

RESEARCH

Confirmation of Quantitative Trait Loci for Ethanol Sensitivity in Long-Sleep and Short-Sleep Mice

Paul D. Markel,^{1,2,4} Beth Bennett,^{1,3} Mary Beeson,¹ Lena Gordon,¹ and Thomas E. Johnson^{1,3}

¹Institute for Behavioral Genetics and ²Department of Psychology, University of Colorado, Boulder, Colorado 80309; ³Department of Veterans Affairs Alcohol Research Center, Denver, Colorado 80220

Initial insensitivity to alcohol is a strong predictor of human alcoholism, a widespread and heritable health problem. The Long Sleep and Short Sleep lines of mice were developed by genetic selection for high or low alcohol sensitivity. We have identified seven quantitative trait loci (QTLs) specifying differences in alcohol sensitivity using intercross progeny from these selected strains. These QTLs (*Lore1–Lore7*) together account for ~60% of the total genetic variance for this trait. This represents the first report of linkages for genes influencing alcohol action in any mammalian system using stringent, genome-wide mapping criteria.

At least 10 million Americans are reported to have drinking problems. Alcoholism is a complex trait with a significant genetic component and heterogeneous architecture in family studies, suggesting that different genes may contribute to the segregation of alcoholism in different families (Devor and Cloninger 1989). Alcohol sensitivity is an important predictor of alcoholism: Sons of alcoholic fathers are at an elevated risk for alcoholism and are less sensitive to the effects of ethanol than are sons of nonalcoholic fathers (Schuckit 1985, 1994; Newlin and Thomson 1990; Pollock 1992). Moreover, Schuckit (1994) has shown that insensitivity to ethanol is a highly significant predictor of future alcohol abuse in young men, independent of familial alcoholism. It is possible that initial insensitivity to ethanol is a predisposing factor for alcoholism. Thus, a first step toward the understanding of alcoholism may involve the discovery of those genes that influence initial ethanol sensitivity.

The considerable experimental difficulties in studying the genetics of human alcoholism are circumvented by several rodent models developed by genetic selection for various aspects of alcohol action (Crabbe et al. 1994a). Long-Sleep (LS) and Short-Sleep (SS) mice are selected lines that differ 18-fold in ethanol sensitivity, measured as the duration of loss of righting reflex (LORR) after a hypo-

notic dose of ethanol (McClearn and Kakihana 1981). This differential response to ethanol is mediated primarily by the central nervous system (Heston et al. 1974; Erwin et al. 1976; Howerton et al. 1983) and very little by the rate of ethanol metabolism (Smolen et al. 1986; Smolen and Smolen 1989). LORR is ~40% heritable and is specified by approximately seven quantitative trait loci, or QTLs (Dudek and Abbott 1984; DeFries et al. 1989; Markel et al. 1995a).

RESULTS

Genetic Strains and Crosses

We have focused on identifying QTLs that influence differential duration of LORR between LS and SS mice (Johnson et al. 1992; Markel et al. 1996b). Previously, we identified 12 provisional QTLs for LORR in the LS × SS recombinant inbred strains (RIs) (Markel et al. 1996). Only one of these (chromosome 2, 80 cM) exceeded confirmational threshold ($P > 0.0002$; 11). In this report we use a large sample of intercross progeny (F_2) derived from reciprocal crosses of the inbred LS (ILS) and the inbred SS (ISS) strains to confirm or disprove provisional QTLs from the RIs (Belknap 1992; Johnson et al. 1992; Neumann 1992) as well as identify new QTLs in a genome-wide screen.

This sample of ILS × ISS F_2 mice ($n = 1072$) was characterized using a repeated measures design

⁴Corresponding author. Present address: Millennium Pharmaceuticals, Inc., Cambridge Massachusetts 02139. E-MAIL markel@mpi.com; FAX (617) 374-9379.

QTLs FOR ALCOHOL SENSITIVITY

wherein LORR was measured on each individual mouse twice separated by ~1 week (Markel et al. 1995a,b); mean LORR did not differ significantly between trials. The use of averaged individual scores resulted in a 39% reduction in environmental variance in the measure of LORR (Markel et al. 1995a), which enhances the resolution of genetic variance for QTL mapping (Falconer 1989).

Genetic Mapping

Ninety-two of the longest and 92 of the shortest sleeping F₂ mice, based on averaged scores, were chosen for genotyping. The 12 QTLs provisionally identified in LS × SS RI strains (Markel et al. 1996) were tested by assessing simple sequence length polymorphism (SSLP; Dietrich et al. 1992) marker genotypes in these regions of the genome (Table 1) using MapMaker/QTL (Lander et al. 1987). Only two provisional QTLs identified previously in the LS × SS RIs were confirmed. These two QTLs had lod scores in F₂ mice of 5.4 and 5.8 near the position of the maxima identified in the RI screen; these values exceed those required for whole genome scans (Belknap 1992; Neumann 1992; Lander and Schork 1994; Lander and Kruglyak 1995). The 1-lod-support interval for each of these QTLs included the locus identified in the RI analysis. These confirmed QTLs are on chromosomes 1 and 2 and show maximum lod scores of 5.4 and 6.6 at 54 and 85 cM, respectively (Table 2; Fig. 1). One other QTL, on chromosome 18, was confirmed but at a lower level

of certainty (lod = 1.8, corresponding to a value of 5% error over all tests) (Belknap 1992; Neumann 1992).

Genome Scan

We then conducted a whole genome scan at an average spacing of 20 cM using SSLP markers in the longest and shortest sleeping F₂ mice. Because we used selected strains for the progenitors, the test of significance is one-sided and the likelihood for each QTL found exceeded the recommended threshold that corresponds to a genome-wide significance level of 5% (lod ≥ 3.4; Lander and Schork 1994; Lander and Kruglyak 1995). Three new QTLs were detected with lod scores higher than 3.4 (Table 2). Together with the three QTLs mentioned above and one weak QTL identified previously on chromosome 7 (Markel and Corley 1994), seven QTLs have been identified. These have been named *Lore1–Lore7* (Table 2) for loss of righting induced by ethanol. Assuming a strictly additive model for QTL effect, these seven loci account for 59% of the genetic variance in LORR. The total effect explained by these seven QTLs is >130 min of the 170-min difference in LORR between ILS and ISS parental strains.

Blood Ethanol Concentration

We also attempted to confirm the provisional QTLs

Table 1. Tests of Provisional QTLs for LORR in ILS × ISS Intercross

RI (LS × SS)				F ₂ (ILS × ISS) (local max. lod)			
Chr.	cM	t	P	lod ^a	max. ^b	cM	(± 1 lod)
1	45	3.14	0.002	5.3	5.4	54	(43–59)
1	99	1.98	0.03	1.0	—	—	—
2	80	4.04	0.0002	5.9	6.6	85	(78–95)
3	74	2.01	0.03	0.4	1.0	31	(cen.–70)
4	56	1.92	0.03	1.2	1.5	72	(39–tel.)
4	74	2.00	0.03	1.5	1.5	72	(39–tel.)
5	72	1.89	0.04	1.0	—	—	—
6	46	2.33	0.02	1.4	1.5	48	(7–70)
12	43	2.18	0.02	1.6	3.1	17	(10–26)
13	25	2.23	0.02	1.5	2.3	10	(cen.–13)
16	41	—2.26	0.02	0.6	1.4	57	(40–tel.)
18	16	2.06	0.03	1.2	1.8	41	(24–tel.)

^alod score from F₂ analysis at site of maximum from RI analysis.

^blod score at site of maximum in the local genetic region from F₂ analysis (n = 184).

Table 2. QTLs for Ethanol-Induced LORR (from Genome Scan)

QTL	Chr.	cM	Max. lod	1-lod interval ^a	Effect (min)	Percent V_G ^b	No. of mice	Method ^c	
								RI	F ₂
<i>Lore1</i>	1	54	5.4	43–59	20.9	10.2	186	*	***
<i>Lore2</i>	2	85	6.6	78–95	25.2	14.2	186	*	***
<i>Lore3</i>	8	59	3.4	44–71	19.0	7.7	186		**
<i>Lore4</i>	11	49	6.5	44–56	25.0	12.2	186		***
<i>Lore5</i>	15	46	4.0	32–55	19.0	8.4	186		**
<i>Lore6</i>	18	41	1.8	24–tel.	12.6	3.5	186	*	*
<i>Lore7</i>	7	50	1.7	25–61	10.8	2.7 ^d	1063	* ^d	* ^e
Total					133.6	59.1			

^aPeak lod \pm 1 lod interval. Map positions from chromosome committee reports.^b(V_G) Genetic variance.^c(*) Suggestive; (**) significant; (***) highly significant (Lander and Kruglyak 1995).^dDefries et al. (1989).^eMarkel and Corley (1994).

for blood ethanol concentration at awakening (BEC) found in LS \times SS QTL mapping (Markel et al. 1996a). BEC, which is independent of ethanol dose for each strain, measures CNS sensitivity to ethanol and serves to rule out pharmacokinetic (metabolic) aspects of alcohol action: SS mice wake with higher BEC than LS mice (Smolen et al. 1986). Although LS and SS mice may differ in their ethanol metabolism, this difference is primarily attributable to dose and route of administration (Smolen et al. 1986), both of which were constant in this study. Of the 10 tested, only 1 provisional QTL reached suggestive levels of confirmation in F₂ (data not shown). In part, this is because only ~40% of the mice were also tested for BEC but is consistent with the findings for confirmation of LORR.

A whole genome search for QTLs for BEC revealed six provisional QTLs in the ILS \times ISS F₂ mice, with lod scores between 2.0 and 3.0. These provisional QTLs are located on chromosomes 2, 8, 9, 11, 14, and 15. One of these QTLs (chromosome 2, 60 cM) was detected previously in the study of the LS \times SS RIs (Markel et al. 1996b). These provisional QTLs together explain a range of 184 mg% for differential brain levels of ethanol at awakening and ~50% of the genetic variance. These provisional QTLs for BEC must be confirmed in independent samples, which are currently under way.

Reliability of the Measure for Alcohol Sensitivity

We also addressed whether the strategy of duplicate

assessments of LORR on each F₂ mouse led to a noticeable increase in lod scores (Fig. 1). The lod scores of both LORR₁ and LORR₂ are similar for most loci and not very different from the lod derived from mapping the average. *Lore2* is an exception in that a significantly higher lod score was obtained in mapping LORR₁ than in mapping LORR₂. This locus may determine tolerance such that the effect of alcohol would not be observed as readily at the second time point because the expression of this gene has been modified by prior exposure.

Epistasis

Applying segregation analysis, a small proportion of genetic variance for LORR was attributed to an epistatic effect among these ILS \times ISS F₂ mice (Markel et al. 1995a). Using two-way analyses of variance, nonlinear interactions between each confirmed LORE QTL and each unlinked marker (including other QTLs) were examined. After adjusting probability values for multiple test using a Bonferroni correction (Markel et al. 1996b), no significant epistatic effects were detected between LORR QTLs.

DISCUSSION

This report represents the first general linkage analysis of alcohol action in any mammalian system using stringent, genome-wide mapping criteria. We have determined approximate map positions for

QTLs FOR ALCOHOL SENSITIVITY

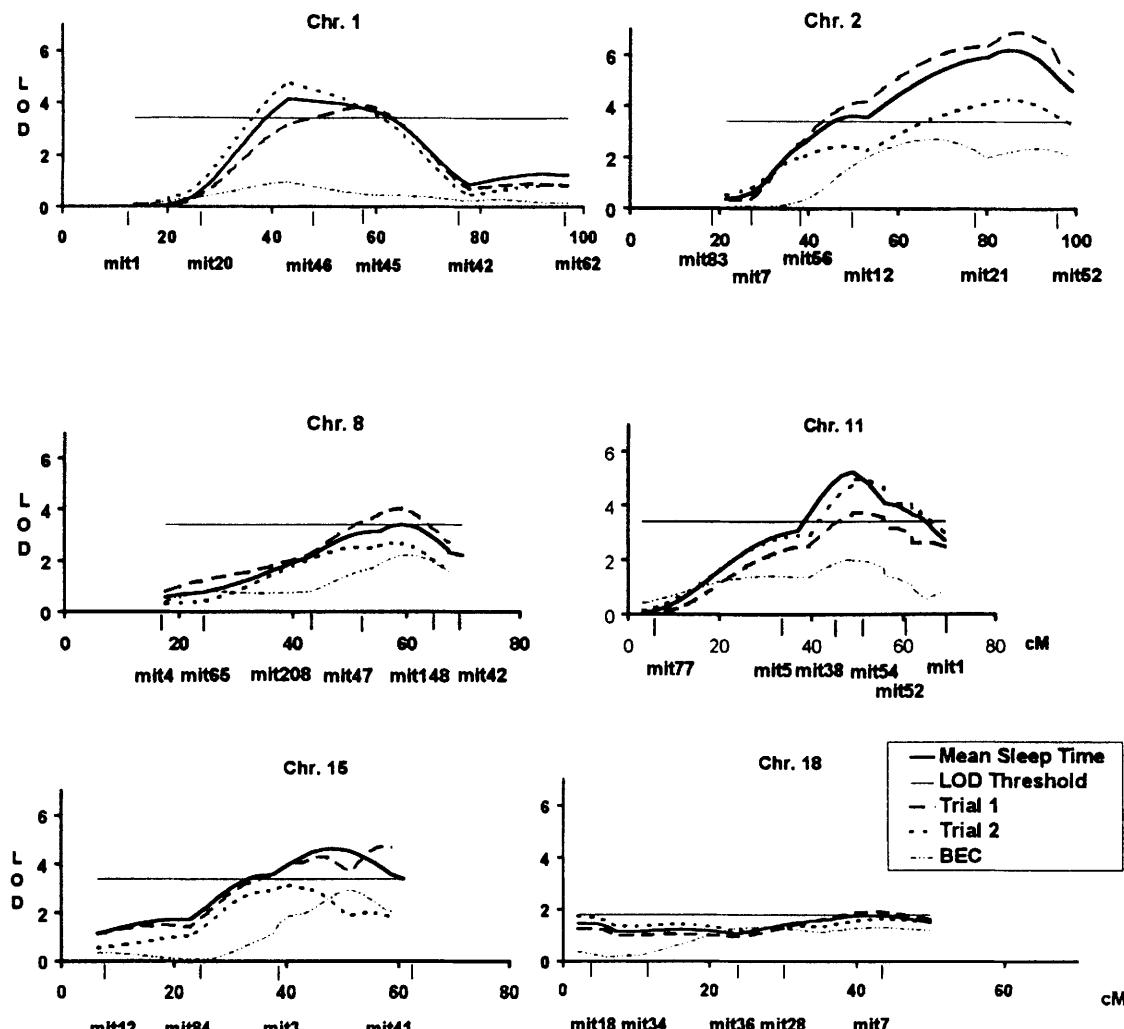


Figure 1 Likelihood maps for QTLs on chromosomes 1, 2, 8, 11, 15, and 18. The lod score is shown on the vertical axis; lod of 3.4 (shown as a horizontal line) is the cutoff for statistical significance in all but chromosome 18, which was confirmed at a lod of 1.8. Genetic distance along the chromosome is plotted on the horizontal axis; markers used for the assessment are also indicated. The thick line is for average sleep time. The two dotted lines show lod scores for sleep time 1 and sleep time 2. A map of BEC is also shown.

seven QTLs that influence individual differences for hypnotic sensitivity to ethanol. The identification of these QTLs for ethanol sensitivity is a valuable step toward identifying genes involved in the etiology of alcoholism and alcohol disorders.

Although several groups have used various RI series for the provisional mapping of QTLs influencing several ethanol related responses (McClearn et al. 1992; Belknap et al. 1993; Crabbe et al. 1994b; Rodriguez et al. 1995; Markel et al. 1996a), our F_2 results strongly encourage confirmation or disproof of provisional QTLs from RI-mapping experiments using large F_2 stocks.

Our findings suggest that initial characteriza-

tion in RIs can identify major QTLs, such as those on chromosomes 1 and 2, with relative certainty. Nevertheless, three QTLs (*Lore3*, *Lore4*, and *Lore5*) were not detected in RI mapping even though these loci each explain 7%–12% of the variance. This is consistent with earlier power calculations (Belknap 1992; Neumann 1992; Markel et al. 1996a), suggesting that QTLs explaining <25% of the variance in the RI panels of this size would not be detected with any degree of certainty. Equally important is the fact that of 12 other provisional QTLs identified with a probability value between 0.05 and 0.01 (Markel et al. 1996a), only 1 was confirmed in the F_2 replication. The use of RIs alone is likely to produce

MARKEL ET AL.

mostly false-positive associations unless very stringent statistical criteria are applied.

LORR and BEC are each a measure of ethanol sensitivity. In the LS × SS RI the correlation between LORR and BEC is -0.79 (Smolen and Smolen 1989); in our F_2 it is -0.50 . We asked whether the *Lore* QTLs all show the expected negative relationship with BEC (Table 3; Fig. 1). *Lore1* is the only exception to the general finding of significant negative relationships between LORR and BEC. One explanation could be that the *Acrd* and *Acrg* genes, which are in the candidate region for *Lore1* and code for the acetylcholine receptor subunits delta and gamma, respectively, are not expressed in the CNS but in the peripheral nervous system. Instead, *Lore1* may mediate the righting response, in part via peripheral motor control; such nicotinic receptors have been shown to be sensitive to alcohol (Gage et al. 1975; Aiken et al. 1991; Murrell et al. 1991).

Several candidate genes mapped previously can now be explored for possible direct involvement in the etiology of alcohol sensitivity. A number of candidate genes lie within the 1-lod support interval for the *Lore* QTLs (Fig. 2). The gene for the high-affinity neurotensin (NT) receptor (*Ntsr*) is located at ~ 80 cM on chromosome 2 very near the maximum lod for *Lore2*, and this locus also shows consistent correlation ($P < 0.05$) with the observed NT receptor density in the frontal cortex and striatum in the LS × SS RI panel (V.G. Erwin, P.D. Markel, T.E. Johnson, and B.C. Jones, in prep.). Four of the QTLs reported here (*Lore1*, *Lore2*, *Lore4*, and *Lore5*) fall in virtually the same location as QTLs from the LS × SS RI panel for NT receptor density (V.G. Erwin, P.D. Markel, T.E. Johnson, and B.C. Jones, in prep.). NT treatment

enhances hypnotic sensitivity to ethanol in a genotype-dependent manner in the LS and SS lines, and both NT levels and NT receptor density vary widely among the LS × SS RI strains. Densities of the high-affinity NT receptors in the entorhinal cortex, the frontal cortex, and the striatum are positively correlated with ethanol-induced sleep time (Erwin et al. 1993).

The α -adrenergic $\beta 2$ receptors, via G proteins, activate adenylyl cyclase (AC), which has been studied intensively as a biochemical marker for vulnerability to alcoholism (Saito et al. 1987). Ethanol exposure can reduce AC activity as much as 35% in LS mice while effecting no change in SS mice (Wand and Levine 1991). Two candidate genes in this pathway are located at or near *Lore2*; these are *Adra2b*, the $\alpha 2$ adrenoreceptor subtype 2b at 71 cM, and *Gpcr8*, the G-protein-coupled receptor 8, at 73 cM.

Prodynorphin (Pdyn) and inducible nitric oxide synthase (Nos2) have been implicated in alcohol action; these are candidate genes near *Lore2* and *Lore3*, respectively. Other candidate genes of general pharmacological interest are suggested by the mapping data. For example, a serotonin transporter gene (Htt) was found near *Lore4* on chromosome 11. *Cchl1a3* (calcium channel L, type 1A3 subunit) is within the support region for *Lore1*. *Cyp11b-1,-2* (steroid-11- β -hydroxylase) is a candidate for *Lore5* on chromosome 15.

METHODS

Propagation and Characterization of Mice

All animals were developed and obtained from the Institute for Behavioral Genetics, (Boulder, CO). All experiments were

Table 3. Summary of Findings for *Lore* Genes

QTL	Chr.	LORR		BEC		Individual LORR ^a			
		max. (lod)	effect (min)	max. (lod)	effect (mg%)	(lod)	effect (min)	(lod)	effect (min)
<i>Lore1</i>	1	5.4	22	0.8	N.S. ^b	3.9	20	4.8	21
<i>Lore2</i>	2	6.6	25	2.0	-27	6.9	24	4.3	22
<i>Lore3</i>	8	3.4	19	2.2	-39	4.0	20	2.7	16
<i>Lore4</i>	11	6.5	25	2.0	-37	3.7	23	5.0	24
<i>Lore5</i>	15	4.0	19	3.0	-19	4.3	20	3.2	16
<i>Lore6</i>	18	1.8	12	1.3	N.S. ^b	1.9	11	1.6	13
<i>Lore7</i>	7	1.7	11	2.7	N.S. ^b	2.0	10	1.4	11

^aPeak lod for first and second measurements of LORR (see text).

^b(N.S.) Not significant.

QTLs FOR ALCOHOL SENSITIVITY

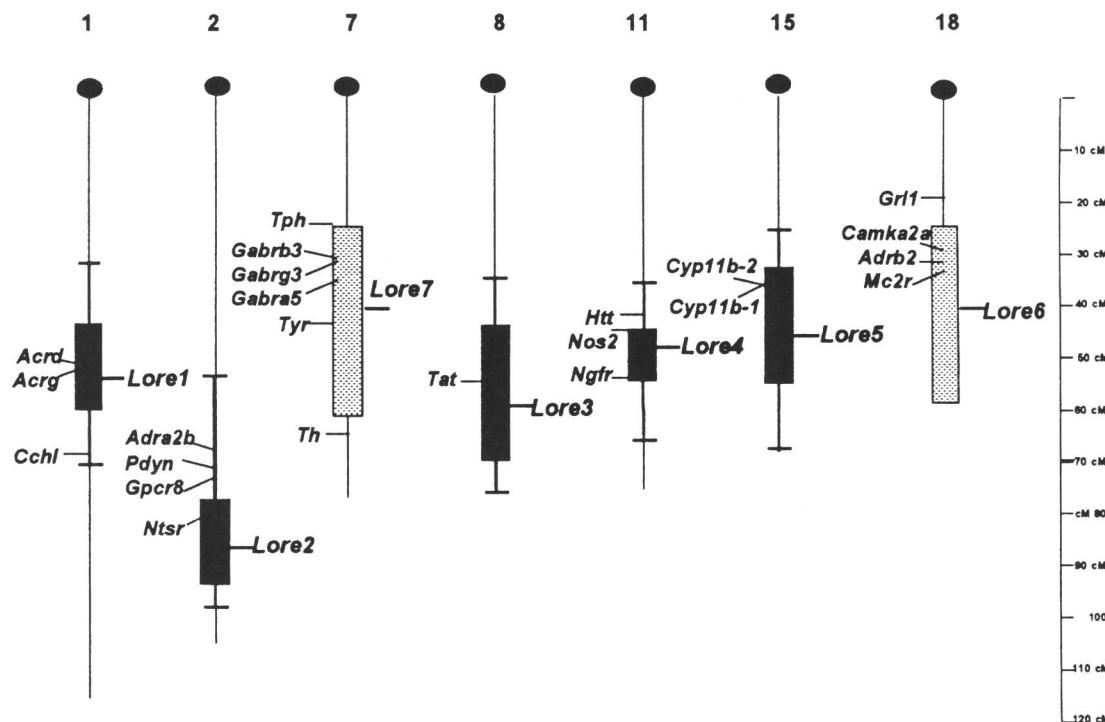


Figure 2 *Lore1–Lore6*. The thick bar shows a 1-lod support interval; thin lines show the 2-lod support interval. Some candidate genes are also shown: (Acrd) Acetyl choline receptor subunit δ ; (Acrg) acetyl choline receptor subunit γ ; (Cchl) calcium channel L, type 1A3 subunit; (Adra2b) a₂ adrenoreceptor subtype 2 b; (Pdyn) prodynorphin; (Gpcr8) G-protein-coupled receptor; (Ntsr) high-affinity neurotensin receptor; (Tph) tryptophan hydroxylase; (Gabrb3) GABA β 3; (Gabrg3) GABA γ 3; (Gabra5) GABA α 5; (Tyr) tyrosinase (C, or albino) (Th) tyrosine hydroxylase; (Tat) tyrosine aminotransferase; (Htt) serotonin 5-hydroxytryptamine transporter; (Nos2) inducible nitric oxide synthase; (Ngfr) nerve growth factor receptor; (Cyp11b-1) steroid 11-b-hydroxylase; (Cyp11b-2) steroid 11-b-hydroxylase; (Grl1) glucocorticoid receptor-1; (Camk2a) calmodulin kinase 2a; (Adrb2) adrenergic receptor b2; (Mc2r) melanocortin 2 receptor.

reviewed and approved by the proper institutional committees.

ILS and ISS mice were developed by 20 generations of sib matings of LS or SS, respectively. At generation 30, ILS females were crossed with ISS males (L/S), and the reciprocal crosses (S/L) were performed to produce the F₁ generation. F₂ mice were produced by four crosses: L/S \times L/S, L/S \times S/L, S/L \times L/S, and S/L \times S/L. Mice were maintained on a 12-hr light/dark cycle and were given food and water ad libitum (Wayne diet by Teklad). Mice were tested at 55–65 days of age (trial 1) and again 7–10 days later (trial 2). A 4.1 grams/kg dose of ethanol [20% (wt/vol) solution in saline] was injected intraperitoneally between 9:00 a.m. and 1:00 p.m., 2–6 hr after the start of the light cycle. Mice were injected in the same order on both trials. Duration of LORR was determined by placing mice on their backs in a Plexiglas trough after injection. The start of LORR was considered as the minute when a mouse, after being placed on its back, was no longer able to right itself at least three times within 1 min, and the duration of LORR was the time until spontaneous righting occurred at least three times within 1 min. An LORR score of zero was assigned if no LORR was seen within 15 min after injection. Mice were excluded for leaky or subcutaneous injections. Mice were returned to

cages after testing and were subsequently sacrificed for molecular analyses.

DNA Marker Analysis

Some methods were reported previously (Markel and Johnson 1994). Primer pairs that amplify SSLPs were obtained commercially from Research Genetics (Huntsville, AL). The forward primer for each marker pair was labeled at the 5' end with [γ -³³P]ATP. A typical labeling cocktail for 40 reactions was 4.48 mM in the forward primer and contained 5.0 units of T4 polynucleotide kinase (New England Biolabs), and 1.7 ml of [γ -³³P]ATP (sp. ac. 2000 Ci/MM; Dupont/NEN) in 10 ml of kinase buffer. Reactions were incubated at 37°C for 30 min and stopped by incubation at 65°C for 10 min. Reaction cocktails (all ingredients minus template) were prepared manually and contained the components from the labeling reactions—0.25 units of Taq polymerase (Perkin Elmer Cetus)—and were 112 nM for each primer, 200 mM for each nucleotide, 2.0 mM for MgCl₂ (Promega), 10 mM for Tris-HCl at pH 8.4, and 50 mM for KCl. Aliquots (5 μ l) of the reaction cocktail were transferred using the P20 pipetting tool of the Biomek 1000 Work-

MARKEL ET AL.

station (Beckman) into each reaction tube (Perkin Elmer). Aliquots of 5 µl of target DNA at 4 ng/ml were added with the P20 pipetting tool giving 20 ng of genomic DNA in a final volume of 10 ml. PCRs were initiated by 3 min at 94°C, followed by incubation for 30 cycles of 30 sec at 94°C, 20–30 sec at 55°C, and 40–60 sec at 72°C on a Perkin Elmer Cetus 9600 Thermocycler. PCR products were diluted with the addition of 7 µl of a stop-dye solution (95% formamide, 20 mM EDTA, 0.05% bromophenol blue, 0.05% xylene cyanol in water). This solution was incubated in a 65°C heating block for 2–5 min. Samples were electrophoresed on 5% polyacrylamide denaturing gels (Sequagel; National Diagnostics) for 1.5–2 hr at 50 W. Gels were fixed to filter paper, overlayed with Saran Wrap (Dow Chemical), and dried on a Bio-Rad gel dryer (model 583) for 2 hr at 80°C. Dried gels were exposed directly to film (XAR 5, Kodak) overnight at room temperature. Autoradiographs were scored independently by at least two individuals.

Linkage Analysis

QTLs were mapped using the transformation mean LORR⁶⁷, Trial 1⁵⁴, and Trial 2⁶⁹ (Markel et al. 1996b). Genetic maps were generated using Mapmaker/EXP version 3.0. QTLs were detected in scans using MapMaker/QTL version 1.1. In virtually all cases, the genetic maps produced with the ILS × ISS F₂ mice correlated very well with those published on-line (Mouse Locus Information, <http://www.informatics.org/locus.html>).

ACKNOWLEDGMENTS

We thank E. Thomas and J. Salazar for careful animal husbandry; G. Hume and J. Costello for technical assistance; A.C. Collins, J.C. DeFries, D. Deitrich, V.G. Erwin, D.W. Fulker, A. Harris, B. Tabakoff, and J. Wehner for helpful advice and manuscript review. This work was supported by grants from the National Institutes of Health (AA-03527, AA-08940), the Denver Veteran's Administration Alcohol Research Center, by a Research Scientist Development Award to T.E.J. (AA00195), and by a training grant for P.D.M. from the National Institute of Mental Health (MH-16880). The mapping data described in this paper will be submitted to the Mouse Genome Database at the Jackson Laboratory.

The publication costs of this article were defrayed in part by payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 USC section 1734 solely to indicate this fact.

REFERENCES

- Aiken, S.P., J.L. Gleitsman, and J.J. McArdle. 1991. Tolerance to alcohol at the neuromuscular junction of long-sleep and short-sleep mice. *Alcohol* **8**: 207–209.
- Belknap, J.K. 1992. Empirical estimates of Bonferroni corrections for use in chromosome mapping studies with the BXD recombinant inbred strains. *Behav. Genet.* **22**: 677–684.
- Belknap, J.K., P. Metten, M.L. Helms, L.A. O'Toole, S. Angel-Gade, J.C. Crabbe, and T.J. Phillips. 1993. Quantitative trait loci (QTL) applications to substance abuse: Physical dependence studies with nitrous oxide and ethanol in BXD mice. *Behav. Genet.* **23**: 213–222.
- Crabbe, J.C., J.K. Belknap, and K.J. Buck. 1994a. Genetic animal models of alcohol and drug abuse. *Science* **264**: 1715–1723.
- Crabbe, J.C., J.K. Belknap, S.R. Mitchell, and L.I. Crawshaw. 1994b. Quantitative trait loci mapping of genes that influence the sensitivity and tolerance to ethanol-induced hypothermia in BXD recombinant inbred mice. *J. Pharmacol. Exp. Ther.* **49**: 456–461.
- DeFries, J.C., J.R. Wilson, V.G. Erwin, and D.R. Petersen. 1989. LSXSS recombinant inbred strains of mice: Initial characterization. *Alcoholism: Clin. Exp. Res.* **13**: 196–200.
- Devor, E.J. and C.R. Cloninger. 1989. Genetics of alcoholism. *Annu. Rev. Genet.* **23**: 19–36.
- Deitrich, W., H. Katz, S.E. Lincoln, H.-S. Shin, J. Friedman, N. Dracopoli, and E.S. Lander. 1992. A genetic map of the mouse suitable for typing intraspecific crosses. *Genetics* **131**: 423–447.
- Dudek, B.C. and M.E. Abbott. 1984. A biometrical genetic analysis of ethanol response in selectively bred long-sleep and short-sleep mice. *Behav. Genet.* **14**: 1–19.
- Erwin, V.G., W.D.W. Heston, G.E. McClearn, and R.A. Deitrich. 1976. Effect of hypnotics on mice genetically selected for sensitivity to ethanol. *Pharmacol. Biochem. Behav.* **4**: 679–683.
- Erwin, V.G., R. Radcliffe, B. Hinkle, and B.C. Jones. 1996. Genetic-based difference in neuropeptidyl receptor levels and receptors in brains of LSXSS mice. *Peptides* **14**: 821–828.
- Falconer, D.S. 1989. *Introduction to quantitative genetics*, 3rd ed. Longman Scientific and Technical, John Wiley and Sons, Inc., New York, NY.
- Gage, P.W., R.N. McBurney, and G.T. Schneider. 1975. Effects of some aliphatic alcohols on the conductance change caused by a quantum of acetylcholine at the toad end-plate. *J. Physiol.* **244**: 409–429.
- Heston, W.D.W., V.G. Erwin, S.M. Anderson, and H. Robbins. 1974. A comparison of the effects of alcohol in mice selectively bred for differences in ethanol sleep-time. *Life Sci.* **14**: 365–370.
- Howerton, T.C., M.F. O'Connor, and A.C. Collins. 1983. Differential effects of long-chain alcohols in long- and short-sleep mice. *Psychopharmacology* **79**: 313–317.
- Johnson, T.E., J.C. DeFries, and P.D. Markel. 1992. Mapping quantitative trait loci for behavioral traits in the mouse. *Behav. Genet.* **22**: 635–653.
- Lander, E.S. and L. Kruglyak. 1995. Genetic dissection of complex traits: Guidelines for interpreting and reporting linkage results. *Nature Genet.* **11**: 241–247.

QTLs FOR ALCOHOL SENSITIVITY

- Lander, E.S. and N.J. Schork. 1994. Genetic dissection of complex traits. *Science* **265**: 2037–2048.
- Lander, E.S., P. Green, J. Abrahamson, A. Barlow, M. Daley, S. Lincoln, and L. Newburg. 1987. MAPMAKER: An interactive computer package for constructing primary genetic linkage maps of experimental and natural populations. *Genomics* **1**: 174–181.
- Markel, P.D. and R.P. Corley. 1994. A multivariate-analysis of repeated measures: Linkage of the albino locus (Tyr) to a QTL influencing ethanol-induced anesthesia in mice. *Psych. Genet.* **4**: 205–210.
- Markel, P.D. and T.E. Johnson. 1994. Initial characterization of STS markers in the LSXSS series of recombinant inbred strains. *Mamm. Genome* **5**: 199–202.
- Markel, P.D., J.C. DeFries, and T.E. Johnson. 1995a. Ethanol-induced anesthesia in inbred strains of long-sleep and short-sleep mice: A genetic analysis using repeated measures. *Behav. Genet.* **25**: 67–73.
- _____. 1995b. Use of repeated measures in an analysis of ethanol-induced righting reflex in inbred long-sleep and short-sleep mice. *Alcoholism Clin. Exp. Res.* **19**: 299–304.
- Markel, P.D., B. Bennett, M.A. Beeson, L. Gordon, V.J. Simpson, and T.E. Johnson. 1996a. Strain distribution patterns for genetic markers in the LSXSS recombinant-inbred series. *Mamm. Genome* **7**: 408–412.
- Markel, P.D., D.W. Fulker, B. Bennett, R.P. Corley, J.C. DeFries, V.G. Erwin, and T.E. Johnson. 1996b. Quantitative trait loci for ethanol sensitivity in the LSXSS recombinant inbred strains: Interval mapping *Behav. Genet.* **26**: 447–458.
- McClearn, G.E. and R. Kakihana. 1981. Selective breeding for ethanol sensitivity: Short-sleep and long-sleep mice. In *Development of animal models as pharmacogenetic tools* (ed. G.E. McClearn, R.A. Deitrich, and V.G. Erwin), pp. 147–159. DHHS Publication No. [ADM] 81–113, U.S. Government Printing Office, Washington, D.C.
- McClearn, G.E., R. Plomin, G. Gora-Maslak, and J.C. Crabbe. 1992. The gene chase in behavioral science. *Psych. Science* **2**: 222–229.
- Murrell, R.D., M.S. Braun, and D.A. Haydon. 1991. Actions of n-alcohols on nicotinic acetylcholine receptor channels in cultured rat myotubes. *J. Physiol.* **437**: 431–448.
- Neumann, P.E. 1992. Inference in linkage analysis of multifactorial traits using recombinant inbred strains of mice *Behav. Genet.* **22**: 665–676.
- Newlin, D.B. and J.B. Thomson. 1990. Alcohol challenge with sons of alcoholics: A critical review and analysis. *Psych. Bull.* **3**: 383–402.
- Pollock, V.E. 1992. Meta-analysis of subjective sensitivity to alcohol in sons of alcoholics. *Am. J. Psychiatry* **149**: 1534–1538.
- Rodriguez, L.A., R. Plomin, D.A. Blizzard, B.C. Jones, and G.E. McClearn. 1995. Alcohol acceptance, preference, and sensitivity in mice. II. Quantitative trait loci mapping analysis using BXD recombinant inbred strains. *Alcoholism Clin. Exp. Res.* **19**: 367–373.
- Saito, T., J.M. Lee, P.L. Hoffman, and B. Tabakoff. 1987. Effects of chronic ethanol treatment of the beta-adrenergic receptor-coupled adenylate cyclase system of mouse cerebral cortex. *J. Neurochem.* **48**: 1817–1822.
- Schuckit, M.A. 1985. Ethanol-induced changes in body sway in men at high alcoholism risk. *Arch. Gen. Psychiatry* **42**: 375–379.
- _____. 1994. Low level of response to alcohol as a predictor of future alcoholism. *Am. J. Psychiatry* **151**: 184–189.
- Smolen, A. and T.N. Smolen. 1989. Reproducibility of ethanol elimination rates in Long-sleep and Short-sleep mice. *J. Stud. Alcohol* **50**: 519–524.
- Smolen, A., M.J. Marks, T.N. Smolen, and A.C. Collins. 1986. Dose and route of administration alter the relative elimination of ethanol by Long-sleep and Short-sleep mice. *Alcoholism: Clin. Exp. Res.* **10**: 198–204.
- Wand, G.S. and M.A. Levine. 1991. Hormonal tolerance to ethanol is associated with decreased expression of the GTP-binding protein, G_s, and adenylyl cyclase activity in ethanol-treated LS mice. *Alcoholism Clin. Exp. Res.* **15**: 705–710.

Received September 30, 1996; accepted in revised form December 17, 1996.



Confirmation of quantitative trait loci for ethanol sensitivity in long-sleep and short-sleep mice.

P D Markel, B Bennett, M Beeson, et al.

Genome Res. 1997 7: 92-99

Access the most recent version at doi:[10.1101/gr.7.2.92](https://doi.org/10.1101/gr.7.2.92)

References This article cites 36 articles, 3 of which can be accessed free at:
<http://genome.cshlp.org/content/7/2/92.full.html#ref-list-1>

Creative Commons License This article is distributed exclusively by Cold Spring Harbor Laboratory Press for the first six months after the full-issue publication date (see <http://genome.cshlp.org/site/misc/terms.xhtml>). After six months, it is available under a Creative Commons License (Attribution-NonCommercial 3.0 Unported License), as described at <http://creativecommons.org/licenses/by-nc/3.0/>.

Email Alerting Service Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article or [click here](#).

To subscribe to *Genome Research* go to:
<http://genome.cshlp.org/subscriptions>
