

# A novel mutation (a886g) in exon 5 of FGFR2 in members of a family with Crouzon phenotype and plagiocephaly

Daniela Steinberger, Hartmut Collmann, Bernhard Schmalenberger, Ulrich Müller

## Abstract

We identified a novel mutation in members of a family with signs of Crouzon syndrome and plagiocephaly. In affected members of the family an A→G transition was found at position 886 in exon 5 of the fibroblast growth factor receptor 2 (FGFR2) gene. The base change results in the replacement of a lysine by glutamic acid in Ig-like loop III of FGFR2. The unusual finding of plagiocephaly in these Crouzon patients may either be the result of the type of mutation or because of genetic and environmental factors that affect the phenotype in addition to the mutated FGF receptor.

(J Med Genet 1997;34:420-422)

Keywords: craniosynostosis; Crouzon syndrome; plagiocephaly; FGFR2

Crouzon syndrome is a clinically defined craniofacial dysostosis characterised by ocular proptosis owing to shallow orbits, hypertelorism, and craniosynostosis. Craniosynostosis is commonly caused by premature closure of the coronal, lambdoid, and sagittal sutures. Neurological symptoms observed in Crouzon syndrome include mental retardation, seizures, and optic atrophy.<sup>1</sup>

Crouzon syndrome is transmitted as an autosomal dominant trait with variable expression. Molecular genetic analyses have identified mutations in the gene coding for fibroblast growth factor receptor 2 (FGFR2) on chromosome 10 (10q26) as the underlying cause of the disorder.<sup>2</sup> All mutations observed in Crouzon syndrome to date have been detected in the two exons (exons 5 and 7) of the gene that encodes the Ig-like chain III (IIIa and IIIc) of the receptor.<sup>3-7</sup> The mutations probably interfere with normal FGF binding.

Mutations in the FGFR2 gene have also been observed in the clinically defined craniosynostosis syndromes of Apert, Pfeiffer, and Jackson-Weiss.<sup>8-13</sup> With the possible exception of Apert syndrome,<sup>8, 14</sup> it is not possible to correlate the type of FGFR2 mutation with the clinical condition. In fact, identical mutations have been described in Crouzon and Pfeiffer syndromes and in Crouzon and Jackson-Weiss syndromes.<sup>9, 10</sup> Thus the clinically distinct syndromes are extremes of a spectrum of cranial malformations with the eponymous syndromes at opposite ends of this spectrum.

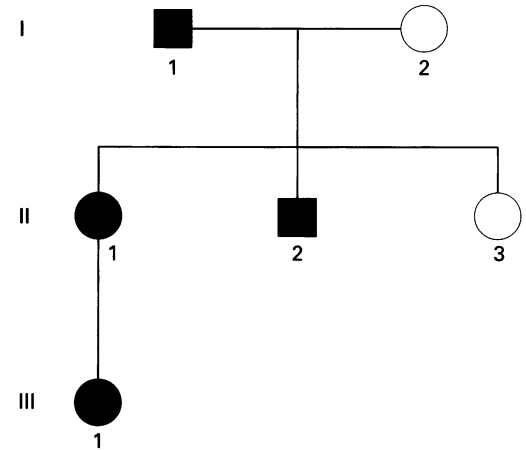


Figure 1 Pedigree of the family studied.

Here we describe a novel mutation in exon 5 of FGFR2 in a family with Crouzon syndrome and plagiocephaly. Since plagiocephaly is not normally a finding in clinically defined Crouzon syndrome, the present study supports the idea that eponymous craniosynostosis syndromes are phenotypic extremes of FGFR mutations rather than nosological entities.

EDTA blood was obtained from all subjects shown in the pedigree (fig 1) except III.1 and DNA was extracted according to standard procedures. III.1 was diagnosed prenatally on DNA extracted from amniotic fluid cells. Despite the detection of a mutation, the parents decided against termination of pregnancy. Primers used for the amplification of exon 5 of FGFR2 were those described by Slaney *et al*<sup>15</sup> (primer A) and Park *et al*<sup>16</sup> (primer B). Single strand conformation polymorphism (SSCP) analysis<sup>17</sup> of the exon 5 amplification products was performed as described previously.<sup>18</sup> Amplification products from those members of the family that had shown a band shift on SSCP analysis were sequenced directly using sequenase (Amersham) according to standard procedures.<sup>19</sup> Both strands were sequenced in all cases.

A pedigree of the three generation family described is given in fig 1. There were three affected males and one affected female.

I.1 was a 47 year old man with mild manifestations of Crouzon syndrome, including hypertelorism, divergent strabismus, and midface hypoplasia. At the age of 37 years no sutural remnants were seen on radiographs, nor were there signs of raised intracranial pressure. He did not have any neurological or ophthalmol-

Institut für  
Humangenetik der  
Justus-Liebig-  
Universität,  
Schlangenzahl 14,  
D-35392 Giessen,  
Germany  
D Steinberger  
U Müller

Neurochirurgische  
Klinik der Universität  
Würzburg,  
Josef-Schneider-Strasse  
11, D-97080 Würzburg,  
Germany  
H Collmann

Tuttlinger Strasse 7,  
94034 Passau,  
Germany  
B Schmalenberger

Correspondence to:  
Professor Müller.

Received 29 August 1996  
Revised version accepted for  
publication 27 November  
1996

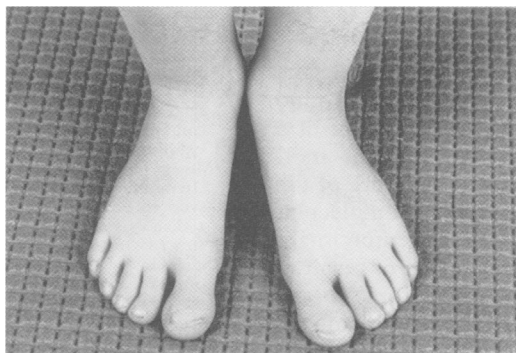


Figure 2 Feet of II.2. Note broadened big toes.

logical abnormalities. Both hands and feet were normal.

II.1 was the 21 year old daughter of I.1 with severe manifestations of Crouzon syndrome. She developed headaches and seizures at 4 years of age. Bilateral optic atrophy and right amaurosis were first observed at 7 years. Cranial findings at the age of 7 included hypertelorism, mild proptosis, brachycephaly, and discrete anterior right plagiocephaly. She had midface hypoplasia and a long philtrum. Radiography showed pansynostosis, increased digital markings, and fusion of cervical vertebrae 2 and 3. Surgery was performed at the age of 7 to alleviate increased intracranial pressure. Seizures recurred at 13 years. Mental performance was normal. There were no abnormalities of either hands or feet.

II.2 was the 9 year old brother of II.1 with severe manifestations of Crouzon syndrome. He was first seen at 1 year and presented with mild anterior plagiocephaly. Radiography showed right unilateral synostosis of the coronal suture. At this stage, the remaining sutures were normal. At 5 years bilateral optic atrophy was diagnosed. Pansynostosis was detected on radiography. Owing to severely increased intracranial pressure, surgery was performed. He did not develop seizures and his psychomotor development was normal. At present, hypertelorism, mild proptosis, and midfacial hypoplasia are striking. There were

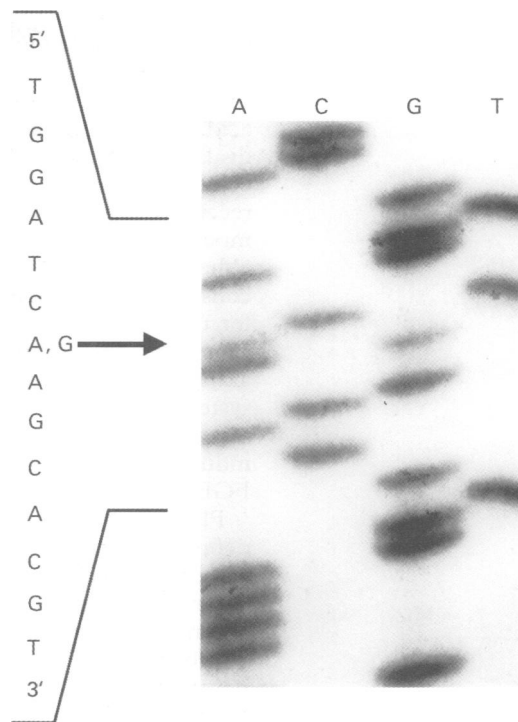


Figure 4 Sequence analysis of exon 5 in II.1. The same A→G transition was observed in all affected but not in unaffected members of the family. The mutation results in the replacement of a lysine by glutamic acid.

no abnormalities of either hands or feet. His big toes were somewhat broadened (fig 2).

II.3 was the unaffected sister of II.1 and II.2.

III.1 was the 6 month old son of II.1. At 6 weeks of age he presented with severe dolichocephaly, anterior plagiocephaly, proptosis, and hypertelorism (fig 3).

Radiography showed premature closure of the sagittal and the right coronal suture resulting in right anterior plagiocephaly. At surgery at 5 months of age, synostosis was found of both coronal sutures in addition to the sagittal suture. There were early signs of premature closure of the lambdoid sutures. Intracranial pressure was slightly increased. There were no neurological abnormalities. Hands and feet were normal.

SSCP analysis of exon 5 showed band shifts in all affected but not in unaffected members of the pedigree in fig 1 (not shown). An A→G transition at position 886 (codon 292) was found by sequencing of exon 5 in the affected subjects (fig 4). Both the wild type and the mutated nucleotide were equally pronounced on cycle sequencing in all affected subjects. There is no evidence of mosaicism in any of the patients. The base change results in the replacement of a lysine by glutamic acid in the first third of Ig-like chain III of FGFR2. The exchange of the basic amino acid lysine for the acidic glutamic acid probably alters structural integrity and thus the function of the receptor.

Although the mutation at codon 292 described here has not previously been recognised in autosomal dominant craniosynostosis, many point mutations at different positions of exons 5 and 7 of FGFR2 are known.<sup>7-13</sup> These two exons code for Ig-like chain III of FGFR2 that is required for ligand binding.<sup>20, 21</sup> A muta-



Figure 3 Patient III.1 at 6 weeks of age. Note plagiocephaly. For further details see text.

tion in exon 7 at amino acid position 342 has been most frequently observed in FGFR associated craniosynostosis.<sup>4</sup> It results in the replacement of a cysteine by another amino acid, thus interfering with the formation of a disulphide bridge within the receptor molecule. This in turn affects the structure of the receptor as was shown by molecular modelling.<sup>22</sup> Alteration of the receptor might either interfere with ligand binding or result in constitutive activation of the receptor. Support for the latter notion comes from experiments examining the common FGFR2 mutation at amino acid position 342 in a *Xenopus* oocyte system. Here constitutive activation of the mutated receptor was found that resulted in the induction of mesoderm even in the absence of FGF.<sup>23</sup>

Plagiocephaly is not a finding in clinically defined Crouzon syndrome, which is characterised by symmetrical premature fusion of sutures. The asymmetrical synostosis of the coronal suture observed in the present patients may be the result of the phenotypic effect of the novel mutation in FGFR2 described here. According to this notion the mutation would give rise to a milder phenotype. This, however, applies to patient I.1 only. The remaining patients were quite severely affected and required surgery to alleviate increased intracranial pressure. Therefore, we suggest that in addition to the type of mutation, additional genetic (for example, determinants of side of premature closure of coronal suture) and environmental factors cause the unusual findings in the present family. Support for this notion comes from the observation of great phenotypic variation in the manifestations of FGFR associated craniosynostosis even within affected members of the same family.<sup>24</sup>

This work was supported by the Deutsche Forschungsgemeinschaft (Ste 770/1-1).

- 1 Jones KL. *Smith's recognizable patterns of human malformation*. 4th ed. Philadelphia: Saunders, 1988.
- 2 Reardon W, Winter RM, Rutland P, et al. Mutations in the fibroblast growth factor receptor 2 gene cause Crouzon syndrome. *Nat Genet* 1994;8:98-103.
- 3 Oldridge M, Wilkie AOM, Slaney SF, et al. Mutations in the third immunoglobulin domain of the fibroblast growth factor receptor 2 gene in Crouzon syndrome. *Hum Mol Genet* 1995;4:1077-82.

- 4 Steinberger D, Mulliken JB, Müller U. Predisposition for cysteine substitutions in the immunoglobulin-like chain of FGFR2 in Crouzon syndrome. *Hum Genet* 1995;96:113-15.
- 5 Gorry MC, Preston RA, White GJ, et al. Crouzon syndrome: mutations in two spliceforms of FGFR2 and a common point mutation shared with Jackson-Weiss syndrome. *Hum Mol Genet* 1995;4:1387-90.
- 6 Meyers GA, Day D, Goldberg R, et al. FGFR2 exon IIIa and IIIc mutations in Crouzon, Jackson-Weiss, and Pfeiffer syndromes: evidence for missense changes, insertions, and a deletion due to alternative RNA splicing. *Am J Hum Genet* 1996;58:491-8.
- 7 Steinberger D, Mulliken JB, Müller U. Crouzon syndrome: previously unrecognized deletion, duplication and point mutation within FGFR2 gene. *Hum Mutat* 1996;8:386-90.
- 8 Wilkie AOM, Slaney SF, Oldridge M, et al. Apert syndrome results from localized mutations of FGFR2 and is allelic with Crouzon syndrome. *Nat Genet* 1995;9:165-72.
- 9 Jabs EW, Li X, Scott AF, et al. Jackson-Weiss and Crouzon syndromes are allelic with mutations in the fibroblast growth factor receptor. *Nat Genet* 1994;8:275-9.
- 10 Rutland P, Pulleyn LJ, Reardon W, et al. Identical mutations in the FGFR2 gene cause both Pfeiffer and Crouzon syndrome phenotypes. *Nat Genet* 1995;9:173-6.
- 11 Muenke M, Schell U, Hehr A, et al. A common mutation in the fibroblast growth factor receptor 1 gene in Pfeiffer syndrome. *Nat Genet* 1994;8:269-74.
- 12 Lajeunie E, Ma HW, Bonaventure J, Munnich A, LeMerrer M. FGFR2 mutations in Pfeiffer syndrome. *Nat Genet* 1995;9:108.
- 13 Schell U, Hehr A, Feldman GJ, et al. Mutations in FGFR1 and FGFR2 cause familial and sporadic Pfeiffer syndrome. *Hum Mol Genet* 1995;4:323-8.
- 14 Moloney DM, Slaney SF, Oldridge M, et al. Exclusive paternal origin of new mutations in Apert syndrome. *Nat Genet* 1996;13:48-53.
- 15 Slaney SF, Oldridge M, Hurst JA, et al. Differential effects of FGFR2 mutations on syndactyly and cleft palate in Apert syndrome. *Am J Hum Genet* 1996;58:923-32.
- 16 Park WJ, Theda C, Maestri NE, et al. Analysis of phenotypic features and FGFR2 mutations in Apert syndrome. *Am J Hum Genet* 1995;57:321-8.
- 17 Orita M, Iwahana H, Kanazawa H, et al. Detection of polymorphisms of human DNA by gel electrophoresis as single-strand conformation polymorphisms. *Proc Natl Acad Sci USA* 1989;86:2766-70.
- 18 Kostrzewa M, Burck-Lehmann U, Müller U. Autosomal dominant amyotrophic lateral sclerosis: a novel mutation in the Cu/Zn superoxide dismutase-1 gene. *Hum Mol Genet* 1994;3:2261-2.
- 19 Ausubel FM, Brent R, Kingston RE, et al. *Current protocols in molecular biology*. New York: John Wiley, 1994.
- 20 Miki T, Bottaro DP, Fleming TP, et al. Determination of ligand-binding specificity by alternative splicing: two distinct growth factor receptors encoded by a single gene. *Proc Natl Acad Sci USA* 1992;89:246-50.
- 21 Zimmer Y, Givol D, Yayon A. Multiple structural elements determine ligand binding of fibroblast growth factor receptors. *J Biol Chem* 1993;268:7899-903.
- 22 Gray TE, Eisenstein M, Shimon T, Givol D, Yayon A. Molecular modeling based mutagenesis defines ligand binding and specificity determining regions of fibroblast growth factor receptors. *Biochemistry* 1995;34:10325-33.
- 23 Neilson KM, Friesel RE. Constitutive activation of fibroblast growth factor receptor-2 by a point mutation associated with Crouzon syndrome. *J Biol Chem* 1995;270:26037-40.
- 24 Steinberger D, Reinhartz T, Unsöld R, Müller U. FGFR2 mutation in clinically nonclassifiable autosomal dominant craniosynostosis with pronounced phenotypic variation. *Am J Med Genet* 1996;66:81-6.



## A novel mutation (a886g) in exon 5 of FGFR2 in members of a family with Crouzon phenotype and plagiocephaly.

D Steinberger, H Collmann, B Schmalenberger and U Müller

*J Med Genet* 1997 34: 420-422  
doi: 10.1136/jmg.34.5.420

---

Updated information and services can be found at:  
<http://jmg.bmj.com/content/34/5/420>

---

### Email alerting service

*These include:*

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

---

### Notes

---

To request permissions go to:  
<http://group.bmj.com/group/rights-licensing/permissions>

To order reprints go to:  
<http://journals.bmj.com/cgi/reprintform>

To subscribe to BMJ go to:  
<http://group.bmj.com/subscribe/>