AMA0076, a Novel, Locally Acting Rho Kinase Inhibitor, Potently Lowers Intraocular Pressure in New Zealand White Rabbits with Minimal Hyperemia

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PURPOSE. To determine whether ROCK inhibition for the treatment of glaucoma can be improved by using novel, locally acting Rho kinase (ROCK) inhibitors, such as AMA0076, that lower IOP without inducing hyperemia.

METHODS. On-target potency of AMA0076 was compared with other ROCK inhibitors (Y-27632 and Y-39983) and conversion of AMA0076 into its functionally inactive metabolite was evaluated in rabbit eye tissues. Human trabecular meshwork (HTM) cell morphology, actin filaments, and focal adhesion were studied in vitro after exposure to AMA0076. The effect of AMA0076 on IOP was investigated in normotensive rabbits and a new, acute hypertensive rabbit model. Intraocular pressure lowering efficacy of AMA0076 was compared with pharmacologic treatments. Hyperemia after single topical dosing of AMA0076 and Y-39983 was scored.

RESULTS. AMA0076 and Y-39983 showed similar on-target potency. AMA0076 was most stable in aqueous humor and converted into its metabolite in other eye tissues. Exposure of HTM cells to AMA0076 led to significant and reversible changes in cell shape and a decrease in actin stress fibers and focal adhesions. Both AMA0076 and Y-39983 provided an equivalent IOP control. Compared with latanoprost and bimatoprost, AMA0076 was more potent in preventing the IOP elevation in the acute hypertensive rabbit model. The degree of hyperemia was significantly lower in rabbits treated with AMA0076 then with Y-39983.

CONCLUSIONS. AMA0076 is a locally acting ROCK inhibitor that is able to induce altered cellular behavior of HTM cells. Administration of AMA0076 effectively reduces IOP in ocular normotensive and acute hypertensive rabbits without causing distinct hyperemia.

Keywords: ROCK inhibitors, IOP lowering, glaucoma
trabecular meshwork (TM) and as such increase conventional outflow facility. Preclinical data indicate that ROCK inhibitors such as Y-27632 (trans-4-[1R]-1-aminoethoxy-N-4-pyridin-1-yl cyclohexanecarboxamido dihydrochloride), Y-39985 (4-[(1R)-1-aminoethoxy]-N-[1H]-pyrrolo[2,3-b]pyridin-4-ylbenzamide hydrochloride), and Fasudil (1-(5-isouquinolinylsulfonyl)-homopiperazine dihydrochloride) alter the contractility of the TM cells, thereby lowering IOP by facilitating conventional outflow via the relaxation of cells in the TM. However, preclinical and clinical studies evaluating ROCK inhibitors have reported mild to severe conjunctival hyperemia, which is most likely due to mechanism-based smooth muscle cell relaxation in conjunctival blood vessels. As a result, most ROCK inhibitors display a narrow therapeutic window. Therefore, Amakem Ophthalmics (Diepenbeek, Belgium) developed a product pipeline of soft ROCK inhibitors. In general, soft drugs, also known as antedrugs or locally acting drugs, are biologically active compounds that are designed to undergo metabolic inactivation by controlled conversion of the parent molecule into a predictable, nontoxic metabolite. AMA0076 is a novel, potent ROCK inhibitor resulting from Amakem Ophthalmics efforts toward the development of ROCK inhibitors such as Y-27632 and Y-27632, using a radiometric method. The on-target activity of AMA0076 was determined and compared with Y-39983 and Y-27632, using a radiometric protein kinase assay (1 μM ATP). The kinetics of ROCKII inhibition by AMA0076 were characterized by varying the concentration and compared with the known ROCK inhibitor, Y-39983. Moreover hyperemia was scored after single topical administration and compared with the known ROCK inhibitor, Y-39983.

**Materials and Methods**

**In Vitro On-Target Activity Measurements**

The on-target activity of AMA0076 was determined and compared with Y-39983 and Y-27632, using a radiometric protein kinase assay (1 μM ATP). The kinetics of ROCKII inhibition by AMA0076 were characterized by varying the concentrations of ROCKII (aa5-554), peptide substrate (long S6 kinase substrate peptide) and ATP, and measuring inhibition at concentrations of ROCKII. The on-target activity of AMA0076 was characterized by varying the concentration and compared with the known ROCK inhibitor, Y-39983.

**Animals**

Male New Zealand White (NZW) rabbits (animal facility, KU Leuven, Leuven, Belgium) aged 12 to 14 weeks and weighing 2 to 3 kg, were used in all studies. All animals were conducted in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research and were approved by the ethical committee for the use of animals at KU Leuven.

**Stability in Eye Tissues**

Eye tissues (cornea, conjunctiva, sclera, and AH) were collected from (NZW) rabbits (animal facility, KU Leuven), snap frozen and stored at –80°C. On the day of the experiments, tissues were divided into equal pieces and weighted. Next, all samples were immersed into freshly prepared glutathione bicarbonate Ringer’s (GBR) buffer oxygenated with O2/CO2 (95:5) to pH 7.4, and were spiked with AMA0076 to a final concentration of 20 μM. After 4-hours incubation at 37°C samples were taken at fixed time points and put into ice-cold ACN/H2O (Merck, Darmstadt, Germany) containing the internal standard warfarin (Fluka; Sigma-Aldrich, Steinheim, Germany), for protein precipitation. The remnant of compound was determined by liquid chromatography-tandem mass spectrometry (LC/MS-MS) using Analyst Software (AB Sciex, Nieuwerkerk aan den IJssel, The Netherlands). The LC/MS-MS apparatus consists of two parts: the LC part (LC20AD; Shimadzu, Duisburg, Germany) and the MS-MS connected to the LC part (3200 Q TRAP LC/MS/MS system; AB Sciex, Applied Biosystems, Nieuwerkerk aan den IJssel, The Netherlands). A calibration curve of AMA0076 was made in GBR buffer to calculate the remaining concentration. Based on the data and the weights of the tissues of at least three independent experiments, the rate of metabolism was calculated, assuming first order decay, and specific enzymatic activity was presented as picomole per minute per milligram ± SEM.

**Culture of Human Trabecular Meshwork (HTM) Cells**

Human TM cells were obtained from ScienCell Research Laboratories (Carlsbad, CA) and cultured in poly-L-lysine coated T75 flasks (ScienCell Research Laboratories). Cells were maintained at 37°C in a humidified atmosphere of 5% CO2. Medium was replaced every 2 days and once confluent, cells were passaged in a one-fourth ratio using the trypsin-EDTA method. Briefly, cells were incubated with 0.25% trypsin-EDTA (Gibco, Carlsbad, CA) for 5 minutes at 37°C and 5% CO2. Afterwards, cells were centrifuged for 5 minutes at 312 g and plated. Cells between the third and sixth passage were used in all experiments.

**Effect on Human TM Cell Morphology**

Confluent cultures of human TM cells were incubated with AMA0076 1 μM or dimethyl sulfoxide (DMSO) at similar concentration as vehicle. Cultures were observed with an inverted microscope (Primo Vert Zeiss, Oberkochen, Germany) and photographed before application and 60 minutes after administration of AMA0076. The solution was replaced with full medium and incubated again for 15 hours to examine whether the effects of AMA0076 on human TM cells were reversible.

**Actin and Vinculin Staining**

Human TM cells were plated on cover glasses in a 12-well plate at a density of 20 × 10^5 cells per well. When cells reached semi confluence, 1 μM of AMA0076 was added and DMSO was used as vehicle. One hour after drug exposure, cells were fixed with 1% paraformaldehyde (PFA) for 20 minutes at room temperature (RT). F-actin and focal adhesions were stained using the FAK 100 actin cytoskeleton/focal adhesion staining kit (Millipore, Billerica, MA) according to the following protocol. Briefly, cells were permeabilized using 0.1% Triton X-100 in PBS for 5 minutes at RT and rinsed with PBS. Blocking solution (1% bovine serum albumin; Roche Diagnostics GmbH, Mannheim, Germany) was applied for 30 minutes. The cells were incubated with antivinculin (focal adhesions; 1:300 in blocking solution) at RT for 1 hour and rinsed with PBS. Next, cells were...
incubated at RT for 1 hour with AlexaFluor 488-conjugated goat anti-mouse (1:200) and tetramethylrhodamine-5-(and-6)-isothiocyanate (TRITC)-phalloidin (F-actin, 1:100), both diluted in PBS. Cells were washed and mounted with Prolong Gold antifade reagent with 4′, 6-diamino-2-phenylindole (DAPI; Invitrogen, Carlsbad, CA). Fluorescence was visualized with a confocal laser scanning microscope (FV 1000; Olympus, Melville, NY). To determine whether the effects of AMA0076 on human TM cells were reversible, the drug solution was replaced with full medium and followed by incubation for another 15 hours.

IOP Measurements
An application tonometer (Tono-Pen; Medtronic Solan, Jacksonville, FL) was used to monitor IOP and three recordings per eye were averaged. Before the start of each IOP experiment the OCU-FILM tip cover was renewed and the tonometer was calibrated. The rabbit eyes were anesthetized by topical instillation of 0.4% oxybuprocaine (Unicaaine; Théa Pharma, Schaffhausen, Switzerland) before every IOP measurement.

Ocular Normotensive NZW Rabbits
In a first set of experiments the IOP-lowering efficacy of single topical administration of AMA0076 at concentrations of 0.1%, 0.3%, and 0.5% was tested in ocular normotensive NZW rabbits. These results were compared with Y-39983 (0.1%, 0.3%, and 0.5%) and to the PGA latanoprost (Xalatan 0.005%; Pfizer, New York, NY). The contralateral eyes of the AMA0076/Y-39983-treated rabbits received vehicle (H2O/ polyethylene glycol [PEG]). Intraocular pressure measurements were performed immediately before and every hour until 8 hours after single administration of the compounds. Latanoprost was administered at two different time points in two separate experiments. In the first experiment, latanoprost was administered the evening before IOP measurements. In a second experimental set-up, we also tested the effect of latanoprost when dosed in the morning on the day of IOP measurements (as AMA0076 and Y-39983). In both experiments contralateral eyes were treated with saline (0.9% NaCl; B. Braun Medical, Bethlehem, PA). In the above experiments all IOP measurements were performed between 8 AM and 5 PM. Furthermore, the effect of AMA0076 0.3% and latanoprost on IOP was investigated during the night. Compounds were administered topically three times at 0 (9 AM), 3 (12 AM) and 6 (3 PM) hours after Viscoat injection. Saline was administered to the contralateral eye of rabbits treated with latanoprost and timolol, whereas vehicle (H2O/PEG) was used as control in the AMA0076 treated rabbits.

In a second experiment, a head to head comparison between AMA0076 and the PGAs, latanoprost and bimatoprost (Lumigan 0.3 mg/mL; Allergan, Westport, Ireland) was performed. AMA0076 was instilled three times at the day of the experiment. Prostaglandin analogues were administered the evening before induction of hypertension, to have their maximal effect at the time of the IOP elevation challenge. Intraocular pressure was measured immediately before injection of Viscoat and every hour until 8 hours after inducing OH.

Hyperemia Scoring
New Zealand White rabbits were used to investigate the hyperemic effect after single dosing of AMA0076, Y-39983, and vehicle. Eyes were photographed using a digital camera (EOS 5D Mark II [100-mm lens]; Canon, Inc., Tokyo, Japan) before, 1, 4, and 8 hours after topical instillation of the compounds. Hyperemia was scored using a scoring system developed by Alcon Research, in which a number is given to the area of vasodilatation: 0 was normal appearance of vessels at limbus and on rectus muscle; 1 was enlargement of vessels normally visible at limbus and on rectus muscle; 2 was branching of vessels at limbus, new vessels are visible; 3 was new vessels visible in open bulbar conjunctival areas; and 4 was diffuse redness in open bulbar conjunctival areas. The numbers 0 and 1 indicate normal scores; 2, 3, and 4 represent adverse events.

Statistical Analysis
Data at individual time points were analyzed using the Student’s t-test for independent samples. The time course of IOP changes was analyzed using ANOVA for repeated measurements (GraphPad Prism 5.0; GraphPad Software, Inc., San Diego, CA). P values smaller than 0.05 were considered statistically significant. Data were presented as mean ± SEM.

RESULTS
AMA0076 is a Potent, Reversible, and ATP-Competitive ROCK Inhibitor
Since AMA0076 is a novel ROCK inhibitor, we first determined its on-target potency and compared it with other known ROCK inhibitors (Y-39983 and Y-27632). Analysis showed that AMA0076 is a potent inhibitor of ROCKI and ROCKII with similar potencies on both isoforms (Table 1). AMA0076 was equipotent to Y-39983 and approximately 30-fold more potent than Y-27632 (Table 1). As AMA0076 and Y-39983 showed similar on target potency for ROCKII, we used Y-39983 as a reference ROCK inhibitor for all in vivo studies.

Inhibition of ROCKII by AMA0076 was competitive with respect to ATP and non-competitive compared with long 86 kinase peptide substrate (Table 2). Inhibition was time-independent and decreased by dilution of the reaction mixture, suggesting reversible binding (data not shown).
AMA0076 differs from other ROCK inhibitors by its soft drug properties. The conversion of AMA0076 into its functionally inactive metabolite was evaluated in NZW rabbit eye tissues (cornea, conjunctiva, and sclera) and AH. The observed degradation rates were converted into specific activities for each tissue, in order to provide a suitable comparison (Table 3). Low specific activity (0.13 pmol min⁻¹ mg⁻¹) was found in the collected AH samples. Higher specific activities were found in all tested tissues. Cornea displayed the highest specific activity, followed by conjunctiva and sclera (4.81, 1.93, and 1.25 pmol min⁻¹ mg⁻¹, respectively). Overall, these values suggest that significant conversion of AMA0076 occurs in vivo in all three tissues, confirming its soft drug properties.

**AMA0076 is Stable in Aqueous Humor and Metabolized in Rabbit Eye Tissues**

Next, the effect of AMA0076 on TM cells was tested in vitro. It has been previously shown that ROCK inhibitors (Y-27632, Y-39983, and HA1077) induce morphologic changes in TM cells. Therefore, we exposed semiconfluent cultures of human TM cells to AMA0076. Treatment with AMA0076 induced cell–cell separation (retraction) and thinning of the cells after 60 minutes. Replacement of drug solution with full medium resulted in recovery of normal cell morphology (Fig. 1A). To examine whether these morphologic changes were due to cytoskeletal rearrangements, the effect of AMA0076 on the cytoskeleton was investigated by immunofluorescent staining. In control cells, actin filaments assembled into large radial bundles associated with focal adhesions (Fig. 1B, left panel). Exposure to AMA0076 induced distinct alterations in the actin filament organization and focal adhesions of human TM cells (Fig. 1B, middle panel). After treatment with AMA0076, cells lost most of their actin bundles and residual actin filaments became associated with the cell periphery rather than with focal adhesions. Exposure to AMA0076 caused a significant decrease in vinculin expression at the focal adhesion points. All these cytoskeletal changes returned to normal after compound replacement with full medium (Fig. 1B, right panel). There was no effect of the vehicle on general morphology or actin/vinculin staining of HTM cells (data not shown).

Taken together, these data indicate that ROCK inhibition by AMA0076 modifies TM cell morphology.

**Single Administration of AMA0076 Effectively Lowers IOP in Normotensive NZW Rabbits**

We then used an in vivo animal model to test the IOP-lowering efficacy of AMA0076. In normotensive NZW rabbits, a single topical administration of the compound significantly decreased IOP compared with control eyes in a dose-dependent manner (Figs. 2A–C). The IOP reduction was significant already 1 hour post dosing, and was sustained for 6 to 8 hours, after which IOP returned to baseline. The peak IOP-lowering efficacy was observed 2 hours post dosing for the 0.5% and 0.3% concentrations, and 4 hours post dosing for the 0.1% concentration. Maximum IOP reduction compared with baseline values after a single dose of AMA0076 at a concentration of 0.5%, 0.3%, and 0.1% was 48% ± 0.32 (P = 0.03), 39% ± 0.41 (P = 0.003), and 23% ± 0.17 (P = 0.0006), respectively (Fig. 2D). A single administration of Y-39983 also significantly lowered IOP compared with vehicle-treated eyes (Figs. 2E–G). Maximum IOP reduction was reached 2 hours after instillation with Y-39983 at 0.5%, 0.3%, and 0.1% (Fig. 2H). Compared with baseline values, IOP was reduced by 38% ± 0.55 (P = 0.01), 34% ± 0.34 (P = 0.01), and 27% ± 0.57 (P = 0.07), respectively. Noteworthy, an attenuated IOP-lowering effect was also observed in the contralateral eye of animals treated with Y-39983. This contralateral effect was not observed in the AMA0076 treated animals. Since this might lead to an underestimation of the IOP reduction by Y-39983, the maximum IOP-lowering effect in all experiments was expressed as compared with baseline.

Finally, we compared the IOP-lowering efficacy of AMA0076 to latanoprost. We tested the effect of latanoprost on IOP after two different time points of administration. First, latanoprost was administered the evening before IOP measurements because, as in humans, latanoprost acts as a produg when administered topically to the rabbit eye. Indeed, the cornea serves as a slow release depot and supplies the active form of latanoprost to the anterior segment during an extended period of time. Next, we also tested the effect of latanoprost when dosed the day of the IOP measurements. In both experiments, administration of latanoprost to ocular normotensive NZW rabbits did not reduce the IOP measured during the day (Figs. 3A, 3B). Compared with contralateral eyes, AMA0076 and latanoprost were able to significantly reduce IOP measured at night, with the peak IOP-lowering effect of both agents being 25.25% and 22.16%, respectively (Figs. 3C, 3D). In conclusion, the efficacy of AMA0076 was comparable with latanoprost when IOP measurements were performed at night. However, in contrast to latanoprost, AMA0076 reduced IOP also during the day with a potency equal to Y-39983.
AMA0076 Is More Potent Than Prostaglandin Analogues in an Acute Hypertensive Rabbit Model

To further test the efficacy of AMA0076 against PGAs we set up a hypertensive rabbit model. A single intracameral injection of viscoelastic material (Viscoat) in NZW rabbit eyes resulted in a rapid IOP increase reaching peak pressures of 30 mm Hg approximately 5 hours after injection (Fig. 4A). Twenty-four hours after injection of Viscoat, IOP consistently returned to pre-injection levels. Injection of saline did not influence IOP.

In a first experiment, the IOP-lowering agents, latanoprost and timolol, were evaluated in the hypertensive rabbit model. Latanoprost significantly prevented IOP rise after injection with Viscoat compared with saline-treated eyes (Fig. 4B), whereas timolol did not lead to pronounced IOP lowering in this model (Fig. 4C). These data confirm that this hypertensive model is suitable to evaluate compounds that directly target outflow, rather than compounds that decrease AH production.

Topical administration of AMA0076 significantly and dose-dependently prevented the IOP rise compared with vehicle-
treated eyes (Fig. 4D). AMA0076 0.1% attenuated the IOP rise induced by the injection of Viscoat compared with the control eye (overall $P < 0.0001$; ANOVA), whereas topical administration of AMA0076 0.3% and 0.5% completely inhibited IOP increase (overall $P < 0.0001$; ANOVA). Finally, a comparison between AMA0076, latanoprost, and bimatoprost revealed that AMA0076 was significantly more potent in preventing the IOP elevation in this ocular hypertensive model (overall $P < 0.0001$; ANOVA; Fig. 5).

Overall, these data show that AMA0076 effectively lowered IOP in an acute hypertensive rabbit model and was more potent than PGAs.

**FIGURE 2.** Effect of single topical administration of AMA0076 and Y-39983 on IOP in normotensive NZW rabbits during the day. Contralateral eyes were treated with vehicle ($n = 5$). (A–C) Time course of IOP reduction after topical administration of AMA0076 (0.1%, 0.3%, and 0.5%) or vehicle. (*$P < 0.05$, **$P < 0.01$, and ***$P < 0.001$ compared with the vehicle group at each time point). (D) Maximum IOP reduction (%) after topical administration of AMA0076 compared with baseline IOP before administration. (E–G) Time course of IOP reduction after topical administration of Y-39983 or vehicle. (*$P < 0.05$, **$P < 0.01$, and ***$P < 0.001$ compared with the vehicle group at each time point). (H) Maximum IOP reduction (%) after topical administration of Y-39983 compared with baseline IOP before administration. Intraocular pressure measurements were performed between 8 AM and 5 PM.
Minimal Conjunctival Hyperemia With AMA0076

ROCK inhibitors are known to induce mild to severe transient conjunctival hyperemia. Therefore the hyperemic effect of AMA0076 0.3% and Y-39983 0.3% was investigated in ocular normotensive NZW rabbits. While vehicle-treated eyes did not show distinguishable hyperemic effects, single administration of Y-39983 induced significant hyperemia in the NZW rabbits. In contrast, AMA0076 treatment only caused very mild hyperemia. Table 4 and Figure 6 represent the degree of hyperemia after single topical administration of AMA0076, Y-39983, and vehicle before, 1, 4, and 8 hours after dosing. Thus, compared with Y-39983, AMA0076 induced minimal conjunctival hyperemia in NZW rabbits.

DISCUSSION

Our results demonstrate that AMA0076, a novel, locally acting ROCK inhibitor significantly reduced IOP after topical administration. In contrast to other ROCK inhibitors, AMA0076 did not cause significant hyperemia at a dose that resulted in strong IOP reduction.

Exposure of human TM cells to AMA0076 led to a decrease in actin bundles and focal adhesions, and thereby induced changes in cell morphology. This finding is in line with the reported effects of other ROCK inhibitors Y-27632, Y-39983, and HA1077. Overall, ROCK inhibitors induce cytoskeletal rearrangements and loss of cell–extracellular matrix interactions in TM cells. These modifications might lead to increased inter trabecular pores in vivo and consequently decrease resistance of AH outflow through the TM, leading to a reduced IOP.

Compared with Y-39983, AMA0076 showed similar on target potencies on ROCKII. The IOP-lowering effect of both ROCK inhibitors was comparable at the tested doses. However, after instillation of Y-39983, an attenuated IOP-lowering effect was also observed in the contralateral vehicle-treated eye. This finding was not observed with AMA0076 and might be explained due to its soft properties. While the efficacy of ROCK inhibitors is promising, they are associated with significant hyperemia due to conjunctival vasodilatation. Soft ROCK inhibitors like AMA0076 undergo rapid degradation toward a predefined, functionally inactive metabolite. Such compounds might reduce side effects and widen the therapeutic window of ROCK inhibitors. We therefore compared the hyperemic effect of Y-39983, with AMA0076 that is metabolized in rabbit eye tissues. Interestingly, the hyperemic effect of a single topical ocular administration of AMA0076 in NZW rabbits was very mild and significantly lower compared with Y-39983 treatment. The mechanism(s) underlying this phenomenon are yet to be fully elucidated. But it is possible that AMA0076 remaining on the ocular surface is rapidly degraded to a functionally inactive metabolite.
metabolized and therefore less likely to reach the conjunctival blood vessels to cause hyperemia. However, once AMA0076 gets through the cornea, the remaining is more stable in AH. The relevance of this observation is significant. Several ROCK inhibitors have already been abandoned in clinical testing, and hyperemia is often suggested to be one of the reasons.

Prevention of hyperemia possibly led to suboptimal dosing with those ROCK inhibitors, resulting in limited IOP lowering. The highest dose of Y-39983 tested in phase 2 trials was 0.1%. Our results show that maximal IOP lowering is clearly not reached at this dose in NZW rabbits. The ability to dose sufficiently high without causing unacceptable levels of hyperemia is considered to be critical for ROCK inhibitors to be acceptable as IOP-lowering medication for the treatment of glaucoma.

As PGAs are an important treatment modality for glaucoma, we compared the IOP-lowering effect of AMA0076 to PGAs. In our study, latanoprost was unable to reduce IOP in ocular normotensive NZW rabbits during the day. This finding is in line with several previous reports but contradicts with
others, which did show an IOP-lowering effect of latanoprost in normotensive rabbits. The IOP-lowering effect of latanoprost in rabbits still remains a matter of debate. The reason for the difference may be attributed to variations in method and timing of measurement as well as differences in baseline IOP. This is also reflected by our data. We were only able to reveal an IOP-lowering effect of latanoprost during the night indicating that, in rabbits, the effect of latanoprost depends on the time of day. Rabbits have a clear circadian rhythm of IOP; low during the light phase and high during the dark phase. Furthermore, the observed IOP-lowering effect of latanoprost solely during the time of the day when IOP is highest suggests that the effect on IOP of latanoprost may depend more on baseline IOP than on the time of treatment. This finding confirms that only compounds which target AH outflow are able to prevent the IOP rise in this model. This is not unexpected, since AH suppressants presumably further increase AH viscosity in this model, and therefore have no IOP-lowering effect. Overall, our findings show that the induced IOP elevation can be counteracted by outflow facilitating drugs, and therefore this model can be used to test the efficacy of compounds that act via this mechanism. Gupta et al. also reported the IOP-lowering effect of different IOP-lowering drugs in several hypertensive rabbit models. The IOP-lowering effect of pilocarpine, timolol, and latanoprost was investigated in a water-loading and steroid-induced OH model. Latanoprost was found to be the most effective in both models. Compared with our model the steroid-induced model is more chronic; however, chronic administration of steroids is associated with a high mortality rate, which was not the case after Viscoat injection. In the water-loading model the IOP increase is very acute and lasts only for 2 hours, whereas in our model a consistent level of OH was induced for 4 hours.

Our study indicates that AMA0076 was more potent in counteracting the IOP elevation compared with latanoprost and bimatoprost in the hypertensive rabbit model. This might be explained by the different mechanisms of action of the two compound classes. Clinical and animal studies of AH dynamics have reported that PGAs reduce IOP predominantly by increasing uveoscleral outflow and to a lesser extent trabecular outflow facility. Conventional outflow is considered the main route and is believed to be regulated by the cellular behavior and contractility of TM cells. Since ROCK

### Table 4. Scoring of the Hyperemic Effect of AMA0076, Y-39983, and Vehicle After Single Dosing

<table>
<thead>
<tr>
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<th>Before Dosing (0 h)</th>
<th>1 h</th>
<th>4 h</th>
<th>8 h</th>
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<tr>
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<td>0.60</td>
<td>0.20</td>
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<tr>
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*n = 5 NZW rabbits/compound. Overall P = 0.002 (AMA0076 versus Y-39983).
inhibitors are expected to have a stronger effect on the TM, they have the potential to more effectively lower IOP.24,35

In summary, the present study shows that AMA0076 is a potent ROCK inhibitor with a similar IOP-lowering efficacy as Y-39983 and latanoprost, the latter only when dosed at night. Compared with PGAs, AMA0076 was more potent in preventing the IOP elevation in the acute hypertensive model. Due to its soft drug behavior, AMA0076 clearly has an improved tolerability profile with respect to hyperemia, making optimal dosing possible. This increased tolerability window makes AMA0076 a promising new candidate for the treatment of glaucoma.

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