

Influence of anaesthetic drugs on immune response: from inflammation to immunosuppression

DG Colucci¹, NR Puig¹, R Hernandez-Pando^{2*}

Abstract

Introduction

The immune system protects us from infections through coordinate action of its components. Patients and animals undergoing anaesthesia and surgery show alterations in the immune response. In this paper, we review the effects of some of the most common anaesthetic drugs on the immune system, in particular the effects within 24 hours after their administration, describing their effects on cells and cytokines of the innate and acquired immune system.

Discussion

As it is difficult to isolate the effects of anaesthetic drugs in the case of surgery, diverse *in vitro* studies with human immune cells or *in vivo* with animal models have been used to study the effect of anaesthetic drugs on the immune system. These studies have demonstrated diverse effects, such as changes in immune cell counts and functionality, and on the secretion patterns of diverse cytokines affecting the inflammatory response in the postoperative period.

Conclusion

Effects of anaesthetic drugs on the immune system are clinically important because the amount and function of the immune cells, as well as the balance between pro- and anti-inflammatory cytokines secretion, are related to postoperative infections and tissue injury.

* Corresponding author
Email: rhdezpando@hotmail.com

¹ Facultad de Ciencias Médicas, Instituto de Inmunología, Universidad Nacional de Rosario, Argentina

² Departamento de Patología Experimental, Instituto Nacional de Ciencias Médicas y Nutrición Dr. Salvador Zubiran, Mexico

Introduction

The immune system is vital for survival because our environment has plenty of potentially deadly microbes and immune system protects us from infectious pathogens. The immune system recognises and eliminates pathogens with the induction of innate and then adaptive immune responses. Innate immunity, also called natural or native immunity, is the firstline of defence and refers to protective mechanisms that are present even before infection. Principal components of innate immunity are epithelial membranes that block the entry of microbes, phagocytic cells (neutrophils and macrophages), dendritic cells, natural killer (NK) cells, and several plasma proteins, including the complement system. Most important cellular reactions of innate immunity are inflammation—the process in which phagocytic cells are recruited and activated to eliminate microbes—and virus elimination, mediated by dendritic and NK cells. Adaptive immunity, also called acquired or specific immunity, consists of mechanisms that are induced by microbes and are capable of specifically recognising microbial and nonmicrobial molecules called antigens. The adaptive immune system consists of lymphocytes and their products, including antibodies and cytokines. The receptors of lymphocytes are much more diverse than those of the innate immune system, and they are capable of recognising a vast array of foreign substances. There are two types of adaptive immunity: humoral immunity, which is mediated by B lymphocytes and their secreted antibodies, which protect against extracellular microbes and

their toxins, and cell-mediated or cellular immunity, which is mediated by T lymphocytes which mainly protect against intracellular microbes. Both types of acquired immunity are linked by a broad family of proteins called cytokines, which play an important role in immune cell activation, regulation and communication.

Patients or experimental animals submitted to anaesthesia and surgical procedures suffer diverse immunological alterations, which are difficult to determine if they are induced by anaesthetic drugs or by surgical procedure stress. Anaesthetics comprise a heterogeneous group of drugs whose mechanism of action is not yet fully clarified. It is known that anaesthetic drugs can alter synaptic transmission by two mechanisms. A nonspecific membrane perturbing action and a specific action on membrane receptors, primarily acting as an agonist on the inhibitory GABA_A¹ receptor, which is the major inhibitory nervous system receptor. Indeed, the specific mechanisms by which anaesthetic drugs affect the immune system still remain unclear, but anaesthetic drugs induce analgesia affecting the transmission of nerve impulses, and they modulate surgical stress by acting on the hypothalamus–pituitary–adrenal axis, affecting catecholamines and glucocorticoid secretion². Thus, part of the effect of anaesthetics on the immune system would be due to its action on the well-known immunomodulatory effect of glucocorticoids³.

Diverse *in vitro* experiments with human immune cells⁴, *ex vivo*⁵⁻⁷, *in vivo*⁸⁻¹⁰ or animal models have been used to study the effect of anaesthetic drugs on the immune system.

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

FOR CITATION PURPOSES: Colucci DG, Puig NR, Hernandez Pando R. Influence of anaesthetic drugs on immune response: from inflammation to immunosuppression. OA Anaesthetics 2013 Dec 30;1(3):21.

These studies have demonstrated diverse effects, such as changes in immune cell counts and functionality, and effect on the secretion patterns of diverse immune mediators, affecting the inflammatory response through the cytokine release in the postoperative period³. These effects are clinically important because the balance between the pro- and anti-inflammatory cytokines secretion is related to postoperative infections and tissue injury^{11,12}. Here, we review the effects of some of the most common anaesthetic drugs (Table 1) on the immune system. Although some studies have showed effects of anaesthetic drugs on immune response even several days after their administration^{9,13,14}, we will focus mainly on the effects within 24 hours after procedure, describing their effects on cells and cytokines of the innate and acquired immune systems.

Discussion

The authors have referenced some of their own studies in this review. The protocols of these studies have been approved by the relevant ethics committees related to the institution in which they were performed. Animal care was in accordance with the institution guidelines

Effects of anaesthetic drugs on immune cells

Anaesthetic drugs generally induce an increase in leucocytes counts^{9,13-17} exerting diverse effects on each of different immune cell subpopulation (Table 2). In the next section, most

significant functions of each immune cell type and the effect of specific anaesthetic drugs are described.

Neutrophils

Neutrophils are the most abundant population of circulating leucocytes. These cells are significant participants in the earliest phase of the inflammatory response. Neutrophils rapidly migrate to sites of infection, where they identify, ingest (phagocytosis) and destroy microbes (respiratory burst, lysosome degranulation). The neutrophil cytoplasm contains granules with microbicidal substances. Activated neutrophils release cytokines [tumour necrosis factor (TNF)- α , interleukin (IL)-1 β , IL-8 and transforming growth factor (TGF)- β], prostaglandins, thromboxanes and leukotrienes. Anaesthetic drugs affect both count and functionality of neutrophils. Thiopental and midazolam inhibit phagocytosis; and propofol, isoflurane, bupivacaine and lidocaine inhibit both phagocytosis and respiratory burst^{16,18}. Propofol and desflurane increase neutrophil counts in peripheral circulating blood^{13,14}, although several studies have reported contradictory effects of sevoflurane on neutrophil count^{13,14,19}.

Mononuclear phagocytes

The mononuclear phagocytic system consists of cells whose primary function is phagocytosis and plays central role in both the innate and adaptive immune responses. Monocytes are circulating cells which are incompletely differentiated until they migrate to tissues, where they

mature and become macrophages. Despite their primary function being phagocytosis, these cells also produce and release cytokines that stimulate inflammation (IL-1, IL-6, IL-12 and TNF- α). Anaesthetic drugs affect both monocyte circulating levels and functionality. Sevoflurane decreases circulating monocytes¹⁰ without affecting their phagocytic activity²⁰. Experiments carried out with halothane have contributed to understand the tissue specialisation of macrophages. Halothane does not have an effect on spleen macrophage phagocytic activity²¹, but enhance peritoneal macrophage phagocytosis and respiratory burst⁶. Intravenous anaesthetics, such as propofol, midazolam and thiopental, inhibit both phagocytosis and respiratory burst^{15,18}. Halothane and lidocaine decrease mononuclear cell counts by enhancing their apoptosis¹⁰.

Lymphocytes

This subpopulation of leucocytes has a common cell precursor and has several subtypes: NK cells, T cells (CD3, CD4 and CD8, and B cells (CD19) among others. Anaesthetic drugs exert effects on these cells. Some studies have reported a decrease in lymphocyte counts after anaesthesia with propofol^{7,14}. Inhalational anaesthetics induced different effects on lymphocytes. Some studies have showed an increase in lymphocyte counts after sevoflurane, halothane and desflurane anaesthesia^{6,13,19}, while others showed lower counts^{10,20}. Sevoflurane and isoflurane induced lymphocyte apoptosis²² *in vitro*. Ketamine²³ and local anaesthetics, such as bupivacaine and lidocaine²⁴, inhibited proliferation after mitogen stimuli. These are the reported effects of anaesthetic drugs in each lymphocyte subtype.

Natural killer cells

NK cells are large lymphocytes with abundant cytoplasmic granules and specific membrane markers. They are participants of the innate immune response, and essentially recognise

Table 1 Classification of anaesthetics

General anaesthetics		Local anaesthetics
Intravenous anaesthetics	Inhalational anaesthetics	
Ketamine	Halothane	Lidocaine
Thiopental	Sevoflurane	Bupivacaine
Propofol	Desflurane	Levobupivacaine
Fentanyl	Isoflurane	
Remifentanyl	Enflurane	
Midazolam		
Opioids		

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

FOR CITATION PURPOSES: Colucci DG, Puig NR, Hernandez Pando R. Influence of anaesthetic drugs on immune response: from inflammation to immunosuppression. OA Anaesthetics 2013 Dec 30;1(3):21.

Table 2 Anaesthetic drugs and their effect on immune cells

Immune cells	Effect of anaesthetic drugs
Leucocytes	Sevoflurane ↑ cell counts ¹³ Desflurane ↑ cell counts ¹³ Propofol ↑ cell counts ¹⁴ Isoflurane ↑ cell counts ¹⁴ Levobupivacaine ↑ cell counts
Neutrophils	Thiopental ↓ phagocytosis ^{16,18} Propofol ↓ phagocytosis and respiratory burst and ↑ cell counts ^{13,14,16,18} Isoflurane ↓ phagocytosis and respiratory burst ^{16,18} Bupivacaine ↓ phagocytosis and chemotaxis ^{16,18} Lidocaine ↓ phagocytosis and chemotaxis ^{16,18} Desflurane ↑ cell counts ^{13,14}
Mononuclear phagocytes	Sevoflurane ↓ monocyte counts ¹⁴ Halothane ↑ peritoneal macrophage phagocytosis and respiratory burst ⁴ ↓ cell counts ¹⁰ Propofol ↓ phagocytosis and respiratory burst ^{15,18} Midazolam ↓ phagocytosis and respiratory burst ^{15,18} Thiopental ↓ phagocytosis and respiratory burst ^{15,18} Lidocaine ↓ cell counts ¹⁰
Lymphocytes	Propofol ↓ cell counts ^{7,14} Sevoflurane ↓ cell counts ²² Isoflurane ↓ cell counts ²² Ketamine ↓ proliferation ²³ Bupivacaine ↓ proliferation ²⁴ Lidocaine ↓ proliferation ²⁴
NK cells	Propofol ↑ cell counts ^{7,14} , ↓ cytotoxic activity ⁷ Sevoflurane ↑ cell counts ¹³ Desflurane ↑ cell counts ¹³ Isoflurane ↓ cell counts ²⁵ Halothanes ↓ cytotoxic activity ⁷
CD4 Helper T lymphocytes	Halothane ↓ cell counts ²¹ Sevofluranes ↓ cell counts ¹³ , ↓ Th1 and ↑ Th2 ¹⁷ Isoflurane ↓ cell counts ²⁵ Propofol ↑ cell counts ¹⁴ and ↓ Th1 ¹⁷
CD8 Cytotoxic T lymphocytes	Propofol ↓ cell counts ^{14,21} Sevoflurane ↓ cell counts ^{14,21} Halothane ↓ cell counts ^{14,21} Isoflurane ↑ cell counts ^{13,25} Desflurane ↑ cell counts ^{13,25} Isoflurane ↑ apoptosis ²⁴
CD19 B lymphocytes	Isoflurane ↑ cell counts ^{13,25} Desflurane ↑ cell counts ^{13,25} Halothanes ↑ antibody titre ⁸ Sevoflurane ↑ Primary and secondary immune responses ^{5,8,9,20,21,27} Opioids ↓ proliferation and antibody production ¹⁵

virus-infected and stressed cells, and respond by direct cell killing and secretion of inflammatory cytokines [interferon (IFN)- γ]. An antibody

receptor on the NK surface, called CD16 (Fc γ RIIIa), recognises the Fc regions of IgG1 and IgG3 antibodies. Thus, NK cells kill target cells that have

been coated with antibody molecules (antibody-dependent cell-mediated cytotoxicity). The anaesthetic drug propofol induces an increase in NK

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

FOR CITATION PURPOSES: Colucci DG, Puig NR, Hernandez Pando R. Influence of anaesthetic drugs on immune response: from inflammation to immunosuppression. OA Anaesthetics 2013 Dec 30;1(3):21.

Cytokines	Effect of anaesthetic drugs
IL-1	Propofol ↓ plasmatic levels ¹⁵ Ketamine ↓ plasmatic levels ¹⁵ Thiopental ↓ plasmatic levels ¹⁵ Isoflurane ↑ plasmatic levels ²⁸ Sevoflurane ↓ plasmatic levels ¹⁵ Desflurane ↓ plasmatic levels ¹⁵ Opioids ↓ plasmatic levels ¹⁵ Lidocaine ↓ release ⁵ Bupivacaine ↓ release ²⁹
IL-6	Propofol ↑ plasmatic levels ^{14,18,29,31} Ketamine ↓ plasmatic levels ¹⁵ Thiopental ↓ plasmatic levels ¹⁵ Remifentanil ↑ plasmatic levels ³⁰ Fentanil ↑ plasmatic levels ³⁰ Sevoflurane ↑ plasmatic levels ¹⁴ Isoflurane ↑ plasmatic levels ^{4,28,31} Opioids ↓ plasmatic levels ¹⁵ Levobupivacaine ↑ plasmatic levels ²⁹ Bupivacaine ↑ plasmatic levels ⁴
IL-8	Ketamine ↓ plasmatic levels ¹⁵ Propofol ↑ plasmatic levels ³¹ Thiopental ↓ plasmatic levels ¹⁵ Midazolam ↓ plasmatic levels ¹⁵ Isoflurane ↑ plasmatic levels ³¹ Propofol <i>in vitro</i> ↓ release from neutrophils ¹⁸
IL-10	Ketamine ↑ plasmatic levels ¹⁵ Propofol ↑ plasmatic levels ^{14,15} Thiopental ↑ plasmatic levels ¹⁵ Fentanil ↑ plasmatic levels ³⁰ Remifentanil ↑ plasmatic levels ³⁰ Isoflurane ↑ plasmatic levels ⁴ Sevoflurane ↑ plasmatic levels ¹⁴ Bupivacaine ↑ plasmatic levels ⁴
TNFα	Ketamine ↓ plasmatic levels ¹⁵ Propofol ↓ plasmatic levels ¹⁵ Thiopental ↓ plasmatic levels ¹⁵ Fentanil ↑ plasmatic levels ³⁰ Remifentanil ↑ plasmatic levels ³⁰ Isoflurane ↑ plasmatic levels ^{4,25} Sevoflurane ↓ plasmatic levels ¹⁵ Enflurane ↓ plasmatic levels ¹⁵ Opioids ↓ plasmatic levels ¹⁵ Bupivacaine ↑ plasmatic levels ⁴

cell count and decreases cytotoxic activity^{7,14}. Inhalational anaesthetics such as sevoflurane and desflurane enhance NK cell counts¹³ and isoflurane decreases them²⁵. Halothane reduces NK cytotoxic activity⁷.

T lymphocytes (CD3)

T lymphocytes are produced in the bone marrow and their precursors migrate to and mature in the thymus. Two major T-cell subsets are helper T lymphocytes (CD4) and cytotoxic

T lymphocytes (CD8), which express in the membrane of the CD3 molecule that is part of the antigen receptor complex. Both CD4 and CD8 are involved in adaptive immune response. The effects of anaesthetic drugs depend on the T cell type. Isoflurane, sevoflurane and halothane enhance both CD4 and CD8 apoptosis not only by upregulation of the Fas/FasL system but also by affecting the expression of other anti-apoptotic and pro-apoptotic factors^{10,22,26}.

Helper T lymphocyte (CD4): CD4 lymphocytes are key mediators directing the adaptive immune response either to a cellular response (Th1) or humoral response by activating B cells (Th2). Repeated anaesthesia with halothane²¹ and desflurane¹³ decreases CD4 cells. Propofol increases cells counts¹⁴, while, depending on the model, sevoflurane may increase¹⁴ or decrease^{14,20} CD4 cell counts. Regarding the type of response, propofol decreases Th1 and sevoflurane diminishes Th1 and enhances Th2 responses¹⁷.

Cytotoxic T lymphocytes (CD8): The main function of these cells of the adaptive immune response is their specific cytotoxic activity against infected cells with intracellular organisms or neoplastic cells. CD8 cells are activated by cytokines (IL-2 and IFN-γ) secreted by Th1 lymphocytes. Anaesthesia with propofol, sevoflurane and halothane decreases CD8 cell numbers^{14,21}, while isoflurane and desflurane increases CD8 cell numbers^{13,25}.

B lymphocytes (CD19)

B cells are able to recognise soluble antigens by membrane antibodies. This recognition induces B-cell activation, proliferation and differentiation to plasma cells that secrete antibodies. The effect on the number of B lymphocytes depends on the drug type. Isoflurane and desflurane increase the cell numbers^{13,25}, while sevoflurane and propofol can increase^{13,14,27} or decrease^{18,20} it. Anaesthetics also affect B lymphocyte

Table 4 Effects of anaesthetic drugs on adaptive immune response cytokines

Cytokines	Effect of anaesthetic drugs
IL-2	Propofol ↓ production ²⁹ Ketamine ↓ release ¹⁵ Morphine ↓ plasmatic levels ¹⁵
IFN-γ	Propofol ↑ plasmatic levels ¹⁵ Thiopental ↓ plasmatic levels ¹⁵ Remifentanyl ↓ plasmatic levels ¹⁵ Opioids (morphine) ↓ plasmatic levels ¹⁵
IL-4	Thiopental ↓ plasmatic levels ¹⁵ Morphine ↓ plasmatic levels ¹⁵
TGF-β	Sevoflurane ↑ plasmatic levels ¹⁴ Morphine ↑ plasmatic levels ¹⁵

activation. Halothane increases the antibody titre⁸, and, as sevoflurane does, it increases the primary and secondary responses^{5,8,9,20,21,27}. Opiates decrease B-cell proliferation and antibody production¹⁵.

Effect of anaesthetic drugs on cytokines

Many interactions and effector functions of leucocytes are mediated by short-acting secreted mediators called cytokines, a heterogeneous group of molecules mainly produced by leucocytes, although, under certain conditions, they are also mediated by other cell types. Cytokine expression is highly regulated, and cell activation is necessary for their synthesis to exert their biological activity. They can act in autocrine, paracrine or endocrine way and their functions are also pleiotropic and redundant. Some of them are considered as innate immune response regulators and others regulate the adaptive immune response. In the following section, we describe the effects of anaesthetic drugs on cytokines; in fact, these are the better studied effects of anaesthetics on the immune system.

Cytokines of innate immunity

Cytokines such as IL-1, IL-6, IL-8, IL-10 and TNF-γ are produced and released mainly by cells of the innate immune response, such as activated

macrophages and monocytes. Lymphocytes and endothelial cells can also produce these cytokines (Table 3). The production of these cytokines can be affected by anaesthetic drugs.

IL-1: It is produced by monocytes, macrophages, dendritic and NK cells. There are two forms: IL-1α and IL-1β. IL-1 has significant inflammatory effects, such as histamine release induction, fever and the synthesis of acute phase proteins. Isoflurane induces high mRNA expression, and enhances IL-1 levels²⁸. Anaesthetic drugs such as ketamine, thiopental, sevoflurane, enflurane, propofol and opiates decrease IL-1 plasmatic levels¹⁵. *In vitro*, local anaesthetics such as lidocaine and bupivacaine inhibit IL-1 release²⁹.

IL-6: It is produced mainly by monocytes, macrophages and T lymphocytes; and among other functions, it stimulates acute phase protein synthesis. IL-6 also affects adaptive immune response, stimulates antibodies production and enhances IL-2. Anaesthetic drugs such as propofol, remifentanyl, fentanyl, sevoflurane, isoflurane, levobupivacaine and bupivacaine increase IL-6 levels^{4,14,18,28-32}, while ketamine, thiopental and opiates decrease IL-6 concentration¹⁵. The mechanism is not clear, but it is known that isoflurane increases mRNA expression²⁸ and sevoflurane enhances the level of the alarmin homobox protein 1³².

IL-8: It is actually a chemokine produced by leucocytes that stimulates neutrophil chemotaxis and degranulation. Propofol and isoflurane increase IL-8³¹, whereas ketamine, thiopental and midazolam inhibit its liberation¹⁵. *In vitro* experiments have demonstrated that propofol inhibits neutrophil IL-8 liberation¹⁸.

IL-10: It is produced by monocytes, macrophages, T-regulatory cells and B lymphocytes. It is the main anti-inflammatory cytokine because it inhibits the synthesis of IFN-γ, TNF-α, IL-2 and IL-12. IL-10 also induces IgG synthesis. Many anaesthetic drugs such as ketamine, thiopental, propofol, fentanyl, remifentanyl, isoflurane, sevoflurane and bupivacaine increase IL-10 levels^{4,14,15,30}.

TNF-α: It is produced by NK cells, T and B lymphocytes, mastocytes, monocytes and macrophages in response to bacterial antigens. TNF-α is the main pro-inflammatory cytokine and is responsible for septic shock. This cytokine favours extravasation with endothelial cell stimuli to produce IL-8 and adhesion molecules. Isoflurane raises mRNA²⁸ expression and increases TNF-α levels, and is the same with fentanyl, remifentanyl and bupivacaine^{4,25,28,30}. Sevoflurane, enflurane, ketamine, thiopental, propofol and opiates inhibit its liberation from mononuclear cells¹⁵.

Cytokines that regulate adaptive immune response

T CD4 lymphocyte activation, proliferation and differentiation after antigenic stimuli lead to an adaptive immune response towards Th1 or Th2 immunity. Th1 lymphocytes produce IL-2, IFN-γ and TNF-α, while Th2 lymphocytes mainly produce IL-4, IL-10 and IL-13 cytokines (Table 4). In fact, some of these cytokines are produced by the innate and adaptive immune responses; those that are specific for the adaptive response are affected by anaesthetic drugs.

IL-2: It is released by activated T lymphocytes (CD4 and CD8) after

antigenic stimuli. IL-2 is the main inducer of lymphocyte proliferation (T, B and NK cells). IL-2 also enhances natural (by NK cells) and specific (by CD8) cytotoxicity and increases MHC type II molecule expression. Anaesthetic drugs, through different mechanisms, decrease its effects. Ketamine inhibits its release¹⁵, morphine diminishes its levels¹⁵ and propofol suppresses IL-2 production²⁹.

IFN- γ : This cytokine is produced mainly by Th1, CD8 and NK lymphocytes. IFN- γ activates macrophages and inhibits Th2 differentiation, so it is considered a proinflammatory cytokine. *In vitro*, propofol increases its concentration¹⁵, whereas thiopental and remifentanyl decrease its concentration. The same effect has been observed during chronic opioid administration, such as morphine¹⁵.

IL-4: It is produced by Th2 cells, NK lymphocytes and mast cells. IL-4 promotes Th2 differentiation and inhibits Th1 response. IL-4 also blocks the action of IL-1 inducing the production of IL-1Ra. IL-4 is related to parasitic infections and allergic processes as it enhances IgE production. Thiopental inhibits IL-4 liberation, while morphine increases it¹⁵.

TGF- β : It is produced by activated T-regulatory lymphocytes and macrophages. TGF- β has immunosuppressive effects because it inhibits the synthesis of IFN- γ , TNF- α , IL-1 and IL-2, and it also inhibits NK and CD8 cytotoxic activity. Sevoflurane¹⁴ increases its levels and after chronic administration, morphine¹⁵ favours its production by lymphocytes, monocytes and macrophages.

Effect of anaesthetic drugs on glucocorticoids

Glucocorticoids (GCs) are significant immunomodulatory hormones. They bind to specific receptors that translocate to the nucleus and modulate cytokine expression. The GC-receptor complex binds to and inactivates nuclear factor- κ B (NF- κ B), one of the most important immune transcription

factors. In human blood mononuclear cells in culture, GCs strongly decrease the production of IL-1, TNF- α , IL-2, IL-3, IL-4, IL-5, IL-10, IL-12, IFN- γ , IL-6 and IL-8³. Sevoflurane and desflurane increase Gc plasmatic level¹³, while propofol decreases it¹⁴.

NF- κ B regulates T- and B-lymphocyte activation, and pro-inflammatory cytokine production and release, as well as adhesion molecule expression. NF- κ B is located in the cytoplasm in an inactivated form; after specific external signal, it is activated, translocated to the nucleus and promotes cytokine synthesis. Anaesthetic drugs such as ketamine, propofol and morphine inhibit cytokine production by blocking NF- κ B activation¹⁵.

Conclusion

Several immune functions are modified after anaesthetic drugs are administered by direct or indirect effects on stress responses. Significant activities such as phagocytosis, respiratory burst, proliferation and cell count are modified after anaesthetic procedures. Anaesthesia also affects the immune response by suppressing or by releasing different cytokines, affecting the inflammatory response. Thus, the type of anaesthetic drug is important to consider the surgical procedures for animal models because they are able to affect diverse immune system functions and should be taken into account when choosing the anaesthetic drug. Regarding the clinical practice, it has been reported that some alterations in the immune system persist several days after the end of the anaesthetic exposure. However, the post-anaesthetic immunological complications are rare in patients with proper immune system function; while in patients with certain immunodeficiency, the choice of appropriate pain therapy should be carefully selected, considering that the interaction between anaesthesia and the immune system can lead to complications.

This is the case in HIV-infected patients that in addition to the effects of anaesthetics on the immune system, it is also important to consider the interactions between antiretroviral drugs and anaesthetics. Antiretrovirals increase or decrease the activity of liver enzymes shortening or lengthening, respectively, the effects of anaesthetic drugs. This subject is also important in other conditions, such as in cancer, considering that immunosuppression induced by anaesthetic drugs enhances progression of metastasis after tumour removal surgery. NK cells, through the release of IFN- γ and their cytotoxic activity, are significant factors that contribute for the elimination of neoplastic cells; anaesthetic drugs that affect these functions provoke decreased IFN- γ release or cytotoxic activity impairing the elimination of tumour cells. This is a situation that must be considered in each patient, which includes trying to find the best anaesthetic technique that leads to a better outcome.

Abbreviations list

GC, glucocorticoids; IL, interleukin; INF, interferon; NF- κ B, nuclear factor- κ B; NK, natural killer; TGF, transforming growth factor; TNF, tumour necrosis factor.

References

1. Wu J, Harata N, Akaike N. Potentiation by sevoflurane of the gamma-aminobutyric acid-induced chloride current in acutely dissociated CA1 pyramidal neurones from rat hippocampus. *Br J Pharmacol*. 1996 Nov;119(5):1013–21.
2. Graziola E. Influencia del estrés anestésico-quirúrgico sobre la distribución y función de los leucocitos. *Rev Arg Anest*. 2002;60:387–90. In Spanish.
3. Brattsand R, Linden M. Cytokine modulation by glucocorticoids: mechanisms and action in cellular studies. *Aliment Pharmacol Ther*. 1996 Feb;10(Suppl. 2): 81–90.
4. Amin OAI, Salah HE. The effect of general or spinal anaesthesia on pro- and anti-inflammatory intracellular cytokines in patients undergoing appendectomy

- using flowcytometric method. *Egypt J Anaesth.* 2011 Apr;27(2):121–5.
5. Puig NR, Elena GA, Barragán J, Comba JO, Amerio N. Oxygen tensio-associated changes on secondary immune response in halothane or isoflurane anesthetized mice. *Acta Anesthesiol Scand.* 1995 Oct;39(7):945–8.
 6. Colucci D, Harvey G, Gayol MC, Elena G, Puig N. Halothane anesthesia in mice: effect on the phagocytic activity and respiratory burst of peritoneal macrophages. *Neuroimmunomodulation.* 2011 Sep;18(1):11–8.
 7. Miyata T, Kodama T, Honma R, Nezu Y, Harada Y, Yogo T, et al. Influence of general anesthesia with isoflurane following propofol induction on natural killer cell cytotoxic activities of peripheral blood lymphocytes in dogs. *J Med Vet Sci.* 2013 Jul;75(7):917–21.
 8. Puig N, Elena GA, Barragán J, Comba JO, Amerio N. Halothane-associated enhancement of the secondary immune response to sheep erythrocytes in mice: cell transfer studies. *Acta Anesthesiol Scand.* 1993 Oct;37(7):647–51.
 9. Elena G, Amerio N, Ferrero P, Bay ML, Valenti J, Colucci D, et al. Effects of repetitive sevoflurane anesthesia on immune response, select biochemical parameters and organ histology in mice. *Lab Anim.* 2003 Jul;37(3):193–203.
 10. Simeonova GP, Slovov E, Usunov R, Halacheva K, Dinev DN. Increased apoptosis of peripheral blood mononuclear cells (PBMC) during general and epidural anaesthesia in dogs. *Vet Res Commun.* 2008 Dec;32(8):619–26.
 11. Hildebrand E, Pape HC, Krettek C. The importance of cytokines in the posttraumatic inflammatory reaction. *Unfallchirurg.* 2005 Oct;108(10):793–4,796–803.
 12. Lin E, Calvano SE, Lowry SF. Inflammatory cytokines and cell response in surgery. *Surgery.* 2000 Feb;127(2):117–26.
 13. Cocelli LP, Ugur MG, Karadasli H. Comparison of effects of low-flow sevoflurane and desflurane anesthesia on neutrophil and T-cell. *Curr Therapeut Res.* 2012 Feb/Apr;73(1–2):41–51.
 14. Schneemilch CE, Ittenson A, Ansorge S, Hachemberg T, Bank U. Effect of 2 anesthetic techniques on the postoperative proinflammatory and anti-inflammatory cytokine response and cellular immune function to minor surgery. *J Clin Anesth.* 2005 Nov;17(7):517–27.
 15. Lisowska B, Szymańska M, Nowacka E, Olszewska M. Anesthesiology and the cytokine network. *Postepy Hig Med Dosw(Online).* 2013 Aug;67:761–9.
 16. Heine J, Jaeger K, Osthaus A, Weingaertner N, Münte S, Piepenbrock S, et al. Anaesthesia with propofol decreases FMLP-induced neutrophil respiratory burst but not phagocytosis compared with isoflurane. *Br J Anaesth.* 2000 Sep;85(3):424–30.
 17. Inada T, Yamanouchi Y, Jomura S, Sakamoto S, Takahashi M, Kambara T, et al. Effect of propofol and isoflurane anaesthesia on the immune response to surgery. *Anaesthesia.* 2004 Oct;59(10):954–9.
 18. Rizzo A, Campanile D, Spedicato M, Minoia G, Sciorsci RL. Update on anesthesia and the immune response in newborns delivered by cesarian section. *Immunopharmacol Immunotoxicol.* 2011 Dec;33(4):581–5.
 19. Morisaki H, Aoyama Y, Shimada M, Ochiai R, Takeda J. Leukocyte distribution during sevoflurane anaesthesia. *Br J Anaesth.* 1998 Apr;80(4):502–3.
 20. Puig NR, Ferrero P, Bay ML, Hidalgo G, Valenti J, Amerio N, et al. Effects of sevoflurane general anesthesia: immunological studies in mice. *Int Immunopharmacol.* 2002 Jan;2(1):95–104.
 21. Elena G, Puig NR, Bay ML, Urizar L, Barragán J, Comba J, et al. Inhalatory anaesthetic (halothane) associated changes in the immune response in mice. *Int J Immunopharmacol.* 1997 Nov-Dec;19(11–12):699–707.
 22. Matsuoka H, Kurosawa S, Horinouchi T, Kato M, Hashimoto Y. Inhalation anaesthetics induce apoptosis in normal peripheral lymphocytes in vitro. *Anesthesiology.* 2001 Dec;95(6):1467–72.
 23. Bellin B, Rusabrov Y, Shapira Y, Royblat L, Greemberg L, Yardeni IZ, et al. Low-dose ketamine affects immune responses in human during the early postoperative period. *Br J Anaesth.* 2007 Oct;99(4):522–7.
 24. Ramus GV, Cesano L, Barbalonga A. Different concentration of local anaesthetics have different modes of action on human lymphocytes. *Agents Actions.* 1983 Jun;13(4):333–41.
 25. Brand JM, Kirchner H, Poppe C, Schmucker P. The effects of general anesthesia on human peripheral immune cell distribution and cytokine production. *Clin Immunol Immunopathol.* 1997 May;83(2):190–4.
 26. Delogu G, Moretti S, Antonucci A, Marcellini S, Masciangelo R, Famularo G, et al. Apoptosis and surgical trauma. dysregulated expression of death and survival factors on peripheral lymphocytes. *Arch Surg.* 2000 Oct;135(10):1141–7.
 27. Colucci D, Ferrero P, Ferreyra P, Elena G, Puig NR. Effects of sevoflurane anesthesia on the immune response and biochemical parameters in mice. Comparison between single exposure and repeated anesthesia. *Rev Esp Anestesiol Reanim.* 2003 Apr;50(4):170–5.
 28. Wu X, Lu Y, Dong Y, Zhang G, Zhang Y, Xu Z, et al. The inhalational anaesthetic isoflurane increases levels of proinflammatory TNF α , IL-6 and IL-1 β . *Neurobiol Aging.* 2012 Jul;33(7):1364–78.
 29. Žura M, Kozmar A, Šakić K, Malenica B, Hrgovic Z. Effect of spinal and general anesthesia on serum concentration of pro-inflammatory and anti-inflammatory cytokines. *Immunobiology.* 2012 Jun;217(6):622–7.
 30. Ke JJ, Zhan J, Feng XB, Wu Y, Rao Y, Wang YL. A comparison of the effect of total intravenous anaesthesia with propofol and remifentanyl and inhalational anaesthesia with isoflurane on the release of pro- and anti-inflammatory cytokines in patients undergoing open cholecystectomy. *Anesth Intens Care.* 2008 Jan;36(1):74–8.
 31. Mazoti MA, Braz MG, de Assis Golim M, Braz LG, Dias NH, Salvadori DM, et al. Comparison of inflammatory cytokine profiles in plasma of patients undergoing otorhinological surgery with propofol or isoflurane anesthesia. *Inflamm Res.* 2013 Oct;62(10):879–85.
 32. Manganelli V, Signore M, Pacini I, Misasi R, Tellam G, Garofalo T, et al. Increased HMBG1 expression and release by mononuclear cells following surgical/anesthesia trauma. *Crit Care.* 2010 Nov;14(6):R197.
 33. Bajwa SJ, Kulshrestha A. The potential anesthetic threats, challenges and intensive care considerations in patients with HIV infection. *J Pharm Bioallied Sci.* 2013 Jan;5(1):10–6.
 34. Deegan CA, Murray D, Doran P, Ecimovic P, Moriarty D, Buggy DJ. Effect of anaesthetic technique on oestrogen receptor-negative breast cancer cell function in vitro. *Br J Anaesth.* 2009 Nov;103(5):685–90.

Licensee OA Publishing London 2013. Creative Commons Attribution License (CC-BY)

FOR CITATION PURPOSES: Colucci DG, Puig NR, Hernandez Pando R. Influence of anaesthetic drugs on immune response: from inflammation to immunosuppression. *OA Anaesthetics* 2013 Dec 30;1(3):21.