



Carnevale Neto, F., Guaratini, T., Pio, C., Callegari Lopes, J. L., Gates, P., & Lopes, N. P. (2017). Effect of charge generation in ESI source on the neutral aromatic elimination mechanism in xanthophylls. *Revista Semioses*, 10(2), 70-76. DOI: 10.15202/1981996x.2016v10n2p70

Publisher's PDF, also known as Version of record

License (if available):
CC BY

Link to published version (if available):
[10.15202/1981996x.2016v10n2p70](https://doi.org/10.15202/1981996x.2016v10n2p70)

[Link to publication record in Explore Bristol Research](#)
PDF-document

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
<http://www.bristol.ac.uk/pure/about/ebr-terms>

EFFECT OF CHARGE GENERATION IN ESI SOURCE ON THE NEUTRAL AROMATIC ELIMINATION MECHANISM IN XANTHOPHYLLS

EFEITO DA GERAÇÃO DE CARGAS NA FONTE DE ESI SOBRE O MECANISMO DE ELIMINAÇÃO AROMÁTICA NEUTRA EM XANTOFILAS

FAUSTO CARNEVALE NETO

Núcleo de Pesquisas em Produtos Naturais e Sintéticos (NPPNS),
Departamento de Física e Química, Faculdade de Ciências Farmacêuticas de Ribeirão Preto,
Universidade de São Paulo, Ribeirão Preto, SP, Brasil.

THAIS GUARATINI

Núcleo de Pesquisas em Produtos Naturais e Sintéticos (NPPNS),
Departamento de Física e Química;
Departamento de Bioquímica, Instituto de Química,
Faculdade de Ciências Farmacêuticas de Ribeirão Preto,
Universidade de São Paulo, Ribeirão Preto, SP, Brasil.

PIO COLEPICOLA

Núcleo de Pesquisas em Produtos Naturais e Sintéticos (NPPNS),
Departamento de Física e Química, Faculdade de Ciências Farmacêuticas de Ribeirão Preto,
Universidade de São Paulo, Ribeirão Preto, SP, Brasil.

JOÃO LUIS CALLEGARI LOPES

Departamento de Bioquímica, Instituto de Química,
Universidade de São Paulo, São Paulo, SP, Brasil.

PAUL J. GATES

School of Chemistry, University of Bristol,
Cantock's Close, Bristol, BS8 1TS, United Kingdom.

NORBERTO P. LOPES

Departamento de Bioquímica, Instituto de Química,
Universidade São Paulo, São Paulo, SP, Brasil.
npelopes@fcfip.usp.br

1. ABSTRACT

Carotenes and xanthophylls are natural pigments with high economic relevance in chemical, cosmetics, food and pharmaceutical industries. Systematic studies of carotenoid fragmentation pathways have demonstrated that the neutral elimination of aromatic ring from the polyene chain by electrocyclization reaction produced diagnostic ions to rapidly identify their presence in mixture. However, carotenes and xanthophylls also showed the ability to produce both protonated and radical molecular species, opening up multiple acid-base

and/or redox fragmentation, which hamper carotenoids elucidation. Here we investigate the ionization and fragmentation of the radical/protonated and sodiated parent masses $[M]^{+•}$ / $[M+H]^+$ and $[M+Na]^+$ of two natural xanthophylls (canthaxanthin and fucoxanthin) and the synthetic apo- β -carotene using a Fourier-transform ion-cyclotron resonance mass spectrometer (FTICR MS). The MS/MS analysis showed that sodium adduct yielded better fragmentation of the diagnostic aromatic ring elimination in a simpler MS/MS spectra, whereas molecular ion and protonated molecule resulted in a multitude of fragments involving additional

charge-remote fragmentations and direct cleavages of the conjugated π -system by retro-ene and vinyl-allyl reactions. The results suggested that Na^+ promoted electrocyclic aromatic ring elimination by assisting the correct orbital conformation of the polyene chain, giving clearly fragments to the unambiguous determination of carotenoids in biological samples.

Keywords: Carotenoids. Xanthophylls. Pericyclic rearrangement. Electrocyclization. Aromatic ring elimination. FTICR MS.

1. INTRODUCTION

Carotenoids are 40-carbon isoprene derivatives with a conjugated polyene chain - usually up to 15 double bonds. These molecules are widely distributed in nature where they play a key role as antioxidants, singlet oxygen quenchers, membrane stabilisers and light harvesters in photosynthetic organisms.^[1-5]

Several analytical methods have been developed to detect and quantify carotenoids based on their chemical reactivity and light-absorbing properties.^[6, 7] Liquid chromatography coupled with photodiode array (LC-UV) is by far the most common method, but the spectra similarity demand a complete LC resolution to assure their identity.^[6, 8] Liquid chromatography coupled with tandem mass spectrometry (LC-MS/MS) can improve identification of carotenoids by providing unique structural features in small amounts of very low concentration samples.^[6, 9]

Carotenoids have been analyzed by MS using different ionization methods including electron impact (EI), fast atom bombardment (FAB), matrix-assisted laser desorption/ionization (MALDI), electrospray (ESI), atmospheric pressure chemical ionization (APCI).^[9-12]

Recently, it was reported the full fragmentation of 16 carotenes and xanthophylls by electrospray ionization tandem mass spectrometry (ESI-CID-MS/MS) and nanospray tandem mass spectrometry (nanoESI-CID-MS/MS) where it was demonstrated that fragmentation of protonated carotenoids produced a multitude of fragments derived from charge-remote fragmentations and pericyclic rearrangements, such as electrocyclic and retro-ene eliminations dependent on cis-trans isomerization and the formation of several

conjugated polyene carbocation intermediates.^[13]

Additionally, previous studies had indicated that carotenoids and xanthophylls can be ionized by a radical process due to lower oxidation potentials (<1.0 V versus $\text{Ag}/\text{AgCl}/\text{sat}$), forming molecular ion besides protonated molecule in ESI (positive mode) source.^[14-16] This ability to produce radical molecular species by loss of one or two electrons open up multiple fragmentation routes based on distinguished gas-phase mechanisms - acid-base and/or redox reactions - which hamper structural elucidation.^[13]

Despite all information regarding carotenoids ionization and fragmentation, their structural diversity, the presence of *cis/trans*-isomers and the formation of radical molecular ion still make identification of carotenoids by LC-MS, a challenge.

In order to rapidly identify carotenoids in mixture, this work proposes to develop a novel method focused on the specific neutral elimination of aromatic ring in carotenoids.

Generally, the presence of carotenoids can be achieved based on the detection of toluene (92 u) and xylene (106 u) neutral losses.^[13, 17, 18] However, ionization of carotenes and xanthophylls into $[\text{M}]^{+\bullet}$, $[\text{M}+\text{H}]^+$ and/or $[\text{M}+\text{Na}]^+$ results in multiple fragmentation pathways and an unclear generation of diagnostic aromatic ring neutral losses, competing with other charge-remote fragmentations and pericyclic rearrangements, such as retro-ene eliminations.^[13]

Here we investigate the ionization and fragmentation of the radical/protonated and sodiated parent masses $[\text{M}]^{+\bullet}/[\text{M}+\text{H}]^+$ and $[\text{M}+\text{Na}]^+$ of two natural xanthophylls, canthaxanthin (**1**) and fucoxanthin (**2**) and the synthetic apo- β -carotene (**3**), figure 1, using an Fourier-transform ion-cyclotron resonance mass spectrometer (FTICR MS) focused on the systematic neutral losses of toluene (92 u) and xylene (106 u).

capacity to form radical, protonated and/or sodiated molecules, the understanding of the multiple fragmentation pathways with distinguished gas-phase mechanisms from the selective collision induced dissociation (CID) analysis of $M^{+\bullet}$, $[M+H]^+$ and $[M+Na]^+$ can give rise to characteristic MS/MS profiles and enhanced the detection of target ions, in our case, the neutral losses of 92 and 106 u.^[9, 13, 20]

According to the major product ions from tandem mass spectra (available in the Support-

ing Information and are summarized in Table 1), all molecules produced distinct fragment structures for $[M+H]^+$ and $[M+Na]^+$, with sodium adduct yielding better fragmentation of the diagnostic aromatic elimination in a simpler MS/MS spectra, whereas the protonated molecules resulted in a multitude of fragments involving additional charge-remote fragmentations and direct cleavages of the conjugated π -system by retro-ene and vinyl-allyl reactions.

Table 1. Tabulated data for the positive ion CID-MS/MS product ions from both the ESI-FTICR analysis of the protonated and sodiated carotenoids.

Carotenoid	observed m/z	Electrocyclic reaction**	other abundant ions
1	$[M+H]^+$	473, 459, 441	203, 215, 255, 269, 293
	$[M+Na]^+$	495, 481	277
2	$[M+H]^+$	549, 531	391, 355, 275, 213, 193, 149
	$[M+Na]^+$	589	449, 357, 327, 221, 215, 175
3	$[M+H]^+$	-	337, 291, 235, 221, 171, 157, 119
	$[M+Na]^+$	391	327, 295, 202, 189, 165, 152, 115

** Ions represent elimination of toluene (92 u), xylene (106 u) and possible additional losses of water.

In the case of canthaxanthin, a xanthophyll with one keto group in each ionone ring, the $[M+H]^+$ produced the neutral elimination of toluene and xylene at low intensities (m/z 473 and 459, respectively), whereas these diagnostic frag-

ments were the main ions of $[M+Na]^+$ (m/z 495 as base peak and 481), as depicted in figure 2. Additionally, the protonated form showed ions related to dehydration and cleavage of polyene chain.

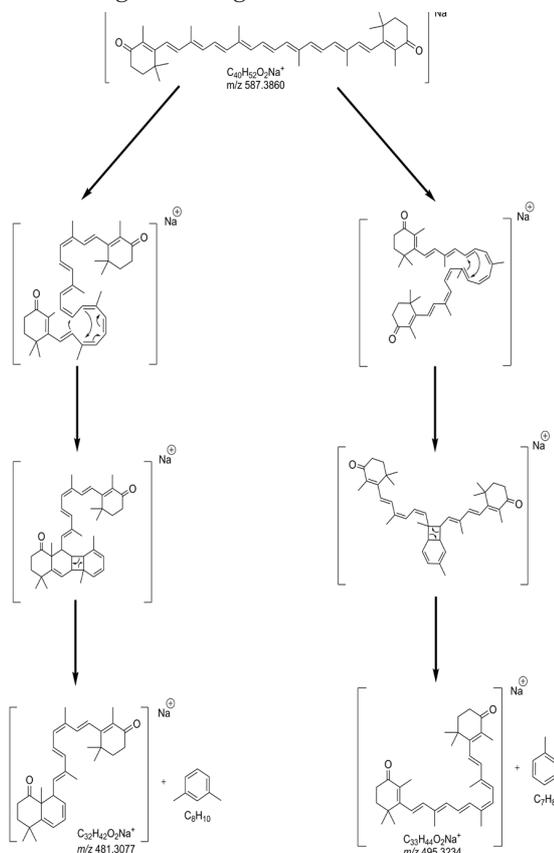


Figure 2. Proposed neutral losses of toluene and xylene from sodiated canthaxanthin adduct.

Similar results were obtained for fucoxanthin, a xanthophyll with an unusual allenic bond besides epoxy, hydroxyl and carbonyl moieties. The major fragments of $[M+H]^+$ were related to dehydration and the neutral loss of

acetic acid (60 u) by 1,4 elimination, while the aromatic ring were not observed. On the other hand, $[M+Na]^+$ showed the neutral loss of 92 u (m/z 589) although at low intensity, as indicated in figure 3.

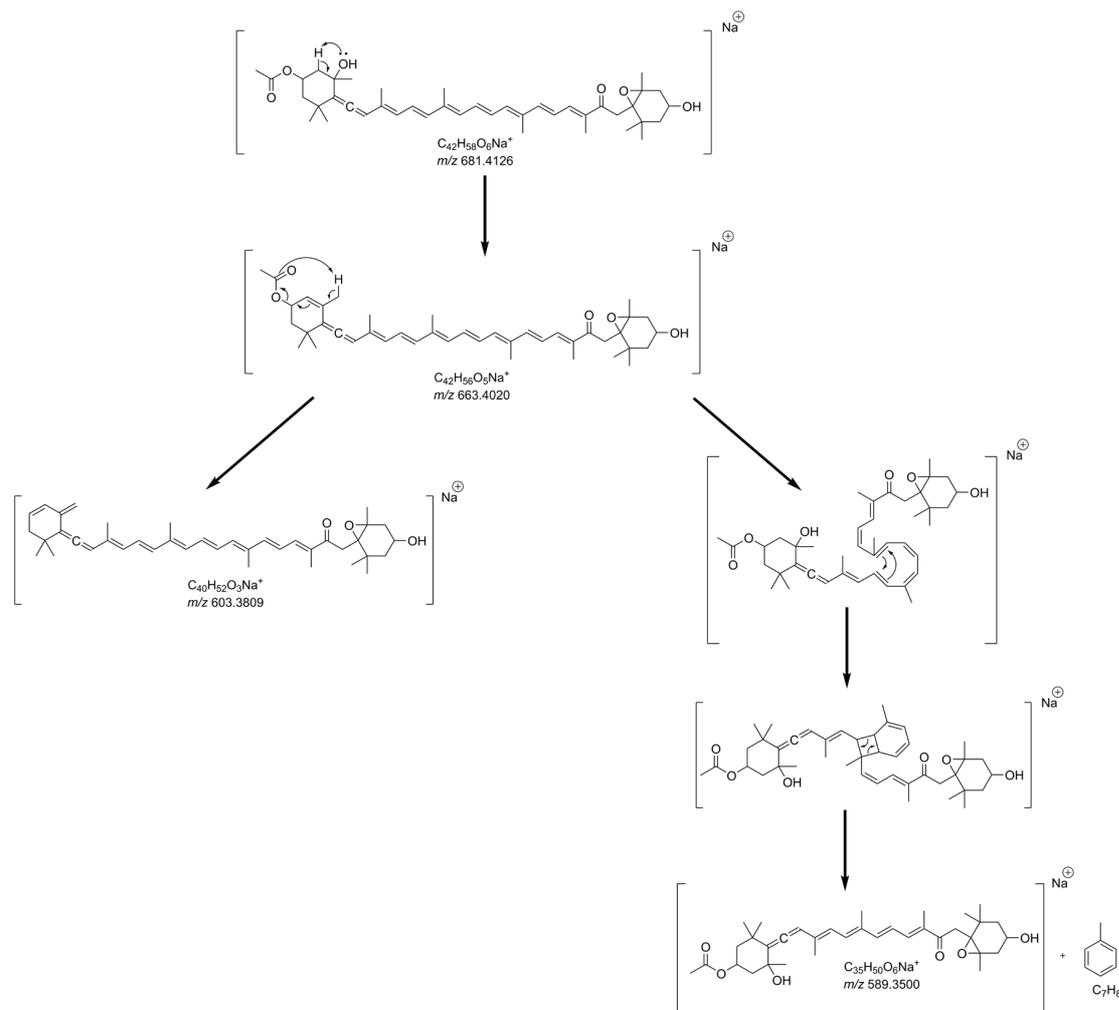


Figure 3. Proposed neutral losses of toluene from sodiated fucoxanthin adduct.

Ethyl-8'-apo- β -carotene-8'-oate also generated different MS/MS spectra. For $[M+H]^+$ several fragments were observed; however none represented characteristic aromatic ring eliminations. In case of $[M+Na]^+$, represented in figure 4, the elimination of toluene (m/z 391) is clearly detected in a simpler MS/MS spectrum.

Coordination reactions have already been described to enhance the detection of target substances and to understand fragmentation

pathways using different ions such as silver, sodium, cobalt, nickel, copper, manganese, zinc, calcium, barium, potassium, lithium and others.^[21, 22] The chemical interaction of the metal atom through formation of metal complexes with ions generated in ESI source induces variations in the geometry and symmetry of coordinated molecules which can affected the intensity and number of fragment ions. In carotenoids, the neutral losses of 92 and 106 u, explained

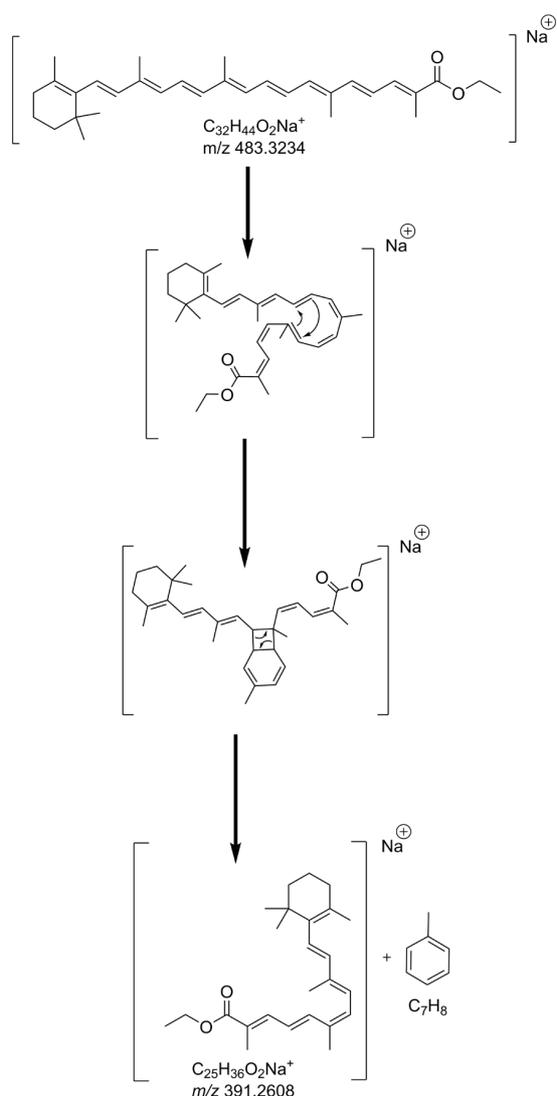


Figure 4. Proposed neutral losses of toluene from sodiated ethyl-8'-apo- β -carotene-8'-oate adduct.

by a sequential $8\pi/6\pi$ electron electrocyclization, required the correct orbital orientation and γ -hydrogen distribution, achieved through carbocation formation and cis-trans π -system conversion.^[13]

According to the results, the coordination between sodium and the carotenoids suggested that the characteristically distorted tetrahedral geometry favoured the electrocyclic elimination, e.g., the spatial requirements to the symmetry allowed disrotatory ring-closure of the cyclo-octatetraene observed in the electrocyclization followed by formation of a four-membered ring intermediate.^[13, 22]

Additionally, previous studies indicated

that an increase in the metal ionic radius led to decrease in metal electronegativity and, consequently, an increase in the metal-ligand average distance. In case of $[M+Na]^+$, the sodium's electronegativity and ionic radius permits the attraction of the electrons to start the electrocyclic elimination, but not to produced significantly more fragment ions through retro-ene eliminations of the polyene chain.

4. CONCLUSIONS

This study described the ionization and fragmentation of carotenoids standards in FTICR MS, showing that sodium adduct parent masses $[M+Na]^+$ gave much simpler MS/MS spectra than $[M]^+$ and $[M+H]^+$ parents. Coordination with sodium resulted in diagnostic aromatic ring eliminations from polyene chain, whereas protonated molecules generated a multitude of fragment ions, mainly direct cleavages of the conjugated π -system by retro-ene and vinyl-allyl reactions. These findings indicated that sodium ionic radius and electronegativity are related to the intensity and number of fragment ions and demonstrate its ability to induce electrocyclic aromatic ring elimination by assisting the correct orbital conformation of the polyene chain, giving clearly fragments to the unambiguous determination of carotenoids in biological samples. The results suggested that Na^+ promoted electrocyclic aromatic ring elimination by assisting the correct orbital conformation of the polyene chain, giving clearly fragments to the unambiguous determination of carotenoids in biological samples.

5. ACKNOWLEDGEMENTS

The authors thanks to Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP - N.P.L. grant n^o 2014/50265-3 and F.C.N. grant n^o 2014/12343-2), CAPES (Coordenação de Aperfeiçoamento de Pessoal de Nível Superior) and CNPq/INCT-if (Conselho Nacional de Desenvolvimento Científico e Tecnológico) for fellowships and financial support.

REFERENCES

- R. Edge, D. McGarvey, T. Truscott. *The carotenoids as anti-oxidants—a review. Journal of Photochemistry and Photobiology B: Biology* **1997**, 41, 189.

- **E. Pinto, L. H. Catalani, N. P. Lopes, P. Di Mascio, P. Colepicolo.** *Peridinin as the major biological carotenoid quencher of singlet oxygen in marine algae Gonyaulax polyedra.* *Biochemical and biophysical research communications* **2000**, 268, 496.
- **M. Havaux.** *Carotenoids as membrane stabilizers in chloroplasts.* *Trends in Plant Science* **1998**, 3, 147.
- **N. E. Holt, D. Zigmantas, L. Valkunas, X.-P. Li, K. K. Niyogi, G. R. Fleming.** *Carotenoid cation formation and the regulation of photosynthetic light harvesting.* *Science* **2005**, 307, 433.
- **A. J. Young, G. M. Lowe.** *Antioxidant and prooxidant properties of carotenoids.* *Archives of Biochemistry and biophysics* **2001**, 385, 20.
- **S. Rivera, R. Canela-Garayoa.** *Analytical tools for the analysis of carotenoids in diverse materials.* *Journal of Chromatography A* **2012**, 1224, 1.
- **S. M. Rivera Vélez.** *Guide for Carotenoid Identification in Biological Samples.* *Journal of natural products* **2016**.
- **T. Guaratini, R. Vessecchi, E. Pinto, P. Colepicolo, N. P. Lopes.** *Balance of xanthophylls molecular and protonated molecular ions in electrospray ionization.* *Journal of mass spectrometry* **2005**, 40, 963.
- **S. M. Rivera, P. Christou, R. Canela-Garayoa.** *Identification of carotenoids using mass spectrometry.* *Mass spectrometry reviews* **2014**, 33, 353.
- **P. Crupi, R. A. Milella, D. Antonacci.** *Simultaneous HPLC-DAD-MS (ESI+) determination of structural and geometrical isomers of carotenoids in mature grapes.* *Journal of mass spectrometry* **2010**, 45, 971.
- **T. Maoka.** *Carotenoids in marine animals.* *Marine drugs* **2011**, 9, 278.
- **R. B. van Breemen, L. Dong, N. D. Pajkovic.** *Atmospheric pressure chemical ionization tandem mass spectrometry of carotenoids.* *International journal of mass spectrometry* **2012**, 312, 163.
- **F. Carnevale Neto, T. Guaratini, L. Costa-Lotufo, P. Colepicolo, P. J. Gates, N. P. Lopes.** *Re-investigation of the fragmentation of protonated carotenoids by electrospray ionization and nanospray tandem mass spectrometry.* *Rapid Communications in Mass Spectrometry* **2016**, 30, 1540.
- **T. Guaratini, P. J. Gates, E. Pinto, P. Colepicolo, N. P. Lopes.** *Differential ionisation of natural antioxidant polyenes in electrospray and nanospray mass spectrometry.* *Rapid Communications in Mass Spectrometry* **2007**, 21, 3842.
- **R. Vessecchi, A. M. Crotti, T. Guaratini, P. Colepicolo, S. Galembeck, N. Lopes.** *Radical ion generation processes of organic compounds in electrospray ionization mass spectrometry.* *Mini-Reviews in Organic Chemistry* **2007**, 4, 75.
- **T. Guaratini, R. L. Vessecchi, F. C. Lavarda, P. M. M. Campos, Z. Naal, P. J. Gates, N. P. Lopes.** *New chemical evidence for the ability to generate radical molecular ions of polyenes from ESI and HR-MALDI mass spectrometry.* *Analyst* **2004**, 129, 1223.
- **T. Guaratini, N. P. Lopes, E. Pinto, P. Colepicolo, P. J. Gates.** *Mechanism for the elimination of aromatic molecules from polyenes in tandem mass spectrometry.* *Chemical communications* **2006**, 4110.
- **S. K. Bijttebier, E. D'Hondt, N. Hermans, S. Apers, S. Voorspoels.** *Unravelling ionization and fragmentation pathways of carotenoids using orbitrap technology: a first step towards identification of unknowns.* *Journal of Mass Spectrometry* **2013**, 48, 740.
- **N. J. Coughlan, B. D. Adamson, K. J. Catani, U. Wille, E. J. Bieske.** *Ion Mobility Unlocks the Photofragmentation Mechanism of Retinal Protonated Schiff Base.* *The Journal of Physical Chemistry Letters* **2014**, 5, 3195.
- **Y. Weesepeel, J. P. Vincken, R. M. Pop, K. Liu, H. Gruppen.** *Sodiation as a tool for enhancing the diagnostic value of MALDI-TOF/TOF-MS spectra of complex astaxanthin ester mixtures from Haematococcus pluvialis.* *Journal of Mass Spectrometry* **2013**, 48, 862.
- **D. B. Silva, E. D. Rodrigues, G. V. J. da Silva, N. P. Lopes, D. C. R. de Oliveira.** *Post-column sodiation to enhance the detection of polyacetylene glycosides in LC-DAD-MS analyses: an example from Bidens gardneri (Asteraceae).* *Talanta* **2015**, 135, 87.
- **N. P. Lopes, F. A. Almeida-Paz, P. J. Gates.** *Influence of the alkali metal cation on the fragmentation of monensin in ESI-MS/MS.* *Revista Brasileira de Ciências Farmacêuticas* **2006**, 42, 363.