

Supplementary Information

**A glucose-responsive complex polymeric micelle
enabling repeated on-off release and insulin protection**

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1-1 Synthesis of *t*-Boc-aminoethyl 2-bromoisobutyrate (BEBIB)

BEBIB was synthesized similar to ref.¹ A solution of *t*-Boc-aminoethyl alcohol (1.61 g, 10 mmol) and triethylamine (1.11 g, 11 mmol) in methylene chloride (50 mL) was cooled in an ice bath. Into the solution was added 2-bromoisobutyryl bromide (2.29 g, 10 mmol) in methylene chloride (20 mL). The reaction was slowly warmed to room temperature and stirred for 4 h. The salt was filtered off and the reaction mixture was extracted sequentially with water and saturated sodium bicarbonate solution. The organic phase was dried over magnesium sulfate. Solvent was evaporated and the crude product was purified by column chromatography using heptane/diethyl ether (80/20, w/w) as an eluent to get a pale-yellow liquid. Then it was dissolved and crystallized in a little bit of *n*-hexane at 4 °C to give 2.6 g of BEBIB as a white acicular crystal (yield 84%). ¹H-NMR (CDCl₃) δ (ppm): 1.45 (e, 9 H), 1.95 (a, 6 H), 3.44-3.46 (c, 2 H), 4.23-4.25 (b, 2 H), 4.78 (d, H) .

1-2 Synthesis of Monoamino-terminated poly(*N*-isopropylacrylamide) (PNIPAM₁₃₀-NH₂)

Firstly *N*-*t*Boc-PNIPAM was synthesized by ATRP of *N*-isopropylacrylamide using BEBIB as initiator and CuCl/Me₆TREN as catalyst. The detailed process is introduced as follows. CuCl (110 mg, 1.1 mmol) and Me₆TREN (276 mg, 1.2 mmol) were introduced into the reaction flask and then DMF (4 mL) was added. The sample was first stirred with ultrasonic followed by the addition of *N*-isopropylacrylamide (11.3 g, 0.1 mol) dissolved in H₂O (8 mL) and DMF (6 mL), and then degassed by two freeze-thaw cycles. Subsequently, BEBIB (217 mg, 0.7 mmol) dissolved in DMF (2 mL) was introduced into the flask and the mixed solution was degassed by freeze-thaw again. Polymerization was performed at 35 °C for 3 h and monomer conversion is about 75%. The *N*-*t*Boc-PNIPAM was purified by passing through a Al₂O₃ Column to remove the copper catalyst and then the sample was deposited in 10-fold excess of cold diethyl ether. The precipitate of *N*-*t*Boc-PNIPAM was then filtered under vacuum and dried in vacuum at room temperature.

To obtain PNIPAM-NH₂, *N*-*t*Boc-PNIPAM was firstly dissolved in CH₃OH followed by adding excess K₂CO₃ solid to totally substitute the Br- to CH₃O- group. After stirring at room temperature for 12 h, CH₂Cl₂ was added following the evaporation of

CH₃OH to dissolve the polymer. Then the polymer was treated with excess trifluoroacetic acid under vigorous stirring to remove the *t*-Boc protection group. Finally, after dialyzed against NaOH aq at pH 10 for three days and deionized water for another 4 days in a dialysis bag with a molecular cutoff of 3500 D, 7 g of PNIPAM-NH₂ was obtained by lyophilizing the solution. The DP was calculated as 130 from GPC (THF as the eluent).

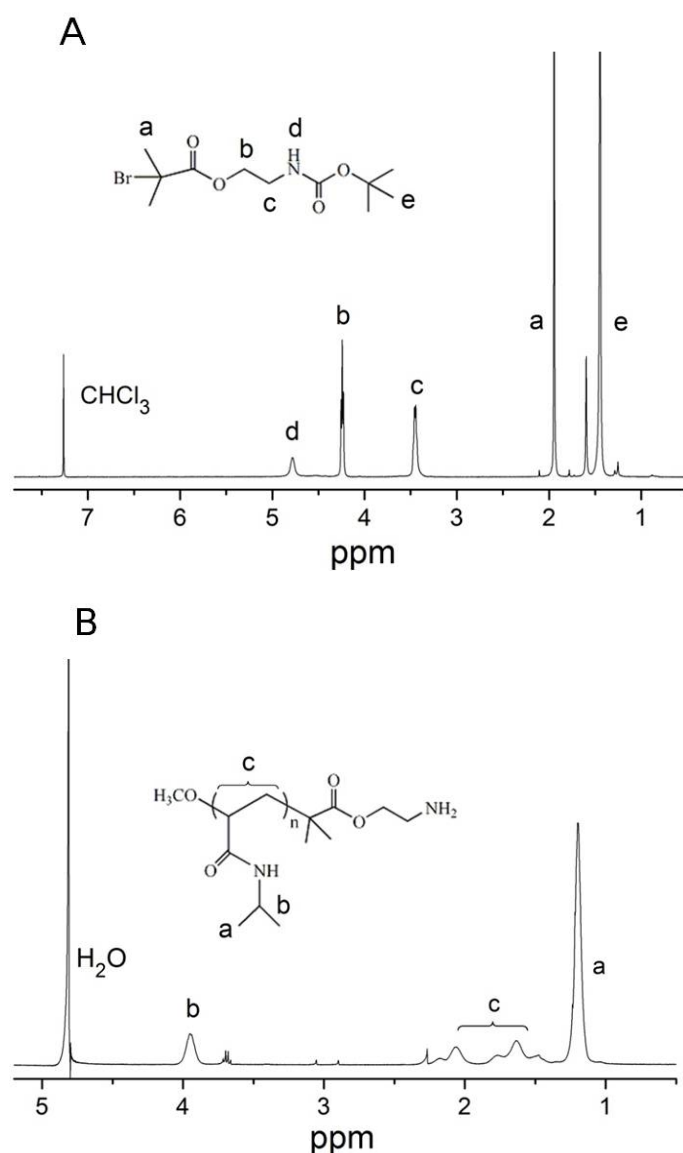


Figure S1. ¹H-NMR spectra of BEBIB in CDCl₃ (A) and PNIPAM₁₃₀-NH₂ in D₂O (B).

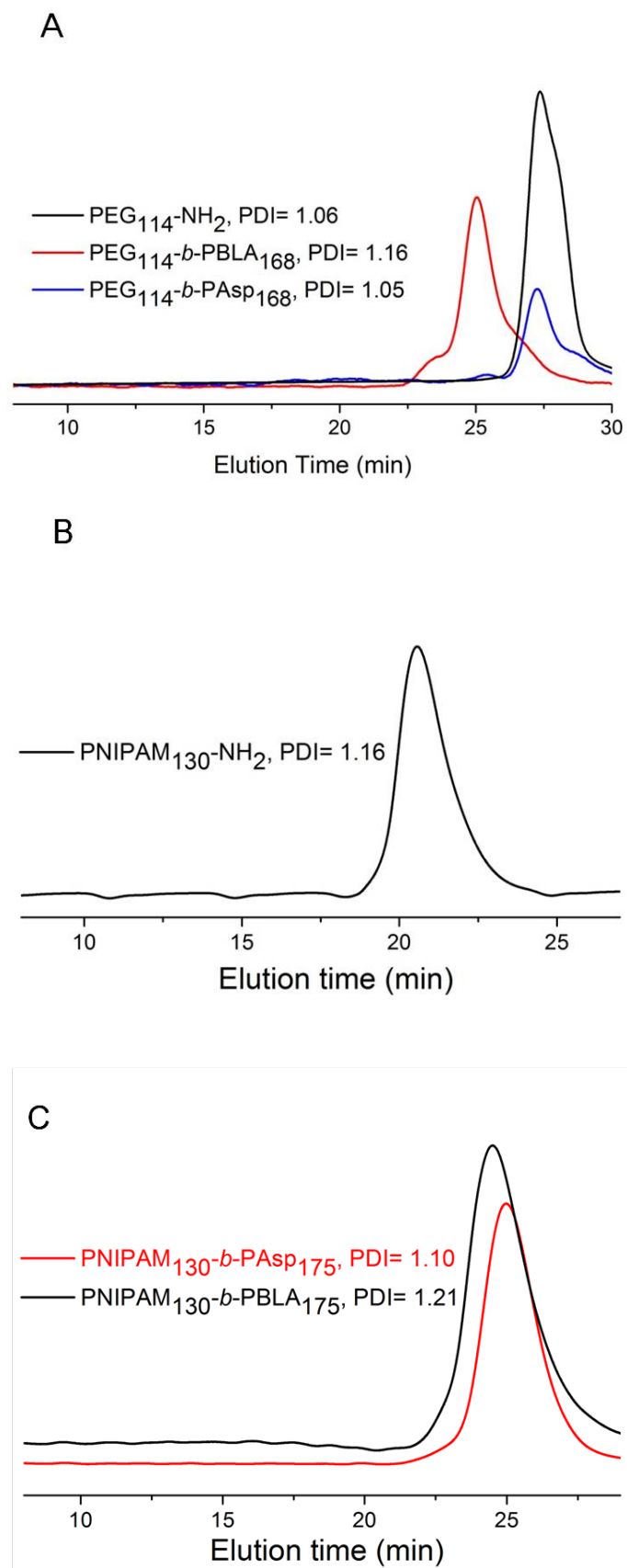


Figure S2. (A) GPC profiles of PEG₁₁₄-NH₂, PEG₁₁₄-*b*-PBLA₁₆₈ and PEG₁₁₄-*b*-PAsp₁₆₈ in DMF. (B) GPC profile of PNIPAM₁₃₀-NH₂ in THF. (C) GPC

profiles of PNIPAM₁₃₀-*b*-PBLA₁₇₅ and PNIPAM₁₃₀-*b*-PAsp₁₇₅ in DMF.

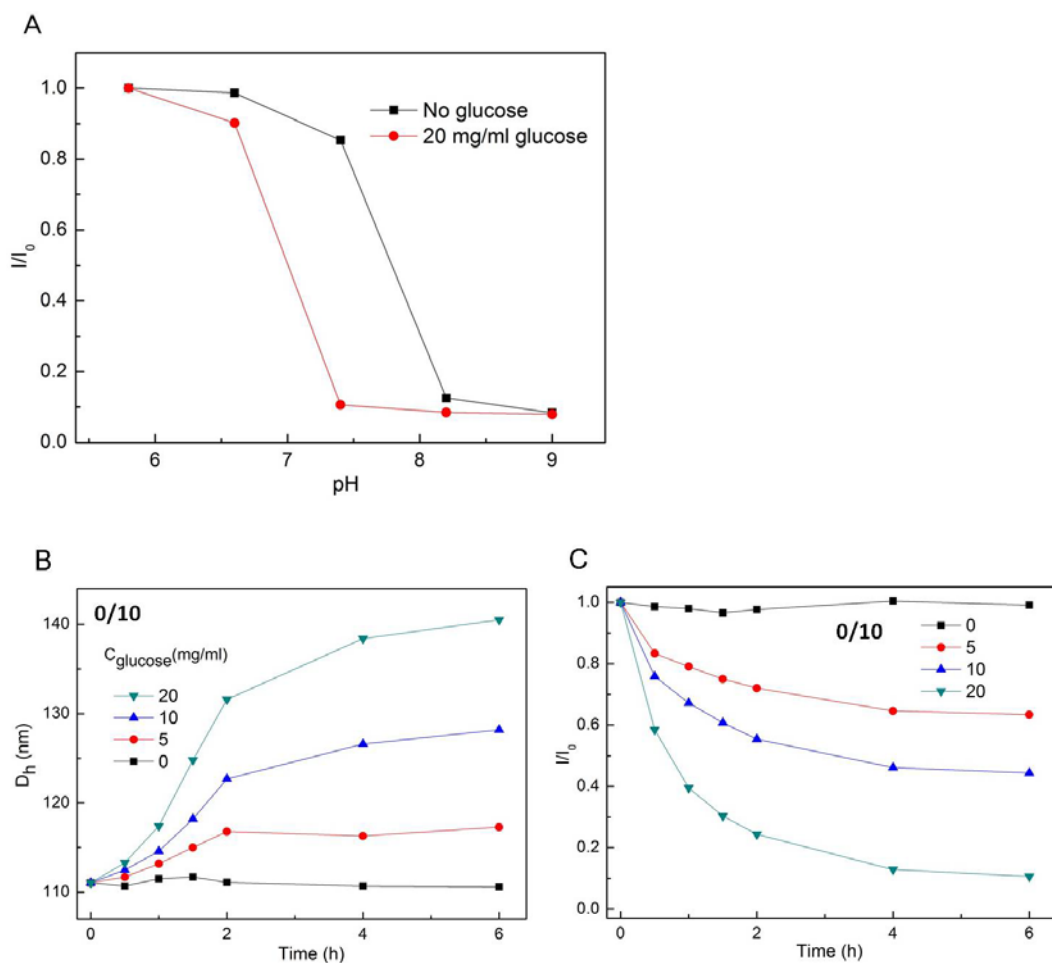
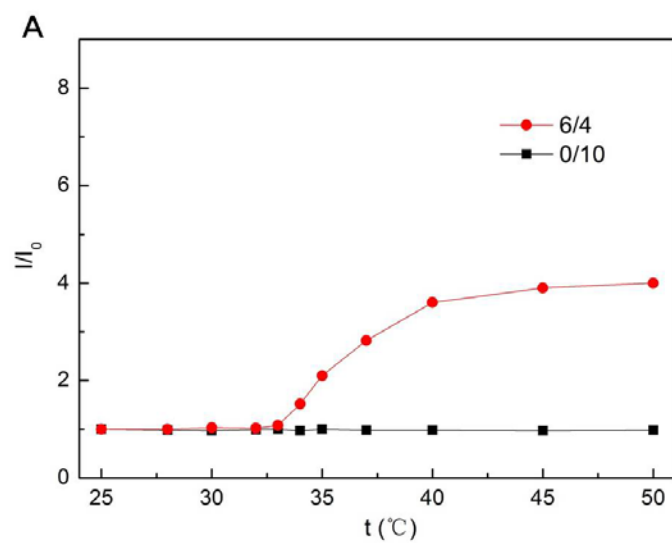


Figure S3. Glucose-responsiveness study of 0/10 PM. (A) pH-dependent Normalized light scattering intensity change with and without addition of glucose solution. Time-dependent hydrodynamic diameter (B) and normalized scattered intensity (C) changes at pH 7.4 and 37 °C under various glucose concentrations, where the scattering angle is 90°.

Table S1. DLS and SLS measurements of PMs with different $W_{\text{PNIPAM}}/W_{\text{PEG}}$ at 25 and 37 °C. *a* D_h represents the apparent average hydrodynamic diameter measured at the scattering angle of 90°.

$W_{\text{PNIPAM}}/W_{\text{PEG}}$	t (°C)	D_h^a (nm)	R_h (nm)	R_g (nm)	R_g/R_h
0/10	25	111.5	65.9	58.0	0.88
	37	111.0	65.8	58.0	0.88
2/8	25	102.4	60.3	53.7	0.89
	37	97.6	58.2	49.5	0.85
4/6	25	109.8	63.1	56.2	0.89
	37	102.9	59.8	48.4	0.81
6/4	25	111.6	62.5	55.6	0.89
	37	99.4	57.6	44.9	0.78
8/2	25	105.4	61.4	55.3	0.90
	37	724.2	-	-	-



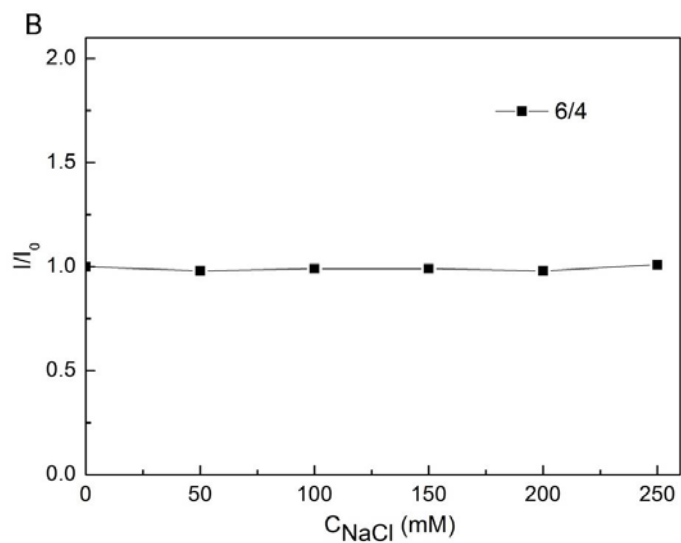
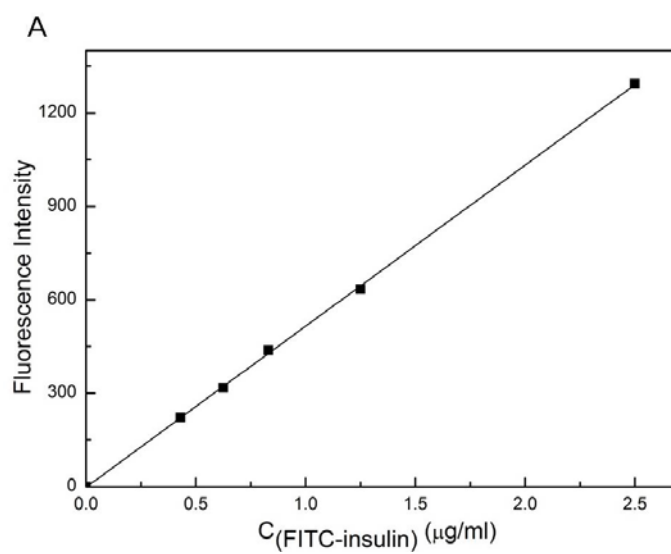


Figure S4. Structure characterization of PMs. (A) Temperature-dependent normalized scattered intensity of 0/10 and 6/4 PMs. (B) salt concentration-dependent normalized scattered intensity of 6/4 PMs.



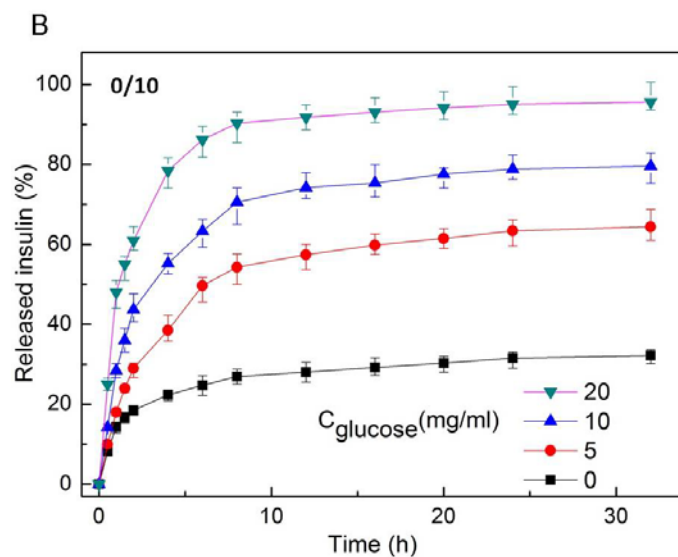


Figure S5. (A) Fluorescence intensity at various concentrations of FITC-insulin. (B) Release profiles of FITC-insulin from 0/10 PMs upon varying concentrations of glucose.

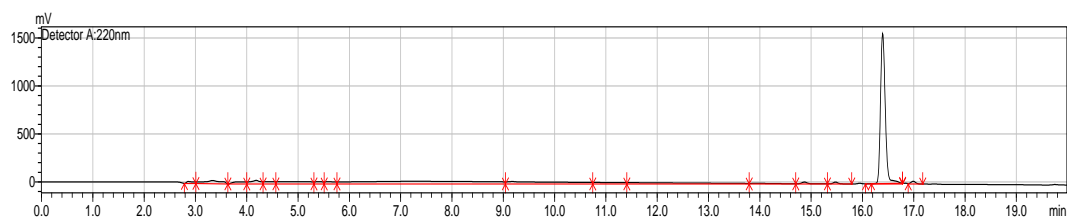


Figure S6. HPLC profile of insulin at 35 °C (mixed solvent: acetonitrile and 0.1% trifluoroacetic acid)

References:

- 1 Z. Yang, S. Y. Zheng, W. J. Harrison, J. Harder, X. X. Wen, J. G. Gelovani, A. Qiao and C. Li, *Biomacromolecules*, 2007, **8**, 3422-3428.