



Scholars Research Library

Der Pharmacia Lettre, 2012, 4 (3):919-926
(<http://scholarsresearchlibrary.com/archive.html>)



Pathological observations on the treatment of oral sub mucous fibrosis of curcumin gels in animal models

N. Sanjeev Kumar¹, D. Vijaybhaskar^{2*}, K. Purushotham Rao² and S. Pratima¹

¹Dept. of Pathology, M. R. Medical College, Gulbarga (Karnataka) – India

²Dept. of Pharmaceutics, H. K. E. S's College of Pharmacy, Gulbarga (Karnataka) – India

ABSTRACT

Mucoadhesive gels were prepared for the treatment of oral sub mucous fibrosis, which provide effect for extended periods of time. Stress was given for improvised local action of the drug with the addition of mucoadhesive polymer in the formulation. Curcumin was taken as a model drug as it exhibits profound antitumeric & antimutogenic activity. The semisolid preparations comprised of stabilizer like sodium metabisulphite, muco retention / mucoadhesive polymers like HEC, NaCMC and equal mixture of HEC & NaCMC, and were subjected for various physicochemical parameters like pH, spreadability, drug content uniformity, extrudability, viscosity & I.R. studies. In-vitro drug release studies were carried out in phosphate buffer (6.4 pH). Stability studies were also done at room temperature for a period of eight weeks. Among three formulations (formulation code F₁ containing equal mixture of NaCMC & HEC as base, formulation code F₂ containing NaCMC as base and formulation code F₃ containing HEC as base), the formulation containing equal mixture of NaCMC & HEC as base showed good in-vitro release and good adhesion to oral mucosae. IR studies showed that there was no drug-excipient interaction. The in-vivo studies were carried out in two phases using 18 mice with the permission of ethical committee under the supervision and help of staff, Department of Pathology, M.R. Medical College, Gulbarga. In first phase oral sub mucous fibrosis was induced in mice using marketed Gutkha preparation and formulating into a mucoadhesive gel form and applying to mice oral mucosa with the help of cotton bud for a period of 6 months. In second phase, treatment was carried out following the above method using curcumin formulation. The tissue samples collected for 1, 3 & 6 months induction period & 1, 3 & 6 months of treatment period on 6 months oral sub mucous fibrosis induced mice. Histopathological observations reported that there was considerable induction of oral sub mucous fibrosis and excellent treatment results on curcumin usage. The results of the present study of mucoadhesive semi-solid drug design for the treatment of oral sub mucous fibrosis will be useful for drug industry for the benefit of patients suffering from oral sub mucous fibrosis.

Key Words: Mucoadhesive, semi solid preparations, curcumin, oral sub mucous fibrosis.

INTRODUCTION

Oral submucous fibrosis is a condition reported mainly from India and is seen in 33% to 40% of patients with oral cancer. In early stages vesicles or fibrous bands are present on the labial mucosa associated with pigment changes. In later stages, the mucosa become stiff, causing difficulty in opening the mouth. Histologically, the mucosa varies from atrophic to normal. A characteristic feature is a prominent sub epithelial eosinophilic band. The juxtaepithelial connective tissue is amorphous and non bundular as against the normal undulated bundular collagen [1,2,3]. The main cause for Oral submucous fibrosis are chewables like gutkha, tobacco, pan masalas, areca nut [4]. A thorough literature survey has been carried out on the proposed topic and most prominent references found are Hastak K, Lubri N, Jakhi SD, More C, John A, Bhaishasa SD, Bhide SV, (1997), studied the effect of turmeric oil and

turmeric oleoresin on cytogenetic damage in patients suffering from OSMF. *In vitro* studies on the effect of alcoholic extracts of turmeric, turmeric oil and turmeric oleoresin, on the incidence of micronuclei in lymphocytes from normal healthy subjects showed that the test compounds did not cause any increase in the number of micronuclei as compared with those found in untreated controls [5]. Katharia S K, Singh S P, K Kulshrcshtha V K studied the effect of placenta extract in management of Oral submucous fibrosis and stated that there was significant improvement in mouth opening, colour of oral mucosa and reduction of fibrous bands [6]. Krishna Prasad N S, Sarasija Suresh developed a simple and easy method of estimation of curcumin based on the solubility of curcumin in methanol [7]. Sarasija Suresh, Shobha Rani R Hiremath, Praveen S, Aney Thomas prepared and characterized curcumin gels using a bioadhesive polymer like pluronic F-127 for local application as topical therapeutic system [8]. Dr. Paranjothy K L K, Dr. Thampi formulated zinc sulphate gel using a Sodium Carboxy Methyl Guar as oral gel for mouth ulcers [9]. Pandey S, Pai M, Singh U V, Udupa N prepared huccal mucoadhesive films and mucoadhesive gels of captopril using Hydroxy propyl methyl cellulose, ethyl cellulose and carbopol. The drug release pattern was higher with formulations containing carbopol [10], Uma Devis, Gancsh M, Mohanta G P & Manavalan R designed and evaluated tetracycline hydro chloride gels. The tetracycline gels formulated with hydroxy propyl methyl cellulose and carbopol showed increase in drug release with increase in polymer concentration [11].

There is no effective treatment for OSMF and there is a need for drug research for its treatment in any type of dosage form. In the present study an attempt has been made to develop mucoadhesive semisolid preparations of Curcumin for oral application directly on to the inflamed site to produce local action, using mucoadhesive hydrophilic polymers like hydroxy ethyl cellulose (HEC), Sodium carboxy methyl cellulose (NaCMC) [12].

MATERIALS AND METHODS

Curcumin was procured from Sami Labs, Bangalore, Glycerin from Ranbaxi Lab Ltd. Chandigarh, Sodium carboxy methyl cellulose and Hydroxy ethyl cellulose were purchased from S. D. Fine Chem. Ltd., Mumbai, Sodium meta bisulphite and methanol from Qualigens Mumbai.

Preparation of formulation:

Three semisolid formulations were prepared consisting of Sodium carboxy methyl cellulose, hydroxy ethyl cellulose and equal mixture of Sodium carboxy methyl cellulose and hydroxy ethyl cellulose as polymers. Prehydrated polymer samples for 12 hours were dissolved in 85 ml of distill water on constant stirring for about one hour. Then added 15 ml of ethanolic curcumin solution. On continues stirring, glycerine, sodium meta bisulphite were dissolved in the above Polymer-Drug solution (Tab 1).

Evaluation of physicochemical parameters:

The prepared formulation were subjected for various physicochemical parameters such as spreadability, extrudability, pH, viscosity, Mucoadhesive, drug content estimation .

Spreadability:

Spreadability was determined by an apparatus suggested by Muttimer et al., which was suitability modified in the laboratory and used for the study. It consisted of a wooden block which was provided by a pulley at one end (Tab 2).

Extrudability:

The formulation under study was filled in a clean, lacquered [13,14] aluminum collapsible one-ounce tube with a nasal tip of 5 mm opening extrudability was then determined by measuring the amount of ointment, cream and gels extruded through the tip when a constant load of 1 Kg. was placed on the pan were collected and weighed. The percentage of ointment, cream and gel extruded was calculated: recorded and grades were allotted (+++ Good; ++ Fair; + Poor) (Tab-2).

Determination of Viscosity:

All the products formulated in the semi-solid form were subjected to viscosity studies. Instrument used to measure viscosity is Brookfield digital viscometer (Tab-2).

Determination of pH:

Weigh accurately 5 ± 0.1 gm. of the cream in a 100 ml. beaker, add 45 ml. of water and dispersed the cream in it. Determine the pH of the suspension at 27°C using the pH meter (Tab-2).

Determination of Drug Content Uniformity:

Drug content uniformity was carried out by taking 5 gm sample of prepared formulation and subjected for analytical assay to calculate the drug present in the sample using UV spectrophotometer at λ_{\max} 430 nm. The drug content was uniform in all formulations (Tab-2).

Mucoadhesive Studies:

The glass plates are coated with the polymer and suspended from a microbalance. The glass plate is immersed in a temperature controlled mucous solution. The force required to pull the plate out of solution is determined under constant experimental conditions. A number of methods use liquid adhesive mass for evaluation. Duration of mucosal adhesion i.e., the time span required until the adhesive patch completely loses its adhesive contact with the mucosa was measured. The results are given in the (Tab-2).

Drug polymer interaction studies:

The studies were carried out using IR method with the help of Perkin-elmer 1615 spectrophotometer to check the possible drug polymer interaction (fig-1).

Evaluation of Drug Release:

Release of the curcumin from various semisolid preparation was studied by applying the permeation apparatus as directed by Fitter *et al*.

In-vivo Studies:

The *in-vivo* studies were carried out in mice with the permission of ethical committee under the supervision and help of staff, Department of Pathology, M. R. Medical College, Gulbarga.

The *in-vivo* studies were carried out in two phases using 18 mice.

- ❖ Induction of OSMF in animals for a period of six months.
- ❖ Treatment of OSMF on the induced animals.

Induction of OSMF:

18 (eighteen) Swiss male albino mice weighing 25 - 30 gms were selected for the experimental design in the present work OSMF was induced with the causative ingredients of marketed brands of gutkhas. The gutkha powder was pulverized with the help of mortar and pestle and passed through Sieve No. 200. Mucoadhesive gel formulations containing 1% gutkha powder prepared in the laboratory were applied with the help of cotton bud on to the buccal mucosa of the animals for a period of 6 months. During the induction period the animals were without water and food for a period of 6 hours and other times with regular food and water. To study the effect of induction a punch biopsy technique was used by sacrificing the animals using skin punch biopsy forceps (No. 5). The biopsy sample of buccal mucosa collected in normal saline vials of 3 animals was subjected for histopathological slide preparation and study of observation. The similar procedure was followed to check the induction after 3 months and 6 months. A biopsy sample of buccal mucosa of 3 healthy animals, were collected and set aside for comparative purpose.

Ingredients of Gutkha:

Betelnuts, Catechu, Lime, Cardamom, Menthol, Tobacco, Natural perfumes, Sandal oil species & flavours.

Formula used to prepare muco adhesive semi solid preparation gutkha of marketed product

Ingredients	Quantity
Gutkha (Sieve No. 200)	1.0 gms.
Polymer	4 gms
Glycerine	2.0 gms.
Sodium Meta bisulphite	0.5 gms.
Distilled water q.s. (ml.)	100 gms.

Procedure details:

85 ml of distilled water was taken in 250 ml glass beaker. Then add the polymer, glycerine and the preservative (Sodium metabisulphite) and mix with a glass rod. Cover the beaker with a glass plate and keep aside for 24 hrs. For hydration of the polymer. Then add gutkha powder to 15 ml of water. This solution was added slowly to the hydrated base and mixed using a Remi stirrer at 100 rpm.

Treatment:

After six (6) months of induction study. The remaining nine animals were tested for the purpose of treatment of OSMF, 1% curcumin muco adhesive gel prepared in the laboratory was used. The curcumin gel was applied on to the oral cavity of buccal mucosa in mice with the help of cotton bud and the procedure followed for application as

used in induction method. For histopathological observations of treatment, the biopsy samples were collected on 3 animals each after 1 month, 3 months & 6 months. Unlike in induction process, the slides of smears of the biopsy sample were processed for comparative evaluation of treatment.

Stability studies:

The formulations were then packed in the collapsible tube and stored at room temperature for 8 weeks and studied for viz., spreadability, extrudability, pH, drug content, viscosity (Tab-3).

RESULTS AND DISCUSSION

The gels were subjected to physical evaluations such as viscosity, extrudability, spreadability, pH, drug content uniformly and results are shown in Table-2. During our physico-chemical evaluation studies all the formulations were within pH range. Drug content estimation, drug present in formulation F₁, F₂ & F₃ were found to be 99.87, 99.63, 99.27. The formulation F₁ (containing equal mixture of sodium carboxy methyl cellulose & Hydroxy ethyl cellulose as base) showed good mucoadhesion for 32 minutes and formulation F₂ (containing sodium carboxy methyl cellulose as base) showed poor mucoadhesion for 26 minutes. Mucoadhesive curcumin gels were evaluated for drug polymer interaction by infrared spectral studies. After comparing the spectra i.e., absorption bands of pure drug with the spectra of formulations, the absorption bands of the pure drug were retaining in the formulations without undergoing any interaction with the polymers. *In-vitro* drug release from curcumin gels was studied and as per *in-vitro* data obtained in F₁, F₂ and F₃ at the end of 120 min, percent cumulative drug release was 25.31%, 24.21% and 22.56% respectively. In our present investigation of stability studies, all formulations did not segregate, ferment or physically deteriorate during normal condition of storage and use (Tab-3). The three formulations were planned for *in-vivo* studies using mice as model animal. The present study tries to focus the array of histomorphological changes in oral mucosa of albino mice after oral application of gutkha and histomorphological changes in already OSMF induced albino mice after oral application of curcumin and to see whether curcumin can heal OSMF [15]. In first phase of histopathological studies of OSMF induction in mice there was gross change of mucosa is observed and increased significance seen the use of gutkha gel from 1 month application to 6 months applications. In second phase of treatment part of OSMF using prepared curcumin semi-solid preparation and encouraging results were observed. There is a marked reduction (more than 50%) of OSMF seen from the histopathological studies on the specimen samples taken after 1 month, 3 month and 6 months (fig 2). The results of the present study of mucoadhesive semi-solid drug design for the treatment of OSMF will be useful for drug industry for the benefit of patients suffering from OSMF.

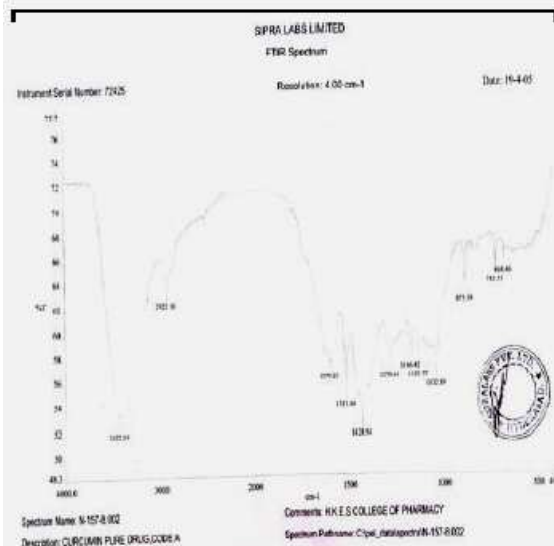
Table-1: Formulae used to prepare mucoadhesive gels

Sl. No.	Ingredients	Formulation F1 (NaCMC+HEC)	Formulation F2 (NaCMC)	Formulation F3 (HEC)
1.	Curcumin	1.0 gm	1.0 gm	1.0 gm
2.	HEC	2.0 gms	-	4.0 gms
3.	NaCMC	2.0 gms	4.0 gms	-
4.	Glycerine	2.0 gms	2.0 gms	2.0 gms
5.	Sodium meta bisulphite	0.5 gms	0.5 gms	0.5 gms
6.	Ethanol	15 ml.	15 ml.	15 ml.
7.	Distilled water q.s. (ml)	100 gms	100 gms	100 gms

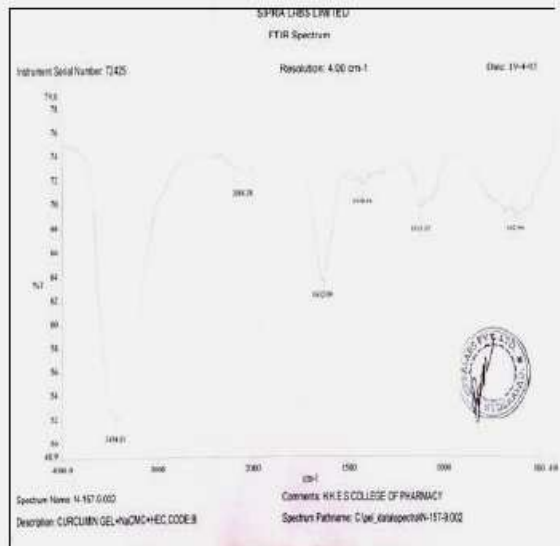
Table-2: Characterization of prepared formulations

Sl. No.	Formulation No.	Spreadability (Sec.)	Extrudability	Viscosity (CPS)	pH	Drug Content (%)	Duration of Mucosal Adhesion (min.)
1.	F ₁	14.12	+++	2.6 x 10 ⁵	6.7	99.87	32
2.	F ₂	22.86	+++	2.9 x 10 ⁵	6.5	99.63	26
3.	F ₃	24.00	++	3.2 x 10 ⁵	6.9	99.27	30

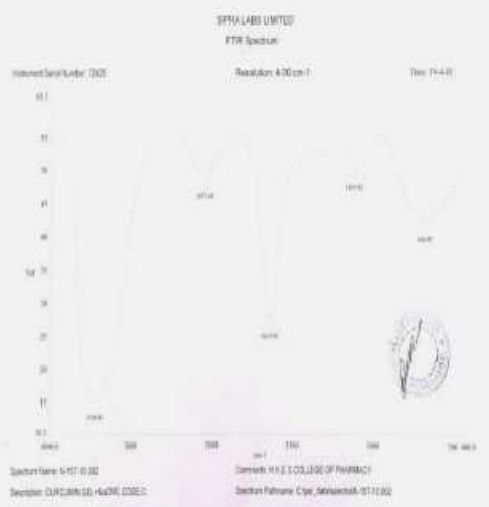
FIGURE-1 INFRARED SPECTRA OF MUCOADHESIVE CURCUMIN ANTI TOBACCO SEMISOLID FORMULATIONS AND PURE DRUG



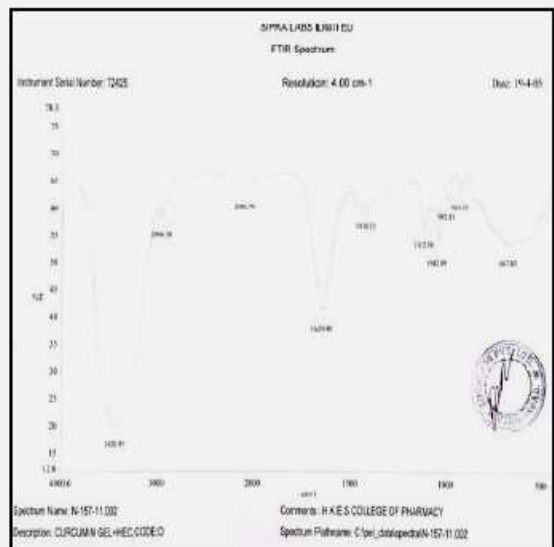
I.R. Spectra of Pure Drug



I.R. Spectra of formulation F1 containing equal mixture of Sodium carboxy methyl cellulose & Hydroxy ethyl cellulose as base and curcumin

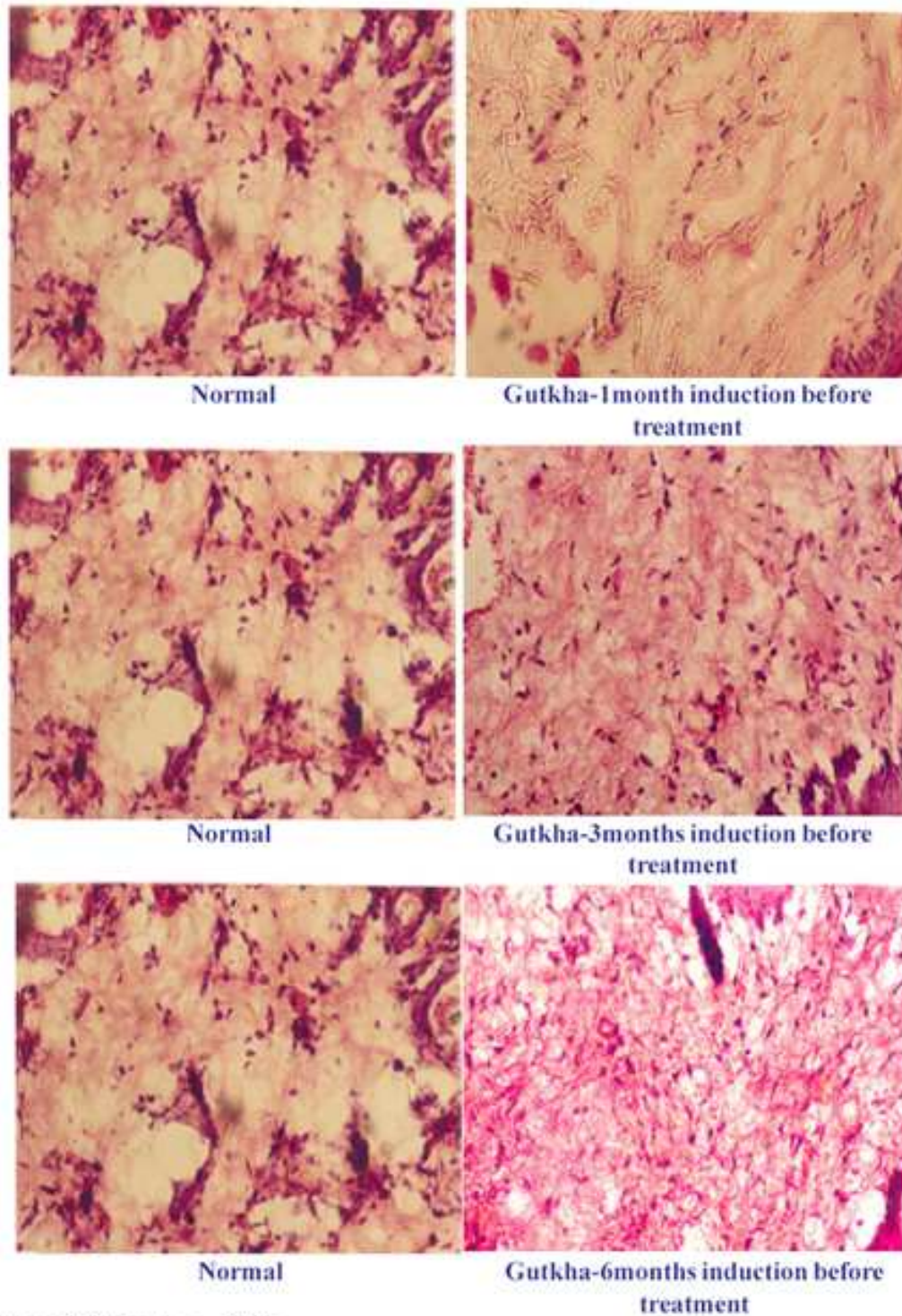


I.R. Spectra of formulation F2 containing Sodium carboxy methyl cellulose as base and curcumin



I.R. Spectra of formulation F3 containing Hydroxy ethyl cellulose as base and curcumin

Fig – 2: Section of oral mucosa of mice after induction of OSMF with Gutkha at different periods of time



Magnification = 200x
Stain used = Haematoxylin eosin

Fig – 3: Section of Oral mucosa of mice after treatment of OSMF with turmeric at different periods of time

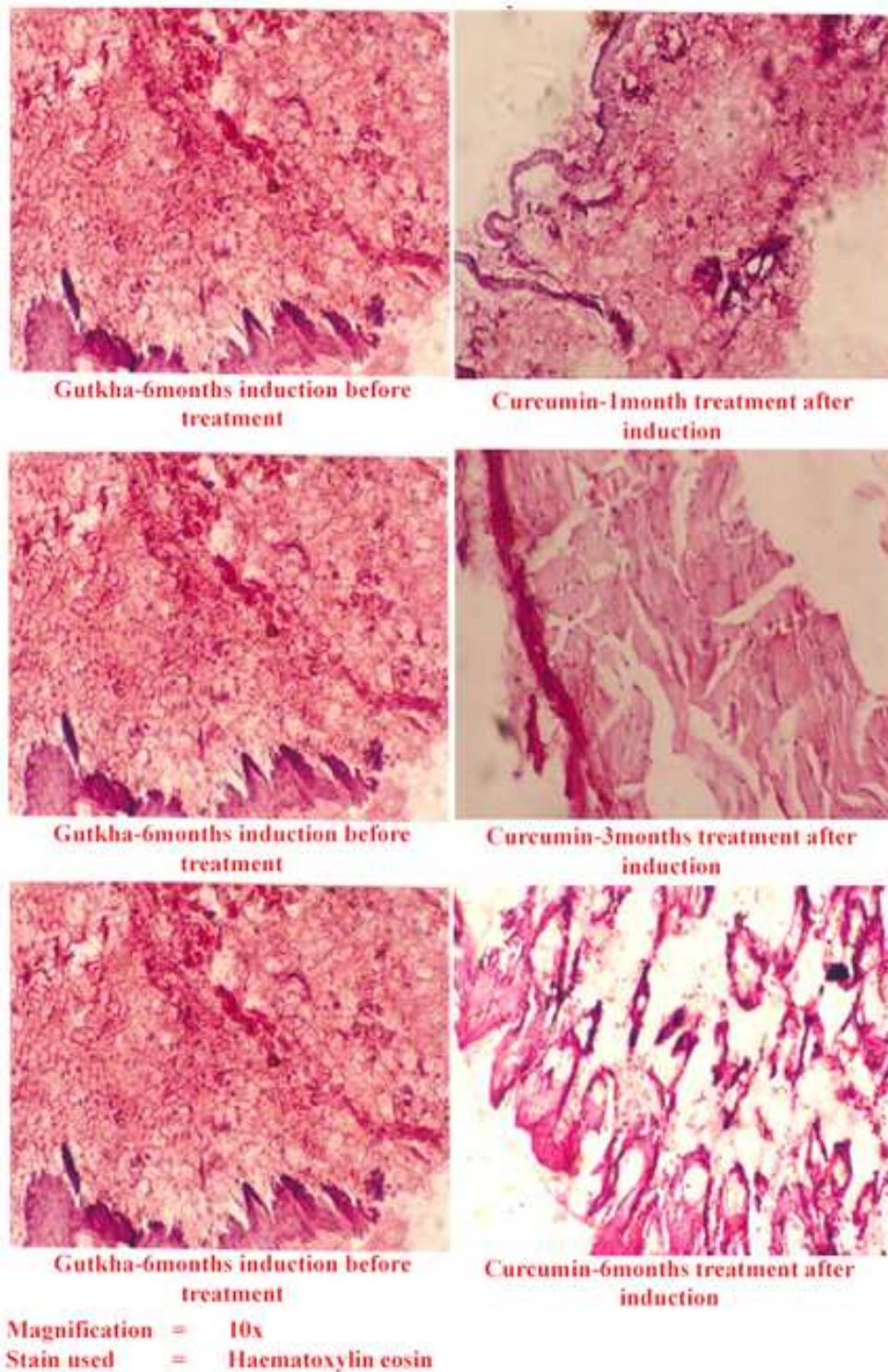


Table-3: Stability studies data of F₁, F₂ and F₃

Storage Temp.	Time of Analysis	Spreadability (Sec.)			Extrudability			pH			Viscosity (CPS)			Drug Content (%)		
		F1	F2	F3	F1	F2	F3	F1	F2	F3	F1	F2	F3	F1	F2	F3
Room Temperature	1 st Week	14.12	22.86	24.00	+++	+++	++	6.7	6.5	6.9	2.6 x 10 ⁵	2.9 x 10 ⁵	3.2 x 10 ⁵	99.89	99.63	99.27
	2 nd Week	14.12	22.86	24.00	+++	+++	++	6.7	6.5	6.8	2.6 x 10 ⁵	2.9 x 10 ⁵	3.1 x 10 ⁵	99.89	99.63	99.26
	3 rd Week	14.11	22.85	23.98	+++	+++	++	6.6	6.5	6.8	2.5 x 10 ⁵	2.9 x 10 ⁵	3.2 x 10 ⁵	99.88	99.62	99.25
	4 th Week	14.11	22.84	23.97	+++	+++	++	6.5	6.5	6.9	2.5 x 10 ⁵	2.9 x 10 ⁵	3.2 x 10 ⁵	99.88	99.63	99.26
	5 th Week	14.11	22.85	23.98	+++	+++	++	6.6	6.4	6.8	2.4 x 10 ⁵	2.8 x 10 ⁵	3.1 x 10 ⁵	99.88	99.62	99.27
	6 th Week	14.12	22.85	23.98	+++	+++	++	6.6	6.4	6.9	2.4 x 10 ⁵	2.8 x 10 ⁵	3.1 x 10 ⁵	99.87	99.62	99.26
	7 th Week	14.10	22.84	23.98	++	++	++	6.5	6.4	6.8	2.5 x 10 ⁵	2.8 x 10 ⁵	3.1 x 10 ⁵	99.88	99.63	99.27
	8 th Week	14.10	22.84	23.98	++	++	++	6.6	6.4	6.8	2.5 x 10 ⁵	2.8 x 10 ⁵	3.1 x 10 ⁵	99.87	99.62	99.25

Acknowledgements

The authors are thankful to principal, H. K. E. College of Pharmacy, Gulbarga for providing necessary laboratory facilities to carry out this work with great ease and precision and also thanks to SAMI Labs, Bangalore for supplying Curcumin as gift sample for the research work and also thanks to Sipra Lab, Hyderabad for providing facilities of IR spectral analysis for prepared formulations.

REFERENCES

- [1] Harsh Mohan, "The aetiology of Oral sub mucous fibrosis" "A text book of pathology" 4th edition, J P Publication, 2000; New Delhi, 325 pp.
- [2] Bailey & Love's, "The aetiology of Oral sub mucous fibrosis" "Short practice of surgery" 2nd edition, Arnold Publishers, 2000; 639 pp.
- [3] Canniff J P Harvey, *Int. J of Oral surgery*, 1981, 10 (1), 163-167 pp
- [4] Halliday, Janet Anne, Robertson, Steven, "Oral transmucosal delivery", Controlled Therapeutics (Scotland) Ltd., 2002, www.uspto.gov/patft/index.html
- [5] Katharia S K, Singh S P, Kulshreshtha V K, *Ind. J of Pharmacology*, 1992, 24(6), 181-183 pp.
- [6] Krishna Prasad NS, Sarasija Suresh, *Indian Drugs*, 1997, 34(4), 227-228pp.
- [7] Sarasija Suresh, Shobha Rani R Hiremath, Praveen S, Aney Thomas, *Indian Drugs*, 1999, 36(5), 326-328pp.
- [8] Dr. Paranjothy K L K, D Thampi, *Indian Drugs*, 1995, 32(4), 186-188pp.
- [9] Pandey S, Pai M, Singh U V, Udupa N, *Pharmag*, 2001, 13(1,2), 12-15pp.
- [10] Uma Devi, Ganesh M, Mohanta G P & Manavalan R, *Indian Drugs*, 2002, 39(10), 552-553pp.
- [11] Hastak K, Lubri N, Jakhi SD, More C, John A, Bhaisasa SD, Bhide SV, *Cancer-Lett*, 1997, 116(2), 265-269.
- [12] Sethi PD, "Qualitative analysis of drug in pharmaceutical formulation", CSBC Publishing, 201, 220, 221pp.
- [13] Rameshacharya Bala R, Vani G. & Rao Madhusudhan Y, *Drug development & industrial pharmacy*, 1999, 25 (5), 685-690pp.
- [14] Huang S, Ling T, Wu H, *Carcinogenesis*, 1997, 15(2), 91-96pp.
- [15] Culling FA, "A book of Histopathological & Histochemical techniques", Butter work & co. publication, Great Britain, 3rd edn. 1975, 5-50pp.