

# Renal Ischemia Regulates Marinobufagenin Release in Humans

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**Abstract**—Cardiotonic steroids, including marinobufagenin, are a group of new steroid hormones found in plasma and urine of patients with congestive heart failure, myocardial infarction, and chronic renal failure. In animal studies, partial nephrectomy induces marinobufagenin elevation, cardiac hypertrophy, and fibrosis. The objective of this study is to test the effect of renal ischemia on marinobufagenin levels in humans with renal artery stenosis (RAS). To test this, plasma marinobufagenin levels were measured in patients with RAS of the Prospective Randomized Study Comparing Renal Artery Stenting With or Without Distal Protection, non-RAS patient controls who were scheduled for coronary angiography, and normal healthy individuals. Marinobufagenin levels were significantly higher in patients with RAS compared with those of the other 2 groups. Multivariate analysis shows that occurrence of RAS is independently related to marinobufagenin levels. In addition, renal artery revascularization by stenting partially reversed marinobufagenin levels in the patients with RAS ( $0.77 \pm 0.06$  nmol/L at baseline;  $0.66 \pm 0.06$  nmol/L at 24 hours; and  $0.61 \pm 0.05$  nmol/L at 1 month). In conclusion, we have found that marinobufagenin levels are increased in patients with RAS, whereas reversal of renal ischemia by stenting treatment reduces marinobufagenin levels. These results suggest that RAS-induced renal ischemia may be a major cause of marinobufagenin release. (*Hypertension*. 2010;56:00-00.)

**Key Words:** renal artery stenosis ■ hypertension ■ cardiotonic steroids ■ marinobufagenin ■ renal artery stenting

Cardiotonic steroids (CTSs) are a group of steroid hormones that have been found recently in mammals including humans.<sup>1</sup> There is evidence demonstrating that these compounds can be synthesized endogenously and possess identical structures as their plant- and amphibian-originated counterparts.<sup>2,3</sup> In humans, an endogenous CTS, marinobufagenin (MBG), was isolated and identified from the urine of myocardial infarction patients<sup>4</sup> and from uremic plasma.<sup>2</sup> Both in vitro and in vivo studies demonstrated that epinephrine, angiotensin II, and adrenocorticotropic hormone could induce adrenal cortical cells to release endogenous CTS.<sup>5-7</sup> Other factors that can stimulate CTS include physical exercise,<sup>8</sup> hypoxia,<sup>9</sup> and behavioral stress.<sup>10</sup>

Renal artery stenosis (RAS) is a major cause for secondary hypertension in the United States.<sup>11</sup> Importantly, RAS-induced hypertension has a 3 times higher incidence of adverse cardiovascular (CV) events than those with essential hypertension when matched with equivalent blood pressure.<sup>12-14</sup> It has been reported that CTS levels increase in patients and animals with volume-expanded hypertension or preeclampsia,<sup>15,16</sup> as well as renal failure.<sup>2</sup> We have recently demonstrated in animal models that partial nephrectomy increases plasma MBG levels and induces hypertension and

cardiac fibrosis.<sup>17</sup> Neutralization of MBG by active immunization against an MBG-albumin conjugate attenuates the pathological cardiac fibrosis in rats.<sup>18,19</sup> The objective of the current study was to test whether renal ischemia induced by RAS alters MBG levels in humans.

## Methods

### Subjects

All of the subjects provided written informed consent from a protocol approved by an institutional review board. RAS subjects were from the Prospective Randomized Study Comparing Renal Artery Stenting With or Without Distal Protection (RESIST), which was conducted by the University of Toledo Clinical Coordinating Center.<sup>20</sup> The inclusion criteria were patients with hypertension and  $\geq 1$  RAS of  $\geq 50\%$  and  $< 100\%$ , treatable with stenting. The primary exclusion criteria were a systolic blood pressure  $> 200$  mm Hg or diastolic blood pressure  $> 120$  mm Hg on the day of randomization, age  $< 18$  years, pregnancy, dialysis, kidney transplant, kidney size  $< 8$  cm, restenosis, or stroke, major surgery, congestive heart failure, major trauma, and myocardial infarction within a short period of time of planned enrollment. All of the patients successfully received stenting treatment. Patient control subjects were adult patients who have a history of hypertension or angina scheduled for coronary angiography and no RAS. Normal healthy control subjects were healthy individuals (age  $> 18$  years) who have no history of hypertension, angina, or RAS.

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This trial has been registered at [www.clinicaltrials.gov](http://www.clinicaltrials.gov) (identifier NCT00234585).

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## Blood Sample Collection

All of the peripheral venous blood samples were collected in lithium heparin plasma separator tubes, spun at 1000g for 15 minutes, and the obtained plasma samples were stored at  $-80^{\circ}\text{C}$  until analysis. RAS patients also had samples collected at 24 hours and 1 month poststenting during the RESIST.

## Measurement of Plasma MBG

The plasma sample extraction was based on the method described before.<sup>21</sup> The disposable Sep-Pak C-18 columns (Waters) were activated by 10 mL of 100% acetonitrile and washed once with 5 mL of distilled water. Plasma samples (0.5 mL) were then loaded onto the columns. After another washing step with 5 mL of distilled water, each column was eluted with 7 mL of 20% acetonitrile followed by 7 mL of 80% acetonitrile. The eluates were combined, lyophilized, and resuspended in Tris-buffered saline buffer (50 mmol/L of Trizma, 150.0 mmol/L of NaCl, and 7.7 mmol/L of  $\text{NaN}_3$  [pH 7.4]). The concentration of MBG was then determined using a competitive ELISA based on a 4G4 anti-MBG murine monoclonal antibody.<sup>22</sup> Briefly, 100  $\mu\text{L}$  of MBG standards or sample eluates were mixed with 100  $\mu\text{L}$  of anti-MBG monoclonal antibody (1:1000 dilution in Tris-buffered saline buffer with 1.00% BSA and 0.25% Tween 20). The mixture was then added to a MBG-thyroglobulin-coated and 1% BSA-blocked ELISA plate. After 1 hour of incubation, plates were washed 3 times, and secondary antimouse antibody conjugated with alkaline phosphatase (1:10 000 dilution in Tris-buffered saline buffer with 1.00% BSA and 0.25% Tween 20) was added and incubated for another 1 hour. A fluorescent signal amplifier, FDP (fluorescein diphosphate, tetraammonium salt), the substrate of alkaline phosphatase from ANASpec, was used to detect the signals after washing out the secondary antibody. The sample MBG concentrations were calculated based on the standard curve using the MBG compound. MBG was purified from parotid secretion of a *Bufo marinus* toad, and MBG-thyroglobulin was synthesized as reported previously.<sup>23</sup> The secondary antimouse antibody was purchased from Sigma.

## Other Laboratory Analyses

The baseline biomarkers were measured as described before.<sup>20</sup> Glomerular filtration rate (GFR) was calculated from the modified Modification of Diet in Renal Disease equation<sup>20,24</sup> and was used as the primary measure of renal function.

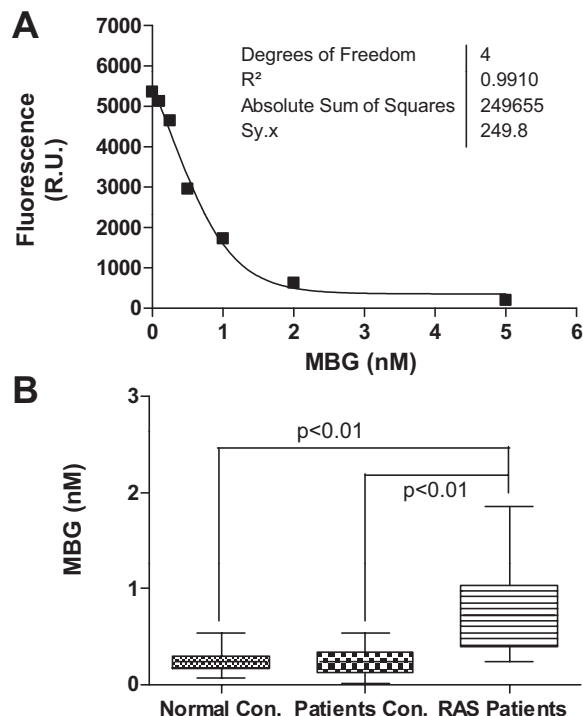
## Statistical Analysis

Study data are presented as continuous (mean  $\pm$  SE) and categorical data. Because the MBG data are not normally distributed in RAS patients, the Kruskal-Wallis test was used for the analysis to detect the significance among the RAS patients and the control subjects. Repeated-measure ANOVA was used to test the changes among the baseline, 24-hour, and 1-month poststenting for the RAS patients. We also performed a Fisher least significant difference procedure to detect the pairwise difference. Multivariate analysis was conducted using linear logistic regression. All of the analyses were performed with SAS 9.1 or JMP software (SAS Inc).

## Results

### RAS-Induced Renal Ischemia Increases Plasma MBG Levels

To test whether RAS-induced renal ischemia increases plasma MBG levels, we first compared the plasma MBG concentration in RAS patients with that in normal healthy individuals (age: 30 to 55 years and no hypertension, angina, or RAS history). The result demonstrated that MBG levels are significantly higher in RAS patients ( $0.77 \pm 0.06$  nmol/L,  $n=49$ , versus  $0.25 \pm 0.02$  nmol/L,  $n=26$ , in control subjects;  $P<0.01$ ; Figure 1). Because MBG levels were also found elevated in patients with myocardial infarction and hyperten-



**Figure 1.** Plasma MBG levels in RAS patients and control subjects. The MBG concentration was measured in plasma samples from RAS patients or control subjects as described in the Methods section. A, Representative standard curve of MBG measurement using purified MBG compound; B, distribution and mean MBG levels in normal healthy controls (Normal con.), hypertensive patient controls (Patients con.), and patients with RAS (RAS patients).

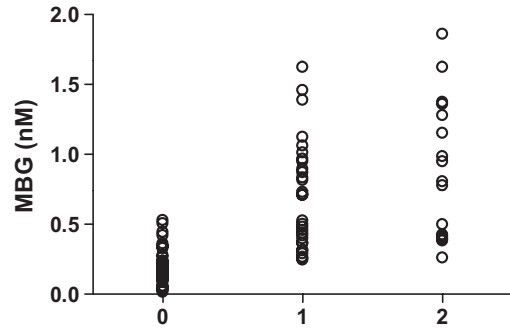
sion,<sup>4,15</sup> we then compared 60 non-RAS hypertensive patients who were scheduled for coronary angiography as an additional control group to test whether renal ischemia specifically contributes to the increased MBG in RAS patients. To eliminate the confounding factors, we compared the basic characteristics of the RAS and non-RAS patients, including their age, sex, medical history, blood pressure, kidney function, medications, and other risk factors. As shown in Table 1, the MBG concentration is significantly higher in RAS patients than in non-RAS patient controls ( $0.77 \pm 0.06$  versus  $0.20 \pm 0.06$  nmol/L;  $P<0.01$ ). Other basic characteristics of the RAS and non-RAS patients were listed in Table 1. Among these variables, the age, systolic blood pressure, presence of hypertension, GFR, use of diuretics, and use of angiotensin-converting enzyme inhibitor (ACEi)/angiotensin II receptor blocker (ARB) were significantly different between the 2 groups, and serum creatinine was close to significance. To test whether these factors are independently associated with the MBG elevation, we performed the multivariate analysis. All of the variables in Table 1 with  $P \leq 0.1$  were included in the multivariate analysis. As shown in Table 2, the occurrence of RAS and use of ACEi/ARB are independently associated with the increased plasma MBG levels in the multivariate model. For patients with RAS, the plasma MBG levels are significantly higher in patients receiving ACEi/ARB treatment than in patients without receiving ACEi/ARB treatment ( $0.93 \pm 0.08$  nmol/L,  $n=26$ , versus  $0.63 \pm 0.08$

**Table 1. Basic Characteristics of RAS Patients and Non-RAS Patient Controls**

Variables	Non-RAS Patients, (n=60)	RAS Patients, (n=49)	P, Non-RAS vs RAS
Plasma levels of CTS			
MBG, nM	0.20±0.06	0.77±0.06	<0.01
Demographic characteristics			
Age, y	61.4±1.5	70.5±1.3	<0.01
Women, n (%)	28 (46)	30 (61)	0.18
White, n (%)	52 (87)	45 (92)	0.54
BMI	31.1±1.1	28.9±0.8	0.10
Systolic BP, mm Hg	131±2	159±5	<0.01
Diastolic BP, mm Hg	77±1	76±2	0.57
Heart rate, bpm	68±1	67±2	0.63
Indications for treatment, n (%)			
Hypertension	43 (72)	48 (98)	<0.01
Angina	24 (40)	23 (47)	0.56
Laboratory values			
Serum creatinine, mg/dL	0.98±0.03	1.11±0.06	0.07
MDRD GFR	78.3±4.4	64.7±3.7	0.02
Risk factors, n (%)			
CAD	32 (53)	34 (69)	0.08
Diabetes mellitus	15 (25)	11 (22)	0.82
History of smoking	31 (52)	34 (69)	0.08
Medications, n (%)			
ACEi/ARB	18 (31)	23 (47)	0.08
Diuretics	14 (24)	23 (47)	0.02

BMI, body mass index; BP, blood pressure; CAD, coronary artery disease; MDRD GFR, GFR using modification of diet in renal disease. Values are mean±SE or No. (%) of patients.

nmol/L, n=23;  $P<0.05$ ). Figure 2 shows that MBG levels in patient plasma samples are correlated with the severity of the RAS. The average MBG concentrations are 0.20±0.06 nmol/L in patients without RAS, 0.69±0.07 nmol/L in



**Figure 2.** Distribution of plasma MBG levels in patients categorized by the severity of RAS (0 indicates patients without RAS; 1, patient with unilateral RAS; 2, patients with bilateral RAS or RAS patients with only a solitary kidney). The average MBG concentrations are 0.20±0.06 nmol/L in patients without RAS, 0.69±0.07 nmol/L in patients with unilateral RAS (n=32), and 0.88±0.12 nmol/L in patients with bilateral RAS (n=16), respectively.

patients with unilateral RAS (n=32), and 0.88±0.12 nmol/L in patients with bilateral RAS (n=16), respectively.

**Reversal of Renal Ischemia by Stenting Reduces MBG Levels**

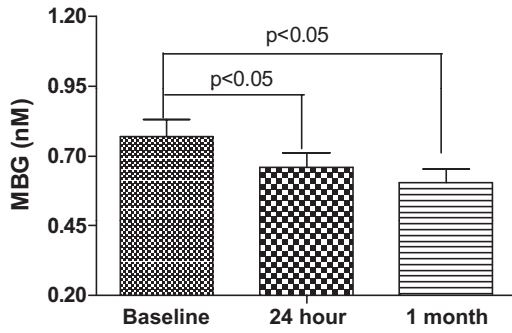
To further confirm that renal ischemia is a cause of MBG elevation in RAS patients, we measured the plasma MBG levels of RAS patients at 24 hours and 1 month after the stenting. A total of 49 available paired samples were tested. The result demonstrated that MBG levels decreased after stenting (0.77±0.06 nmol/L baseline versus 0.66±0.05 nmol/L at 24 hours and 0.60±0.05 nmol/L at 1 month;  $P<0.05$ ; Figure 3). MBG levels at 24 hours and 1 month were significantly lower than the baseline levels, but no further reduction was seen from 24 hours to 1 month. Because the RESIST patients were randomized into 4 groups (control group, Angioguard-only group, abciximab-only group, and Angioguard plus abciximab group) before receiving the renal artery stenting treatment, we also compared the changes of MBG and found no significant differences between these groups (Figure 4).

To test whether MBG levels were related to renal function, we analyzed the creatinine concentrations in plasma samples

**Table 2. Multivariate Analysis for Predictors of Plasma MBG Levels**

Model	Unstandardized Coefficients		Standardized Coefficients, $\beta$	t	Sig.	95% CI for B	
	B	SE				Lower Bound	Upper Bound
Constant	0.364	0.447		0.814	0.418	-0.524	1.252
Age	3.667×10 <sup>-5</sup>	0.003	0.001	0.012	0.990	-0.006	0.006
BMI	-0.005	0.005	-0.077	-0.950	0.345	-0.015	0.005
Serum creatinine	-0.214	0.150	-0.188	-1.424	0.158	-0.512	0.085
MDRD GFR	0.000	0.002	-0.024	-0.182	0.856	-0.004	0.003
Systolic BP	0.001	0.001	0.103	1.262	0.210	-0.001	0.004
RAS_severity	0.348	0.047	0.633	7.478	0.000	0.255	0.440
ACEi/ARB	0.152	0.065	0.178	2.351	0.021	0.024	0.281
Diuretics	0.009	0.066	0.010	0.136	0.892	-0.122	0.140

Dependent variable is Baseline\_MBG. BMI indicates body mass index; systolic BP, systolic blood pressure; RAS, renal artery stenosis; MDRD GFR, GFR using modification of diet in renal disease; Sig., significance.

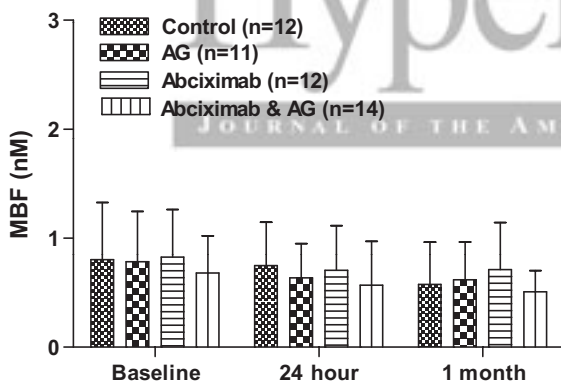


**Figure 3.** MBG levels at baseline, 24 hours, and 1 month after renal artery stenting in RAS patients. The MBG levels were measured in the plasma samples collected from patients at 3 time points. The baseline sample was collected immediately before the patient receiving the renal artery stenting, and the poststenting samples were collected at 24 hours and 1 month after receiving the renal artery stenting, respectively.

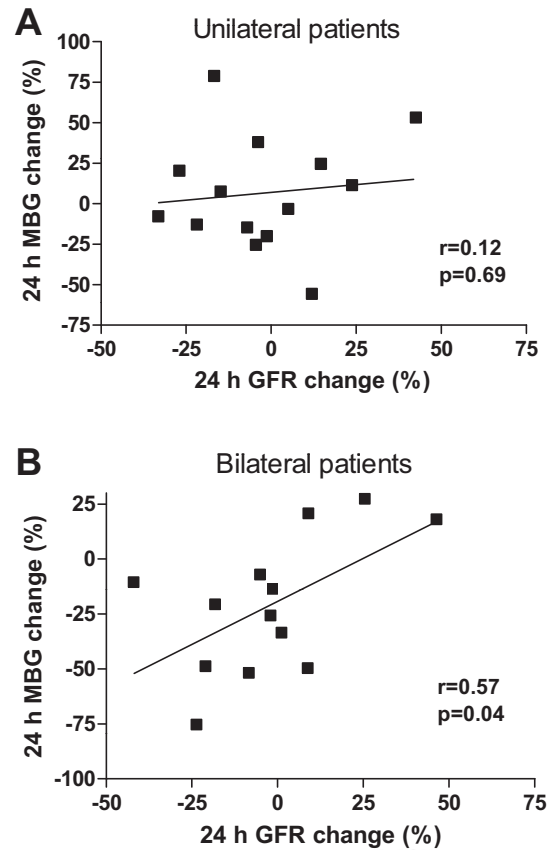
and calculated the GFR from RAS patients at baseline, 24 hours poststenting, and 1 month poststenting. As shown in Figure 5, MBG changes after stenting correlated with the GFR changes in patients with bilateral RAS ( $r=0.57$ ;  $P<0.05$ ) but not in patients with unilateral RAS ( $r=0.12$ ;  $P>0.05$ ).

### Discussion

In the current study we observed that patients with RAS appear to have elevated plasma levels of MBG when contrasted against either healthy adults or a comparator group of hypertensive patients (Figure 1 and Table 1). MBG is a bufadienolide type of CTS originally found from parotid secretion of the *Bufo marinus* toad.<sup>23</sup> It has been demonstrated that MBG can also be synthesized in animal adrenal glands<sup>25</sup> and in cultured adrenocortical cells.<sup>26</sup> Like other CTSs, MBG inhibits Na/K-ATPase activity and may poten-



**Figure 4.** MBG levels in the randomized groups of RESIST at baseline, 24 hours, and 1 month. RAS patients were randomly assigned into 4 groups before receiving the renal artery stenting, as described previously.<sup>20</sup> The control group received neither Angioguard nor abciximab; the Angioguard-only group (AG) received Angioguard during the stenting procedure; the abciximab-only group (Abciximab) received abciximab treatment before the stenting procedure; and the Angioguard and abciximab group (Abciximab & AG) received both abciximab treatment before stenting and the Angioguard during the stenting procedure. The plasma MBG concentrations were analyzed in the above 4 groups at their baseline and 24 hours and 1 month poststenting.



**Figure 5.** Correlation between MBG and GFR changes in RAS patients after renal artery stenting. The percentage changes of MBG and GFR were calculated using the following formula: 24 hour change (%)=(24 hour value–baseline value)/baseline value×100%. Negative values indicate a decrease in MBG or GFR after stenting. A, Correlation of 24-hour change between MBG and GFR in patients with unilateral RAS; B, correlation of 24-hour change between MBG and GFR in patients with bilateral RAS.

tially regulate the sodium reabsorption and kidney function in conditions of high-salt loading or plasma volume expansion.<sup>27,28</sup> Increased levels of CTS, including MBG, have been reported in patients with hypertension, myocardial infarction, and heart failure,<sup>4,29,30</sup> as well as in patients with end-stage renal disease and chronic renal diseases.<sup>2,31</sup> There is evidence from the current study showing that renal ischemia induced by RAS is causally related to increased plasma MBG in these patients. First, we found a good correlation between the severity of RAS and the plasma MBG levels in the RAS patients, as shown in Figure 2. Importantly, reversal of renal ischemia by stenting reduced the plasma MBG levels in these patients. Finally, these results are consistent with our previous animal experiments. For example, in partial nephrectomy-induced renal ischemia animals and in salt-loaded animals, both the plasma MBG level and the urine MBG excretion increased.<sup>17,19,32</sup>

The mechanism(s) linking renal ischemia to increased concentration of MBG in humans has not been characterized. Because the MBG was found mainly synthesized in the adrenal gland,<sup>25</sup> renal ischemia may trigger the release of hormones from the kidney or pituitary that, in turn, stimulates

the release of MBG from the adrenal gland. Specifically, elevation of angiotensin II has been reported to regulate CTS release from the adrenal cortex.<sup>6,8,27</sup> Despite this, the current study found that RAS patients on ACEi/ARB treatment had higher plasma MBG levels compared with the patients without ACEi/ARB treatment. It is not known whether reduced GFR in these patients has any effects on the MBG excretion, which may merit further studies to measure the 24-hour urine MBG excretion in the RAS patients.

On the other hand, MBG has been found to have a natriuretic effect in animal models with salt-induced volume expansion.<sup>32,33</sup> MBG, as well as other CTS compounds, can induce the protein endocytosis of the kidney proximal tubule Na/K-ATPase.<sup>34,35</sup> Reduced Na/K-ATPase protein and activity on the basolateral membrane of kidney proximal tubules blunt the sodium reabsorption and, therefore, increase natriuresis. The current study has not shown an independent association between the renal function (plasma creatinine level or GFR) and the baseline MBG levels. However, as shown in Figure 5, the reduction of MBG at 24 hours after renal artery stenting correlated with a decrease in GFR in patients with bilateral RAS but not in patients with unilateral RAS. We hypothesize that an acute reduction of MBG may affect the kidney function in these patients. However, it requires further study to explain the mechanisms involved in this effect.

Elevated levels of MBG may help explain the relationship between RAS and CV events. Wollenweber et al<sup>36</sup> described a 6-year CV event-free survival of 53%, with risk related to the severity of the renal stenosis. Several others have suggested that the risk of adverse CV events is high and occurs in excess of the hypertension severity.<sup>37–39</sup> More recently, a significant difference in 4-year survival was seen between those with incidental RAS compared with those without, with a graded effect on mortality, according to the severity of RAS.<sup>40</sup> In RAS patients specifically, renal dysfunction is associated with increased CV event rates and increased mortality.<sup>41,42</sup> Ventricular dysfunction and overt congestive heart failure are common in patients with RAS, just as RAS is common in patients with congestive heart failure.<sup>43</sup> The elevation of endogenous CTS has now been linked with a variety of CV and renal disease settings.<sup>44–49</sup> Animal experiments using rats and mice have demonstrated that renal ischemia induced by partial nephrectomy increases MBG and causes diastolic dysfunction and cardiac fibrosis.<sup>17–19</sup> Importantly, the cardiac fibrosis seen in such animals can be prevented by immunization against MBG, whereas infusion of MBG results in a similar pathological lesion. Our result of MBG elevation may indicate that MBG is an important contributor to the increased CV events in RAS patients.

### Perspectives

The current study shows that renal ischemia is associated with high levels of a circulating endogenous CTS, MBG. Recent work in this area demonstrates that CTSs are likely important intermediaries in the linkage between chronic kidney disease and the development of cardiac hypertrophy and fibrosis. Thus, the MBG elevation may in part attribute to the high CV events in patients with RAS, and the measure-

ment of plasma MBG levels may serve as a biomarker for the cardiorenal syndrome.

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### References

- Schoner W, Scheiner-Bobis G. Role of endogenous cardiotoxic steroids in sodium homeostasis. *Nephrol Dial Transplant*. 2008;23:2723–2729.
- Komiyama Y, Dong XH, Nishimura N, Masaki H, Yoshika M, Masuda M, Takahashi H. A novel endogenous digitalis, telocinobufagin, exhibits elevated plasma levels in patients with terminal renal failure. *Clin Biochem*. 2005;38:36–45.
- Komiyama Y, Nishimura N, Munakata M, Mori T, Okuda K, Nishino N, Hirose S, Kosaka C, Masuda M, Takahashi H. Identification of endogenous ouabain in culture supernatant of PC12 cells. *J Hypertens*. 2001;19:229–236.
- Bagrov AY, Fedorova OV, Dmitrieva RI, Howald WN, Hunter AP, Kuznetsova EA, Shpen VM. Characterization of a urinary bufodienolide Na<sup>+</sup>,K<sup>+</sup>-ATPase inhibitor in patients after acute myocardial infarction. *Hypertension*. 1998;31:1097–1103.
- Laredo J, Hamilton BP, Hamlyn JM. Ouabain is secreted by bovine adrenocortical cells. *Endocrinology*. 1994;135:794–797.
- Laredo J, Shah JR, Lu ZR, Hamilton BP, Hamlyn JM. Angiotensin II stimulates secretion of endogenous ouabain from bovine adrenocortical cells via angiotensin type 2 receptors. *Hypertension*. 1997;29:401–407.
- Fedorova OV, Anderson DE, Bagrov AY. Plasma marinobufagenin-like and ouabain-like immunoreactivity in adrenocorticotropin-treated rats. *Am J Hypertens*. 1998;11:796–802.
- Bauer N, Muller-Ehmsen J, Kramer U, Hambarchian N, Zobel C, Schwinger RH, Neu H, Kirch U, Grunbaum EG, Schoner W. Ouabain-like compound changes rapidly on physical exercise in humans and dogs: effects of  $\beta$ -blockade and angiotensin-converting enzyme inhibition. *Hypertension*. 2005;45:1024–1028.
- De Angelis C, Haupt GT Jr. Hypoxia triggers release of an endogenous inhibitor of Na<sup>+</sup>-K<sup>+</sup>-ATPase from midbrain and adrenal. *Am J Physiol*. 1998;274:F182–F188.
- Weidemann H, Salomon N, Avniti-Sagi T, Weidenfeld J, Rosen H, Lichtstein D. Diverse effects of stress and additional adrenocorticotrophic hormone on digitalis-like compounds in normal and nude mice. *J Neuroendocrinol*. 2004;16:458–463.
- Derx FH, Schalekamp MA. Renal artery stenosis and hypertension. *Lancet*. 1994;344:237–239.
- Davis BA, Crook JE, Vestal RE, Oates JA. Prevalence of renovascular hypertension in patients with grade III or IV hypertensive retinopathy. *N Engl J Med*. 1979;301:1273–1276.
- Losito A, Fagugli RM, Zampi I, Parente B, de Rango P, Giordano G, Cao P. Comparison of target organ damage in renovascular and essential hypertension. *Am J Hypertens*. 1996;9:1062–1067.
- Simon N, Franklin SS, Bleifer KH, Maxwell MH. Clinical characteristics of renovascular hypertension. *JAMA*. 1972;220:1209–1218.
- Lopatin DA, Ailamazian EK, Dmitrieva RI, Shpen VM, Fedorova OV, Doris PA, Bagrov AY. Circulating bufodienolide and cardenolide sodium pump inhibitors in preeclampsia. *J Hypertens*. 1999;17:1179–1187.
- Vu HV, Ianosi-Irimie MR, Pridjian CA, Whitbred JM, Durst JM, Bagrov AY, Fedorova OV, Pridjian G, Puschet JB. Involvement of marinobu-

- fagenin in a rat model of human preeclampsia. *Am J Nephrol.* 2005;25:520–528.
17. Kennedy DJ, Vetteth S, Periyasamy SM, Kanj M, Fedorova L, Khouri S, Kahaleh MB, Xie Z, Malhotra D, Kolodkin NI, Lakatta EG, Fedorova OV, Bagrov AY, Shapiro JI. Central role for the cardiotonic steroid marinobufagenin in the pathogenesis of experimental uremic cardiomyopathy. *Hypertension.* 2006;47:488–495.
  18. Elkareh J, Kennedy DJ, Yashaswi B, Vetteth S, Shidyak A, Kim EG, Smaili S, Periyasamy SM, Harii IM, Fedorova L, Liu J, Wu L, Kahaleh MB, Xie Z, Malhotra D, Fedorova OV, Kashkin VA, Bagrov AY, Shapiro JI. Marinobufagenin stimulates fibroblast collagen production and causes fibrosis in experimental uremic cardiomyopathy. *Hypertension.* 2007;49:215–224.
  19. Kennedy DJ, Elkareh J, Shidyak A, Shapiro AP, Smaili S, Mutgi K, Gupta S, Tian J, Morgan E, Khouri S, Cooper CJ, Periyasamy SM, Xie Z, Malhotra D, Fedorova OV, Bagrov AY, Shapiro JI. Partial nephrectomy as a model for uremic cardiomyopathy in the mouse. *Am J Physiol Renal Physiol.* 2008;294:F450–F454.
  20. Cooper CJ, Haller ST, Colyer W, Steffes M, Burket MW, Thomas WJ, Safian R, Reddy B, Brewster P, Ankenbrandt MA, Virmani R, Dippel E, Rocha-Singh K, Murphy TP, Kennedy DJ, Shapiro JI, D'Agostino RD, Pencina MJ, Khuder S. Embolic protection and platelet inhibition during renal artery stenting. *Circulation.* 2008;117:2752–2760.
  21. Bagrov AY, Fedorova OV, Austin-Lane JL, Dmitrieva RI, Anderson DE. Endogenous marinobufagenin-like immunoreactive factor and Na<sup>+</sup>, K<sup>+</sup> ATPase inhibition during voluntary hypoventilation. *Hypertension.* 1995;26:781–788.
  22. Fedorova OV, Simbirtsev AS, Kolodkin NI, Kotov AY, Agalakova NI, Kashkin VA, Tapiłskaya NI, Bzhelyansky A, Reznik VA, Frolova EV, Nikitina ER, Budny GV, Longo DL, Lakatta EG, Bagrov AY. Monoclonal antibody to an endogenous bufadienolide, marinobufagenin, reverses preeclampsia-induced Na/K-ATPase inhibition and lowers blood pressure in NaCl-sensitive hypertension. *J Hypertens.* 2008;26:2414–2425.
  23. Bagrov AY, Roukoyatkina NI, Pinaev AG, Dmitrieva RI, Fedorova OV. Effects of two endogenous Na<sup>+</sup>,K<sup>(+)</sup>-ATPase inhibitors, marinobufagenin and ouabain, on isolated rat aorta. *Eur J Pharmacol.* 1995;274:151–158.
  24. Stevens LA, Coresh J, Greene T, Levey AS. Assessing kidney function—measured and estimated glomerular filtration rate. *N Engl J Med.* 2006;354:2473–2483.
  25. Fedorova OV, Talan MI, Agalakova NI, Lakatta EG, Bagrov AY. Endogenous ligand of  $\alpha(1)$  sodium pump, marinobufagenin, is a novel mediator of sodium chloride-dependent hypertension. *Circulation.* 2002;105:1122–1127.
  26. Dmitrieva RI, Bagrov AY, Lalli E, Sassone-Corsi P, Stocco DM, Doris PA. Mammalian bufadienolide is synthesized from cholesterol in the adrenal cortex by a pathway that is independent of cholesterol side-chain cleavage. *Hypertension.* 2000;36:442–448.
  27. Bagrov AY, Shapiro JI, Fedorova OV. Endogenous cardiotonic steroids: physiology, pharmacology, and novel therapeutic targets. *Pharmacol Rev.* 2009;61:9–38.
  28. Fedorova OV, Doris PA, Bagrov AY. Endogenous marinobufagenin-like factor in acute plasma volume expansion. *Clin Exp Hypertens.* 1998;20:581–591.
  29. Liu ZQ, Ma AQ, Zhang L, Yang DY. Intra-cellular electrolyte changes and levels of endogenous digoxin-like substance within the plasma in patients with congestive heart failure. *Int J Cardiol.* 1990;27:47–53.
  30. Manunta P, Maillard M, Tantardini C, Simonini M, Lanzani C, Citterio L, Stella P, Casamassima N, Burnier M, Hamlyn JM, Bianchi G. Relationships among endogenous ouabain,  $\alpha$ -adducin polymorphisms and renal sodium handling in primary hypertension. *J Hypertens.* 2008;26:914–920.
  31. Gonick HC, Ding Y, Vaziri ND, Bagrov AY, Fedorova OV. Simultaneous measurement of marinobufagenin, ouabain, and hypertension-associated protein in various disease states. *Clin Exp Hypertens.* 1998;20:617–627.
  32. Fedorova OV, Lakatta EG, Bagrov AY. Endogenous Na,K pump ligands are differentially regulated during acute NaCl loading of Dahl rats. *Circulation.* 2000;102:3009–3014.
  33. Bagrov AY, Fedorova OV, Dmitrieva RI, French AW, Anderson DE. Plasma marinobufagenin-like and ouabain-like immunoreactivity during saline volume expansion in anesthetized dogs. *Cardiovasc Res.* 1996;31:296–305.
  34. Liu J, Periyasamy SM, Gunning W, Fedorova OV, Bagrov AY, Malhotra D, Xie Z, Shapiro JI. Effects of cardiac glycosides on sodium pump expression and function in LLC-PK1 and MDCK cells. *Kidney Int.* 2002;62:2118–2125.
  35. Periyasamy SM, Liu J, Tanta F, Kabak B, Wakefield B, Malhotra D, Kennedy DJ, Nadoor A, Fedorova OV, Gunning W, Xie Z, Bagrov AY, Shapiro JI. Salt loading induces redistribution of the plasmalemmal Na/K-ATPase in proximal tubule cells. *Kidney Int.* 2005;67:1868–1877.
  36. Wollenweber J, Sheps SG, Davis GD. Clinical course of atherosclerotic renovascular disease. *Am J Cardiol.* 1968;21:60–71.
  37. Isles C, Main J, O'Connell J, Brown I, Findlay J, Stewart R, Wilkinson R. Survival associated with renovascular disease in Glasgow and Newcastle: a collaborative study. *Scott Med J.* 1990;35:70–73.
  38. Sheps SG, Osmundson PJ, Hunt JC, Schirger A, Fairbairn JF II. Hypertension and renal artery stenosis: seral observations on 54 patients treated medically. *Clin Pharmacol Ther.* 1965;6:700–709.
  39. Valentine RJ, Clagett GP, Miller GL, Myers SI, Martin JD, Chervu A. The coronary risk of unsuspected renal artery stenosis. *J Vasc Surg.* 1993;18:433–439; discussion 439–440.
  40. Conlon PJ, Little MA, Pieper K, Mark DB. Severity of renal vascular disease predicts mortality in patients undergoing coronary angiography. *Kidney Int.* 2001;60:1490–1497.
  41. Dorros G, Jaff M, Mathiak L, Dorros II, Lowe A, Murphy K, He T. Four-year follow-up of Palmaz-Schatz stent revascularization as treatment for atherosclerotic renal artery stenosis. *Circulation.* 1998;98:642–647.
  42. Johansson M, Herlitz H, Jensen G, Rundqvist B, Friberg P. Increased cardiovascular mortality in hypertensive patients with renal artery stenosis: relation to sympathetic activation, renal function and treatment regimens. *J Hypertens.* 1999;17:1743–1750.
  43. MacDowall P, Kalra PA, O'Donoghue DJ, Waldek S, Mamtara H, Brown K. Risk of morbidity from renovascular disease in elderly patients with congestive cardiac failure. *Lancet.* 1998;352:13–16.
  44. Bagrov AY, Agalakova NI, Kashkin VA, Fedorova OV. Endogenous cardiotonic steroids and differential patterns of sodium pump inhibition in NaCl-loaded salt-sensitive and normotensive rats. *Am J Hypertens.* 2009;22:559–563.
  45. Bagrov YY, Manusova NB, Frolova EV, Egorova IA, Kashkin VA, Tapiłskaya NI, Fedorova OV, Bagrov AY. Endogenous sodium pump inhibitors, diabetes mellitus and preeclampsia preliminary observations and a hypothesis. *Pathophysiology.* 2007;14:147–151.
  46. Dostanic-Larson I, Van Huysse JW, Lorenz JN, Lingrel JB. The highly conserved cardiac glycoside binding site of Na,K-ATPase plays a role in blood pressure regulation. *Proc Natl Acad Sci U S A.* 2005;102:15845–15850.
  47. Fridman AI, Matveev SA, Agalakova NI, Fedorova OV, Lakatta EG, Bagrov AY. Marinobufagenin, an endogenous ligand of  $\alpha$ -1 sodium pump, is a marker of congestive heart failure severity. *J Hypertens.* 2002;20:1189–1194.
  48. Hamlyn JM, Ringel R, Schaeffer J, Levinson PD, Hamilton BP, Kowarski AA, Blaustein MP. A circulating inhibitor of (Na<sup>+</sup> + K<sup>+</sup>)ATPase associated with essential hypertension. *Nature.* 1982;300:650–652.
  49. Schoner W. Salt abuse: the path to hypertension. *Nat Med.* 2008;14:16–17.

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