



## **Sesquiterpene Lactones of Iranian Compositae Family (*Astraceae*); Their Chemical Constituents and Anti-plasmodial Properties of Tehranolide (A.Review)**

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<http://dx.doi.org/10.13005/ojc/330506>

(Received: May 06, 2017; Accepted: June 30, 2017)

### **ABSTRACT**

Sesquiterpene lactones constitute a large and diverse group of biologically active plant chemical that have been identified in the several plant families. The pharmacological properties of interest involving the sesquiterpene lactones are their antibacterial, antifungal and anti-plasmodial activities. Indeed, assessing the biological activities of the sesquiterpene lactones found in plants and their essential oil is of great medicinal importance because they could potentially be utilized as therapeutic agents for the treatment of such infections.

**Keywords:** Iranian Compositae Family (*Astraceae*), Germacranolides, Eudesmanolides, Guaianolides, Elemanolides, Constituents and Biological Activities.

### **INTRODUCTION**

Sesquiterpene lactones are one of the most prevalent and biologically significant classes of secondary metabolite present, and as such have subject to a number of studies.

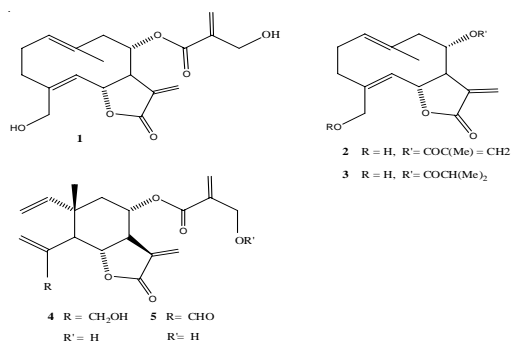
Sesquiterpene lactones are a group of secondary metabolites found across the plant kingdom comprising a large group of over 5500 known compounds<sup>1</sup> being most prevalent in the family *Astraceae*. Sesquiterpenoids are typically located in laticifers, which are specialized secretory

cells in most of the *Astraceae*, but can also be found within the vacuoles of other cell types in the plant, specifically when produced in response to biotic stresses. They are one of the main constituents of latex in latex producing plants, and they are frequently potent antimicrobial agents as well as antifeedants to chewing insects and birds. They also have a range of other effects such as allelopathy, stimulation of germination in the parasitic plant *Orobanchae*<sup>2</sup>.

In fact, The Iranian compositae (*Astraceae*) family has yielded a considerable amount of new, interesting sesquiterpene lactones.

**Chemical constituents*****Onopordon leptolepis* DC.**

*Onopordon leptolepis* DC. Growing in Iran has not been investigated before. The aerial parts also contain Onopordopicrin 1, and two new germacranolides (2 and 3), closely related to 2<sup>3</sup>.

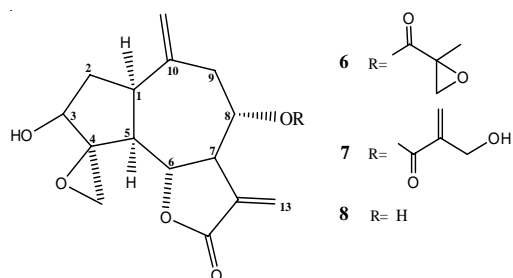


**Fig. 1. Chemical structures of Sesquiterpene lactones from *Onopordon leptolepis* DC.1-5.**

The investigation of the polar fractions of the aerial parts of *Onopordon leptolepis* afforded two new elemnolides, their structures being elucidated by spectroscopic methods and by partial synthesis starting with Onopordopicrin<sup>4</sup>.

**A Guaianolide from *Jurinea carduiformis* Boiss.**

The aerial parts of *J. carduiformis*. afforded, in addition to Repin(6)<sup>5</sup> and Janerin(7)<sup>7</sup>, small amounts of a further lactone 8, the structure of which was deduced from the H<sup>1</sup>-NMR data especially by comparison with the spectra of 6 and 7<sup>7</sup>.

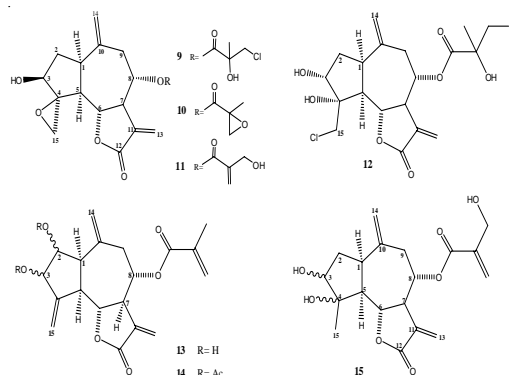


**Fig. 2. Chemical structures of Sesquiterpene lactones from *Jurinea carduiformis* Boiss.6-8**

**Guaianolides from *Acroptilon repens* DC.**

The aerial part of *A. repens* (*Centaurea picris*) DC. has been investigated several times<sup>8</sup>. Two guaianolides were isolated, chlorohyssopifolin C (9)<sup>8</sup> and Repin (10)<sup>9</sup>. A reinvestigation afforded, in addition to these lactones, Janerin (11)<sup>7</sup>, chlorohyssopifolin A (12)<sup>10</sup> and two other lactones, which are the closely

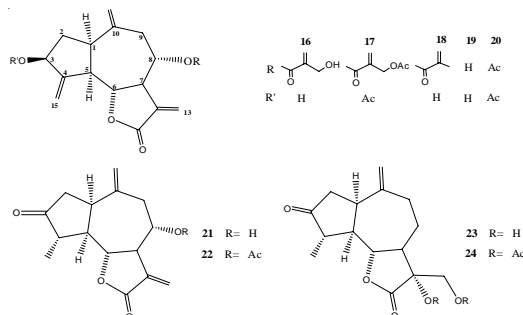
related guaianolides 13 and 15. The structures followed from the H<sup>1</sup>-NMR data, especially if compared with those of 10-12. Acetylation of 13 gave the diacetate 14; its H<sup>1</sup>-NMR data showed that the stereochemistry at C-5, through C-8 was the same as that of 9-12, which the presence of a C-4 methylene group was indicated by two broadened signals at  $\delta$  5.68 and 5.43 ppm in the spectra of 13<sup>11</sup>.



**Fig. 3. Chemical structures of Sesquiterpene lactones from *Acroptilon repens* DC.9-15**

**Guaianolides from *Centaurea behen* L.**

*Centaurea behen* L. native in Iran had not been investigated chemically. The aerial parts of this plant afforded several sesquiterpene lactones, the guaianolides cynaropicrin (16)<sup>12</sup>, arguerin B (18)<sup>13</sup>, desacylcynaropicrin (19)<sup>14</sup>, grosshemin (21)<sup>15</sup> and minor amounts of the ketone 23, which is closely related to solstitialin A, the absolute configuration of which had been established<sup>16</sup>. Structure 23 could only be isolated as its diacetate 24, which still was mixed with the acetate of 21. The latter, however, could be separated from 24 after transformation to the corresponding pyrazoline derivative. The structures of 16, the corresponding



**Fig. 4. Chemical structures of Sesquiterpene lactones from *Centaurea behen* L.16-24**

diacetate 17, 18, 19 and 21 were elucidated by their  $H^1$ -NMR data<sup>17</sup>.

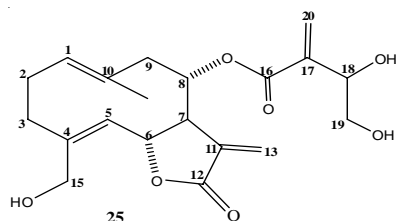


Fig. 5. Chemical structure of Sesquiterpene lactone from *Centaurea brugueriana* DC. 25.

### Chemical Constituents of *Centaurea brugueriana* DC.

Cinicin a germacranolide has been isolated from chloroform extract of the aerial parts of *Centaurea brugueriana* DC. (Compositae)<sup>18</sup>.

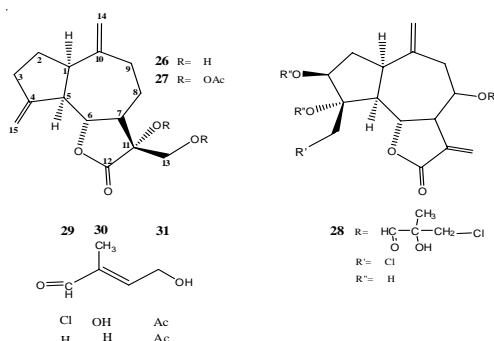


Fig. 6. Chemical structure of Sesquiterpene lactone from *Centaurea imperialis* Hausskn. ex Bomm. 26-31.

### *Centaurea imperialis*

The aerial parts of *Centaurea imperialis* afforded three new guaianolides, 3-desoxysolstitialin A and two derivatives of centaurepensin<sup>19</sup>.

### Guaianolides from *Centaurea kandavanensis* Wagenitz.

From the large genus *Centaurea* (Compositae) numerous different types of constituents, especially guaianolides, have been reported<sup>20</sup>. We now have investigated a species which grows on the mountains near Kandavan.

The polar fraction afforded two crystalline compounds, the guaianolides 32 and 34. The molecular formula of 32 was  $C_{17}H_{20}O_5$  as followed from the mass spectrum. The  $H^1$ -NMR spectrum showed the typical signals of methylene lactones. Furthermore the presence of two additional exomethylene groups, acetate and a hydroxyl

group could be deduced from the spectrum which was in part close to that zaluzanin C acetate<sup>21</sup>.

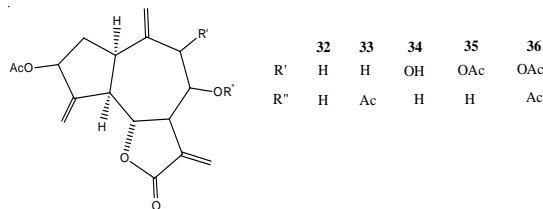


Fig. 7. Chemical structure of Sesquiterpene lactone from *Centaurea kandavanensis* Wagenitz. 32-36.

The more polar lactone 34, Molecular formula  $C_{17}H_{20}O_6$ , obviously was the 9  $\alpha$ -hydroxy derivative of 32. This investigation shows again that guaianolide derived from zaluzanin C may be characteristic for large parts of the genus *Centaurea*<sup>22</sup>.

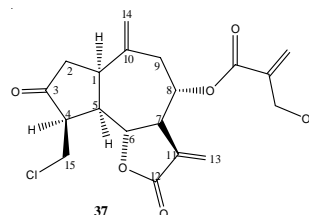


Fig. 8. Chemical structure of Sesquiterpene lactone from *Aegopordon berarioides* Boiss. 37.

### A New Guaianolide from *Aegopordon berarioides* Boiss.

The aerial parts of *A. berarioides* afforded, in addition to lupeol, taraxasterol, sitosterol-3-O-glucoside and cynaropicrin and a new guaianolide, the structure of which was deduced by high-field  $H^1$ -NMR spectroscopy. The plant materials were collected in west of Kerman, Iran<sup>23</sup>.

### Sesquiterpene Lactones and Eudesmane Derivatives from *Onopordon carmanicum* (Bomm.) Bomm.

The aerial parts of *Onopordon carmanicum* (Bomm.) Bomm afforded in addition to Onopordopicrin and two related esters the epoxide of Onopordopicrin, a new elemene derivative, two eudesmanolides and two eudesmane derivatives which most likely are the precursors of the latter lactones. The structures were elucidated by highfield NMR spectroscopy<sup>24</sup>.

The genus *Onopordon* (Compositae, tribe Cynareae) is placed together with the large genera *Cousinia*, *Saussurea* and *Jurinea* in the subtribe Carduinae. Taxonomically this genus is closely

related to *Cousinia*, while the position of *Jurinea* and *Saussurea* is uncertain<sup>25</sup>. So far from the genus *Onopordon* in addition to widespread compounds several C<sub>17</sub>-acetylenes<sup>26</sup> and the germacranolide Onopordopicrin<sup>27, 28, 4, 3</sup> as well as closely related lactones<sup>4,3</sup> have been reported.

Similar 15-hydroxyl germacranolides with an 8 $\alpha$ -acyloxy group have been isolated from *Jurinea* species. This type of sesquiterpene lactone seems to be characteristic for a group of genera in the Cynareae. They have been reported from *Centaurea*, *Arctium* and *Cnicus* species. However, lactones with the same substitution pattern with an additional hydroxyl group at C-14 are reported from *Dicoma* species (tribe Mutisieae)<sup>29</sup>. This type is present also in some *Jurinea* species<sup>30</sup>. From *Cousinia* species so far no lactones are reported. We have studied now a further *Onopordon* species *O. carmanicum* (Born.) Born.

The polar fractions of the extract of the aerial parts of *O. carmanicum* gave as the main constituents Onopordopicrin (40)<sup>27</sup> as well as a complex mixture of sesquiterpene lactones which could be separated by HPLC. In addition to the isobutyrate 1<sup>3</sup> and the corresponding methacrylate 39<sup>3</sup> the 4 $\alpha$ , 5 $\beta$ -epoxide of Onopordopicrin (46), the epimeric aldehydes 41 and 42, the epimeric methyl esters of the corresponding precursors 43 and 44 as well as the elemene 45 were isolated<sup>24</sup>.

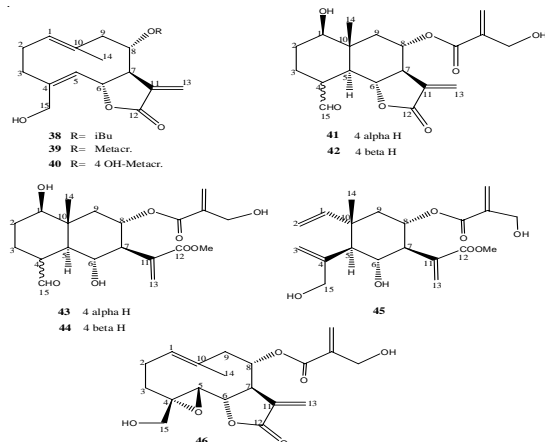


Fig. 9. Chemical structure of Sesquiterpene lactone from *Onopordon carmanicum* born.38-46.

### Sesquiterpene Lactones from *Achillea micrantha* M.B.

The aerial parts of *Achillea micrantha* afforded the eudesmanolides santamain, reynosin, dihydrosantamarin, dihydroerynosin and the germacranolides artemorin, gallicine, dihydroparthenolide as well as a new one, dihydroparthenolide bisepoxide 47. The structures were elucidated by spectroscopy data. From the large genus *Achillea* (compositae-anthemideae) several sesquiterpene lactones were reported<sup>31</sup>. We now have studied *Achillea micrantha* M. B. Careful separation by thin layer and high pressure liquid chromatography of the polar fraction of the column chromatography of the extracted of the aerial parts afforded dihydroparthenolide<sup>32</sup>, santamarin<sup>33</sup>, dihydrosantamarin<sup>34</sup>, dihydroreynosin<sup>33</sup>, reynosin<sup>35</sup>, gallicin<sup>36</sup>, artemorin<sup>37</sup> and a new lactones with molecular formula C<sub>15</sub>H<sub>20</sub>O<sub>4</sub><sup>38</sup>.

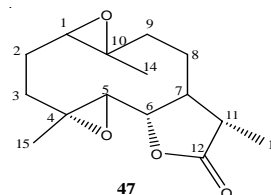


Fig. 10. Chemical structure of Sesquiterpene lactone from *Achillea micrantha* M.B. 47.

### Germacranolides from *Anvillea garcini* (Burn.) DC

The aerial parts of *Anvillea garcini* (Burn.) DC afforded three germacranolides, two of which had not being isolated previously. The structures were elucidated by <sup>1</sup>H-NMR spectroscopy. The configuration of 9-acetoxy parthenolide at C-9 has been revised<sup>39</sup>.

The small genus *Anvillea garcini* (tribe Inuleae, subtribe Inulina) is placed in the Inula group<sup>40</sup>. From *A. garcini* (Burn.) DC flavones<sup>41</sup> and 9 $\alpha$ -hydroxyparthenolide (48)<sup>42</sup> were reported.

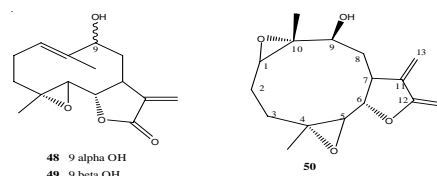


Fig. 11. Chemical structure of Sesquiterpene lactone from *Anvillea garcini* (Burn.)DC 48-50.

An investigation of a sample collected in the South of Iran gave in addition to 9 $\alpha$ -hydroxyparthenolide (48), two further lactones, 49 (the epimer of 48) and 50 (the epoxide of 49). The structures were elucidated by high field proton NMR spectroscopy<sup>39</sup>.

### Sesquiterpene Lactones from *Jurinella moschus* (Halb)

The aerial parts of *Jurinella moschus* afforded the lignane arctigenin, four sesquiterpene lactones, the germacranolides salonitenolide and two new ones as well as a new elemanolide. The structures were elucidated by high field proton-NMR spectroscopy<sup>43</sup>. The small genus *Jurinella* (compositae Cynareae, Carduinae), which is distributed over SW Asia, has hitherto not been a subject for chemical study. The investigation of the aerial parts of *Jurinella moschus* (Halb.) Bobrov afforded salonitenolide (51)<sup>44</sup> and arctigenin (55)<sup>45</sup> as well as three new sesquiterpene lactones, the elemanolide 54, named 20-hydroxyelemajurinelloide<sup>43</sup>.

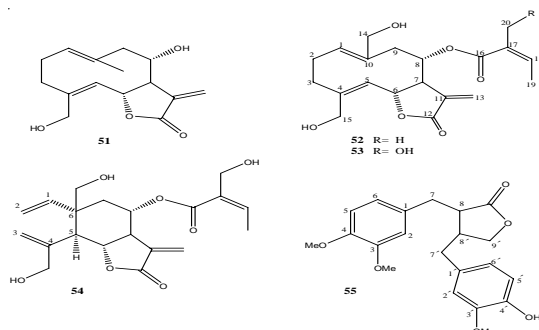


Fig. 12. Chemical structure of Sesquiterpene lactone from *Jurinella moschus* (Halb) 51-55.

The  $^1\text{H}$ -NMR spectra of 51 and 55 were identical with those of authentic material. As the NMR data of the latter have not been reported in the literature we have added then in the Experimental<sup>43</sup>.

### Further Sesquiterpene Lactones from Genus *Dittrichia*

The aerial parts of *Dittrichia graveolens* afforded in addition to compounds isolated previously five new sesquiterpene lactones, two benzoic acid derivatives while *D. viscosa* gave two further derivatives of costic acid. The structures were elucidated by high field NMR spectroscopy<sup>46</sup>.

From the small genus *Dittrichia*, previously a section of *Inula*, one species, *D. viscosa* (L.)Greuter, has been investigated chemically. In addition to costic acid derivatives<sup>47, 48, 49, 50</sup>, the aerial parts of this very widespread species gave sesquiterpene lactones<sup>48, 49, 50</sup> as well as some flavonoids<sup>51</sup>. From the roots, in addition to a thymol derivative<sup>49, 52</sup>, several rare germacranolides were isolated<sup>52</sup>.

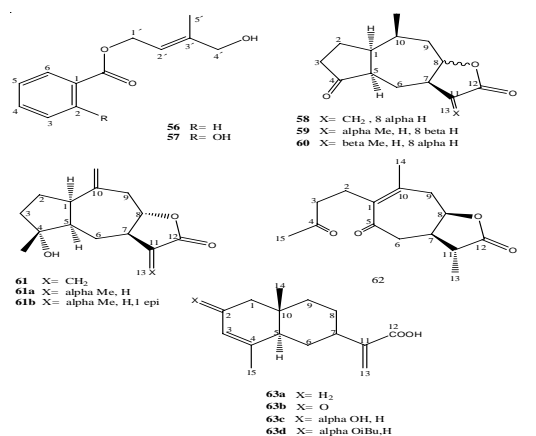


Fig. 13. Chemical structure of Sesquiterpene lactone from Genus *Dittrichia* 56-65.

From *D. graveolens* (Desf.)Greuter, graveolides<sup>53</sup> and pseudoguaianolide without assignment of stereochemistry were reported<sup>54</sup>. We have now reinvestigated a sample from Iran. In addition to known compound, several new sesquiterpene lactones and two unusual benzoic acid derivatives were isolated. From *Dittrichia tenerife* (Canary Islands) similar lactones but also a new costic acid derivative and a rearranged sesquiterpene were isolated 56-65<sup>46</sup>.

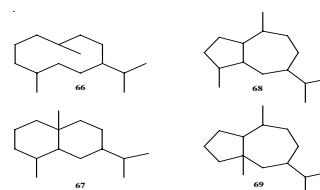
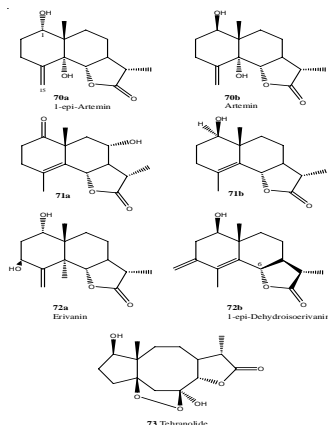


Fig. 14. The frame works Sesquiterpene lactone from *Artemisia diffusa* 66-69.

### Sesquiterpene Lactones from *Artemisia diffusa*

Several reviews on the sesquiterpene lactones of the genus *Artemisia* have appeared in the literature<sup>20</sup> which discuss the taxonomic conclusions to be derived from the distribution of sesquiterpene lactones within *Artemisia* species, the majority of these lactones exhibit one of the four frameworks represented below, i.e. germacrane 66, eudesmane 67, guaiane 68 and pseudoguaiane 69.

Reports dealing with isolation and structure elucidation of sesquiterpene lactones have increased dramatically. Two reasons can be given for the strongly increasing interest in this group of natural products. First, sesquiterpene lactones have been successfully used as markers in biochemical systematic (chemotaxonomy) studies mainly in the Compositae. Secondly, a number of compounds received considerable attention due to various biological activities.



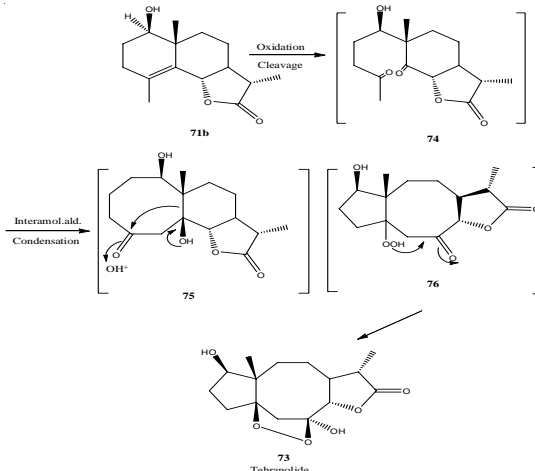
**Fig. 15. Chemical structure of Sesquiterpene lactone from *Artemisia diffusa* 70a-73.**

The genus *Artemisia* is not very uniform and the chemistry is somewhat diverse. However, most species contain sesquiterpene lactone, especially 11-, 13- dihydroderivatives.

The extract of the aerial parts of *A. diffusa* afforded several eudesmanolides (70a, 70b, 71a, 71b, 72a, 72b, 73) and a new type of sesquiterpene lactone with unusual carbon skeleton, an eight-member ring (Tehranolide)(73)<sup>55</sup>.

Most likely this unusual carbon skeleton was formed by oxidative cleavage of the  $\Delta^4$  bond of 71b followed by an internal aldol condensation of the intermediate 5 affording the dihydroxy ketone

75. The latter then could be rearranged to the lactone 76 by attack of  $\text{HO}^-$  followed by acetal formation to give the lactone 73 (Tehranolide).



**Fig. 16. Biosynthesis of Tehranolide**

The extract of the aerial parts of *A. diffusa* collected in the Province of Khorasan (Iran) afforded, in addition to several eudesmanolides a new type of sesquiterpene lactone (Tehranolide) with an endoperoxide group that probably has the same effect as the antimalarial agent artemisinin.

We have already reported the antimalarial properties of the extract of the extract and the fraction which contains sesquiterpene lactones including Tehranolide of the same species (*Artemisia diffusa*)<sup>56, 57, 58</sup>.

Recently Artediffusin (Tehranolide) has been confirmed and considered as a new antimalarial agent<sup>59</sup>.

### Biological Activities

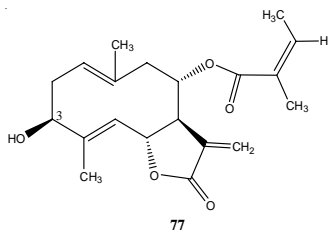
Sesquiterpene lactones are one of the most prevalent and biologically significant classes of secondary metabolite present and hence have been subject to a number of studies. In addition, sesquiterpene lactones have anticarcinogenic, anti-inflammatory capacity<sup>63</sup>. Asteraceous plants are in turn the most diverse and prolific plant family in the world.

To humans, lettuce and chicory (*Lactuca sativa* and *Chicorium inty bus* L.) represent the main dietary source of sesquiterpene lactones, on the basis of the levels of their global consumption.

Casagrande<sup>61</sup> indicates that 11% of Americans studied reach their targets for both fruit and vegetables, though 28% and 32% reached individual targets of two fruit per day and three vegetables per day respectively in 2002 whereas the latest study found only 16% of UK reach their 5 a day target<sup>62</sup>.

Additionally a range of Asteraceous plants are used to impart the bitterness of some alcoholic beverages. Other sources of sesquiterpenoids include spices for example star anise, and herbs, though consumption levels of these are understandably smaller. Traditional medicinal plants can also be a significant source for some populations, as sesquiterpenoids often represent the active ingredient<sup>63, 64, 65</sup>. These medicinal plants are often from the Compositae (*Asteraceae*) family of which "feverfew" (*Tanacetum parthenium* (L.) Sch. Bip.) Yarrow (*Achillia* spp.), and quinghaosu (*A. annua*) in the treatment of malarial type ailments, are among the most commonly used both in historically and in current alternative treatments<sup>66</sup>.

Roman chamomile flower (*Chamomilla romanae flos*) is the inflorescence of *Anthemis nobilis* L. (sometimes known under the name *Chamaemelum nobile* (L.) All.). The plant grows wild in southern and western Europe and also in North Africa. It is cultivated in several European countries, as well as in Egypt and in Argentina.



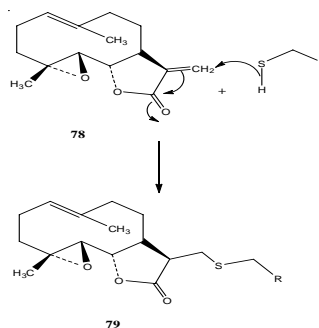
**Fig. 17. Chemical structure of nobilin from *Anthemis nobile* (L.) 77.**

Roman chamomile flowers contain 0.6-2.4% volatile oil, the main components of which are esters, in particular the isobutyl ester, of angelic. Also present in the drug are sesquiterpene derivatives, e.g. the lactone nobilin 77, which has a bitter taste, and flavonoids with a spasmolytic effect, e.g. apigenin 7-glucoside.

Roman chamomile flower- usually taken as a tea- is an herbal remedy for colic and several other complaints<sup>67</sup>.

Parthenolide is a sesquiterpene derivative which is contained in leaves of feverfew, *Chrysanthemum parthenium* Bernh. (Some times known as *Tanacetum parthenium* Sch. Bip.), an herbal remedy used for prophylaxis of migraine. This activity has been proved clinically. Parthenolide is an inhibitor for blood platelet aggregation. Release of 5-hydroxytryptamine (serotonin) accompanies platelet aggregation and has been linked to the onset of migraine<sup>67</sup>.

The  $\alpha$ -methylene butyrolactone function of parthenolide is a Michael acceptor of thiols. It has been suggested that this reaction is responsible for the inhibitory effect of the compound



**Fig. 18. The  $\alpha$ -methylene butyrolactone function of parthenolide is a Michael acceptor of thiols**

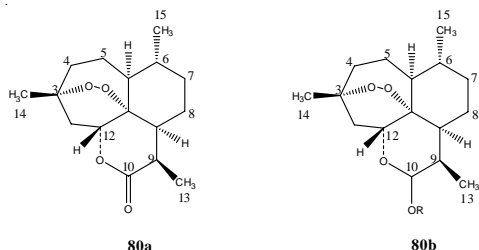
on blood platelet aggregation. Secondly, the inhibitory effects are dose- and time- dependent, and thirdly, treatment of platelet with feverfew extracts or parthenolide causes a dramatic reduction in the number of acid- soluble thiolgroup's present<sup>67</sup>. On the other hand doubts have been raised as to the credibility of this explanation in the clinical situation as parthenolide entering the bloodstream would be rapidly "neutralized" by Michael addition of the thiol residue in glutathione, which is one of the body's main defenses against such compounds (78,79)<sup>67</sup>.

#### Antimalarial Activity

##### Artemisinin and its derivatives

In 1972, a group of Chinese researchers isolated a new anti-malarial drug (+) - artemisinin (1), a sesquiterpene lactone of the amorphene sub

group of cadinene from the hexane extract of a traditional Chinese medicinal plant *Artemisia annua* (*Asteracea*)- a plant which has been used for the treatment of fever and malaria since ancient time<sup>68</sup>.

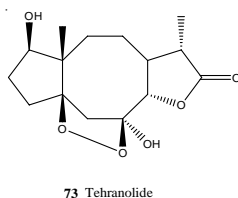


**Fig. 19. Chemical structure of Artemisinin and Dihydroartemisinin (R: H) 80a-80b**

Artemisinin is a sesquiterpene lactone containing an endoperoxide linkage in it. This highly oxygenated sesquiterpene lactone peroxide, unlike most other anti-malarials, lacks nitrogens containing heterocyclic ring systems and was found to be super plusmocidal and blood Schizontocidal agent to conventional anti-malarial drugs, such as chloroquine, quinine etc. against malaria strains, without obvious adverse effects in patients.

#### Tehranolide as a new Antimalarial Candidate

Since the discovery and the use of artemisinin and endoperoxide sesquiterpene lacton, particular attention has been directed to this class of compounds, we have investigated many Iranian *Artemisia* species.



**Fig. 20. Chemical structure of Tehranolide 73.**

*Artemisia aucheri* 69, 70, 71, *A. austriaca* 72, *A. biennis* 73, *A. campestris* 73, *A. deserti* 74, *A. diffusa* 53, 56, 75, *A. gypsacea* 76, 77, *A. haussknechtii* 78, *A. kermanensis* 78, *A. kopetdaghensis* 78, *A. kulbadica* 79, *A. oliveriana* 80, *A. persica* 81, *A. santolina* 77, *A. sieberi* 82, 83, 84, *A. tschernieviana* 73, *A. ciniformis* 85, *A. incana* 85, *A. turanica* 86 and *A. tournefortiana* 71.

From all these species, we discovered an unusual sesquiterpene lactone with endoperoxide group, which we have named Tehranolide. The extract of the aerial parts of *A. diffusa* afforded several eudesmanolide and a new type of sesquiterpene lactone with unusual carbon skeleton, an eight member ring 59.

The anti-malarial activity was determined by using different concentrations including 10, 30, 50 mg/ml-1 of Tehranolide were made in drug vehicle including distilled water, methanol, DMSO and applied for therapy. Percentage of parasitaemia was counted after 24, 48 and 72 h. after treatment for each concentration. Results indicated no effects of low concentration of Tehranolide on parasitaemia, however the concentrations of 10, 30 and 50 mg/ml-1 represented their anti-plasmodial activities. The cytotoxic effects of high concentration occurred by destroying both parasites and RBCs in culture medium. Inhibition concentration of 50% ( $IC_{50}$ ) on plasmodial survival was observed at concentration of 10 mg/ml-1 after 48-72 h. of treatment. It is concluded that, Tehranolide seems to be a promising drug exhibiting good anti-malarial effects in this human malaria *P. falciparum* model in vitro. However, more research is required before Tehranolide can be used for malaria treatment in human cases 87.

#### ACKNOWLEDGMENT

The authors are very thankful to Miss. Mahdiah Ariaee Fard for typing the manuscript.

#### REFERENCES

1. Wedge, D.E.; Galindo, J.C.; Macías, F.A. *Phytochemistry*. **2000**, *53*, 747-757.
2. De Luque, A.P.; Galindo, J.C.G.; Macías, F.A.; Jorrín, J. *Phytochemistry*. **2000**, *53*, 45-50.
3. Rustaiyan, A.; Nazarians, L.; Bohlmann, F. *Phytochemistry*. **1979**, *18*, 883-884.
4. Rustaiyan, A.; Nazarians, L.; Bohlmann, F. *Phytochemistry*. **1979**, *18*, 879-880.
5. Evstratova, R.I.; Rybalko, K.S.; Sheichenko, I. *Chem. Nat. Compd.* **1972**, *8*, 450-457.
6. Rustaiyan, A.; Niknejad, A.; Bohlmann, F.; Schuster, A. *Phytochemistry*. **1981**, *20*, 1154.
7. Evstratova, R.I.; Sheichenko, V.I.; Rybalko, K.S. *Khim. Prir. Soedin.* **1973**, *9*, 161.



8. Evstratova, R.I.; Rybalko, K.S.; Rzazade, R.Y. *Khim. Prir. Soedin.* **1967**, 3, 384.
9. Gonzales, G.A.J.; Bermejo Barrera, J.; Breton Funes, J.L.; Triana, J. *Tetrahedron Lett.* **1972**, 13, 2017-2020.
10. Rustaiyan, A.; Nazarians, L.; Bohlmann, F. *Phytochemistry.* **1981**, 20, 1152.
11. Suchy, M.; Herout, V.; Sorm, F. *Collect. Czech. Chem. Commun.* **1960**, 25, 2777.
12. Gonzales, G.A.J.; Bermejo, J.; Cebrera, I.; Massanet, G.M.; Mansila, H.; Galindo, A. *Phytochemistry.* **1971**, 17, 955-956.
13. Gonzales, A.G.; Bermejo, J.; Massanet, G.M. *Perez, J. Ann. Quim.* **1973**, 69, 1333.
14. Corbela, A.P.; Garriboldi, P.; Jommi, G.; Samek, Z.; Holub, M.; Drozd, B. *Chem. Commun.* **1972**, 386.
15. Thiessen, W.E.; Hope, H. *Acta Crystallogr. Sect.* **1970**, B 26, 554.
16. Rustaiyan, A.; Niknejad, A.; Zdero, C.; Bohlmann, F. *Phytochemistry.* **1981**, 20, 2427-2429.
17. Rustaiyan, A.; Niknejad, A.; Aynehchi, Y. *Planta Med.* **1982**, 44, 185-186.
18. Rustaiyan, A.; Sharif, Z.; Tajarodi, A.; Ziesche, J.; Bohlmann, F. *Planta Med.* **1984**, 2, 193-194.
19. Seaman, F.C. *Bot. Rev.* **1982**, 48, 121-594.
20. Asakawa, Y.; Takemoto, T. *Phytochemistry.* **1979**, 18, 285-288.
21. Rustaiyan, A.; Ardebili, Sh. *Planta Med.* **1984**, 4, 363-364.
22. Izaddoost, M.; Dabiri, M.; Rustaiyan, A. *Fitoterapia.* **1985**, 56, 275-277.
23. Rustaiyan, A.; Ahmadi, B.; Jakupovic, J.; Bohlmann, F. *Phytochemistry.* **1986**, 25, 1659-1662.
24. Dittrich, M. in: *The Biology and Chemistry of the Compositae*, Heywood, V.H., Harborne, J.B. and Turner, B.L., Eds., London: Academic Press, **1977**, 1010.
25. Bohlmann, F.; Burhardt, T.; Zdero, C. *Naturally Occurring Acetylenes*, London: Academic Press, **1973**, 448.
26. Garcia, B.; Skaltsa, H.; Navarro, F.I.; Pedro, J.; Lazari, D. *Phytochemistry.* **1996**, 41, 1113-1117.
27. Miski, M.; Meriçli, A.H.; Mabry, T.J. *Phytochemistry.* **1988**, 27, 1417-1420.
28. Bohlmann, F.; Singh, P.; Jakupovic, J. *Phytochemistry.* **1982**, 21, 2029-2033.
29. Fujimoto, Y.; Kinoshita, T.; Ikekawa, N.; Mungarulire, J. *Phytochemistry.* **1987**, 26, 2593-2595.
30. Trendafilova, A.; Todorova, M.; Mikhova, B.; Vitkova, A.; Duddeck, H. *Phytochemistry.* **2006**, 67, 764-770.
31. Ogura, M.; Cordell, G.A.; Farnsworth, N.R. *Phytochemistry.* **1987**, 17, 957-961.
32. Herz, W.; Lakshmikantham, M.V. *Tetrahedron.* **1965**, 21, 1711-1715.
33. Shafizadeh, F.; Bhadane, N.R.; Morries, S.M.; Kelsey, R.G.; Khanna, S.N. *Phytochemistry.* **1971**, 10, 2745.
34. Yoshioka, H.; Renold, W.; Fischer, N.H.; Higo, A.; Mabry, T.J. *Phytochemistry.* **1970**, 9, 823-832.
35. Gonzales, A.G.; Bermejo, J.; Mansilla, H.; Massanet, G.M.; Cabrera, I.; Amara, J.M.; Galindo, A. *Phytochemistry.* **1977**, 16, 1836-1837.
36. Kelsey, R.G.; Shafizadeh, F. *Phytochemistry.* **1979**, 18, 1591-1611.
37. Rustaiyan, A.; Sharif, Z.; Tajarodi, A.; Sadjadi, A.S. *phytochemistry.* **1987**, 26, 2856-2857.
38. Rustaiyan, A.; Dabiri, M.; Jakupovic, J. *Phytochemistry.* **1986**, 25, 1229-1230.
39. Merxmuller, H.; Leins, P.; Roessler, H. in: *The Biology and Chemistry of the Compositae*, Heywood, V.H.; Harborne, J.B.; Turner, B.L. Eds., London: Academic Press, **1977**, 590.
40. Ulubelen, A.; Mabry, T.J.; Aynekdin, Y. *J. Nat. Prod.* **1979**, 42, 624.
41. Tyson, R.L.; Chang, C.J.; McLaughlin, J.L.; Aynehchi, Y.; Cassidy, J.M. *Experientia.* **1981**, 37, 441.
42. Rustaiyan, A.; Ganji, M.T.; *Phytochemistry.* **1987**, 26, 2857-2859.
43. Zdero, C.; Bohlmann, F. *Phytochemistry.* **1990**, 29, 183-187.
44. Haworth, R.D.; Kelly, W.J. *Chem. Soc.* **1937**, 384.
45. Rustaiyan, A.; Jakupovic, J.; Chau-Thi, T.V.; Bohlmann, F.; Sadjadi, A. *Phytochemistry.* **1987**, 26, 2603-2606.
46. Barbetti, P.; Chiappini, I.; Fardella, G.; Manghini, A. *Planta Med.* **1985**, 51, 471.
47. Ceccherelli, P.; Curini, M.; Marcotullio, M.; Manghini, A. *Phytochemistry.* **1985**, 24, 2987-2989.
48. Doskotch, R.W.; Wilton, J.H.; Harraz, F.M.; Fairchild, E.H.; Huang, C.T.; El-Ferally, F.S. *J. Nat. Prod.* **1983**, 46, 923-929.
49. Daniewski, W.H.; Kroszcyński, W.; Bloszyk, E.; Drozd, D.; Nawrot, J.; Rychlewska, V.; Budesinsky, M.; Holob, M. *Collect. Czech.*

- Chem. Commun.***1986**, *51*, 1710-1721.
50. Grande, M.; Picra, F.; Cuenca, A.; Torres, P.; Bellido, I.S. *Planta Med.***1985**, *51*, 414-419.
  51. Bohlmann, F.; Gupta, R.K. *Phytochemistry*. **1982**, *21*, 1443-1445.
  52. Wang, G.W.; Qin, J.J.; Cheng, X.R.; Shen, W.H.; Shan, L.; Jin, H.Z.; Zhang, W.D. *Expert Opin. Investig. Drugs*. **2014**, *23*, 317-345.
  53. Chiappini, I.; Fardella, G. *Fitoterapia*. **1980**, *51*, 161.
  54. Rustaiyan, A.; Sigari, H.; Jakopovic, J.; Grenz, M. *Phytochemistry*. **1989**, *28*, 2723-2725.
  55. Rustaiyan, A.; Nahrevanian, H.; Kazemi, M. *Pharmacogn. Mag.* **2009**, *4*, 1-7.
  56. Rustaiyan, A.; Nahrevanian, H.; Kazemi, M. *Proc. of BIT's 5th Anniversary Cong. of International Drug Discovery Science and Technology (IDDBST)*, Shanghai, **2007**.
  57. Rustaiyan, A.; Nahrevanian, H.; Kazemi, M. In: *57th International Congress and Annual Meeting of Society for Medicinal Plant and Natural Product Research*, Geneva, **2009**, 16-20.
  58. Rustaiyan, A.; Nahrevanian, H.; Kazemi, M. *Asian J. Chem.***2011**, *23*, 4810-4814.
  59. Merfort, I. *Curr Drug Targets*.**2011**, *12*, 1560-1573.
  60. Casagrande, S.S.; Wang, Y.; Anderson, C.; Gary, T.L. *Am. J. Prev. Med.***2007**, *32*, 257-263.
  61. Rogers, S.; Pryer, J.A. *Public Health Nutr.* **2012**, *15*, 1240-1247.
  62. Canales, M.; Hernández, T.; Caballero, J.; de Vivar, A.R.; Avila, G.; Duran, A.; Lira, R. *J. Ethnopharmacol.***2005**, *97*, 429-439.
  63. Bork, P.M.; Schmitz, M.L.; Kuhnt, M.; Escher, C.; Heinrich, M. *FEBS Lett.***1997**, *402*, 85-90.
  64. Lyß, G.; Knorre, A.; Schmidt, T.J.; Pahl, H.L.; Merfort, I.; *J. Biol. Chem.*, **1998**, *273*, 33508-33516.
  65. Heinrich, M.; Robles, M.; West, J.E.; Ortiz de Montellano, B.R.; Rodriguez, E. *Annu. Rev. Pharmacol. Toxicol.***1998**, *38*, 539-565.
  66. Samuelsson G., and Bohlin L., in *Drugs of Natural Origin. in 6th Ed. A Treatise of Pharmacognosy*, Stockholm, 2009.
  67. Little, D.B.; Croteau, R.B. *Arch. Biochem. Biophys.*, **2002**, *402*, 120-135.
  68. Khorsand Mohammadpoor, S.; Yari, M.; Rustaiyan, A.; Masoudi, S. *J. Essent. Oil Res.***2002**, *14*, 122-123.
  69. Wollenweber, E.; Favre-Bonvin, J.; Waton, H.; Hauteville, M.; Rustaiyan, A. *Phytochemistry*. **1992**, *11*, 105-107.
  70. Rustaiyan, A.; Bamonieri, A.; Raffatrad, M.; Jakupovic, J.; Bohlmann, F. *Phytochemistry*. **1987**, *26*, 2307-2310.
  71. Rustaiyan, A.; Faridchehr, A. *Res. Rev. J. Bot. Sci.***2014**, *3*, 1-9.
  72. Nematollahi, F.; Rustaiyan, A.; Larijani, K.; Nadimi, M. *J. Essent. Oil Res.***2006**, *18*, 339-341.
  73. Kazemi, M.; Dakhili, M.; Rustaiyan, A.; Larijani, K.; Ahmadi, M.A.; Mozaffarian, V. *Pharmacogn. Res.* **2009**, *2*, 120-124.
  74. Rustaiyan, A.; Komeilizadeh, H.; Masoudi, S.; Monfared, A.; Yari, M.; Kardar, M.; Shahgholi A. *J. Sci. Islamic Repub. Iran.* **2000**, *11*, 213-215.
  75. Khazraei- Alizade, Kh.; Rustaiyan, A. *J. Essent. Oil Res.***2001**, *14*, 185-186.
  76. Rustaiyan, A.; Zare, K.; Ganji, M.T.; Sadri, H.A. *Phytochemistry*. **1989**, *28*, 1535-1536.
  77. Rustaiyan, A.; Balalaei, S.; Mohammadi, F.; Masoudi, S.; Yari, M. *J. Essent. Oil Res.***2000**, *12*, 330-332.
  78. Rustaiyan, A.; Tabatabaei-Anaraki, M.; Kazemi, M.; Masoudi, S.; Makipour, P. *J. Essent. Oil Res.* **2009**, *21*, 410-413.
  79. Aghajani, Z.; Kazemi, M.; Dakhili, M.; Rustaiyan, A.; *Nat. Prod. Commun.***2009**, *4*, 1261.
  80. Sanz, J.F.; Rustaiyan, A.; Alberto Marco, J. *Phytochemistry*. **1990**, *29*, 2919-2921.
  81. Rustaiyan, A.; Ameri, A.; Mirjalili, B.F.; Mazloun Ardakani, M.; Hakimi Maybody, M.; Bamoniri, A. *Journal of Sciences (Islamic Azad University)*. **2003**, *13*, 1074-1078.
  82. Weyerstahl, P.; Schneider, S.; Marschall, H.; Rustaiyan, A. *Eur. J. Org. Chem.***1992**, *2*, 111-116.
  83. Weyerstahl, P.; Schneider, S.; Marschall, H.; Rustaiyan, A. *Flavour Fragrance J.***1993**, *8*, 139-145.
  84. Alberto Marco, J.; Sanz-Cervera, J.F.; Sancenon, F.; Jakupovic, J.; Rustaiyan, A.; Mohamadi, F. *Phytochemistry*. **1993**, *35*, 1061-1065.
  85. Rustaiyan, A.; Masoudi, S.; Kazemi, M. *J. Essent. Oil Res.***2007**, *19*, 548-551.
  86. Firouznia, A.; Akbari, M.T.; Rustaiyan, A.; Masoudi, S.; Bigdeli, M.; Tabatabaei-Anaraki, M. *J. Essent. Oil. Bear. Pl.***2007**, *10*, 88-993.
  87. Rustaiyan, A.; Nahrevanian, H.; Zamani, Z.; Taherkhani, M.; Iravani, A. *Res. J. Parasitol.* **2015**, *10*, 73-78.