Effect of Platinum-Based Chemoradiotherapy on Cellular Proliferation in Bone Marrow and Spleen, Estimated by ¹⁸F-FLT PET/CT in Patients with Locally Advanced Non–Small Cell Lung Cancer

Antoine Leimgruber^{1,2}, Andreas Möller³, Sarah J. Everitt^{4–6}, Marine Chabrot¹, David L. Ball^{4,5}, Ben Solomon^{5,7}, Michael MacManus^{4,5}, and Rodney J. Hicks^{1,5}

¹Centre for Molecular Imaging, Division of Radiation Oncology and Cancer Imaging, Peter MacCallum Cancer Centre, East Melbourne, Victoria, Australia; ²Department of Medical Imaging, Centre Hospitalier Universitaire Vaudois and University of Lausanne, Lausanne, Switzerland; ³QIMR Berghofer Medical Research Institute, Herston Queensland, Australia; ⁴Department of Radiation Oncology, Division of Radiation Oncology and Cancer Imaging, Peter MacCallum Cancer Centre, East Melbourne, Victoria, Australia; ⁵The Sir Peter MacCallum Department of Oncology, University of Melbourne, Parkville, Victoria, Australia; ⁶Department of Medical Imaging and Radiation Sciences, Monash University, Clayton, Victoria, Australia; and ⁷Medical Oncology, Peter MacCallum Cancer Centre, East Melbourne, Victoria, Australia

Historically, it has been difficult to monitor the acute impact of anticancer therapies on hematopoietic organs on a whole-body scale. Deeper understanding of the effect of treatments on bone marrow would be of great potential value in the rational design of intensive treatment regimens. 3'-deoxy-3'-18F-fluorothymidine (18F-FLT) is a functional radiotracer used to study cellular proliferation. It is trapped in cells in proportion to thymidine-kinase 1 enzyme expression, which is upregulated during DNA synthesis. This study investigates the potential of ¹⁸F-FLT to monitor acute effects of chemotherapy on cellular proliferation and its recovery in bone marrow, spleen, and liver during treatment with 2 different chemotherapy regimens. Methods: Sixty patients with non-small cell lung cancer underwent concurrent radical chemoradiotherapy to 60 Gy in 6 wk with either cisplatin/etoposide (C/E, n = 28) weeks 1 and 5 or weekly carboplatin/paclitaxel (C/P, n = 32) regimens. ¹⁸F-FLT and ¹⁸F-FDG PET with CT were performed at baseline, week 2 (day 9 for ¹⁸F-FLT and day 10 for ¹⁸F-FDG PET), and week 4 (day 23 for ¹⁸F-FLT and day 24 for ¹⁸F-FDG PET). Visual and semiquantitative standardized uptake value (SUV) measurements were performed in bone marrow outside the radiotherapy field, liver, spleen, and small bowel. These were correlated to blood counts and smears in a subset of patients. Results: The C/E group exhibited a drop in bone marrow $^{18}\mbox{F-FLT}$ uptake at week 2 (median \mbox{SUV}_{max} [maximum SUV] decrease to 31%, 8.7-6.0, P < 0.001), with recovery at week 4, reflecting the absence of chemotherapy between these times. By contrast, the weekly C/P group showed gradually declining bone marrow uptake (P > 0.05). Spleen uptake in both cohorts decreased at week 2, with intense rebound activity at week 4 (SUV_{max} week 4 at 58% above baseline: 2.4-3.8, for C/E, respectively, 30% for C/P: 2.7–3.5, P < 0.001). Liver uptake changed little. ¹⁸F-FLT changes preceded neutrophil count reductions. ¹⁸F-FDG uptake in marrow liver and spleen changed much less than ¹⁸F-FLT. Conclusion: ¹⁸F-FLT imaging may be used to quantify impair-

E-mail: rod.hicks@petermac.org

ment and recovery of bone marrow by specific chemotherapy regimens and may also enable imaging of organ-specific processes such as spleen activation. ¹⁸F-FLT is superior to ¹⁸F-FDG for this purpose. This technology may support novel treatment planning and monitoring approaches in oncology patients.

Key Words: ¹⁸F-FLT; bone marrow; spleen; hematopoiesis; chemotoxicity

J Nucl Med 2014; 55:1075–1080 DOI: 10.2967/jnumed.113.136127

C unctional tracers with whole-body imaging capability constitute the centerpiece of oncologic molecular imaging. A wide field of view facilitates a holistic assessment of disease processes and organ function in the human body because it shows organs and systems beyond the primary lesion. Molecular imaging therefore offers a clinical counterpart to systems-based approaches in fundamental research (1).

Use of combined PET and CT, with the radiotracer ¹⁸F-FDG, has significantly enhanced the management of patients with nonsmall cell lung cancer (NSCLC), including the areas of diagnosis, staging, target delineation for radiation therapy (RT), and therapeutic response monitoring (2).

¹⁸F-FDG is a well-known PET/CT tracer for the imaging of glucose metabolism pathways within tumors, but it also demonstrates variable uptake in the bone marrow. For example, ¹⁸F-FDG uptake is increased in Hodgkin disease in relation to tumor-secreted cytokines (*3,4*), and rebound uptake in the bone marrow has also been well described after cytotoxic chemotherapy (*5*). However, it is unclear to what extent these changes reflect marrow proliferation because ¹⁸F-FDG uptake is multifactorial. In contrast to this, 3'-deoxy-3'-¹⁸F-fluorothymidine (¹⁸F-FLT) is a recent functional tracer more directly linked to proliferation. The trapping of ¹⁸F-FLT in cells is related to the activity and regulation of the cytosol-based thymidine-kinase-1 enzyme (TK1) (*6*). TK1 is upregulated during DNA synthesis. The relationship between this

Received Dec. 26, 2013; revision accepted Mar. 7, 2014.

For correspondence or reprints contact: Rodney J. Hicks, Cancer Imaging, St. Andrew's Place, East Melbourne, Victoria, Australia.

Published online May 27, 2014.

COPYRIGHT © 2014 by the Society of Nuclear Medicine and Molecular Imaging, Inc.

tracer and the cellular cycle and hence, indirectly, the proliferative activity is well-documented (7).

Monitoring the impact of chemotherapy on the hematopoietic system is critical in medical oncology, given the significance of adverse effects induced by immunosuppression and thrombocytopenia (8). The overall toxicity of well-established regimens is thoroughly documented, although dosage schemes and adaptation to parameters such as body surface or renal function remain largely empiric. There are few tools that can offer insights into the toxicity of new therapy regimens or new drugs. Molecular imaging has a role to play and could provide some of the technologies needed to answer these questions and develop personalized chemotherapy schemes. In particular, the time taken for recovery in marrow proliferative function may have important implications for scheduling of chemotherapy.

There have been several recent papers reporting the utility of ¹⁸F-FLT PET/CT in monitoring hematopoietic organs, such as bone marrow and the spleen, in chronic myeloproliferative disorders or within radiotherapy fields (*9–12*), but we are unaware of any reports of time-dependent distribution patterns of ¹⁸F-FLT uptake in organs affected by cytotoxic chemotherapy regimens during treatment. A prospective trial of serial ¹⁸F-FLT and ¹⁸F-FDG PET/CT for monitoring disease response to radical chemoradiotherapy (chemo-RT) in patients with NSCLC is currently ongoing at our center (Registry no. ACTRN12611001283965). In this trial, patients are treated with 1 of 2 chemotherapy regimens (cisplatin/etoposide [C/E] or carboplatin/paclitaxel [C/P]), which have different and well-documented toxicities (*8*). Data from our study therefore presented a unique opportunity to explore these topics further.

Specifically, we hypothesized that distinct radiotracer patterns would be identifiable in response to different chemotherapy regimens. We paid particular attention to the behavior of the bone marrow and spleen, given the renewed interest in the latter in oncology (13, 14). We interrogated the stability of hepatic uptake, critical for qualitative imaging, because the liver has a potential hematopoietic role that may translate as variation of uptake during chemotherapy (15). We further investigated the interrelation between ¹⁸F-FLT and ¹⁸F-FDG in these organs and the potential for ¹⁸F-FLT uptake to anticipate the classic toxicity endpoint of peripheral blood counts and blood smears.

MATERIALS AND METHODS

Patients

Patients with stage I–IIIB NSCLC considered by our multidisciplinary tumor board to be suitable candidates for radical chemo-RT were recruited. Patients with previous thoracic RT or complete macroscopic tumor excision were excluded. Our Institutional Ethics Committee approved the trial, and patients provided written, informed consent.

Chemo-RT

Treatment planning included conformal RT (60 Gy/30 fractions, 6 wk) with concomitant C/E: cisplatin (50 mg/m²) on days 1 and 8 (cycle 1) and 29 and 36 (cycle 2) and etoposide (50 mg/m² daily during cycles 1 and 2—that is, respectively, weeks 1 and 5), or weekly C/P: carboplatin (area under the curve, 2) and paclitaxel (45 mg/m²). Dexamethasone was administered intravenously: 20 mg (C/P) and 12 mg (C/E). Chemotherapy doses were adapted to baseline renal function (electronic glomerular filtration rate). Full blood counts were recorded at baseline and at least before each chemotherapy cycle. Prospective blood sampling with full blood counts and blood smears on the day of each ¹⁸F-FLT PET/CT study was performed in the last 21 patients. Blood smears were scanned on a ScanScope XT slide scanner (20×) (Aperio). Counting of

neutrophils, lymphocytes, and monocytes was performed on eight $10\times$ fields with the ImageScope software (Leica Biosystems).

PET/CT Imaging

Imaging was performed at baseline, week 2 (day 9 for ¹⁸F-FLT and day 10 for ¹⁸F-FDG, after 14 Gy \pm 2 Gy), and week 4 (day 23 for ¹⁸F-FLT and day 24 for FDG, after 38 Gy \pm 4 Gy) of chemo-RT. Scans were acquired on an integrated PET/CT (STE [GE Healthcare] or Biograph [Siemens Medical Solutions]). Patients were weighed and intravenously injected with either 3.7 MBq of ¹⁸F-FLT per kilogram or 4.2 MBq of ¹⁸F-FDG per kilogram. Patients fasted for at least 6 h before ¹⁸F-FDG and rested for at least 1 h before scanning. Patient positioning followed our previously reported method (16). Baseline emission scans were initiated 60 min after injection, and all subsequent scans were scheduled according to the baseline uptake time. Baseline ¹⁸F-FLT scans were obtained from vertex to thighs, and baseline ¹⁸F-FDG scans were obtained from the base of the brain to thighs. Subsequent scans were limited from the thoracic inlet to the mid abdomen to reduce radiation exposure from the CT component by Institutional Ethics Committee request. Approval for ¹⁸F-FLT was granted by the Australian Therapeutic Goods Administration (Cyclotek, Bundoora, Victoria, Australia).

Image Analysis

Visual assessment of the effective radiotherapy field was evaluated on ¹⁸F-FLT scans. The uptake of the spinal bone marrow, spleen, and liver outside the radiation field was compared with the previous scan as increased, stable, or decreased by a reviewer masked to clinical data and chemotherapy regimen. For this purpose, the window width was set from a standardized uptake value (SUV) of 0–9 in all scans.

For all scans, measurements with 3-dimensional volumes of interest (VOIs) were then performed. For the bone marrow, the center of a vertebral body 2 or more vertebral levels below the visible effect of the RT field was selected (9), and mean SUV (SUV_{mean}) was measured with a 3-mL cube VOI with an isocontour 30% maximum SUV (SUV_{max}). Vertebral bodies with discordant uptake in comparison with the adjacent vertebrae or with significant morphologic abnormality such as marked degenerative changes or compression fracture were excluded. For a patient, the same vertebral body was measured in all scans. Spleen and liver uptake was measured with a single VOI centered on the organ, allowing for misregistration and avoiding the superior aspect of the organ potentially prone to direct radiation impact. SUV_{max}, SUV_{mean}, and VOIs and percentage injected dose for the spleen were recorded. For small bowel (jejunum), only 18F-FLT SUVmax was recorded because of peristaltic movement between PET and CT acquisitions. VOIs were placed at least 6 slices (>2 cm) from the limit of the field of view.

Measurements were performed on an Advantage Windows4.0 workstation (GE Healthcare) except for the small bowel, which was measured with Osirix3.9.4 (Osirix Medical Software).

Statistics

Statistical analyses were performed with Prism4.0 (GraphPad Software). Interpatient variations were considered nonnormal, and variations between time points were compared using Wilcoxon matched-pairs with 95% confidence interval statistical tests. Two-tailed tests were to evaluate increase or decrease in uptake and reduce type I statistical errors. Consistency of imaging charters was verified by Bland–Altman analyses (*17*).

RESULTS

Patients and PET/CT Scans

Sixty-seven patients were recruited between March 2009 and June 2013 (Supplemental Table 1; supplemental materials are available at http://jnm.snmjournals.org). Seven patients were



FIGURE 1. (A) Bone marrow ¹⁸F-FLT uptake decreases at week 2 (black arrow) and returns to baseline (white arrow) after week 2 to week 4 chemotherapy break in the C/E group; representative maximum-intensity-projection images from closest-to-median bone marrow SUV_{max} C/E patient. (B) SUV_{max} for C/E group. (C) Bone marrow uptake is nonsignificantly decreasing in C/P group; representative images from closest-to-median bone marrow SUV_{max} C/P patient. (D) SUV_{max} for each C/P patient. Black lines = median SUV_{max}; NS = nonsignificant. **P < 0.01.

excluded after multidisciplinary review of disease burden or withdrew consent before baseline. Sixty patients were retained, 37%(22/60) women, 63% (38/60) men; median age was 67 y in both groups (age range, 31–78 and 54–86 y, respectively). Tumor histology was adenocarcinoma in 45% (27/60), squamous cell carcinoma in 37% (22/60), large cell carcinoma in 8% (5/60), nonspecified in 7% (4/60), and neuroendocrine component in 3% (2/60). Most patients had stage III disease, and radiotherapy included the primary tumor and involved mediastinal and hilar lymph nodes. Bone marrow in the thoracic spine, ribs, and scapulae received direct or scattered radiation from radiotherapy.

Twenty-eight patients (47%) received the cisplatin/etoposide regimen and 32 (53%) a carboplatin-based regimen. Six patients did not complete the study because of disease progression and switched to palliative treatment. One hundred sixty-two and 169 PET/CT studies with ¹⁸F-FLT and ¹⁸F-FDG, respectively, were performed; 2 patients missed ¹⁸F-FLT baseline, and five ¹⁸F-FLT scans were cancelled for tracer unavailability. On average, ¹⁸F-FLT uptake time was 72 min (range, 55–118 min, 9 scans > 90 min). Mean interval between baseline uptake time and other time points was 2.2 min (range, -17 to +32 min, >15 min of difference from baseline in 7 scans). Similar values were observed for ¹⁸F-FDG imaging charters was verified (*17*).

Comparison of ¹⁸F-FLT Patterns in Bone Marrow with Each Chemotherapy Regimen

Visual assessment of images between baseline and week 2 noted decreased uptake in 72% (18/25), stable uptake in 20% (5/25),

and increased uptake in 8% (2/25) of the patients of the C/E cohort (Fig. 1A). For the C/P cohort, 43% (13/30) were decreased, 23% (7/30) stable, and 33% (10/30) increased (Fig. 1C). These results were also reflected by VOI analysis. ¹⁸F-FLT uptake outside the radiotherapy field significantly reduced in the C/E cohort, compared with baseline, with a median fall at week 2 of 31% (8.7/6.0, P < 0.001) but recovery by week 4 (42% increase from week 2, 6.0/8.6, P < 0.001) (Fig. 1B). This result probably reflects the C/E patients not receiving chemotherapy between weeks 2 and 4. However, the temporal pattern of changes in the C/P cohort differed with a nonsignificant fall from baseline to week 2 of 10% (9.1/8.2) and a continuing decreasing trend at week 4 (both P > 0.05) (Fig. 1D). SUV_{mean} measurements showed similar results (data not shown).

Recovery of ¹⁸F-FLT Uptake in Spleen

Qualitative changes in splenic uptake were also apparent during treatment. In particular, splenic uptake tended to fall at week 2 and then increase at week 4 in the C/E cohort; these changes were less apparent in the C/P cohort (Figs. 2A and 2B). Semiquantitative analysis supported this. In the C/E group, ¹⁸F-FLT uptake in the spleen decreased from baseline to week 2

by 33% (2.4/1.6, P < 0.001) but by week 4 had increased, compared with baseline by 58% (2.4/3.8, P < 0.001) and compared with week 2 by 138% (1.6/3.8, P < 0.001) (Fig. 2C). Similarly, in the C/P patients, splenic uptake had also fallen by week 2, with a reduction in SUV_{max} of 26% (2.7/2.0, P < 0.001), followed by an increase at week 4 by 30%, compared with baseline (2.7/3.5, P < 0.001), and 75% (2.0/3.5, P < 0.001), compared with week 2. We verified that there was no significant difference between the patients who had radiation to the left lower lung field and the others (data not shown).

Liver as Reference Organ for ¹⁸F-FLT Imaging

Qualitatively, liver uptake did seem to decrease during both forms of chemotherapy. This fall was significant in the C/E cohort, with a median ¹⁸F-FLT SUV_{max} decrease of 22% from baseline (6.8/5.3, P < 0.01), but the fall of 10% (7.0/6.3) in the C/P group was not statistically significant (Figs. 2A–2C). The recovery in hepatic uptake between week 2 and week 4 was significant in both patient groups (P < 0.05), although the uptake at week 4 remained significantly below baseline in the C/E patients (P < 0.05).

Small Bowel ¹⁸F-FLT Uptake

The mucosa of the small bowel is known to be sensitive to chemotherapy (18). For both chemotherapy cohorts, a significant decrease at week 2 (C/E: -32%, 2.5/1.7, P < 0.001; and C/P: -21%, 2.4/1.9, P < 0.001) and a significant increase from week 2 to week 4 in the C/E cohort only (C/E: +35%, 1.7/2.3, P < 0.001; and C/P: +16%, 1.9/2.2, P > 0.05) was demonstrated in the small bowel (Figs. 2A and 2C).



FIGURE 2. (A) ¹⁸F-FLT uptake in spleen demonstrates significant decrease at week 2 and recovery at week 4, particularly intense for C/E cohort; representative maximum-intensity-projection images from closest-to-median spleen SUV_{max} C/E patient show intense splenic activation at week 4 (white arrow). (B) Representative 8-cm-thickness maximum-intensity-projection image of another C/E patient demonstrates small bowel recovery at week 4 (white arrow). (C) SUV_{max} for each C/E patient; liver and small bowel changes are less prominent but consistent with bone marrow and spleen findings. Black lines = median SUV_{max}; NS = nonsignificant. **P* < 0.05. ****P* < 0.001.

¹⁸F-FLT Changes as Predictor of Decline of Peripheral Neutrophil Count

We prospectively sampled the blood of the last 21 patients recruited (n = 10 C/E and 11 C/P) at the time of ¹⁸F-FLT PET/CT. Daily chemotherapy for 8 d in the C/E cohort induced a more rapid decrease in peripheral blood counts than the 2 d of chemotherapy during the same interval in the C/P cohort (Supplemental Fig. 1). We verified that neutrophil counts did not decrease at week 2 but did at week 4, whereas lymphocyte and monocyte counts dropped markedly at week 2 (P < 0.05 in C/E and C/P groups) (Fig. 3). No recovery of lymphocyte counts was observed by week

4, which was confirmed by evaluating blood smears (Supplemental Fig. 2). Significant recovery of monocyte counts was observed in the C/E group (P < 0.05) whereas only a nonsignificant increase was noted in the C/P group (Fig. 3). We demonstrated no significant correlation between the intensity of ¹⁸F-FLT uptake in the organs and peripheral white blood cell counts (data not shown). Of note, patient P01 had the highest baseline bone marrow uptake and highest neutrophil counts (¹⁸F-FLT SUV_{max}, 16.8; neutrophils, 33.5 × 10⁹/L).

¹⁸F-FLT Changes and Glucose Metabolism

Significant variations of ¹⁸F-FDG SUV_{max} uptake were recorded in the bone marrow and spleen of both patient groups between baseline and week 2 and between week 2 and week 4 in the C/E group only. The variation of median SUV_{max} was less marked than for ¹⁸F-FLT, with a reduction of 15% (2.6/2.2) for bone marrow and 9% (2.4/2.2) for spleen between baseline and week 2 (Supplemental Figs. 3A and 3B). Uptake at week 4 was not significantly different from baseline in either cohort. Finally, median SUV_{max} was approximately 3-fold higher in the bone marrow and 2fold higher in the liver on ¹⁸F-FLT scans ¹⁸F-FDG scans.

DISCUSSION

Of the approximately 400 studies involving ¹⁸F-FLT PET/CT indexed in PubMed, fewer than 5% focus on bone marrow or hematopoiesis. Most studies describe a dramatic drop in ¹⁸F-FLT uptake in irradiated bone marrow (9,10,19). Others have described the relation between ¹⁸F-FLT uptake in bone marrow and chronic myeloproliferative disorders (11) or describe the distribution of proliferative bone marrow in cancer patients (20–22). A relevant study in rats demonstrated bone marrow recovery monitored by ¹⁸F-FLT after bone marrow transplantation (23). In

our study, we demonstrated the capability of ¹⁸F-FLT imaging to monitor chemotherapy injury to hematopoietic organs and to the small bowel mucosa. The findings demonstrate that rapid changes in ¹⁸F-FLT uptake occur in relation to the expression of TK1. The lack of direct correlation with blood counts, however, suggests that the uptake of ¹⁸F-FLT in marrow may be more complex than simply reflecting global proliferation.

A limitation in studying the relationship between ¹⁸F-FLT uptake in hematopoietic organs and chemotherapy in patients is that serial biopsies are neither practical nor likely to be acceptable to patients. Furthermore, peripheral blood counts do not allow the



FIGURE 3. Prospective blood collection at time of each ¹⁸F-FLT PET/CT study demonstrates in both chemotherapy regimens that neutrophils did slightly increase at week 2, at which time C/E patients' bone marrow uptake had decreased. Lymphocytes and monocytes followed change of ¹⁸F-FLT between baseline and week 2, but only monocytes in C/E group showed significant recovery. BL = baseline; NS = nonsignificant. **P* < 0.05.

separation of the contribution of different organs to hematopoiesis or their relative contribution to the composition of whole blood with respect to the different cellular lineages. To circumvent this limitation, we based our analysis on 2 subgroups of chemotherapy regimens. The difference in administration schedule, with cisplatin/etoposide delivered in a dose-intense daily infusion for a week with more than 2 wk of recovery, compared with the fractionated infusion of carboplatin/paclitaxel once a week, seems to have been the main determinant of differences in early ¹⁸F-FLT biodistribution changes. We further cross-correlated ¹⁸F-FLT uptake variation with variation of uptake in the small bowel, because sensitivity to platin-based chemotherapy and in particular decrease in tyrosine kinase activity in the small bowel crypts has been known for more than 2 decades in animals (18). The combination of these 2 approaches allowed us to provide evidence of the relevance of using ¹⁸F-FLT as a likely surrogate of cell-cycle arrest and reactivation during chemotherapy.

All patients were treated according to standard radiotherapy dose and fractionation, indicating that differences between the 2 groups were not dependent on radiotherapy effects. The increase of ¹⁸F-FLT uptake measurements between week 2 and week 4 of the C/E regimen, despite ongoing radiotherapy, is consistent with this hypothesis. The significant impact of radiation on thoracic bone marrow is documented. This impact may have induced complex effects, potentially decoupling peripheral blood counts from ¹⁸F-FLT uptake patterns.

Although we investigated acute changes in blood counts, toxicity effects at or beyond the completion of the treatment are not addressed by our study. In NSCLC during radical chemo-RT, however, oesophagitis and pneumonitis are usually the most significant acute side effects after completion of therapy (24-26).

Bone marrow ¹⁸F-FLT uptake recovery was accompanied by an even more striking recovery in the spleen. The spleen is known to progressively expand in chronic myeloproliferative disorders, with concomitant intense ¹⁸F-FLT uptake (11,27,28). This organ has been rediscovered in recent years for its role in acute nontraumatic insults. These studies in small-animal models of lung carcinoma and in myocardial infarction have demonstrated the role of the spleen as a reservoir for the expansion of bone marrowderived cells and as a supply source for monocytes relocating to the tumor or the infarction site (29,30). A small-animal study has also demonstrated the impact of splenectomy in blood counts recovery early after chemotherapy (31). Interestingly in our study, marked recovery occurs in splenic ¹⁸F-FLT uptake of C/E patients who also show synchronous recovery of circulating monocytes. This work confirmed a specific behavior of the spleen in the setting of acute hematologic toxicity. We further demonstrated that changes in ¹⁸F-FLT uptake, with its marked drop in the bone marrow of C/E patients at week 2, occurred before the fall in peripheral neutrophil counts. This drop indicates a

potential for ¹⁸F-FLT as an imaging surrogate marker of clinically relevant toxicity, similarly to work that indicated peripheral lymphocyte and monocyte counts as predictors of neutropenic events (32,33).

We further demonstrated that the liver showed minimal but significant variation of uptake in the C/E regimen both with ¹⁸F-FLT and with ¹⁸F-FDG tracers. This variation should be accounted for when using this organ as a reference in the visual comparison of iterative studies. However, it is reassuring to note that this would not lead to false-positive response to therapy assessments. Liver uptake did not increase during chemotherapy even with ¹⁸F-FLT, when decreased uptake in other organs led theoretically to increased availability of ¹⁸F-FLT for hepatic glucuronidation. The reliability of the liver as a reference organ in iterative imaging is of prime clinical importance: for example, in lymphomas, in which the value of interim ¹⁸F-FDG scans to assess early response to therapy is the subject of several clinical trials (*34*).

CONCLUSION

This study suggests that ¹⁸F-FLT PET/CT may provide novel insights into acute changes in susceptible organs involved in hematopoiesis. This systems-based approach represents a step toward the study of the individual role of the bone marrow, spleen, and possibly liver in hematologic toxicity. ¹⁸F-FLT appears to be a useful tool to monitor these phenomena and could potentially support the development of new and, ideally, personalized chemotherapy regimens in the future.

DISCLOSURE

The costs of publication of this article were defrayed in part by the payment of page charges. Therefore, and solely to indicate this fact, this article is hereby marked "advertisement" in accordance with 18 USC section 1734. This study is supported by the SICPA Foundation, The CASS Foundation, and NHMRC grant APP1003895. No other potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We kindly thank Prof. Tomas Kron and Marnie Collins for their expertise and Lisa Macfarlane, Mira Liu, and Jenny Trinh for their efficient coordination.

REFERENCES

- Holland JP, Cumming P, Vasdev N. PET of signal transduction pathways in cancer. J Nucl Med. 2012;53:1333–1336.
- Hicks RJ. Role of ¹⁸F-FDG PET in assessment of response in non-small cell lung cancer. J Nucl Med. 2009;50:31S–42S.
- Carr R, Barrington SF, Madan B, et al. Detection of lymphoma in bone marrow by whole-body positron emission tomography. *Blood.* 1998;91:3340–3346.
- Moulin-Romsee G, Hindié E, Cuenca X, et al. ¹⁸F-FDG PET/CT bone/bone marrow findings in Hodgkin's lymphoma may circumvent the use of bone marrow trephine biopsy at diagnosis staging. *Eur J Nucl Med Mol Imaging*. 2010;37: 1095–1105.
- Kostakoglu L, Goldsmith SJ. ¹⁸F-FDG PET evaluation of the response to therapy for lymphoma and for breast, lung, and colorectal carcinoma. *J Nucl Med.* 2003; 44:224–239.
- Bading JR, Shields AF. Imaging of cell proliferation: status and prospects. J Nucl Med. 2008;49:64S–80S.
- Chalkidou A, Landau DB, Odell EW, Cornelius VR, O'Doherty MJ, Marsden PK. Correlation between Ki-67 immunohistochemistry and ¹⁸F-fluorothymidine uptake in patients with cancer: a systematic review and meta-analysis. *Eur J Cancer.* 2012;48:3499–3513.
- Luo J, Leaw SJ, Xu Y, Zheng D. Comparison of cisplatin- and carboplatin-based third-generation chemotherapy in 1,014 Chinese patients with advanced nonsmall-cell lung cancer. *Med Oncol.* 2011;28:1418–1424.
- McGuire SM, Menda Y, Boles Ponto LL, Gross B, Buatti J, Bayouth JE. 3'deoxy-3'-[¹⁸F]fluorothymidine PET quantification of bone marrow response to radiation dose. *Int J Radiat Oncol Biol Phys.* 2011;81:888–893.
- Agool A, Slart RHJA, Thorp KK, et al. Effect of radiotherapy and chemotherapy on bone marrow activity. *Nucl Med Commun.* 2011;32:17–22.
- Agool A, Schot BW, Jager PL, Vellenga E. ¹⁸F-FLT PET in hematologic disorders: a novel technique to analyze the bone marrow compartment. *J Nucl Med.* 2006;47:1592–1598.
- Vanderhoek M, Juckett MB, Perlman SB, Nickles RJ, Jeraj R. Early assessment of treatment response in patients with AML using [¹⁸F]FLT PET imaging. *Leuk Res.* 2011;35:310–316.
- Bronte V, Pittet MJ. The spleen in local and systemic regulation of immunity. *Immunity*. 2013;39:806–818.
- Cortez-Retamozo V, Etzrodt M, Newton A, et al. Angiotensin II drives the production of tumor-promoting macrophages. *Immunity*. 2013;38:296–308.
- López-Guillermo A, Cervantes F, Bruguera M, Pereira A, Feliu E, Rozman C. Liver dysfunction following splenectomy in idiopathic myelofibrosis: a study of 10 patients. *Acta Haematol.* 1991;85:184–188.

- Everitt S, Hicks RJ, Ball D, et al. Imaging cellular proliferation during chemoradiotherapy: a pilot study of serial ¹⁸F-FLT positron emission tomography/computed tomography imaging for non-small-cell lung cancer. *Int J Radiat Oncol Biol Phys.* 2009;75:1098–1104.
- OSI3926g Study Team, Binns DS, Pirzkall A, et al. Compliance with PET acquisition protocols for therapeutic monitoring of erlotinib therapy in an international trial for patients with non-small cell lung cancer. *Eur J Nucl Med Mol Imaging*. 2011;38:642–650.
- Kralovánszky J, Prajda N, Kerpel-Fronius S, Gál F, Kiss F. Comparison of intestinal toxic effects of platinum complexes: cisplatin (CDDP), carboplatin (CBDCA), and iproplatin (CHIP). *Cancer Chemother Pharmacol.* 1988;21: 40–44.
- Menda Y, Boles Ponto LL, Dornfeld KJ, et al. Investigation of the pharmacokinetics of 3'-deoxy-3'-[¹⁸F]fluorothymidine uptake in the bone marrow before and early after initiation of chemoradiation therapy in head and neck cancer. *Nucl Med Biol.* 2010;37:433–438.
- Hayman JA, Callahan J, Herschtal A, et al. Distribution of proliferating bone marrow in adult cancer patients determined using FLT-PET imaging. *Int J Radiat Oncol Biol Phys.* 2011;79:847–852.
- Shields AF, Grierson JR, Dohmen BM, et al. Imaging proliferation in vivo with [F-¹⁸]FLT and positron emission tomography. *Nat Med.* 1998;4:1334–1336.
- Kenny LM. Quantification of cellular proliferation in tumor and normal tissues of patients with breast cancer by [¹⁸F]fluorothymidine-positron emission tomography imaging: evaluation of analytical methods. *Cancer Res.* 2005;65:10104– 10112.
- Awasthi V, Holter J, Thorp K, Anderson S, Epstein R. F-18-fluorothymidine-PET evaluation of bone marrow transplant in a rat model. *Nucl Med Commun.* 2010;31:152–158.
- Gomez DR, Tucker SL, Martel MK, et al. Predictors of high-grade esophagitis after definitive three-dimensional conformal therapy, intensity-modulated radiation therapy, or proton beam therapy for non-small cell lung cancer. *Int J Radiat Oncol Biol Phys.* 2012;84:1010–1016.
- Nijkamp J, Rossi M, Lebesque J, et al. Relating acute esophagitis to radiotherapy dose using FDG-PET in concurrent chemo-radiotherapy for locally advanced non-small cell lung cancer. *Radiother Oncol.* 2013;106:118–123.
- Palma DA, Senan S, Tsujino K, et al. Predicting radiation pneumonitis after chemoradiation therapy for lung cancer: an international individual patient data meta-analysis. *Int J Radiat Oncol Biol Phys.* 2013;85:444–450.
- Spivak JL, Barosi G, Tognoni G, et al. Chronic myeloproliferative disorders. Hematology (Am Soc Hematol Educ Program). 20032003:200–224.
- Tefferi A, Vainchenker W. Myeloproliferative neoplasms: molecular pathophysiology, essential clinical understanding, and treatment strategies. J Clin Oncol. 2011;29:573–582.
- Swirski FK, Nahrendorf M, Etzrodt M, et al. Identification of splenic reservoir monocytes and their deployment to inflammatory sites. *Science*. 2009;325: 612–616.
- Cortez-Retamozo V, Etzrodt M, Newton A, et al. Origins of tumor-associated macrophages and neutrophils. *Proc Natl Acad Sci USA*. 2012;109:2491– 2496.
- Wang Y, Meng Q, Qiao H, Jiang H, Sun X. Role of the spleen in cyclophosphamide-induced hematosuppression and extramedullary hematopoiesis in mice. *Arch Med Res.* 2009;40:249–255.
- Kondo M, Oshita F, Kato Y, Yamada K, Nomura I, Noda K. Early monocytopenia after chemotherapy as a risk factor for neutropenia. *Am J Clin Oncol.* 1999;22:103–105.
- Blay JY, Chauvin F, Le Cesne A, et al. Early lymphopenia after cytotoxic chemotherapy as a risk factor for febrile neutropenia. J Clin Oncol. 1996;14: 636–643.
- Juweid ME, Stroobants S, Hoekstra OS, et al. Use of positron emission tomography for response assessment of lymphoma: consensus of the imaging subcommittee of international harmonization project in lymphoma. J Clin Oncol. 2007;25:571–578.

INDEXT The Journal of NUCLEAR MEDICINE

Effect of Platinum-Based Chemoradiotherapy on Cellular Proliferation in Bone Marrow and Spleen, Estimated by ¹⁸F-FLT PET/CT in Patients with Locally Advanced Non–Small Cell Lung Cancer

Antoine Leimgruber, Andreas Möller, Sarah J. Everitt, Marine Chabrot, David L. Ball, Ben Solomon, Michael MacManus and Rodney J. Hicks

J Nucl Med. 2014;55:1075-1080. Published online: May 27, 2014. Doi: 10.2967/jnumed.113.136127

This article and updated information are available at: http://jnm.snmjournals.org/content/55/7/1075

Information about reproducing figures, tables, or other portions of this article can be found online at: http://jnm.snmjournals.org/site/misc/permission.xhtml

Information about subscriptions to JNM can be found at: http://jnm.snmjournals.org/site/subscriptions/online.xhtml

The Journal of Nuclear Medicine is published monthly. SNMMI | Society of Nuclear Medicine and Molecular Imaging 1850 Samuel Morse Drive, Reston, VA 20190. (Print ISSN: 0161-5505, Online ISSN: 2159-662X)

