

Simultaneous Voltammetric Determination of Dypirone and Paracetamol with Carbon Paste Electrode and Multivariate Calibration Methodology

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Esse artigo descreve a determinação simultânea de dipirona (DIP) e de paracetamol (PAR) utilizando a técnica de Voltametria de Pulso Diferencial (VPD) e eletrodo de pasta de carbono não modificado. Devido à sobreposição dos picos voltamétricos de DIP e PAR, a metodologia de calibração multivariada baseada na Regressão de Mínimos Quadrados Parciais (PLSR) foi proposta. O conjunto de voltamogramas obtidos na presença de ambos os analitos em diferentes concentrações foi pré-processado pelos dados centrados na média. Para escolha do número de componentes principais, um procedimento de validação cruzada foi empregado, sendo que quatro componentes principais foram necessárias para obtenção dos menores valores de PRESS (Prediction Residual Error Sum of Squares). Esse modelo explicou aproximadamente 95,5% da variância do conjunto de dados. Os dados obtidos utilizando-se esse modelo mostraram uma alta correlação entre as concentrações reais e previstas. Entretanto, para baixas concentrações do PAR, os erros relativos aumentaram para 25%. Comparando-se os valores de RMSEP (Root Mean Square of Error Prediction), entre PAR e DIP, foi observado que este foi menor para DIP, provavelmente devido a maior quantidade de informação analítica apresentada pelos voltamogramas desse analito quando comparado ao processo redox de PAR, o qual devido a sua oxidação irreversível apresenta apenas um único pico.

This paper shows the simultaneous electrochemical determination of dypirone (DIP) and paracetamol (PAR) by differential pulse voltammetry technique (DPV) using an unmodified carbon paste electrode. Because of the overlapping of the voltammetric peaks of DIP and PAR, the multivariate calibration methodology based on Partial Least Square Regression (PLSR) was proposed. The data pre-treatment used in this process was mean centering and to choose the principal component number a cross validation procedure was used (leave-one-out). Four principal components were necessary to obtain the lowest PRESS (Prediction Residual Error Sum of Squares). The statistics showed that this model explains approximately 95.5% of the variance from the data set. Using this model, high correlation between real and predicted concentrations was observed. However, for low concentrations of PAR the relative error increased to 25%. Comparing RMSEP (Root Mean Square of Error Prediction) between PAR and DIP, it was observed that it was lower for DIP probably due to higher analytical information in the voltammograms for this analyte when compared to the electrochemical process of PAR, which presented only one potential peak due to its irreversible oxidation.

Keywords: paracetamol, dypirone, carbon paste electrode, differential pulse voltammetry and multivariate calibration methodology

Introduction

The development of analytical techniques for the rapid analysis of pharmaceuticals is important for quality and medical control. Due to this fact, a rigorous method of quality control of pharmaceutical fabrication is demanded.

Many analytical purposes in simultaneous analysis of pharmacological species are based on modern instrumental techniques, such as high performance liquid chromatography (HPLC),¹⁻³ gas chromatography⁴ and capillary electrophoresis,⁵⁻⁷ besides spectrofluorometry and chemiluminescence.⁸⁻¹⁰ However, these techniques are generally expensive and time-consuming, so it becomes very difficult to establish an online system. So, there is a great interest in the development of new analytical

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methods for simultaneous determination of pharmaceuticals without the necessity of a previous separation of the sample components, besides being rapid and with low cost. To overcome these limitations, electrochemical methods, such as voltammetric ones were extensively used for their accuracies, precisions, simplicities and possibilities of analysis without tedious sample pre-treatment.^{11,12}

However, the quantification of two or more components of a pharmaceutical sample using electrochemical techniques is generally a challenge task since most of the active components tend to oxidize or reduce at potentials extremely close which hampers their simultaneous determination. The association of electrochemical methods, such as voltammetric techniques using unmodified carbon electrodes with multivariate calibration methods eliminates the need for previous separations and permits simultaneous determination in the presence of analytical signal interference. The three most commonly used multivariate calibration methods are multiple linear regression (MLR), principal component regression (PCR), and partial least-squares regression (PLSR).¹³⁻¹⁵ These methods constitute a powerful statistical tool for factor analysis and have been mostly applied to the simultaneous multicomponent analysis of mixtures by spectroscopy, chromatography, and voltammetric methods.¹⁶⁻²⁰

Paracetamol or acetaminophen (Figure 1A) is a popular analgesic and antipyretic agent. Its action is similar to aspirin and it is an appropriate alternative for patients who are sensitive to acetylsalicylic acid.²¹ Overdose ingestions of acetaminophen lead to accumulation of toxic metabolites, which cause severe and sometimes fatal hepatotoxicity and nephrotoxicity.^{22,23} This drug is available in different dosage forms of tablets, capsules, suspensions and suppositories and sometimes it is present along with other analgesic agents such as dypirone. For quantification of paracetamol in pharmaceutical products, the American Pharmacopoeia recommends liquid chromatography as the official method

while the Brazilian Pharmacopoeia uses a spectrophotometric technique.^{24,25} Moreover, there are many studies described in the literature using different analytical techniques for determination of this drug.²⁶⁻³⁰ Some electrochemical methods have been used for analysis of paracetamol,³¹⁻³³ specially the voltammetric ones using chemically modified electrodes in order to improve the sensitivity.^{34,35}

Dypirone (Figure 1B), is a water soluble white crystalline powder which also presents analgesic and antipyretic activity. The methods commonly used for dypirone determination in various pharmaceutical formulations are based on its reaction with iodide.²⁵ Spectrophotometric methods such as UV-Vis absorption,³⁶ fluorescence³⁷ and chemiluminescence³⁸ are frequently reported for dypirone determination. However, different from paracetamol, the electrochemical reaction and its determination using electroanalytical techniques has been less investigated.³⁹⁻⁴¹ Matos *et al.*³⁹ proposed a flow injection analysis based on a multi-channel detection system for simultaneous amperometric determination of dypirone, ascorbic acid, dopamine and epinephrine using an array of modified microelectrodes, together with multivariate calibration analysis. The same research group developed a flow cell containing a gold electrode from recordable compact discs for the determination of dypirone in pharmaceutical formulations.³⁹

This paper reports the simultaneous electrochemical determination of dypirone (DIP) and paracetamol (PAR) by differential pulse voltammetry technique (DPV) using an unmodified carbon paste electrode. Because the voltammetric peaks of DIP and PAR overlaps, the multivariate calibration methodology based on Partial Least Square Regression (PLSR) was proposed. The data pre-treatment used in this process was mean centering and to choose the number of principal components a cross validation procedure was used (leave-one-out).

Experimental

Preparation of the carbon paste electrodes

The carbon paste electrode (CPE) was prepared by mixing the analytical grade graphite (Fluka) and about two drops of Nujol[®], added in order to get a homogeneous paste. We used the carbon paste electrode because its cost (it is cheaper than other carbon based electrodes, such as the glassy carbon electrode) and the facility of its preparation. Some initial tests were realized in order to optimize the quantity of mineral oil in the paste. The different electrodes prepared were analyzed in presence of the analytes in a fixed concentration. Best results (with more defined

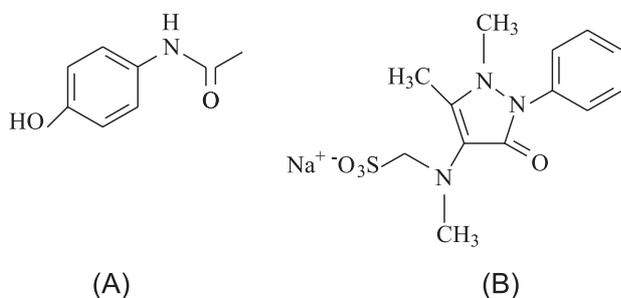


Figure 1. Chemical structures of: (A) paracetamol (*N*-acetyl-*p*-aminophenol, 4-acetamidophenol); (B) dypirone (sodium salt of the 1-phenyl-2,3-dimethyl-4-methyl aminomethane sulfonate-5-pyrazolone).

voltammetric peaks) were obtained with approximately 12% of mineral oil. As the carbon paste electrode had reproducibility problems, we tried to use the same carbon paste prepared for all measurements. The obtained paste was placed in a homemade electrode which consisted of a 1 mm deep cavity in contact with a carbon rod having a 0.5 cm diameter connected to a copper electrode for electrical contact, fused to a plastic tube.

Electrochemical measurements

All the voltammetric measurements (cyclic voltammetry and differential pulse voltammetry) were carried out using a 10 mL capacity electrochemical cell with the conventional three electrode system: a Pt wire as a counter electrode, an Ag/AgCl electrode as the reference and the modified carbon paste electrodes as working electrodes. The potential range analyzed was from -0.4 V to 0.4 V vs. Ag/AgCl, with scan rates in the range between 10 mV s⁻¹ and 100 mV s⁻¹. Solutions of various pH were used, adjusted with HCl and KOH in a 0.5 mol L⁻¹ KCl (different supporting electrolytes were also tested, such as NaCl and LiCl). The response of the electrode in the presence of paracetamol (PAR) and dypirone (DIP) was also studied by cyclic voltammetry in KCl solutions at different pH. Several concentrations of PAR and DIP were used to obtain a calibration curve in the range between 2.5×10^{-5} and 1.5×10^{-3} mol L⁻¹ in a potentiostat / galvanostat Palm Sens 3.7 model, connected to a microcomputer for data acquisition.

Multivariate calibration methodology

A PC/Pentium IV microcomputer equipped with the mathematical software MATLAB for Windows (version 4.2, distributed by MathWorks) and PLS-toolbox (version 1.5, distributed by Eigenvector Research) was employed to obtain the PLSR models. For the processing of the voltammetric data using multivariate calibration, a calibration set consisting of 25 synthetic samples were prepared. These samples were obtained by adding known amounts of working solutions of dypirone and paracetamol in a concentration of 1.0×10^{-2} mol L⁻¹ in NaCl 0.5 mol L⁻¹. The final concentration of these solutions varied between 1.0×10^{-4} and 5.0×10^{-4} mol L⁻¹. All the solutions were prepared with distilled water and all chemicals used were of analytical-reagent grade. The differential pulse voltammograms were obtained in the range of -0.1 and 1.1 V vs. Ag/AgCl.

The experimental design used to develop the multivariate calibration model for dypirone and paracetamol is presented in Figure 2 (●). The following points were obtained in

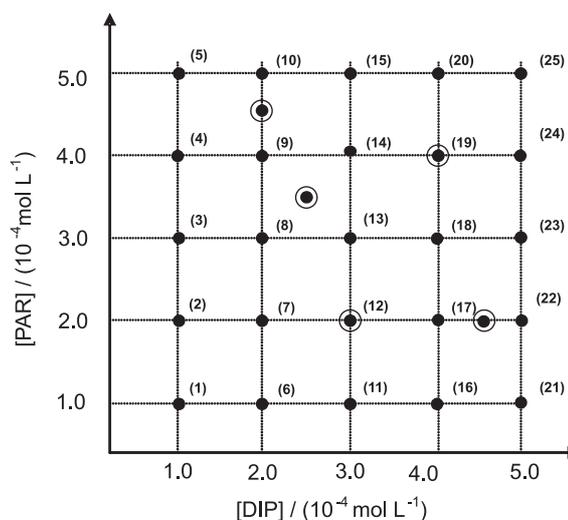


Figure 2. Composition of synthetic mixtures of paracetamol and dypirone: (●) samples used for calibration set; (⊙) samples used for external validation.

duplicate: 1, 3, 5, 11, 13, 15, 21, 23 and 25. The validation set was created by random selection of 5 samples (also obtained in duplicate) from the experimental design, also shown in Figure 2 (⊙).

Results and Discussion

Cyclic voltammetric study of paracetamol and dypirone with carbon paste electrode

Firstly, the electrochemical responses of dypirone (DIP) and paracetamol (PAR) were characterized by cyclic voltammetry using a carbon paste electrode (Figures 3A and 3B).

As can be seen in Figure 3A, PAR exhibits one well defined anodic peak with E_{pa} (anodic peak potential) at 0.77 V vs. Ag/AgCl (pH 6.0) and a poorly defined cathodic peak at $E_{pc} = 0.10$ V which are similar to that observed for an unmodified glassy carbon electrode.⁴² Kissinger *et al.*^{43,44} deeply investigated the electrochemical oxidation of paracetamol through cyclic voltammetric studies. The first reaction step is an electrochemical oxidation involving two electrons and two protons to generate *N*-acetyl-*p*-quinoneimine. All subsequent reaction steps are non-electrochemical, but pH-dependent, processes. For oxidations at pH values higher than 6, the final product is a benzoquinone.

The electrochemical oxidation of DIP presented one reversible pair with $E_{pa_1} = 0.58$ V and $E_{pc_1} = 0.53$ V ($E_{p_{1/2}} = 0.55$ V) and two more irreversible anodic peaks with $E_{pa_2} = 0.43$ V and $E_{pa_3} = 1.00$ V, as shown in Figure 3B. Perez-Ruiz *et al.*⁴¹ also observed the same electrochemical

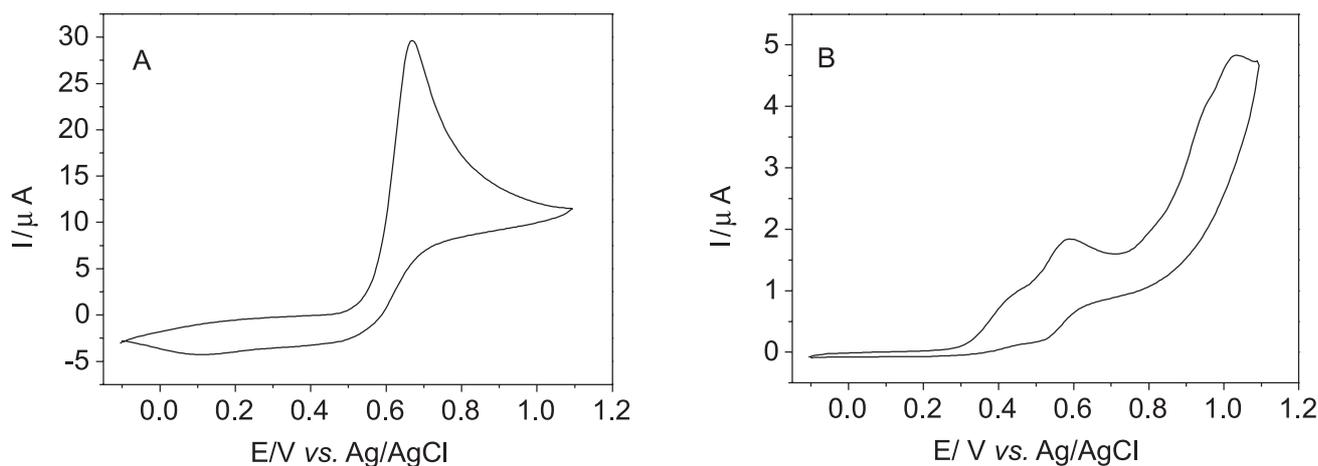


Figure 3. Cyclic voltammograms of (A) paracetamol and (B) dypirone, (concentration of 1.0×10^{-3} mol L $^{-1}$) obtained using a carbon paste electrode in NaCl 0.5 mol L $^{-1}$ (pH 6.0). Potential range of: -0.1 V to 1.1 V vs. Ag/AgCl.

behavior for dypirone using a glassy carbon electrode and a carbon paste electrode respectively. According to Teixeira *et al.*,⁴⁵ the first electrochemical oxidation peak of the dypirone is related to the methylamino-*N*-methanesulphonate group.

The analytical parameters (LOD, Sensitivity, Linear Regression Equation, Linear Range of Concentration) related to the individual determination of DIP and PAR by cyclic voltammetry using the carbon paste electrode are summarized in Table 1. The plots of the anodic current peak against DIP or PAR concentrations were linear in a wide range of concentration, observing a better sensitivity for PAR.

Multivariate calibration methodology

As can be seen from the cyclic voltammetric studies, some redox peaks for DIP and PAR are overlapped which complicates their simultaneous determination by cyclic voltammetry using univariate methods. To overcome this problem, a multivariate calibration methodology based on Partial Least Square Regression (PLSR) using the Differential Pulse Voltammetry (DPV) was proposed. The use of DPV technique was justified by its higher sensitivity (compared to the cyclic voltammetry), which causes better

separation between the redox peaks. Therefore, it minimizes errors for simultaneous determination of the species.

The differential pulse voltammograms for DIP, obtained in the same potential range used for the CV studies, showed four anodic peaks at 0.35; 0.65, 0.82 e 1.0 V vs. Ag/AgCl. In contrast, the DPV of PAR showed only one extremely defined anodic peak at 0.64 V, similar to that obtained by cyclic voltammetry technique (Figure 4).

In the calibration step, a group composed by 25 standard mixtures containing PAR and DIP were analyzed by differential pulse voltammetry (potential scan between -0.1 V and 1.1 V) using a carbon paste electrodes. The current data of the voltammograms (matrix *x*) were correlated with the respective known concentrations of the analytes (PAR and DIP 1.0×10^{-4} mol L $^{-1}$ to 5.0×10^{-4} mol L $^{-1}$; matrix *y*). The data pre-treatment used in this process was mean centering and to choose the principal component number a cross validation procedure was used (leave-one-out). Four principal component were necessary to obtain the lowest RMSECV (Root Mean Squares Error of Cross Validation) showed in Figure 5A. The statistics show that this model explains approximately 95.5% of the variance from the data set and these four latent variables generate the regression coefficients with the evidence that exists an influence of the dypirone in the analytical signal of

Table 1. Analytical parameters obtained in the individual determination of paracetamol (PAR) and dypirone (DIP)

Analyte	LOD / (mmol L $^{-1}$)	Sensitivity / (μ A / mol L $^{-1}$)	Linear Regression Equation	Concentration Linear Range / (mmol L $^{-1}$)
PAR	0.158	2.7×10^4	$Y = -2.144 + 27077.1 X$	0.25 to 2.00
DIP (0.58 V)	0.167	1.6×10^3	$Y = -0.129 + 1550.2 X$	0.25 to 2.00
DIP (1.0V)	0.037	3.0×10^3	$Y = -0.056 + 3008.14 X$	0.25 to 2.00

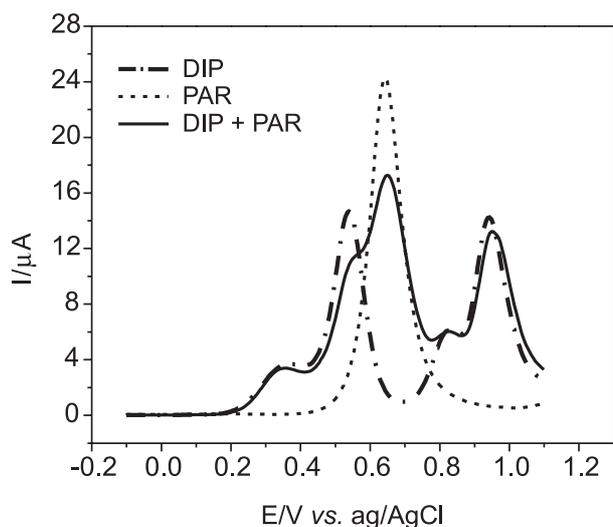


Figure 4. Differential pulse voltammograms of paracetamol (PAR), dypirone (DIP) and a mixture of DIP and PAR (both in concentration of $1 \times 10^{-3} \text{ mol L}^{-1}$) obtained using a carbon paste electrode. Supporting electrolyte: NaCl 0.5 mol L^{-1} (pH 6.0). Potential range of: -0.1 V to 1.1 V vs. Ag/AgCl, scan rate: 25 mV s^{-1} , time pulse: 100 ms , sensibility: $100 \mu\text{A V}^{-1}$.

paracetamol and *vice-versa* (Figure 5B). An important conclusion can be reached by analyzing the number of PLS factors required to adequate this model. It exceeds in two the theoretical limit of two expected factors as related in studies using voltammetric methods.⁴⁶ This could be caused by deviations from linearity because interactions among the electroactive components and competition by the electrode surface.⁴⁷

This model presented high correlation between real and predicted concentrations (Figure 6), and was applied to predict the concentration of five standard mixtures (not used in the calibration step) in duplicate. For low analytes concentrations the relative error increases to 25% for PAR (lower concentration) as shown in Table 2. Comparing the RMSEP (Root Mean Square of Error Prediction) between PAR and DIP was observed that it was lower for DIP, probably due to higher analytical information in the voltammograms (higher number of voltammetric peaks) for this analyte when compared to the electrochemical process of PAR that presented only one potential peak due to its irreversible oxidation.

The prediction results performed in duplicate (Table 2) can be used to determine the precision of multivariate model constructed. The concordance between the predicted concentrations for the duplicate showed similar precision values for both analytes.

These results were compared with univariate calibration methodology. Higher relative errors was obtained (the highest error to PAR = 140% and DIP = -28 %) for analysis of the same five standard mixtures used in the prediction

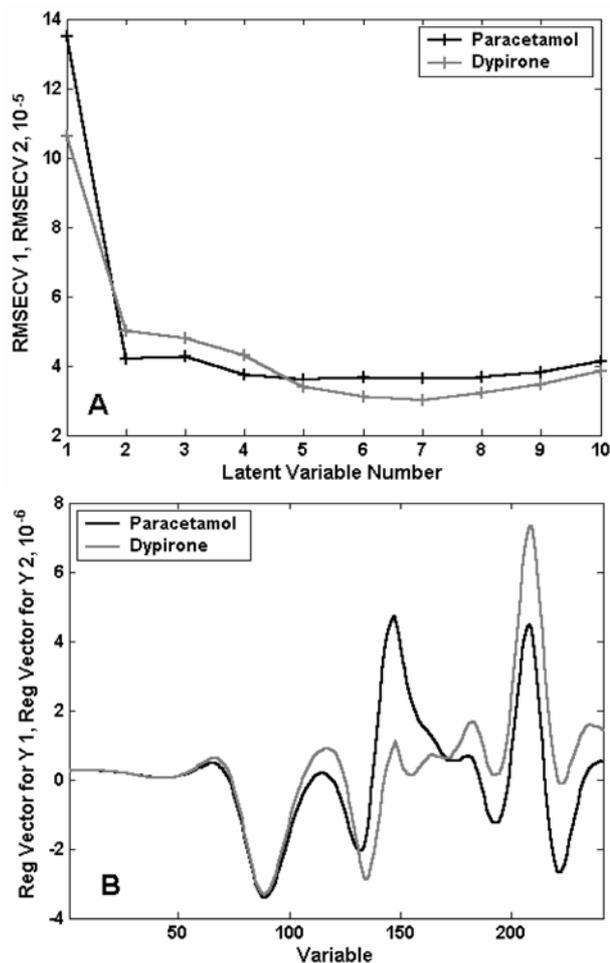


Figure 5. (A) RMSECV for PLSR model to determination of DIP and PAR; (B) Regression Coefficient with four Latent Variables in PLSR model.

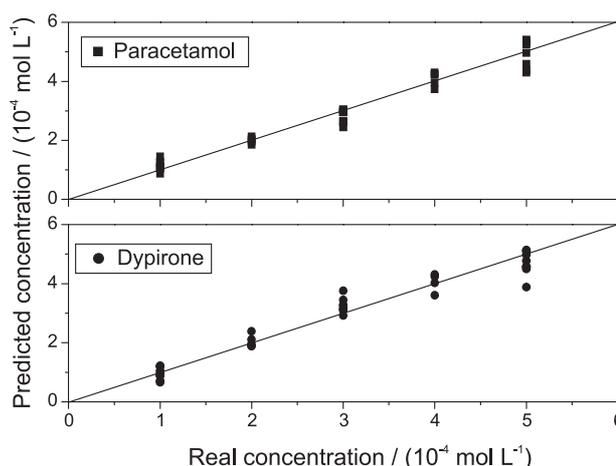


Figure 6. Relationship between predict and real concentrations of PAR and DIP.

step. The multivariate calibration methodology presented results with better prevision capacity for simultaneous determination, but the serious overlapping in the only

Table 2. Simultaneous determination by PLSR of paracetamol (PAR) and dypirone (DIP)

Real Concentration / (mmol L ⁻¹)		Multivariate method			
		Predicted Concentration / (mmol L ⁻¹)		Relative Error (%)	
PAR	DIP	PAR	DIP	PAR	DIP
0.200	0.450	0.224	0.502	11.9	11.5
0.200	0.450	0.252	0.469	25.8	4.3
0.350	0.250	0.393	0.263	12.3	5.2
0.350	0.250	0.400	0.267	14.1	6.8
0.200	0.300	0.212	0.298	6.1	-0.6
0.200	0.300	0.240	0.306	19.9	1.9
0.400	0.400	0.437	0.393	9.2	-1.8
0.400	0.400	0.460	0.406	15.0	1.4
0.450	0.200	0.523	0.202	16.9	1.2
0.450	0.200	0.511	0.205	13.5	2.5
RMSEP (mmol L ⁻¹) *				0.048	0.019
Precision [48]		0.015	0.012		

* RMSEP = $(\sum (y_{\text{pred}} - y_{\text{real}})^2 / n)^{1/2}$.

analytical signal of paracetamol makes it difficult to resolve completely the interference of dypirone. This fact can be observed by the positive relative error for paracetamol determination in all samples of the validation set. This tendency (upper estimate) is less for the dypirone determination due to presence of the two analytical signals (oxidation process).

An alternative to improvement of the obtained results is applying the Artificial Neural Networks (ANN) to the set data. This methodology has been particularly useful in electroanalytical measurements, when the electrode response may behave in a non-linear way.^{20,49}

Conclusions

This paper demonstrated the potentiality of differential pulse voltammetry and the multivariate methodology for simultaneous determination of paracetamol and dypirone, even using the unmodified carbon paste electrode (favorable condition to overlapping of the oxidation and reduction peaks of interest species). PLSR methodology proved to be a powerful tool for simultaneous quantification of PAR and DIP, especially considering the similarity of voltammetric response of analytes. The results obtained were satisfactory, principally at low concentration levels, with prediction of relative errors less than 26% for PAR and 12% for DIP. The methodology proposed eliminates the need for previous separation of these analytes and the modification of the carbon paste electrode to obtain a selective voltammetric response.

References

1. Canada-Canada, F.; Espinosa-Mansilla, A.; de la Pena, A. M.; *J. Sep. Sci.* **2007**, *30*, 1242.
2. Srogi, K.; *Anal. Lett.* **2006**, *39*, 231.
3. Samanidou, V. F.; Evaggelou, E. N.; Papadoyannis, I. N.; *J. Pharm. Biomed. Anal.* **2005**, *38*, 21.
4. Plossl, F.; Giera, M.; Bracher, F.; *J. Chromatogr. A* **2006**, *1135*, 19.
5. Perez-Ruiz, T.; Martinez-Lozano, C.; Tomas, V.; Galera, R.; *J. Pharm. Biomed. Anal.* **2005**, *38*, 87.
6. Alnajjar, A.; AbuSeada, H. H.; Idris, A. M.; *Talanta* **2007**, *72*, 842.
7. Azhagvuel, S.; Sekar, R.; *J. Pharm. Biomed. Anal.* **2007**, *43*, 873.
8. Lopez-Flores, J.; Cordova, M. L. F. D.; Molina-Diaz, A.; *Anal. Chim. Acta* **2005**, *535*, 161.
9. Wolyniec, E.; Niedzwiedzka, U.; Kojlo, A.; *Instrum. Sci. Technol.* **2007**, *35*, 219.
10. Waseem, A.; Yaqoob, M.; Nabi, A.; *Talanta* **2007**, *71*, 56.
11. Carvalho, L. M.; Nascimento, P. C.; Bohrer, D.; Correia, D.; Bairo, A. V.; Pomblum, V. J.; Pomblum, S. G.; *J. Braz. Chem. Soc.* **2007**, *18*, 789.
12. Melo, H. C.; Selegim, A. P. D.; Polito, W. L.; Fatibello-Filho, O.; Vieira, I. C.; *J. Braz. Chem. Soc.* **2007**, *18*, 797.
13. Ni, Y. N.; Wang, Y. R.; Kokot, S.; *Talanta* **2006**, *69*, 216.
14. Matos, R. C.; Angnes, L.; Araújo, M. C. U.; Saldanha, T. C. B.; *Analyst* **2000**, *125*, 2011.
15. Bessant, C.; Saini, S.; *J. Electroanal. Chem.* **2000**, *489*, 76.

16. Santos, P. M.; Sandrino, B.; Moreira, T. F.; Wohnrath, K.; Nagata, N.; Pessoa, C. A.; *J. Braz. Chem. Soc.* **2007**, *18*, 93.
17. Skeika, T.; Marcovicz, C.; Nakagaki, S.; Fujiwara, S. T.; Wohnrath, K.; Nagata, N.; Pessoa, C. A.; *Electroanalysis*, **2007**, *19*, 2543.
18. Apetrei, C.; Gutierrez, F.; Rodriguez-Mendez, M. L.; de Saja, J. A.; *Sensor Actuat B-Chem.* **2007**, *121*, 567.
19. Gonzalez, M. J. G.; Renedo, O. D.; Martinez M. J. A.; *Talanta* **2007**, *71*, 691.
20. Cabanillas, A. G.; Caceres, M. I. R.; Canas, M. A. M.; Burgillos, J. M. O.; Diaz, T. G.; *Talanta* **2007**, *72*, 932.
21. Felix, F. S.; Brett, C. M. A.; Angnes, L.; *J. Pharm. Biomed. Anal.* **2007**, *43*, 1622.
22. Martin, F. L.; McLean, A. E.; *Drug Chem. Toxicol.* **1998**, *21*, 477.
23. Mugford, C. A.; Tarloff, J. B.; *Toxicol. Lett.* **1997**, *93*, 15.
24. *The United States Pharmacopoeia – The National Formulary – USP 23; NF-18*, Twinbrook Parkway: Rockville, 1995, p. 16.
25. *Farmacopéia Brasileira*, 3rd ed., Atheneu Editora: São Paulo, 1977.
27. Burgot, G.; Auffret, F.; Burgot, J. L.; *Anal. Chim. Acta* **1997**, *343*, 125.
28. Moreira, A. B.; Oliveira, H. P. M.; Atvars, T. D. Z.; Dias, I. L. T.; Neto, G. O.; Zagatto, E. A. G.; Kubota, L. T.; *Anal. Chim. Acta* **2005**, *539*, 257.
29. Peng, W.; Li, T.; Li, H.; Wang, E.; *Anal. Chim. Acta* **1994**, *298*, 415.
30. Fatibello-Filho, O.; Lupetti, K. O.; Vieira, I. C.; *Talanta* **2001**, *55*, 685.
31. Boopathi, M.; Won, M. S.; Shim, Y. B.; *Anal. Chim. Acta* **2004**, *512*, 191.
32. Carvalho, R. M.; Freire, R. S.; Rath, S.; Kubota, L. T.; *J. Pharm. Biomed. Anal.* **2004**, *34*, 871.
33. Wangfuengkanagul, N.; Chailapakul, O.; *J. Pharm. Biomed. Anal.* **2002**, *28*, 841.
34. Wang, S. F.; Xie, F.; Hu, R. F. *Sens. Actuators B* **2007**, *123*, 495.
35. Goyal, R. N.; Singh, S. P.; *Electrochim. Acta* **2006**, *51*, 3008.
36. Aburjai, T.; Amro, B. I.; Aiedeh, K.; Abuirjeie, M.; Al-Khalil, S.; *Pharmazie* **2000**, *55*, 751.
37. Perez-Ruiz, T.; Martinez-Lozano, C.; Tomas, V.; Carpena J.; *Microchem. J.* **1993**, *47*, 296.
38. Huang, Y. M.; Zhang, C.; Zhang, X. R.; Zhang, Z. J.; *J. Pharm. Biomed. Anal.* **1999**, *2*, 817.
39. Matos, R. C.; Angnes, L.; Araújo, M. C. U.; Saldanha, T. C. B.; *Analyst* **2000**, *125*, 2011.
40. Munoz, R. A. A.; Matos, R. C.; Angnes, L.; *J. Pharm. Sci.* **2001**, *90*, 1972.
41. Perez-Ruiz, T.; Martinez-Lozano, C.; Tomas, V.; *J. Pharm. Biomed. Anal.* **1994**, *12*, 1109.
42. Wang, S-F.; Xie, F.; Hu, R-F.; *Sens. Actuators B* **2007**, *123*, 495.
43. Miner, D. J.; Rice, J. R.; Riggins, R. M.; Kissinger, P. T.; *Anal. Chem.* **1981**, *53*, 2258.
44. Kissinger, P. T.; Roston, D. A.; Van Benschoten, J. J.; Lewis, J. Y.; Heineman, W. R.; *J. Chem. Educ.* **1983**, *60*, 772.
45. Teixeira, M. F. S.; Marcolino-Júnior, L. H.; Fatibello-Filho, O.; Dockal, E. R.; Cavalheiro, E. T. G.; *J. Braz. Chem. Soc.* **2004**, *15*, 803.
46. Barthus, R. C.; Mazo, L. H.; Poppi, R. J.; *J. Pharm. Biomed. Anal.* **2005**, *38*, 94.
47. Ribero, G. G.; Goicoechea, H. C.; *Talanta* **2003**, *61*, 743.
48. Braga, J. W. B.; Poppi, R. J.; *Quim. Nova* **2004**, *27*, 1004.
49. Palacios-Santander, J. M.; Cubillana-Aguilera, L. M.; Naranjo-Rodríguez, I.; Hidalgo-Hidalgo-de-Cisneros, J. L.; *Chemom. Intell. Lab. Syst.* **2007**, *85*, 131.

Received: September 30, 2007

Web Release Date: April 30, 2008