neural precursors; Dlx5; Green fluorescent protein; Recombinant plasmids

Introduction

Neural precursors are a population of self-renewing cells with the potential to differentiate into multiple lineages. Under appropriate conditions, these cells can undergo asymmetric cell division, giving rise to neurons and neuronal precursors that continue to self-renew (for a review, see Ref. 1). Therefore, these neural precursors can be used to replace injured neurons. However, the molecular mechanisms underlying neural precursor differentiation are unknown, limiting the clinical utilization of neural precursors. This research area is currently under intensive investigation. Elucidating the molecular mechanisms of neural precursor differentiation will greatly facilitate the clinical application of neural precursors to the treatment of central nervous system damage.

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Populations of neural precursors are present in numerous areas of embryonic and adult mouse brains, including the midbrain, hippocampus, subventricular zone (SVZ), and ganglionic eminence. Neural precursors are found in the SVZ of human embryos and adults, but are concentrated in the lateral ventricles of the SVZ during adulthood (2,3). The anterior SVZ (SVZa) contains most of the neural precursors in the SVZ. Starting from the embryonic stage and continuing under the regulation of dorsoventral signaling, SVZa precursors migrate toward the core of the olfactory bulb (OB) via a highly specific pathway, the rostral migratory stream (RMS). The precursors from the central area then scatter toward the granule and peripheral layers, where they differentiate into mature OB interneurons (4). This process of directional migration, proliferation, and differentiation provides continual renewal of OB interneurons (5,6). This renewal is apparent in the neonatal stage and persists in adults (7). Therefore, SVZa neural precursors are an ideal model for the study of neuronal precursor migration and differentiation (8).

A multitude of molecules have been shown to regulate the migration and differentiation of SVZa neural precursors. Our previous studies have indicated that Pax6, BMPs, Wnt-1, and Mash1 play important roles in these processes (9). Recently, Dlx5 was shown to be involved in the development of the central nervous system, especially the forebrain (10). Distal-less homeobox (Dlx) genes belong to a family of highly conserved homeobox transcriptional factors homologous to the *Drosophila* distal gene (for a review, see Ref. 11) and are located on chromosomes in a paired manner. Three pairs have been cloned so far, Dlx1 and 2, Dlx3 and 4, and Dlx5 and 6, all of which are important during forebrain development (for reviews, see Refs. 11 and 12). Among these, Dlx5 is crucially involved in the development of OB neurons. Studies using knock-out mice have shown that Dlx5 plays a critical role in the differentiation of OB interneurons and the axonal contact of OB receptor neurons (13). Dlx5 is also required for the maturation of SVZa precursors in the OB (14). Furthermore, it specifically regulates differentiation of the precursors related to olfactory system development and is therefore essential for neural development after birth (10). It has been hypothesized that Dlx5 may be involved in the migration and differentiation of SVZa precursors. Further studies are required to address the function and molecular mechanisms of Dlx5.

Unfortunately, given that Dlx5 knock-out is lethal, it is not feasible to study Dlx5 function in the neonatal stage, which happens to be the major period for OB interneuron maturation. Therefore, we have been using multiple techniques, such as tissue culture, immunohistochemistry (IHC), recombinant plasmids, RT-PCR, and flow cytometry (FCM), to study the expression and localization of Dlx5 and related homeobox proteins in the brains of neonatal rats. Furthermore, we studied the effect of Dlx5 on neuronal differentiation of cultured SVZa precursors.

Material and Methods

Ethics statement

All experiments were performed with the approval of the Animal Ethics Committee of the Third Military Medical University. All experimental procedures were conducted in accordance with the National Institutes of Health "Guide for the Care and Use of Laboratory Animals" (NIH publication No. 80-23, revised in 1996). All efforts were made to minimize animal suffering and to reduce the number of animals used.

Cell culture

According to the procedures described by Liu et al. (9) and Luskin et al. (15), neonatal Wistar rats (P0) were anesthetized with 0.3% pentobarbital and the SVZa and dorsolateral striatum brain areas were dissected out. We prepared the single cell suspension using mechanical dissociation. After counting living cells with Trypan blue, 1×10^6 cells were seeded into culture flasks and cultured in serum-free medium (DMEM/F12, 1:1) supplemented with B27 (2%, Gibco, USA) and bFGF (20 ng/mL, Sigma, USA). After the formation of primary clones, a single cell suspension was prepared by mechanical dissociation of these primary clones and 1×10^6 cells were again seeded into culture flasks. Cell passage was performed by mechanical dissociation every 3-4 days.

Plasmid construction and transfection

The mouse Dlx5 (mDlx5) fragment was removed from pBluescriptBSK-mDlx5 (a gift from Prof. Giovanni Levi) by *EcoRI* and *KpnI* double digestion. The mDlx5 fragment was then ligated into a pEGFP eukaryotic expression vector (Clontech, Canada) to yield pEGFP-mDlx5, which encodes a fusion protein containing enhanced green fluorescent protein (EGFP) and Dlx5.

SVZa neural precursors (5×10^6) from the fourth passage were transfected with 5 μ g of the pEGFP-mDlx5 plasmid using Nucleofector™ (Amaxa, USA). Some cells were transfected with the pEGFP plasmid without mDlx5 as a negative control. Cells were then seeded onto 6-well plates (1×10^6 cells per well) and cultured in the presence of 10% fetal bovine serum.

Immunohistochemistry staining

Cultured neural precursor cells from the third passage were treated with fetal bovine serum (FBS) for 2 h in order to induce differentiation before immunofluorescent staining. Cells were collected and fixed in 4% paraformaldehyde (PFA) at room temperature for 30 min. Following washing with PBS, the cells were plated onto poly-lysine-coated slides and allowed to dry in a 37°C incubator. Plated cells were incubated in PBS with 5% normal goat serum at room temperature for 30 min. Primary antibodies against neuron-specific enolase (NSE) (1:100, Sigma), glial fibrillary acidic protein (GFAP) (1:100, Sigma), CNP (1:100, Sigma), nestin (1:100, Santa Cruz, USA), Dlx5 (1:100, Santa Cruz), Er81 (1:100, Santa Cruz), or Islet1 (1:100, Santa Cruz) were then added. Primary antibody binding was detected following incubation with fluorescently labeled secondary antibody goat anti-rabbit IgG-Cy3 (1:100 dilution, Sigma) or goat anti-rabbit IgG-FITC (1:100 dilution, Sigma) at 37°C for 30 min. The slides were mounted with 50% glycerol and immediately observed under a fluorescence microscope (Leica, Germany).

To analyze the distribution of Dlx5, Er81 and Islet1 proteins in the brain, neonatal rats were anesthetized with 0.3% pentobarbital and perfused with 10 mL saline and 50 mL pre-chilled 4% PFA. After perfusion, the brains were dissected out, postfixed in 4% PFA for 24 h, and dehydrated in 30% sucrose for 48 h. Transverse slices (20 µm) were then cut with a cryostat and collected on polylysine-coated slides. Primary antibodies against Dlx5 (1:200, Santa Cruz), Er81 (1:200, Santa Cruz) and Islet1 (1:200, Santa Cruz) were used. Immunofluorescent staining of these slices was completed as described above.

RNA extraction and RT-PCR

Total cellular RNA from the first four passages of cultured SVZ neural precursors were extracted using RNAeasy kits (HuaXun Inc., China). The final product of RNA was quantified by absorption and verified by agarose gel electrophoresis. The PCR primer sequences and PCR conditions for rat Dlx5 and Er81 are listed in Table 1. Rat GAPDH was used as an internal standard.

Flow cytometry measurement

Differentially treated neural precursors were digested with 0.25% pancreatin, rinsed with D-Hank's solution and centrifuged three times at 250 g for 5 min. NSE immunofluorescence staining was then performed. The samples were fully mixed and 3 samples from each group were used for FCM detection (type PLUS of FACstar, Becton Dickinson, USA).

Statistical analysis

Data are reported as means ± SD. Statistical analysis was performed by one-way ANOVA. Differences were considered to be statistically significant when $P < 0.05$.

Results

Culture and identification of neural precursors

After 2 days of culture in serum-free medium, most of the primary cells isolated from the SVZa or the dorsolateral striatum died. However, some cells formed multicellular spheres and continued to proliferate. After cell passaging, the number of cells increased rapidly and the cell spheres expanded and became suspended in the medium to form neurospheres. As the cells proliferated, the number and size of neurospheres increased, and they tended to grow adherently (Figure 1A). After induction with 10% FBS for 2 h, the neurospheres started to adhere to the plate and to differentiate (Figure 1B). The peripheral edges of the neurospheres formed short and branchless radial protrusions (Figure 1C). After two additional days of culture, these protrusions became longer and thinner, with more and more branches. Small neurospheres were almost completely differentiated. Immunostaining results confirmed the expression of GFAP, NSE, and CNP (Figure 1D-F). These results suggested that cultured neurospheres were capable of differentiating *in vitro* into neuronal cell types, thus highlighting a basic feature of neural precursors.

Expression of Dlx5, Er81 and Islet1 in neurospheres from the SVZa and dorsolateral striatum

Immunostaining revealed that these neurospheres from the SVZa were positive for Er81, Dlx5, and nestin proteins (Figure 2A-C) but negative for Islet1 expression (Figure 2D). However, the neurospheres from the dorsolateral striatum were positive for Er81, Islet1, and nestin proteins (Figure 2A',C',D') but negative for Dlx5 expression (Figure 2B').

Localized expression of Dlx5, Er81, and Islet1 in SVZ-RMS-OB

Stenman et al. (16) have previously analyzed the expression of Dlx5/6, Er81, and Islet1 at embryonic day E12.5, E16.5 and in P0 mice. Similar to their reports, the IHC results of the present study revealed that Er81 (Figure 3A-C,E,F)

and Dlx5 (Figure 3A'-C',E',F') were widely distributed in various brain areas of neonatal rats (P0), such as the SVZa-RMS-OB, hippocampus, and subcortex. Islet1 was not expressed in the SVZa-RMS-OB, but was expressed in both the dorsolateral and ventral striatum (Figure 3G,H). However, the expression pattern for Er81 and Dlx5 observed here were different from Stenman's results (16). We observed that Er81 was also widely expressed in both the dorsolateral and ventral striatum (Figure 3D), while Stenman et al. (16) reported that Er81 was observed in scattered striatal neurons. In addition, our data showed that Dlx5 was negatively expressed in the striatum (Figure 3C'-D'), while Stenman et al. (16) showed that Dlx proteins were positively expressed in the striatum. Thus, we think that the difference in the expression pattern of Dlx proteins in the striatum might be caused by multiple factors. For example, the Stenman study used specimens and Dlx antibodies from different sources that might have contributed to the differences observed. Furthermore, the expression pattern of Dlx proteins in Stenman's study may reflect the distribution of Dlx1, Dlx2, Dlx3, Dlx4, and Dlx6 in the developing brain, while in the present study it specifically reflected the SVZa-RMS-OB location of the Dlx5 protein in the newborn rat brain.

The effects of mDlx5 on the morphology and differentiation of SVZa neural precursors

Green fluorescence first appeared 4 h after pEGFP-mDlx5 transfection and dramatically increased at 8 h, peaking 24 to 48 h post-transfection and lasting several days. At 24 h post-transfection, total cellular RNA was extracted, and mDlx5 mRNA could be detected by RT-PCR (Figure 4A: the product is 897 bp). At 24 and 48 h post-transfection, the target 58-kDa protein could be detected by Western blot (Figure 4B), indicating that the transfected GFP-mDlx5 fusion protein was expressed robustly. These results indicated that we could use this system to study the role of Dlx5 in SVZa neural precursors further.

The transfected SVZa neural precursors were cultured for 24 h and then stained with NSE-Cy3. As shown in Figure 5, the transfected cells were GFP-positive (Figure 5A) and differentiated to protrusion-bearing cells (Figure 5B). The most GFP-positive cells were positive for NSE-Cy3 (Figure 5C-E), indicating that the neural precursors that expressed mDlx5 successfully differentiated into neurons. However, not all GFP-positive cells were positive for NSE (data not shown). As a negative control experiment, some cells were transfected with an empty pEGFP plasmid that did not contain mDlx5 (Figure 5F). The NSE staining results indicated that only a few scattered GFP-positive cells expressed NSE (Figure 5G,H).

In passages 1 through 3 of the *in vitro* cultured SVZa neural precursors, expression of endogenous Dlx5 (rat Dlx5) mRNA could be detected by RT-PCR (Figure 6, left panel). However, endogenous Dlx5 could no longer be detected in passage 4 precursors (Figure 6, middle panel). In contrast, Er81 mRNA was detected in different passages of SVZa precursors (Figure 6, middle panel). When the exogenous mDlx5 gene was transfected into passage 4 precursors, the expression of mDlx5 and of endogenous rat Dlx5 mRNA appeared after 24 h of culture (Figure 6, right panel).

Next, we used FCM analysis to determine the extent of neuronal differentiation in precursor populations expressing different amounts of Dlx5. As shown in Figure 7, $19.2 \pm 0.8\%$ of the passage 3 precursors with endogenous Dlx5 mRNA could differentiate to NSE-positive neurons. In passage 4 precursors, which did not have Dlx5 mRNA, a significantly smaller fraction of cells (only $8.3 \pm 0.4\%$; $P < 0.05$) differentiated to NSE-positive cells. Intriguingly, in the same background, the neuronal differentiation rate was enhanced to $35.7 \pm 0.6\%$ when mDlx5 was overexpressed. Therefore, we conclude that overexpression of Dlx5 ($P < 0.05$) significantly promoted neuronal differentiation of SVZa precursors.

Discussion

In this study, we showed that the precursors from SVZa were positive for Dlx5 and Er81 but negative for Islet1, while the precursors from the dorsolateral striatum were positive for Er81 and Islet1 but negative for Dlx5. Similar to that reported in a previous investigation (16), the present IHC data indicated that Dlx5 and Er81 are widely expressed across brain areas, such as SVZa-RMS-OB, hippocampus, and subcortex. Islet1, on the other hand, is not expressed in these areas but is localized in the striatum. Interestingly, we observed that Dlx5 was negative but Er81 was positively expressed in the striatum area, again differing from the previous report (16). In addition, the expression of endogenous Dlx5 mRNA in cultured SVZa neural precursors was gradually down-regulated and was completely lost by the fourth passage. Exogenous Dlx5 transfected into passage 4 precursors could stabilize the levels of Dlx5 and promote the neuronal differentiation of cultured SVZa neural precursors. These data demonstrated that Dlx5 plays an important role during neuronal differentiation. Further investigation is required to elucidate the underlying mechanism.

The expression of Dlx5 in SVZa-RMS-OB and SVZa neural precursors

Wichterle et al. (17) studied the migration pathway of embryonic lateral ganglionic eminence (LGE) cells and found that LGE cells continuously migrate to the OB and become the major source of OB interneurons. However, not all LGE cells migrate to the OB; some cells migrate to the striatum and become striatal projection neurons. These results suggest

that there are at least two populations of neural precursors in the LGE. Further studies by Stenman et al. (16) showed that only the cells in the dorsolateral LGE migrate to the OB and differentiate into OB interneurons. These cells have distinct molecular characteristics: they are positive for Dlx and Er81 protein but negative for Islet1. Most of the cells in the ventral lateral LGE migrate toward the striatum and eventually differentiate into striatal interneurons and projection neurons, which are positive for Dlx and Islet1 but negative for Er81. Because the SVZa neural precursors also directionally migrate to the OB and differentiate there, we hypothesized that SVZa neural precursors exhibit the same molecular features as LGE dorsolateral cells. Specifically, we hypothesized that they are also positive for Dlx and Er81 and negative for Islet1.

In the present investigation, we studied the expression of Dlx5 in SVZa-RMS-OB by IHC and found that Dlx5 is widely expressed in a multitude of areas, including the SVZa-RMS-OB, hippocampus, and subcortex. However, Er81 is also widely expressed, not only in these areas but also in the striatum. Hence, Dlx5 and Er81 expression is high and overlaps in the SVZa-RMS-OB. On the other hand, Islet1 is not expressed in the SVZa-RMS-OB, but is expressed in the striatum. The expression of Dlx5 in SVZa-RMS-OB suggests a close relationship between the Dlx5 gene and the process of migration and differentiation of OB interneurons. It also confirms our hypothesis that SVZa precursors and LGE dorsolateral cells possess similar molecular features, i.e, they are positive for Dlx and Er81 and negative for Islet1. Previous studies have shown that Dlx5 mRNA is highly expressed in forebrain subcortex GABA and dopamine neurons as well as in precursors of GABA neurons, but is low in differentiated neurons (11). Dlx5 appears to be preferentially expressed in neural precursors and stem cells. Therefore, the dorsolateral area of the LGE in embryonic stages may be similar to the postnatal SVZa structure. Furthermore, the SVZa neural precursors in neonates may originate from the dorsolateral area of the LGE.

Neural precursors isolated and cultured from the SVZa exhibited features of neural stem cells. Our data demonstrate that they continuously divide and proliferate, form suspended neurospheres in serum-free medium, show positive staining for the stem cell marker nestin (for a review, see Ref. 18), and can differentiate into neural cell types. These results suggest that, indeed, we successfully isolated neural precursors. Immunostaining confirmed that these precursors are positive for Dlx5 and Er81 and negative for Islet1. We performed immunostaining studies on dorsolateral striatal neural precursors to compare the molecular characteristics of SVZa neural precursors and neural precursors from other brain areas. We found that these cells are Islet1 and Er81 positive but Dlx5 negative. These data suggested that detecting Dlx5, Er81, and Islet1 proteins in cultured SVZa neural precursors can facilitate exclusion of the precursors from other regions (at least the dorsolateral striatum). Islet1 and Er81 have been used as markers for striatal and OB interneurons, respectively (16), although their functions are still largely unknown. Our studies showed that Dlx5 is closely involved in the differentiation of OB interneurons from SVZa precursors and may play an important role in SVZa neural precursor differentiation.

Effect of Dlx5 on the neuronal differentiation of SVZa neural precursors

After migration to the OB through the RMS, SVZa neural precursors terminally differentiated into OB interneurons. This process is regulated by multiple signaling events at both the transcriptional and post-transcriptional levels. Long et al. (14) showed that Dlx5 knock-out mice had few OB interneurons, suggesting that Dlx5 plays a major role in the differentiation of OB interneurons and in the synaptogenesis of OB receptor neurons. The neural precursors isolated from the telencephalon of Dlx5^{-/-} mice lost the ability to differentiate into neurons. Therefore, we speculated that Dlx5 is related to the migration and neuronal differentiation of SVZ precursors. Dlx5^{-/-} mice are neonatal lethal and therefore cannot be studied further. Hence, we studied the role of Dlx5 in neuronal differentiation of SVZa neural precursors using recombinant plasmid and *in vitro* culture methods.

Green fluorescence protein is a small stable protein that does not interfere with the function of cells and shows no toxicity, even when overexpressed (19). It is a widely used marker for live fluorescent labeling. Here, we used the EGFP vector to construct the plasmid pEGFP-mDlx5. This permitted us to directly monitor mDlx5 expression in SVZa neural precursors and to detect the cells expressing the target gene. Using this system, we confirmed the expression of mDlx5-GFP fusion protein in transfected SVZa neural precursors by fluorescence microscopy. Because the molecular weight (MW) of GFP is approximately 27 kDa and that of rat Dlx5 is 31 kDa, the fusion protein should be approximately 58 kDa. Our Western blot data demonstrated that the MW of the fusion protein matches the expected MW (Figure 5B), thus further confirming that the recombinant plasmid can be used to study the role of Dlx5 in the migration and differentiation of SVZ neural precursors.

The technique of live fluorescent labeling provided a very useful tool for analyzing the morphology of cells in the central nervous system. Here we studied the morphological changes in SVZa neural precursors following overexpression of GFP-mDlx5, demonstrating that the transfected cells differentiate into neuron-like cells. IHC staining for NSE confirmed that most of the Dlx5-transfected cells differentiated into neurons. We further found that the expression of endogenous

Dlx5 decreased with increasing cell passage and became undetectable by RT-PCR in passage 4 cells. The Er81 expression level was unchanged during subsequent passages. Consistently, when the passage 4 precursors were induced to differentiate, less than 10% of cells successfully differentiated into neurons. This leads us to conclude that Dlx5 levels may play an important role during the neuronal differentiation of SVZa neural precursors. This conclusion is further supported by the previous report that the Dlx genes encode homeodomain transcription factors related to *Drosophila* distal-less gene, and their mammalian homologs, Dlx1, Dlx2, Dlx5, and Dlx6, are expressed in the subcortical forebrain and are involved in the initiation of GABAergic neuron differentiation and neuronal migration (10,11).

We have provided multiple lines of evidence demonstrating that Dlx5 plays an important role in the neuronal differentiation of SVZa neural precursors. First, Dlx5 and Er81 are expressed in SVZa neural precursors; this expression pattern is one of the molecular features of SVZa precursors. In the *in vitro* cultured SVZa neural precursors, Dlx5 expression is gradually lost and can be regained by exogenous Dlx5 transfection, suggesting the existence of auto-regulation and external cue regulation. Taken together, these results demonstrate that Dlx5 plays an important role during neuronal differentiation.

Acknowledgments

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Table 1. Primer sequence and reaction conditions for PCR experiments.

Genes	Sense primer (5'→3')	Anti-sense primer (5'→3')	Renaturation temperature	Amplification length
mDlx5	AGAGTCCCAAGCATCCGATCC	CCAGCACAACACTGTAGTCCC	56.8°C	897 bp
rDlx5	TTATGCGGACTACGGCTACG	GGCAGGTGGGAATTGATTGA	52.3°C	543 bp
Er81	AAGGGTCCCAGGCAGTTCTAT	GATGCTCTTCAGGCTCAATCA	52.9°C	229 bp
GAPDH	TGCTGAGTATGTCGTGGAGT	AGTCTTCTGAGTGGCAGTGAT	55.0°C	289 bp

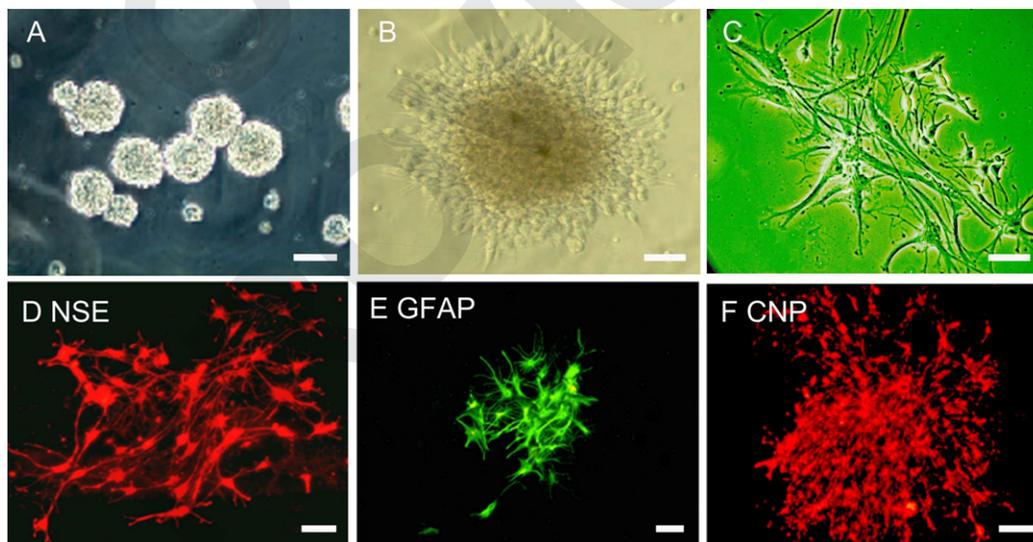


Figure 1. *In vitro* culture of anterior subventricular zone neural precursors. The light microscope images show that the subventricular zone neural precursors form neurospheres in suspension culture and rapidly increase in size and tend to grow together (A). After 2 h of induction with 10% fetal bovine serum, a light microscope image indicates that the neurosphere starts to adhere to the culture plate (B). The neurospheres shown in C are at the furthest periphery. The images of immunohistochemical staining show that the neurospheres were positive for the antibodies against NSE (D), GFAP (E), and CNP (F). NSE = neuron-specific enolase; GFAP = glial fibrillary acidic protein; CNP = _____ . Magnification bars are A = 50 μ m, B and F = 40 μ m, C-E = 20 μ m.

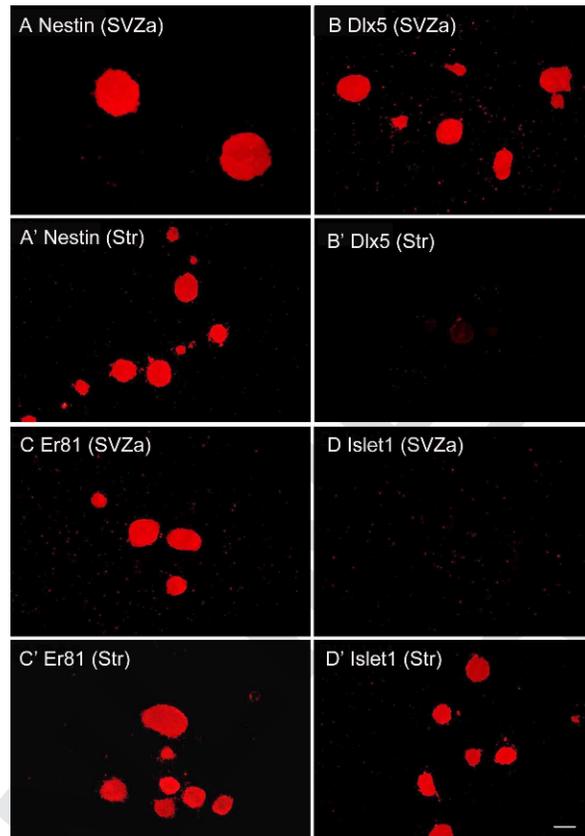


Figure 2. Expression of nestin (A, A'), Dlx5 (B, B'), Er81 (C, C'), and Islet1 (D, D') in the neurospheres from anterior subventricular zone (SVZa) and striatum (Str). The neurospheres from the SVZa are positive for nestin (A), which is a precursor cell marker. They are also positive for Dlx5 (B) and Er81 (C), but negative for Islet1 (D). However, the neurospheres from the striatum are positive for nestin (A'), Er81 (C') and Islet1 (D') but negative for Dlx5 (B'). Magnification bar = 50 μ m for all panels.

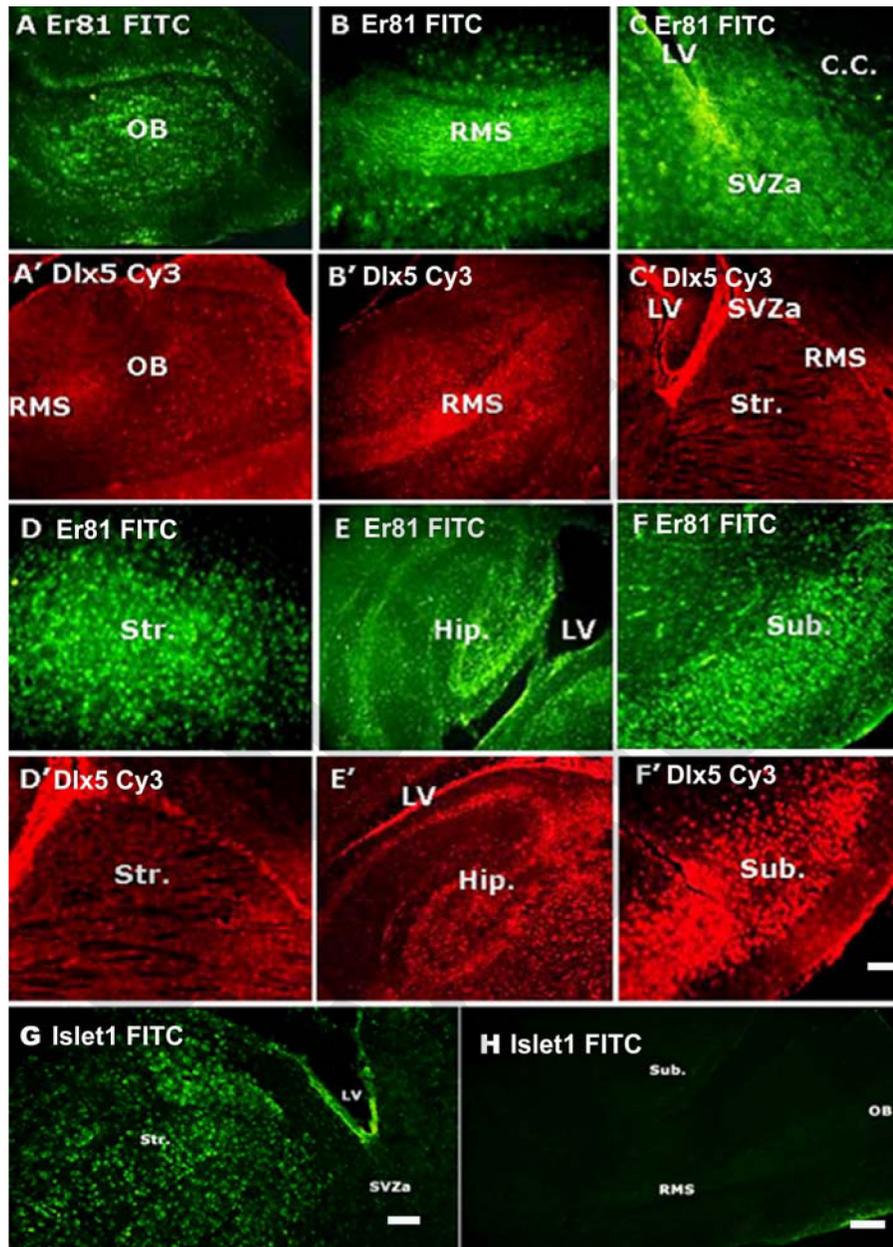


Figure 3. Expression of Er81 (A-F), Dlx5 (A'-F'), and Islet1 (G-H) at P0. Er81 proteins are widely expressed in various brain areas including OB (A), RMS (B), SVZa (C), striatum (D), hippocampus (E), and subcortex (F). Similarly, Dlx5 are also located in the OB (A'), RMS (B'), SVZa (C'), hippocampus (E'), and subcortex (F'), but not in the striatum (C',D'). Islet1 was not expressed in the SVZa, RMS and OB but was expressed in the striatum (Figure 3G,H). OB = olfactory bulb; RMS = rostral migratory stream; SVZa = anterior subventricular zone; LV = lateral ventricle; C.C. = corpus callosum; Str. = striatum; Hip. = hippocampus; Sub. = subcortex. Magnification bars are A-F and A'-F' = 50 μ m, G = 30 μ m, H = 100 μ m.

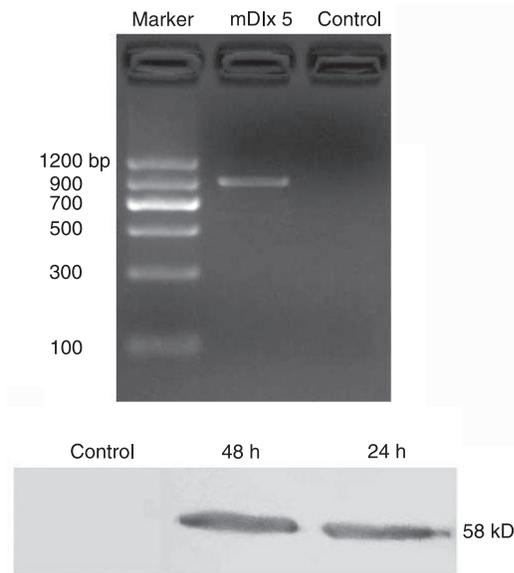


Figure 4. Expression of mouse Dlx5 (mDlx5) in anterior subventricular zone (SVZa) precursors transfected with pEGFP-mDlx5. Agarose gel electrophoresis analysis of RT-PCR showed the expression of mDlx5 RNA in SVZa precursors 24 h after transfection (A). Lane M is the marker, the middle lane (mDlx5) is from cells transfected with pEGFP-mDlx5, and the right lane (control) is from cells without transfection of pEGFP-mDlx5. B, Western blot analysis showed the expression of mDlx5 proteins in SVZa precursors at 24 and 48 h after transfection.

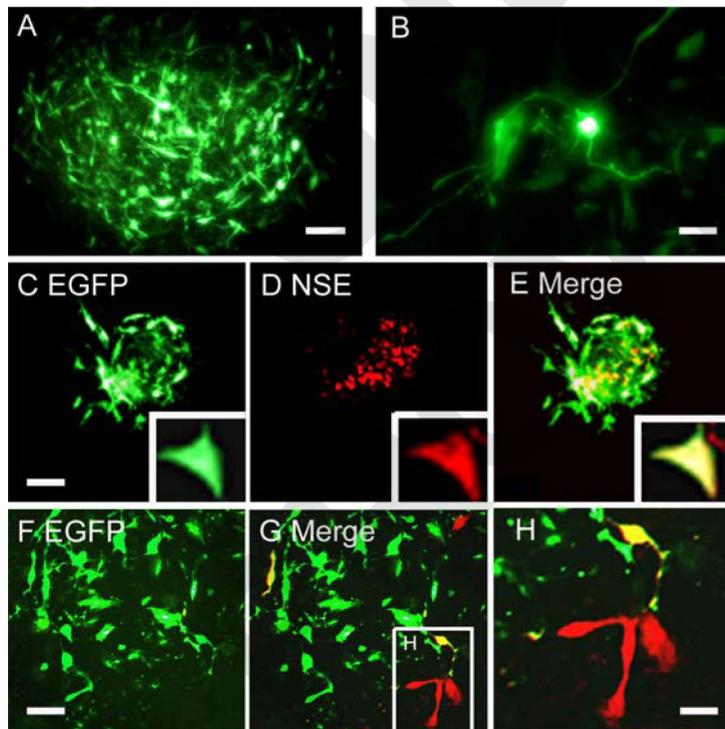


Figure 5. Expression of GFP and NSE in anterior subventricular zone (SVZa) neural precursors transfected with pEGFP-mDlx5. A, After 24 h of transfection with pEGFP-mDlx5, the majority of SVZa neurospheres expressed GFP. B, Representative GFP-positive SVZa neural precursors. Double-staining with GFP (C) and NSE-Cy3 (D) reveals that most of GFP-positive neurospheres/cells were differentiated into neurons (E). However, cells transfected with the pEGFP plasmid without mDlx5 (F) show low rates of NSE staining (G,H). GFP = glial fibrillary protein; NSE = neuron-specific enolase. Magnification bars are A = 10 μ m, B,H = 30 μ m, C-E = 100 μ m, F,G = 15 μ m.

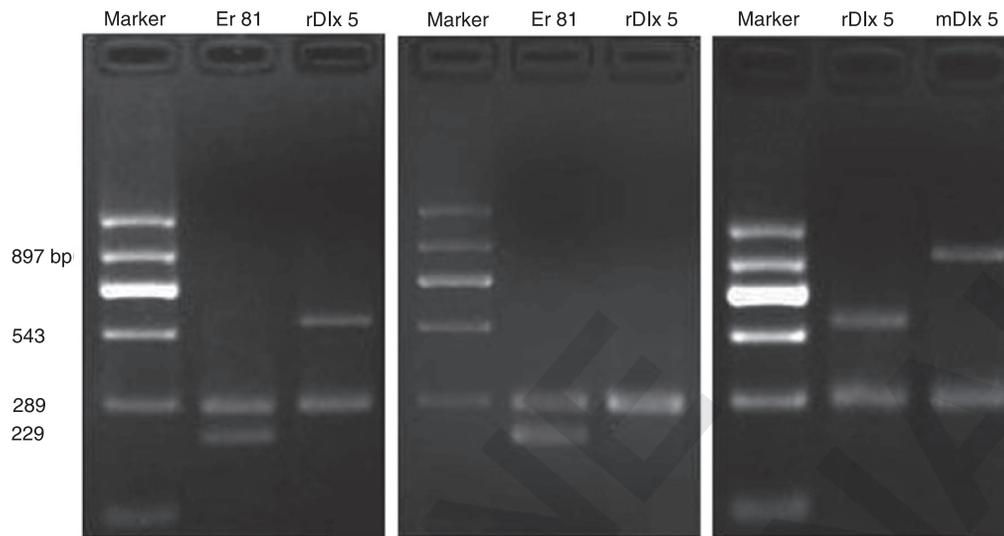


Figure 6. Expression of Dlx5 and Er81 mRNA in anterior subventricular zone (SVZa) precursors. Passage 3 (left panel) and passage 4 (middle panel) of the cultured SVZa precursors were analyzed. Er81 and endogenous Dlx5 (rDlx5) mRNA detection by RT-PCR followed by agarose electrophoresis analysis. The products are 229 and 543 bp, respectively. *Right panel*, Expression of endogenous Dlx5 (rDlx5) and transfected Dlx5 (mDlx5) mRNAs by RT-PCR followed by agarose gel electrophoresis analysis. After transfection with pEGFP-mDlx5, mDlx5 (897 bp) and rDlx5 expression can be detected in passage 4 precursors. Rat GAPDH was used as an internal standard; the product is 289 bp. GAPDH = glyceraldehyde 3-phosphate dehydrogenase.

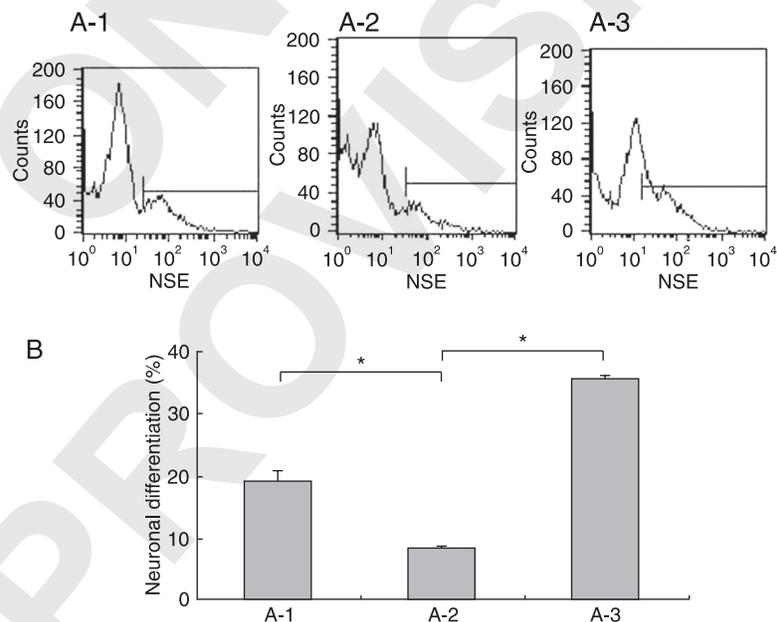


Figure 7. Determination of percent neuronal differentiation in subventricular zone (SVZ) precursors with different Dlx5 levels. *A*, Flow cytometry measurement was used to quantify NSE-positive cells after the anterior SVZ (SVZa) neural precursors differentiated. The cell populations are induced to differentiate from passage 3 of SVZ precursors (A-1) and passage 4 of SVZa precursors (A-2). Passage 4 of SVZ precursors were transfected with pEGFP-mDlx5 (A-3). *B*, Quantification and comparison of the data in *A*. One-way ANOVA confirmed that the differences between A-1 and A-2, A-2 and A-3 groups were significant (* $P < 0.01$, ** $P < 0.001$). NSE = neuron-specific enolase.