

# CD4 T Cells Treated with gp120 Acquire a CD45RO+/CD45RA+ Phenotype

Sergey A. Trushin<sup>1,2</sup>, Gary D. Bren<sup>1</sup>, Andrew D. Badley<sup>\*,1,2</sup>

<sup>1</sup>Division of Infectious Diseases, <sup>2</sup>Program in Translational Immunovirology and Biodefense, Mayo Clinic, Rochester, MN 55905, USA

**Abstract:** HIV-infected patients exhibit quantitative and qualitative defects in CD4 T cells, including having increased numbers of CD4+CD45RO+/CD45RA+ T cells, although it remains unclear how these cells arise. Here we demonstrate that gp120 treatment of activated but not resting primary human CD4 T cells decreases number of cells with single positive CD45RO+/CD45RA- effector memory phenotype while proportionally increasing the subset of cells with double positive CD45RO+/CD45RA+ mixed phenotype. We found that double positive CD45RO+/CD45RA+CD4 T cells preferentially undergo apoptosis while single positive CD45RO+/CD45RA- and CD45RO-/CD45RA+ do not. Blocking gp120-CD4 interaction with sCD4 or inhibition Lck activity reverses gp120 induced increase in double positive CD45RO+/CD45RA+CD4 T cells and subsequently diminishes the apoptosis of double positive CD45RO+/CD45RA+ cells. Altogether these data indicate that gp120 ligation of the CD4 receptor increases the number of double positive CD45RO+/CD45RA+ CD4 T cells which subsequently undergo apoptosis in a CD4 dependent manner.

## INTRODUCTION

One hallmark of HIV pathogenesis is a decline of CD4 T cell number that results from the death of both HIV-1 infected CD4 T cells, as well as uninfected cells. Both HIV expressed proteins, as well as immune activation, contribute to this T cell death [1]. Immune activation leads to an increase in T cell turnover from enhanced proliferation which in turn is associated with high rates of apoptosis [2, 3]. The magnitude of immune activation correlates with the level of HIV viremia [4]. Increased expression of immune activation markers (HLA-DR+, CD38+, CD45RO+, and CD95) also correlates with higher apoptosis rates of CD4 T cells [5]. Importantly, suppression of viral replication with HAART treatment reduces immune activation [6, 7], normalizes expression of activation markers, and decreases CD4 T cell apoptosis [8-10].

As a result of chronic immune activation, central memory and naïve CD4 T cells are constantly recruited into the effector pool [11, 12] resulting in dramatic changes in populations of naïve, effector and central memory CD4 T cells [13]. The mechanisms of how the ratio of these CD4 T cells is altered during HIV infection are not completely understood.

In particular, chronic HIV infection results in an increased number of CD4 T cells with a peculiar double positive CD45RO+/CD45RA+/CD25+CD4+ phenotype, and this subset is further increased following intermittent IL-2 therapy [14]. The origin of this subset was previously ascribed to be transition phase of naïve CD45RO-/CD45RA+CD4 T cells transitioning to a memory CD45RO+/CD45RA-CD4 T cell phenotype [15]. Furthermore, HAART with IL-2 therapy selectively increases the number of activated CD4 T cell expressing CD45RO+/CD25+ [16]. Here we describe a

novel effect of HIV-1 gp120 signaling through CD4 resulting in altered CD45 isoform expression by CD4 T cells.

## MATERIALS AND METHODOLOGY

### Cell Culture and Reagents

This protocol was reviewed and approved by the Mayo institutional review board. CD4+ T cells were isolated from the blood of healthy volunteer blood donors by using RosetteSep CD4 enrichment cocktail (StemCell Technologies, Vancouver, British Columbia, Canada), producing 98% pure CD4+ T cells as determined by flow cytometry. CD4 T cells were maintained in RPMI 1640 supplemented with 10% fetal bovine serum (Invitrogen, Carlsbad, CA), 2 mM L-glutamine, and antibiotics (penicillin 100 U/ml, streptomycin 100 µg/ml) at  $0.5 \times 10^6$  cells/ml. CD4+ T cells used in the various experiments were stimulated with PHA (1 µg/ml) for 24 hours, and then cells were washed twice with RPMI 1640 and maintained in media supplemented with 50 U/ml of IL-2 for 48-72 hours. CD4 T cells were incubated with HIV-1 X4 gp120IIIB (Immuno Diagnostics, Inc. Woburn, MA) or gp120 IIIB pretreated with soluble CD4 (1:2 ratio) (Immuno Diagnostics, Inc. Woburn, MA) at concentrations of 1 µg/ml/ $2 \times 10^6$  cells for 24 hours at 37°C. AMD3100 (NIH AIDS Research and Reference program) was used at 2 µM for 30 minutes at 37°C. PP2 was purchased from CalBiochem (La Jolla, CA). Anti-CD4-PE, anti-CD25-FITC, anti-CD69-PE, anti-CD4PerCP, anti-CD62L-PE, anti-CD45RO-FITC, anti-CD45RA-PE-Cy-7, anti-HLA-DR-PE, AnnexinV-Cy-5, AnnexinV-APC, IgG1κ-PE-Cy7, IgG2a-FITC, IgG1κ-PE and propidium iodine were purchased from BD Biosciences (San Jose, CA).

### Cell Death Analysis and Flow Cytometry

CD4 T cells were untreated or pre-incubated with specific inhibitors and stimulated with either BSA or with soluble gp120IIIB (10µg/ml) overnight. The following day, cell death was analyzed by staining with AnnexinV-Cy-5 and propidium iodine following the manufacturer's instructions

\*Address correspondence to this author at the Division of Infectious Diseases, Mayo Clinic, 200 First Street SW, Rochester, MN 55905, USA; Tel: 507-266-5065; Fax: 507-284-3757; E-mail: badley.andrew@mayo.edu

from BD Biosciences (San Jose, CA). All experiments were performed at least three times.

T cell phenotyping studies were performed by using four-color Flow analysis on FACSCanto cytometer and using FACSDiva 6.0 software. Briefly,  $2 \times 10^6$  cells were resuspended in 200  $\mu$ l of PBS+0.5%BSA, stained with the indicated primary conjugated antibodies for 20 minutes (anti-CD45RO-FITC, anti-CD45RA-PE-Cy-7, anti-CD27 APC and anti-CD62L-PE), washed, fixed and then analyzed. In some cases, for analysis of cell death, cells were stained in binding buffer (140 mM NaCl, 10 mM HEPES/NaOH (pH 7.4), 2.5 mM CaCl<sub>2</sub>) as described above except that anti-CD62L-PE was substituted with AnnexinV-PE.

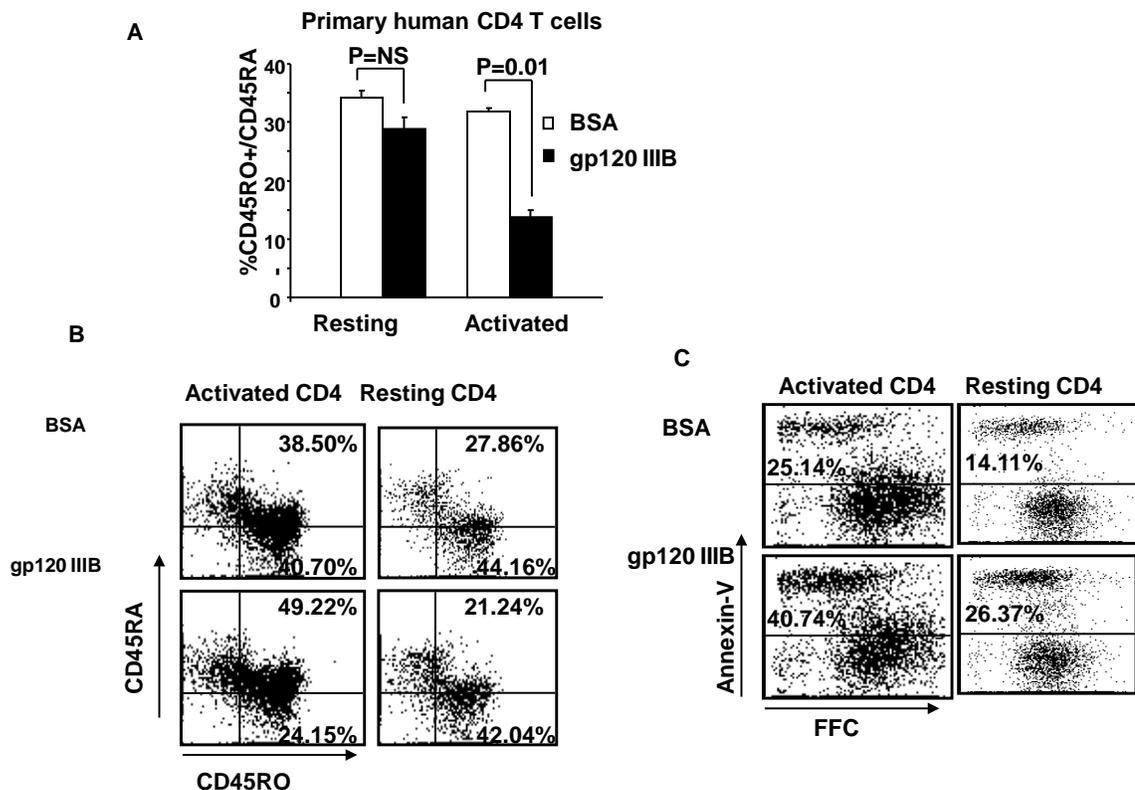
## RESULTS

### Gp120 Decreases CD45RO+ Memory Phenotype in Activated But Not Resting CD4 T cells

We and others have previously demonstrated that gp120 induces death of resting CD4 T cell in a CXCR4-p38 dependent manner [17]. However, the effect of gp120 on activated CD4 T cells is largely unknown. To examine the effect of gp120, resting (CD4+CD25-CD69-HLA-DR-) and activated (CD4+CD25+CD69+HLA-DR+) primary human CD4 T cells were treated with gp120 IIIB (10  $\mu$ g/ml) for 24 hours and then CD4 T cells were analyzed for CD45RO and CD45RA expression by flow cytometry. As shown in Fig. (1A), gp120 treatment results in a 50% decrease in the subset of single positive CD45RO+/CD45RA- (31.85  $\pm$  0.63% with

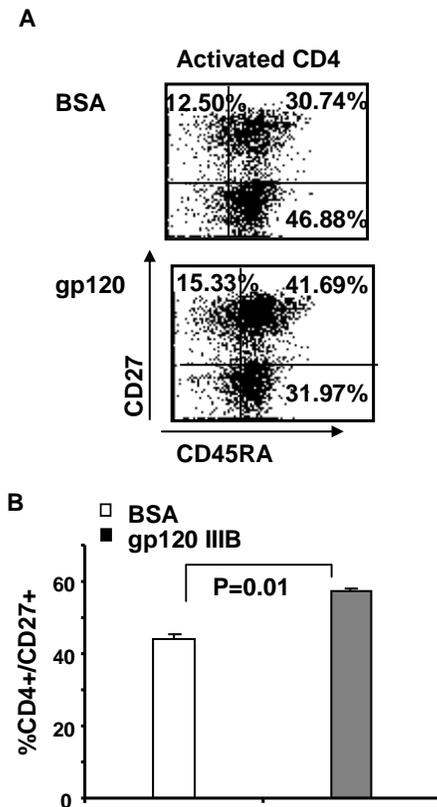
BSA, 13.80  $\pm$  3.73% with gp120,  $p=0.01$ ) memory cells when activated but not resting CD4 T cells are treated with gp120 (34.25  $\pm$  1.1% with BSA, 28.95  $\pm$  1.96% with gp120,  $p=0.08$ ). In parallel, there is a proportional increase in the subset of double positive CD45RO+/CD45RA+ mixed phenotype CD4+ T cells (Fig. 1B) as well as an increase in the proportion of Annexin-V positive CD4 T cells (Fig. 1C). While the increased apoptosis of resting cells following gp120 is due to CXCR4 signaling [17], the mechanisms by which gp120 causes death in activated cells are unknown.

The finding that gp120 treatment of activated cells increases the lymphoid homing marker, CD27 (13.8% of total CD4 T cells) preferentially in double positive CD45RO+/CD45RA+ cells suggests that these cells might be the specific cell type which dies following gp120 treatment of activated CD4 T cells (Fig. 2A, B). We therefore measured absolute number and proportion of Annexin-V positivity of double positive CD45RO+/CD45RA+, single positive CD45RO-/CD45RA+ and CD45RO+/CD45RA- subsets by using four color flow cytometry analysis. First we observed that CD45RO+RA- proportion decreased (39.42  $\pm$  3.44% with BSA, 27.31  $\pm$  6.38% with gp120) while CD45RO+RA+ population increased (35.97  $\pm$  2.70% with BSA, 49.42  $\pm$  10.05% with gp120), suggesting that the former transitioned to the latter (Fig. 3A). Next we observed that only double positive CD45RO+/CD45RA+ CD4 T cell subset contain a significant proportion of Annexin positive cells (~40%) in contrast to single positive CD45RO+/CD45RA- and CD45RO-/CD45RA+ subsets that do not (Fig. 3B). Therefore, gp120 signaling both



**Fig. (1). Gp120 decreases CD45RO+/CD45RA- memory phenotype in activated but not resting primary human CD4 T cells. (A and B)** CD4+ T cells (98% pure) were stimulated with PHA for 24 hours or left rested, and treated with either p120 (IIIB) (1  $\mu$ g/ml) or BSA control (1  $\mu$ g/ml) and analyzed for CD45RO and CD45RA (gated on CD4 population) by flow cytometry. **(C)** Resting and activated primary human CD4 T cells were treated with gp120 (IIIB) as above and analyzed for Annexin-V positive cells. Data is representative of three independent experiments.

activates cells towards a double positive phenotype and subsequently promotes their apoptotic death. Of interest, these Annexin positive CD45R0+/CD45 RA+ cells are also CD27+. Altogether these results indicate that gp120 treatment of activated CD4 T cells result in decrease memory CD45R0+/CD45RA- CD4 T cells due to accumulation of CD45R0+/CD45RA+ /CD27+ T cell subset that subsequently and selectively undergo apoptosis.



**Fig. (2).** Gp120 increases subset of CD27+/CD45RO+/CD4+ T cells. (A) Activated primary human CD4 T cells (98% pure) were treated with gp120 IIIIB and the expression of CD27 and CD45RA were analyzed by flow cytometry. (B) Pooled data from three independent experiments. P values were determined by Student's paired t test.

#### Gp120 Stimulated Accumulation of CD45RO+/CD45RA+ CD4 T Cells Requires CD4 and Lck But Not CXCR4

To investigate whether gp120 effect on CD45R0+/CD45RA+ CD4 T cell is due to ligation to CD4 or CXCR4, we used either soluble CD4 or the CXCR4 inhibitor, AMD3100. Independent experiments verified the activity of AMD3100 by blocking SDF-induced chemotaxis (data not shown). As shown in Fig. (4A, B), inhibition of CD4 ligation by sCD4 [17] blocks the increase in double positive CD45R0+/CD45RA+ memory cells. Consistent with this observation, inhibition of Lck activity with PP2 inhibitor also blocks increase in CD45R0+/CD45RA+ CD4 T cell subset. Conversely inhibition of gp120-CXCR4 interaction with AMD3100 [17] does not block completely gp120 mediated increase of CD45R0+/CD45RA+ CD4 T cells. Therefore, our results demonstrate that gp120 ligation to CD4 and subsequent activation of Lck results in accumulation of dou-

ble positive CD45R0+/CD45RA+ memory CD4 T cells which subsequently and selectively undergo apoptosis in a CD4 dependent and CXCR4 independent manner.

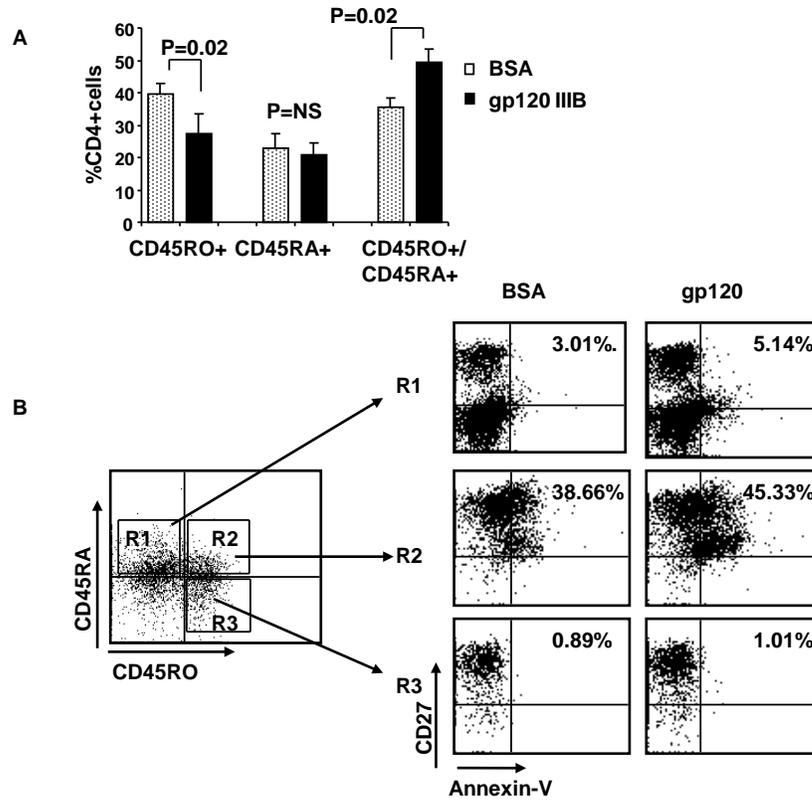
#### DISCUSSION

In the current report we have shown that gp120 treatment of activated CD4 T cells decreases the subset of single positive CD45R0+/CD45RA- CD4 T cells and increases the subset of double positive CD45R0+/CD45RA+ CD4 T cells which selectively undergo apoptosis. These results imply the gp120 may play a role in depletion of memory CD45R0+/CD45RA- CD4 T during HIV.

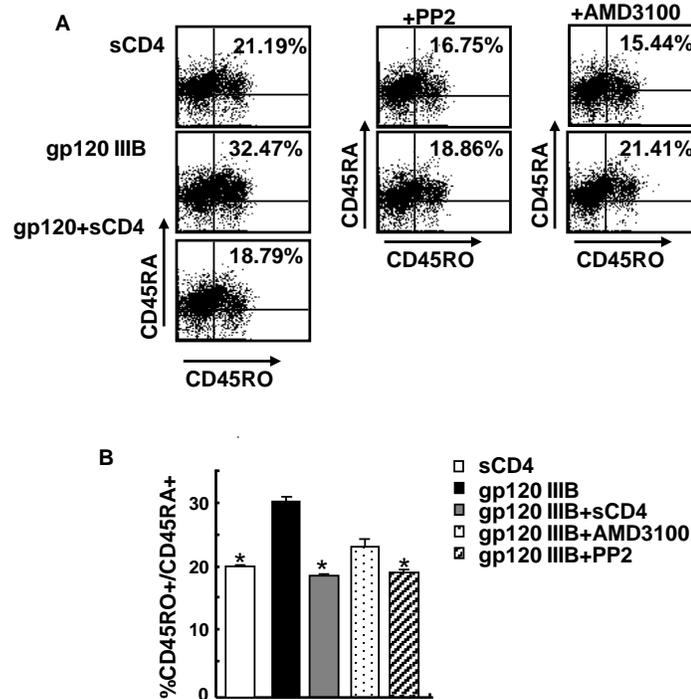
Previously, the CD45R0+/CD45RA+ subset of CD4 T cells was characterized as a transition from naïve to effector phenotype during CD4 priming [15, 18] with increased expression of activation markers [19, 20]. The generation of this subset can occur in two ways: (i) the transition of recently activated of CD45RA+ T naïve cells to CD45R0+/CD45RA+ phenotype [15, 21] and; (ii) re-expression of CD45RA isoform in the absence of Ag stimulation [15]. The fact that only few CD45R0+/CD45RA+ cells express markers of recently activated cells [19, 20] and upregulation of CD45RA+ (up to 10%) by CD45R0+/CD45RA- cells [15] suggest that gp120 rather upregulates CD45RA+ on single positive CD45R0+/CD45RA- memory CD4 T cells rather than promoting CD45RO expression on naïve CD4 T cells. Indeed, increase of double positive CD45R0+/CD45 RA+ CD4 T cells is followed by proportional decrease in CD45R0+/CD45RA- memory but not decrease in naïve CD45RO-/CD45RA+ CD4 T cells.

Others have shown that PBMC stimulated with staphylococcal enterotoxin A (SEA) in the presence of V3-derived gp120 peptides results in increase of CCR5+/CXCR4+/CD45RO+ CD4 T cells followed by increased levels of apoptosis [22]. Our findings are in good agreement with this observation: first, both results demonstrate the gp120 mediated increase in transient state from naïve to activated CD4 T cells; second, both studies observe the increased apoptosis of activated CD4 T cells in the presence of gp120 or V3-derived peptide. However, our results highlight that gp120 priming of previously activated CD4 T cells reverses single positive CD45R0+/CD45RA- effector phenotype to double positive CD45R0+ /CD45RA+ transient phenotype.

The observation that double positive CD45R0+/CD45RA+ preferentially undergo apoptosis presents one potential scenario of how inappropriately activated CD4 T cells die during HIV disease, and why IL-2 augments this process [16]. It is of further interest that gp120 treatment results in apoptosis of CD27+/CD45R0+/CD45RA+ cells. In fact, CD27 expression is rapidly upregulated following TCR stimulation [23] and CD27 signaling is essential for survival of Ag-primed CD4 T cells [24]. Therefore, either the lack of antigen specific signaling or the lack of CD27 signaling may explain the increased apoptosis of double positive CD45R0+/CD45RA+ following gp120 treatment. Finally, the observation that CD4 ligation by gp120 and subsequent activation of Lck is required for generation of double positive CD45R0+/CD45RA+ CD4 T cells is of relevance and suggests that inhibitors of CD4-gp120 interactions may reverse the expansion of double positive CD45R0+/CD45RA+ population and subsequent increased apoptosis during HIV infection.



**Fig. (3).** Gp120 increases subset of double positive CD45RA+/CD45RO+ CD4 T cells that undergo apoptosis. (A) Activated primary human CD4 T cells were treated with gp120 IIIIB and CD45RA+/CD45RO-, CD45RA-/CD45RO+ and double positive CD45RA+/CD45RO+ cells were quantified by flow cytometry. (B) Same as Fig. (3A) except that CD27 expression and Annexin-V positivity were measured by four color flow cytometry. The data is representative of three independent experiments.



**Fig. (4).** CD4 and Lck are required for gp120 mediated increase of double positive CD45RA+/CD45RO+ CD4 T cells. (A) Activated primary human CD4 T cells were left untreated or were pre-treated with either Lck inhibitor, PP2 (5µM) or CXCR4 inhibitor, AMD3100 (2µM) or sCD4 (2µg/ml) followed by gp120. Then number of single positive CD45RA+/CD45RO-, CD45RA-/CD45RO+ and double positive CD45RA+/CD45RO+ cells were analyzed by flow cytometry. (B) The data represents three independent experiments as described above. P values were determined by Student's paired t test. \* = P < 0.05 compared to gp120 alone.

## ACKNOWLEDGEMENTS

**Grant Support:** Dr. Andrew Badley is supported by the following grants: NIH R01 AI62261, R01 AI40384 and a Burroughs Wellcome Award ID #1005160.

## REFERENCES

- [1] Lum J, Badley AD. In: Badley AD, Ed. Cell Death During HIV Infection. Boca Raton: Taylor and Francis Press 2006; pp. 109-126.
- [2] Douek DC, Betts MR, Hill BJ, *et al.* Evidence for increased T cell turnover and decreased thymic output in HIV infection. *J Immunol* 2001; 167: 6663-8.
- [3] Hellerstein M, Hanley MB, Cesar D, *et al.* Directly measured kinetics of circulating T lymphocytes in normal and HIV-1-infected humans. *Nat Med* 1999; 5: 83-9.
- [4] Perelson AS, Neumann AU, Markowitz M, Leonard JM, Ho DD. HIV-1 dynamics *in vivo*: virion clearance rate, infected cell life-span, and viral generation time. *Science* 1996; 271: 1582-6.
- [5] Gougeon ML, Lecoer H, Dulioust A, *et al.* Programmed cell death in peripheral lymphocytes from HIV-infected persons: increased susceptibility to apoptosis of CD4 and CD8 T cells correlates with lymphocyte activation and with disease progression. *J Immunol* 1996; 156: 3509-20.
- [6] Hazenberg MD, Stuart JW, Otto SA, *et al.* T-cell division in human immunodeficiency virus (HIV)-1 infection is mainly due to immune activation: a longitudinal analysis in patients before and during highly active antiretroviral therapy (HAART). *Blood* 2000; 95: 249-55.
- [7] Lempicki RA, Kovacs JA, Baseler MW, *et al.* Impact of HIV-1 infection and highly active antiretroviral therapy on the kinetics of CD4+ and CD8+ T cell turnover in HIV-infected patients. *Proc Natl Acad Sci USA* 2000; 97: 13778-83.
- [8] Badley AD, Dockrell DH, Algeciras A, *et al.* *In vivo* analysis of Fas/FasL interactions in HIV-infected patients. *J Clin Invest* 1998; 102: 79-87.
- [9] Bohler T, Walcher J, Holzl-Wenig G, *et al.* Early effects of antiretroviral combination therapy on activation, apoptosis and regeneration of T cells in HIV-1-infected children and adolescents. *AIDS* 1999; 13: 779-89.
- [10] Giorgi JV, Majchrowicz MA, Johnson TD, Hultin P, Matud J, Detels R. Immunologic effects of combined protease inhibitor and reverse transcriptase inhibitor therapy in previously treated chronic HIV-1 infection. *AIDS* 1998; 12: 1833-44.
- [11] Hazenberg MD, Hamann D, Schuitemaker H, Miedema F. T cell depletion in HIV-1 infection: how CD4+ T cells go out of stock. *Nat Immunol* 2000; 1: 285-9.
- [12] Silvestri G, Feinberg MB. Turnover of lymphocytes and conceptual paradigms in HIV infection. *J Clin Invest* 2003; 112: 821-4.
- [13] McCune JM. The dynamics of CD4+ T-cell depletion in HIV disease. *Nature* 2001; 410: 974-9.
- [14] Sereti I, Martinez-Wilson H, Metcalf JA, *et al.* Long-term effects of intermittent interleukin 2 therapy in patients with HIV infection: characterization of a novel subset of CD4(+)/CD25(+) T cells. *Blood* 2002; 100: 2159-67.
- [15] Hamann D, Baars PA, Hooibrink B, van Lier RW. Heterogeneity of the human CD4+ T-cell population: two distinct CD4+ T-cell subsets characterized by coexpression of CD45RA and CD45RO isoforms. *Blood* 1996; 88: 3513-21.
- [16] Sereti I, Herpin B, Metcalf JA, *et al.* CD4 T cell expansions are associated with increased apoptosis rates of T lymphocytes during IL-2 cycles in HIV infected patients. *AIDS* 2001; 15: 1765-75.
- [17] Trushin SA, Algeciras-Schimmich A, Vlahakis SR, *et al.* Glycoprotein 120 binding to CXCR4 causes p38-dependent primary T cell death that is facilitated by, but does not require cell-associated CD4. *J Immunol* 2007; 178: 4846-53.
- [18] Picker LJ, Treer JR, Ferguson-Darnell B, Collins PA, Buck D, Terstappen LW. Control of lymphocyte recirculation in man. I. Differential regulation of the peripheral lymph node homing receptor L-selectin on T cells during the virgin to memory cell transition. *J Immunol* 1993; 150: 1105-21.
- [19] Baars PA, Maurice MM, Rep M, Hooibrink B, van Lier RA. Heterogeneity of the circulating human CD4+ T cell population. Further evidence that the CD4+CD45RA-CD27- T cell subset contains specialized primed T cells. *J Immunol* 1995; 154: 17-25.
- [20] Prince HE, York J, Jensen ER. Phenotypic comparison of the three populations of human lymphocytes defined by CD45RO and CD45RA expression. *Cell Immunol* 1992; 145: 254-62.
- [21] Wallace DL, Beverley PC. Phenotypic changes associated with activation of CD45RA+ and CD45RO+ T cells. *Immunology* 1990; 69: 460-7.
- [22] Porichis F, Vlata Z, Hatzidakis G, Spandidos DA, Krambovitis E. HIV-1 gp120/V3-derived epitopes promote activation-induced cell death to superantigen-stimulated CD4+/CD45RO+ T cells. *Immunol Lett* 2007; 108: 97-102.
- [23] Watts TH. TNF/TNFR family members in costimulation of T cell responses. *Annu Rev Immunol* 2005; 23: 23-68.
- [24] Hendriks J, Xiao Y, Borst J. CD27 promotes survival of activated T cells and complements CD28 in generation and establishment of the effector T cell pool. *J Exp Med* 2003; 198: 1369-80.

Received: February 26, 2009

Revised: March 6, 2009

Accepted: March 12, 2009

© Trushin *et al.*; Licensee Bentham Open.

This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted, non-commercial use, distribution and reproduction in any medium, provided the work is properly cited.