



White adipose tissue browning and obesity

Xiao-Bing Cui, Shi-You Chen[✉]

Department of Physiology & Pharmacology, University of Georgia, Athens, GA 30602, USA.

Obesity and metabolic disorders are major health concerns worldwide. Although a range of therapies have been developed, these pharmaceutical treatments often have adverse side effects or limited efficacy. Therefore, there is a growing need of novel therapeutics to prevent or treat obesity. Obesity is thought to be caused by an imbalance between energy intake and energy consumption. Increasing energy consumption is considered as a potential therapeutic strategy to treat obesity and its related disorders.

Brown fat is a specialized fat depot characterized by increased energy expenditure and heat production^[1]. Its expansion and/or activation can protect against diet-induced obesity. The finding of active brown fat in human adults has aroused a great interest in the study of adipose tissue browning^[2]. Beige adipocytes that share some common characteristics with brown adipocytes such as high mitochondria content and uncoupling protein 1 (UCP1) expression can be induced in white adipose tissue (WAT)^[3].

The origin of classical brown adipocytes and beige adipocytes are different. Classical brown adipocytes originate from precursor cells in the embryonic mesoderm that express *Myf5* and *Pax7*^[4]. However, the origin of beige adipocytes is still controversial. Although a few studies have shown that beige adipocytes are originated from the trans-differentiation of mature white adipocytes^[5-6], there is a report showing that most of the beige adipocytes are from *de novo* differentiation of a precursor population^[7]. These beige pre-adipocytes are characterized by the expression of Cd137 and transmembrane protein 26 (Tmem26) on the cell surface^[8].

WAT browning is stimulated by a complex hormonal

interplay and numerous environmental factors. WAT is readily converted to beige adipocytes with prolonged cold exposure or β -adrenergic agonist treatment^[3,9]. Exercise has also been reported to increase WAT browning and energy expenditure^[10]. At the molecular level, WAT browning is regulated by multiple factors and signaling pathways. Peroxisome proliferator-activated receptor- γ (PPAR γ) coactivator-1 α (PGC-1 α) is a cold-induced interacting partner of PPAR γ in brown fat^[11]. It is also a master regulator of mitochondrial biogenesis and oxidative metabolism in adipocytes and can induce the expression of UCP1 and other thermogenic components. In addition, fibroblast growth factor 21 (FGF21) acts in an autocrine and paracrine manner to enhance WAT browning^[12]. Bone morphogenetic proteins (BMPs) are members of the transforming growth factor- β (TGF- β) superfamily. BMP4 and BMP7 induce browning of WAT and increase whole body metabolic rate and insulin sensitivity^[13-14]. Blocking of TGF- β /Smad3 signaling protects mice from obesity and diabetes through promoting WAT browning^[15]. Apparently, imbalanced TGF- β /BMPs signaling occurs during WAT browning, although the precise mechanisms and central regulators remain unknown.

Response gene to complement 32 (*RGC-32*) is a TGF- β downstream target gene. Our study shows that high-fat diet induces *RGC-32* expression in adipose tissue. *RGC-32* deficiency increases WAT browning and whole body metabolic rate and thus protects against high-fat diet-induced obesity^[16]. *RGC-32* appears to be involved in the imbalanced TGF- β /BMPs signaling. However, how *RGC-32* interacts with Smad3, BMP4 and BMP7 needs an extensive future investigation.

Although animal studies indicate that increased WAT

[✉] Corresponding author: Shi-You Chen, PhD, Department of Physiology & Pharmacology, The University of Georgia, 501 D.W. Brooks Drive, Athens, Georgia 30602, USA. Tel/Fax: 706-542-8284/706-542-3015; Email: sc229@uga.edu.

Received 12 August 2016, Accepted 10 November 2016, Epub 20 December 2016

CLC number: R575.5, Document code: B

The authors reported no conflict of interests.

browning prevents diet-induced obesity, whether or not the decline of brown fat activity is causally involved in the development of obesity in humans remains to be determined. Therefore, a further understanding of the morphology, development, and metabolism of the classic brown and beige adipocytes is still very important.

Acknowledgments

This work was supported by grants from National Institutes of Health (HL119053 and HL123302).

References

- [1] Harms M, Seale P. Brown and beige fat: development, function and therapeutic potential[J]. *Nat Med*, 2013, 19(10): 1252–1263.
- [2] Heaton JM. The distribution of brown adipose tissue in the human[J]. *J Anat*, 1972, 112(Pt 1): 35–39.
- [3] Cousin B, Cinti S, Morrioni M, et al. Occurrence of brown adipocytes in rat white adipose tissue: molecular and morphological characterization[J]. *J Cell Sci*, 1992, 103(Pt 4): 931–942.
- [4] Seale P, Bjork B, Yang W, et al. PRDM16 controls a brown fat/skeletal muscle switch[J]. *Nature*, 2008, 454(7207): 961–967.
- [5] Himms-Hagen J, Melnyk A, Zingaretti MC, et al. Multilocular fat cells in WAT of CL-316243-treated rats derive directly from white adipocytes[J]. *Am J Physiol Cell Physiol*, 2000, 279(3): C670–C681.
- [6] Barbatelli G, Murano I, Madsen L, et al. The emergence of cold-induced brown adipocytes in mouse white fat depots is determined predominantly by white to brown adipocyte transdifferentiation[J]. *Am J Physiol Endocrinol Metab*, 2010, 298(6): E1244–E1253.
- [7] Wang QA, Tao C, Gupta RK, et al. Tracking adipogenesis during white adipose tissue development, expansion and regeneration[J]. *Nat Med*, 2013, 19(10): 1338–1344.
- [8] Wu J, Boström P, Sparks LM, et al. Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human[J]. *Cell*, 2012, 150(2): 366–376.
- [9] Loncar D. Convertible adipose tissue in mice[J]. *Cell Tissue Res*, 1991, 266(1): 149–161.
- [10] Handschin C, Spiegelman BM. The role of exercise and PGC1alpha in inflammation and chronic disease[J]. *Nature*, 2008, 454(7203): 463–469.
- [11] Puigserver P, Wu Z, Park CW, et al. A cold-inducible coactivator of nuclear receptors linked to adaptive thermogenesis[J]. *Cell*, 1998, 92(6): 829–839.
- [12] Fisher FM, Kleiner S, Douris N, et al. FGF21 regulates PGC-1 α and browning of white adipose tissues in adaptive thermogenesis[J]. *Genes Dev*, 2012, 26(3): 271–281.
- [13] Schulz TJ, Huang TL, Tran TT, et al. Identification of inducible brown adipocyte progenitors residing in skeletal muscle and white fat[J]. *Proc Natl Acad Sci U S A*, 2011, 108(1): 143–148.
- [14] Qian SW, Tang Y, Li X, et al. BMP4-mediated brown fat-like changes in white adipose tissue alter glucose and energy homeostasis[J]. *Proc Natl Acad Sci U S A*, 2013, 110(9): E798–E807.
- [15] Yadav H, Quijano C, Kamaraju AK, et al. Protection from obesity and diabetes by blockade of TGF- β /Smad3 signaling[J]. *Cell Metab*, 2011, 14(1): 67–79.
- [16] Cui XB, Luan JN, Ye J, et al. RGC32 deficiency protects against high-fat diet-induced obesity and insulin resistance in mice[J]. *J Endocrinol*, 2015, 224(2): 127–137.