

Characterization of the antigenic phenotype of α B-crystallin-expressing peripapillary glial cells in the developing chick retina

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Abstract: Radial glia are transdifferentiated into astrocytes within the developing brain and spinal cord. The neural retina contains Müller cells, which are retinal radial glia. Some of the cells that surround the optic nerve head among Müller cells in the chicken retina are called peripapillary glial cells (PPGCs). PPGCs express different molecules compared to typical Müller cells. However, an antigenic PPGC phenotype has not yet been clearly established. In this study, we classified the antigenic PPGC phenotypes and identified the differentiation stages of these cells. At embryonic day (E)8, α B-crystallin-positive PPGCs had a bipolar shape with long processes that traversed entire layers of the retina. Pax2 and vimentin were expressed in α B-crystallin-positive PPGCs. Glial fibrillary acidic protein (GFAP) immunoreactivity was not observed in PPGCs. At E18, α B-crystallin immunoreactivity disappeared from the vitread processes of PPGCs. However, the PPGC cell bodies and ventricular processes contained α B-crystallin protein, and the PPGCs retained the same Pax2-positive/vimentin-positive/GFAP-negative profile as that seen at E8. At post-hatch day 120, α B-crystallin and Pax2 immunoreactivity was not observed, but vimentin and GFAP expression was clearly observed in the presumptive location of the PPGCs. Furthermore, these two proteins overlapped within that location. Considering that vimentin expression is prolonged until the post-hatching period in chicken brain, these findings suggest that Pax2-negative/vimentin-positive/GFAP-positive PPGCs are phenotypically identical to mature astrocytes in this avian species.

Key words: Peripapillary glial cell, α B-crystallin, Pax2, Vimentin, Glial fibrillary acidic protein

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Introduction

Radial glia are cells that serve as scaffolding for the radial migration of neuroblasts in the central nervous system (CNS). After neuroblast migration is complete, radial glia are transformed into astrocytes [1-3]. It is generally accepted that vimentin-positive radial glia are astrocyte precursor cells (APCs). Recently, the lineage of astrocytic cells was

classified by antigenic phenotyping into four distinct stages in the retina of a rodent: Pax2-positive/vimentin-positive/glia fibrillary acidic protein (GFAP)-negative APCs, Pax2-positive/vimentin-positive/GFAP-positive immature perinatal astrocytes, Pax2-positive/vimentin-negative/GFAP-positive mature perinatal astrocytes, and Pax2-negative/vimentin-negative/GFAP-positive astrocytes [4]. Based on these criteria, the degree of maturation of each astrocyte can be determined.

Müller cells are radial glia in the retina. Although Müller cells help in the migration of immature neurons during retinal development, they remain in the retina after migration of ganglion cells ceases. A subpopulation of Müller cells, called peripapillary glial cells (PPGCs), are located adjacent to the optic nerve head (ONH) in the chicken retina [5-7]. PPGCs

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have a similar shape to Müller cells and bear bidirectional processes; the vitread processes extend toward the vitreous body, and the ventricular processes protrude toward the subretinal space. However, PPGCs are dissimilar to Müller cells with regard to antigenic phenotype [7, 8]. For example, glutamine synthetase, a specific Müller cell antigen, is not expressed in PPGCs. We recently reported that α B-crystallin, a lens protein, is exclusively expressed in PPGCs during chick retina development [8]. However, a role for PPGCs in retinal development and the phenotypic characteristics of PPGCs are still unknown. To determine if PPGCs are phenotypically similar to radial glia in the brain, we labeled PPGCs with anti- α B-crystallin antibody and tested the antigenic phenotype of these cells based on the criteria of Chan-Ling et al. [4]. We demonstrated that α B-crystallin-labeled developing PPGCs had the antigenicity of APCs, similar to radial glia in the brain; however, the antigenicity of mature PPGCs was analogous to that of mature astrocytes. We first classified the PPGC antigenic phenotypes and identified the differentiation stage of these cells.

Materials and Methods

Experimental animals

This work was approved by the institutional animal care and use committee of Chungbuk National University (approval no. CBNUA-092-0906-01). Post-hatch day 120 (P120) chickens were used in this experiment as adults. Fertilized eggs (Pulmuwon, Seoul, Korea) were incubated at 38°C in a humid atmosphere until they reached the appropriate embryonic (E) day, according to the Hamburger and Hamilton method [9].

Tissue preparation

Eyes were quickly obtained from decapitated adult chickens and chick embryos, and the cornea, lens, and vitreous body were sequentially removed from each. Tissue was placed in 4% paraformaldehyde in phosphate buffered saline (PBS) for 3 hours. The tissue was cryoprotected by serial sucrose infiltration, embedded in OCT compound, and frozen rapidly. Tissue specimens were cut into 10-20 μ m horizontal sections on a cryostat (CM 3050, Leica, Wetzlar, Germany).

Immunohistochemical staining

Retinal sections were incubated overnight at 4°C with a monoclonal anti- α B-crystallin (1 : 500, Enzo Life Sciences International, Plymouth Meeting, PA, USA) antibody diluted in PBS. Subsequently, immunohistochemistry was performed using the avidin-biotin peroxidase complex method, as described previously [8]. After the monoclonal antibody incubation, the sections were incubated with biotinylated horse anti-mouse IgG (Vector Laboratories, Burlingame, CA, USA) diluted 1 : 400 in PBS. The sections were washed in PBS and incubated with the avidin-biotin-peroxidase complex (Vector Laboratories). Next, the sections were reacted with 0.05% 3,3'-diaminobenzidine and a 0.01% solution of hydrogen peroxide in PBS. The immunoreactions were observed under a multipurpose microscope (DMLB, Leica) with an attached digital camera system (Macrofire, Optronics, East Muskogee, OK, USA).

Double immunofluorescence

For double-immunostaining, mouse anti- α B-crystallin, mouse anti-vimentin (1 : 2, 3CB2, Developmental Studies Hybridoma Bank, University of Iowa, Iowa City, IA, USA), rabbit anti-Pax2 (1 : 100, Covance, Princeton, NJ, USA), and rabbit anti-GFAP (1 : 200, Dako, Glostrup, Denmark) antibodies were used as primary antibodies. For secondary antibodies, Cy2-labeled anti-rabbit IgG (1 : 300, Jackson ImmunoResearch Laboratories, West Grove, PA, USA) and Cy3-labeled anti-mouse IgG (1 : 500, Jackson ImmunoResearch Laboratories) were used. Stained samples were observed under a confocal microscope (TCS SP2 AOBS, Leica).

Results

α B-crystallin expression in PPGCs from E8 to adult

Immunostaining for α B-crystallin was performed to identify morphological changes in the α B-crystallin-positive PPGCs following maturation. At the early developmental stage (E8), α B-crystallin-immunoreactive PPGCs adjacent to the ONH had a bipolar shape with long processes traversing entire layers of the retina (arrowheads in Fig. 1A). At the late developmental stage (E18), α B-crystallin immunoreactivity disappeared in the vitread processes of PPGCs (arrows in Fig. 1B). However, the cell bodies and ventricular processes of the PPGCs contained the α B-crystallin protein (arrowheads in

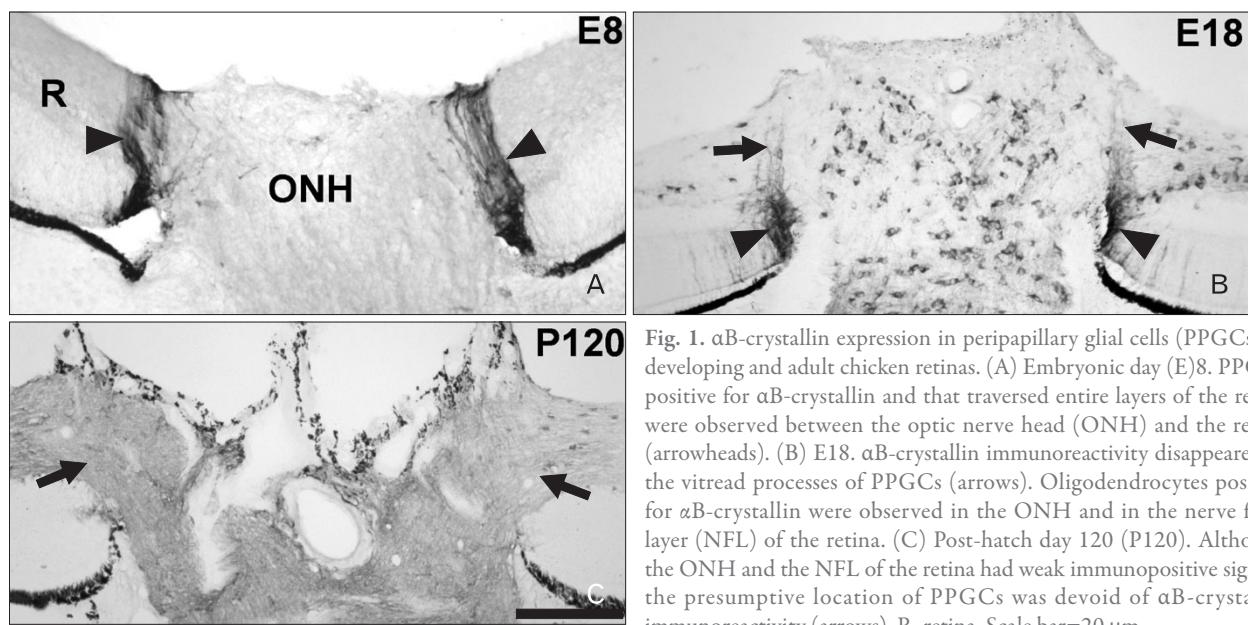


Fig. 1. α B-crystallin expression in peripapillary glial cells (PPGCs) of developing and adult chicken retinas. (A) Embryonic day (E)8. PPGCs positive for α B-crystallin and that traversed entire layers of the retina were observed between the optic nerve head (ONH) and the retina (arrowheads). (B) E18. α B-crystallin immunoreactivity disappeared in the vitread processes of PPGCs (arrows). Oligodendrocytes positive for α B-crystallin were observed in the ONH and in the nerve fiber layer (NFL) of the retina. (C) Post-hatch day 120 (P120). Although the ONH and the NFL of the retina had weak immunopositive signals, the presumptive location of PPGCs was devoid of α B-crystallin immunoreactivity (arrows). R, retina. Scale bar=20 μ m.

Fig. 1B). Additionally, α B-crystallin immunoreactivity was also observed in oligodendrocytes of the ONH and the nerve fiber layer (NFL) of the retina, similar to our previous report [8]. In the adult (P120), α B-crystallin immunoreactivity was not observed in presumptive PPGC locations (arrows in Fig. 1C). Instead, α B-crystallin-immunopositive signals were weakly observed in the ONH and the NFL of the retina.

Pax2, vimentin, and GFAP expression in α B-crystallin-positive PPGCs

Pax2, vimentin, and GFAP expression were observed in α B-crystallin-positive PPGCs to identify the astrocytic phenotypes of PPGCs. At E8 and E18, Pax2 and vimentin were expressed in α B-crystallin-positive PPGCs (Fig. 2A, B, D, E). However, GFAP immunoreactivity was not observed in PPGCs (Fig. 2C, F). Thus, developing PPGCs exhibited a Pax2-positive/vimentin-positive/GFAP-negative profile. Pax2 and vimentin were also expressed in PPGCs in the ONH at these ages. GFAP immunoreactivity was not observed in the PPGCs of the retina and the ONH at E8, suggesting that astrocyte maturation had not yet occurred in that region (Fig. 2C). In contrast to E8, GFAP was highly expressed in the ONH at E18. Interestingly, GFAP expression was not ever observed in α B-crystallin-positive PPGCs (Fig. 2F). In the adult (P120), Pax2 immunoreactivity was not observed in the presumptive location of PPGCs (data not shown). This result indicates that Pax2-expressing immature astrocytes and PPGCs lose their Pax2 antigenicity as they mature. Vimentin

and GFAP were clearly observed in the presumptive location of PPGCs in the retina (dotted areas in Fig. 2G, H), as well as in the ONH. The expression of these two proteins overlapped within each location (dotted area in Fig. 2I). Mature PPGCs at P120 were Pax2-negative/vimentin-positive/GFAP-positive. Fig. 3 depicts a scheme of the astrocytic phenotype of α B-crystallin-positive PPGCs.

Discussion

α B-crystallin expression in PPGCs

Because of the absence of specific molecules for marking PPGCs, it has been difficult to distinguish these cells from surrounding structures using Golgi stains or electron microscopy. Recently, we found that α B-crystallin was exclusively expressed in PPGCs and some oligodendrocytes in the developing chick retina, and reported that α B-crystallin is a useful marker protein for observing PPGCs [8]. The results of the present study show that α B-crystallin expression in PPGCs decreased from E18 and ceased at P120. These findings raise two possibilities. One possibility is that PPGCs are transformed thoroughly into typical astrocytes, including their shape. The other possibility is that PPGC features are retained and only α B-crystallin expression decreases within PPGCs. Based on their data from previous studies, a group of researchers have reported that PPGCs have a bipolar shape with a long vitread process and a short ventricular process at

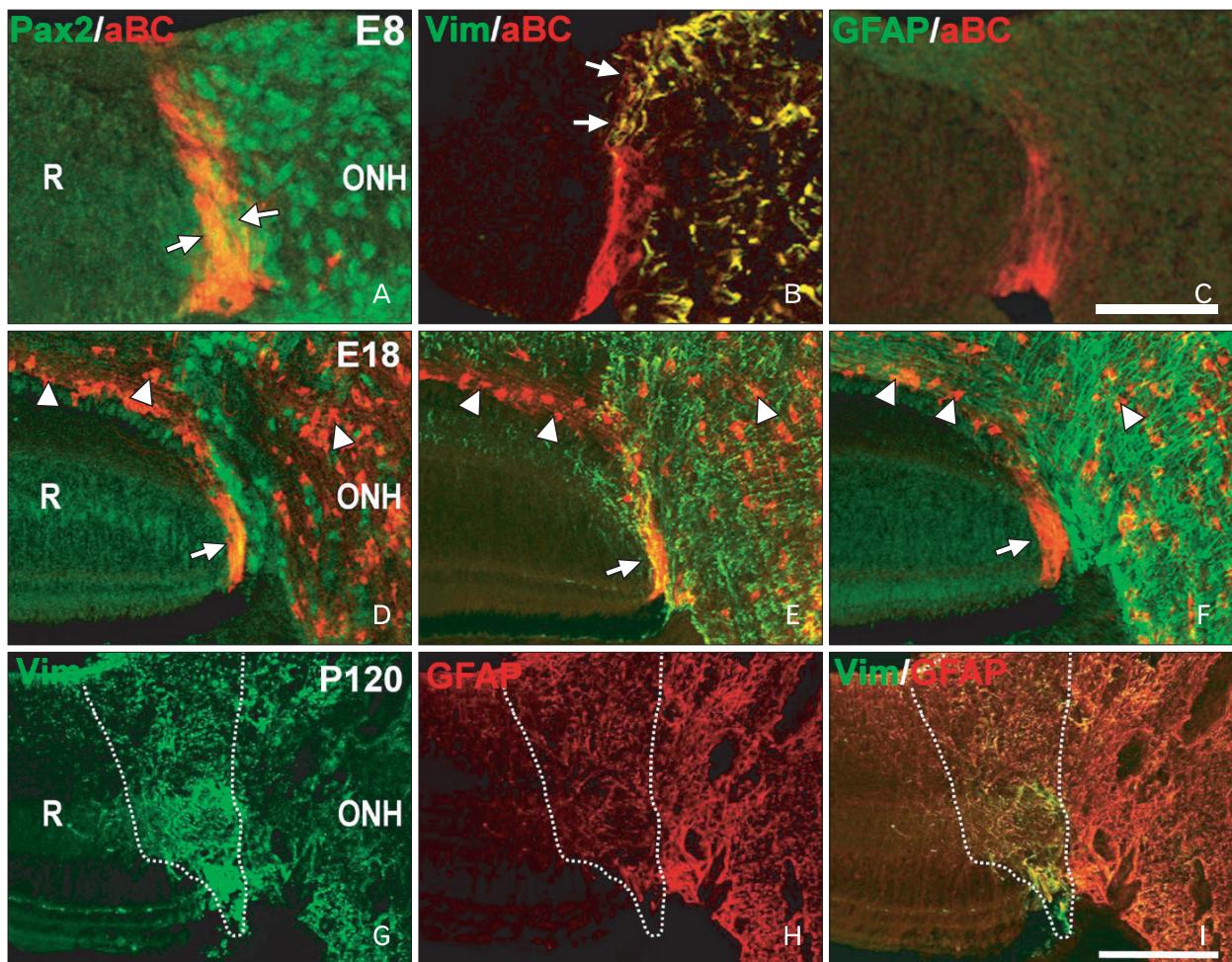


Fig. 2. Pax2, vimentin, and glial fibrillary acidic protein (GFAP) expression in α B-crystallin-positive peripapillary glial cells (PPGCs) in the developing and adult chick retina. (A-C) Embryonic day (E)8. Pax2 (green in A) and vimentin (green in B) immunoreactivities overlapped with α B-crystallin immunoreactivity (red in A and B) in PPGCs. GFAP (green in C) was not expressed in α B-crystallin-positive PPGCs (red in C) in the retina or the optic nerve head (ONH). (D-F) E18. Pax2 (green in D) and vimentin (green in E) were expressed in α B-crystallin-positive PPGCs (red in D and E). A large amount of GFAP (green in F) was expressed in the ONH but not in PPGCs. At this stage, α B-crystallin immunoreactivity was expressed in oligodendrocytes of the ONH and the retina (arrowheads in D-F). (G-I) Post-hatch day 120 (P120). Vimentin and GFAP expression was observed in the presumptive location of PPGCs (dotted areas of G and H). Furthermore, GFAP immunoreactivity overlapped with vimentin immunoreactivity within the same location (dotted area of I). aBC, α B-crystallin; R, retina; Vim, vimentin. Scale bar in C=10 μ m (A-C), I=20 μ m (D-I).

early developmental stages, have lateral processes extending off of the original vitread process at middle stages, and finally have numerous fibrous lateral processes at later embryonic stages and at the adult stage. However, in their report, the PPGC processes did not shorten during maturation [6, 7, 10]. Therefore, the first possibility that PPGCs may be transformed into astrocytes like other radial glia during development is excluded. However, the reason why α B-crystallin is temporally expressed within PPGCs is currently unclear. Given the fact that α B-crystallin is a member of the small heat-shock protein family [11-13] and acts as a molecular chaperone [14, 15], it

is conceivable that α B-crystallin plays a role in the survival of developing PPGCs.

Astrocytic PPGC phenotype

Pax2 is a transcription factor that plays a critical role in determining cell fate in the CNS during development [16]. Recently, it was reported that Pax2-expressing cells are immature astrocytic cells [4, 17, 18]. In the present study, developing PPGCs were Pax2-positive/vimentin-positive/GFAP-negative until hatching. Chan-Ling et al. [4] defined Pax2-positive/vimentin-positive/GFAP-negative

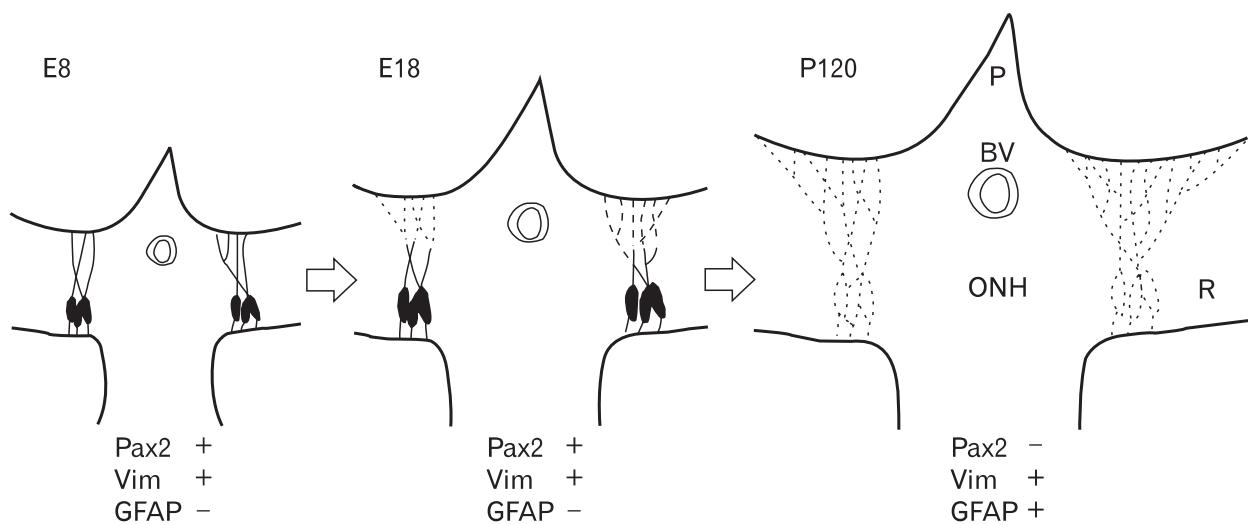


Fig. 3. Schematic drawing showing the astrocytic phenotype of α B-crystallin-positive peripapillary glial cells (PPGCs). Lines indicate α B-crystallin-positive PPGCs and dotted lines indicate the presumptive location of PPGCs devoid of α B-crystallin immunoreactivity. At the early stage (embryonic day [E]8), the morphology of α B-crystallin-immunoreactive PPGCs was bipolar with a Pax2-positive/vimentin-positive/glial fibrillary acidic protein (GFAP)-negative phenotype. At the late stage (E18), α B-crystallin immunoreactivity disappeared within the vitread processes of PPGCs, which had the same antigenic phenotype as E8 cells. In the adult retina (post-hatch day 120 [P120]), a number of secondary processes extended from the primary vitread processes of PPGCs to form a densely fibrous triangular area. Mature PPGCs have a Pax2-negative/vimentin-positive/GFAP-positive phenotype. However, α B-crystallin was not expressed in mature PPGCs. BV, blood vessel; ONH, optic nerve head; P, pecten; R, retina; Vim, vimentin.

cells phenotypically as APCs and demonstrated that APCs are highly proliferative. We injected BrdU into the yolks of E6, E14, and E18 eggs and harvested embryos 6 hours later to examine the proliferative activity of developing PPGCs. Consequently, few BrdU-labeled cells were observed in the presumptive location of PPGCs at all three stages (data not shown). Although PPGCs are phenotypically APCs, they do not show proliferative activity. These data strongly suggest that astrocytes in the ONH are not derived from PPGCs.

Several studies have indicated that astrocytes are absent in the chicken retina [8, 19]. However, a recent study reported the presence of retinal astrocytes in the chick [20]. Therefore, this issue is still controversial. Our result that Pax2-expressing astrocytes were observed in the ONH from E8 to E18, but not in the retina, strongly indicate the absence of retinal astrocytes. Indeed, Pax2 expression was not detected in the ONH or the retina at P120, suggesting that Pax2-positive immature astrocytes completely differentiated into Pax2-negative mature astrocytes in the adult eye. Additionally, Pax2-negative/vimentin-negative/GFAP-positive cells of the rodent and human retina are considered mature astrocytes [4, 21]. Mature PPGCs (Pax2-negative/vimentin-positive/GFAP-positive) also have similar antigenic phenotypes to mature astrocytes, except for vimentin. Considering that vimentin

expression is prolonged until the post-hatching period in the chicken brain [22-24], it seems reasonable to conclude that mature PPGCs are phenotypically identical to mature astrocytes in this avian species.

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References

- Culican SM, Baumrind NL, Yamamoto M, Pearlman AL. Cortical radial glia: identification in tissue culture and evidence for their transformation to astrocytes. *J Neurosci* 1990;10:684-92.
- Gray GE, Sanes JR. Lineage of radial glia in the chicken optic tectum. *Development* 1992;114:271-83.
- Barry D, McDermott K. Differentiation of radial glia from radial precursor cells and transformation into astrocytes in the developing rat spinal cord. *Glia* 2005;50:187-97.

4. Chan-Ling T, Chu Y, Baxter L, Weible Ii M, Hughes S. In vivo characterization of astrocyte precursor cells (APCs) and astrocytes in developing rat retinae: differentiation, proliferation, and apoptosis. *Glia* 2009;57:39-53.
5. Gerhardt H, Schuck J, Wolburg H. Differentiation of a unique macroglial cell type in the pecten oculi of the chicken. *Glia* 1999;28:201-14.
6. Schuck J, Gerhardt H, Wolburg H. The peripapillary glia of the optic nerve head in the chicken retina. *Anat Rec* 2000;259:263-75.
7. Quesada A, Prada FA, Aguilera Y, Espinar A, Carmona A, Prada C. Peripapillary glial cells in the chick retina: a special glial cell type expressing astrocyte, radial glia, neuron, and oligodendrocyte markers throughout development. *Glia* 2004;46:346-55.
8. Kim JY, Sohn HJ, Lee EY, Goo YS, Kim DW, Seo JH. Expression of alphaB-crystallin in the peripapillary glial cells of the developing chick retina. *Neurochem Res* 2011;36:76-82.
9. Hamburger V, Hamilton HL. A series of normal stages in the development of the chick embryo. 1951. *Dev Dyn* 1992;195:231-72.
10. Linser PJ, Irvin CK. Immunohistochemical characterization of delta crystallin-containing retina/optic nerve "boundary" cells in the chick embryo. *Dev Biol* 1987;121:499-509.
11. Klemenz R, Fröhli E, Steiger RH, Schäfer R, Aoyama A. Alpha B-crystallin is a small heat shock protein. *Proc Natl Acad Sci U S A* 1991;88:3652-6.
12. de Jong WW, Leunissen JA, Voorster CE. Evolution of the alpha-crystallin/small heat-shock protein family. *Mol Biol Evol* 1993;10:103-26.
13. Lee DC, Kim RY, Wistow GJ. An avian alpha B-crystallin: non-lens expression and sequence similarities with both small (HSP27) and large (HSP70) heat shock proteins. *J Mol Biol* 1993;232:1221-6.
14. Horwitz J. Alpha-crystallin can function as a molecular chaperone. *Proc Natl Acad Sci U S A* 1992;89:10449-53.
15. Ganea E. Chaperone-like activity of alpha-crystallin and other small heat shock proteins. *Curr Protein Pept Sci* 2001;2:205-25.
16. Soukkarieh C, Agius E, Soula C, Cochard P. Pax2 regulates neuronal-glial cell fate choice in the embryonic optic nerve. *Dev Biol* 2007;303:800-13.
17. Mi H, Barres BA. Purification and characterization of astrocyte precursor cells in the developing rat optic nerve. *J Neurosci* 1999;19:1049-61.
18. Stanke J, Moose HE, El-Hodiri HM, Fischer AJ. Comparative study of Pax2 expression in glial cells in the retina and optic nerve of birds and mammals. *J Comp Neurol* 2010;518:2316-33.
19. Reichenbach A, Stolzenburg JU, Eberhardt W, Chao TI, Dettmer D, Hertz L. What do retinal muller (glial) cells do for their neuronal 'small siblings'? *J Chem Neuroanat* 1993;6:201-13.
20. Rompani SB, Cepko CL. A common progenitor for retinal astrocytes and oligodendrocytes. *J Neurosci* 2010;30:4970-80.
21. Chu Y, Hughes S, Chan-Ling T. Differentiation and migration of astrocyte precursor cells and astrocytes in human fetal retina: relevance to optic nerve coloboma. *FASEB J* 2001;15:2013-5.
22. Dahl D, Crosby CJ, Sethi JS, Bignami A. Glial fibrillary acidic (GFA) protein in vertebrates: immunofluorescence and immunoblotting study with monoclonal and polyclonal antibodies. *J Comp Neurol* 1985;239:75-88.
23. Kálmán M, Székely AD, Csillag A. Distribution of glial fibrillary acidic protein and vimentin-immunopositive elements in the developing chicken brain from hatch to adulthood. *Anat Embryol (Berl)* 1998;198:213-35.
24. Seo JH, Chang JH, Song SH, Lee HN, Jeon GS, Kim DW, Chung CK, Cho SS. Spatiotemporal gradient of astrocyte development in the chick optic tectum: evidence for multiple origins and migratory paths of astrocytes. *Neurochem Res* 2008;33:1346-55.