

## New Alkaloids from the Mediterranean Sponge *Hamigera hamigera*

Wafaa Hassan,<sup>1,3</sup> RuAngelie Edrada,<sup>1</sup> Rainer Ebel,<sup>1</sup> Victor Wray<sup>2</sup> and Peter Proksch<sup>1,\*</sup>

<sup>1</sup> Institut für Pharmazeutische Biologie, Heinrich-Heine-Universität, Universitätsstr 1, Geb. 26.23, 40225 Düsseldorf, Germany

<sup>2</sup> Gesellschaft für Biotechnologische Forschung, Mascheroder Weg 1, 38124 Braunschweig, Germany

<sup>3</sup> Department of Pharmacognosy, Faculty of Pharmacy, Zagazig University, Zagazig, Egypt

\* Author to whom correspondence should be addressed; Fax: +49/211-8111923. E-mail: proksch@uni-duesseldorf.de

Received: 30 April 2004 / Accepted: 27 May 2004 / Published: 25 August 2004

---

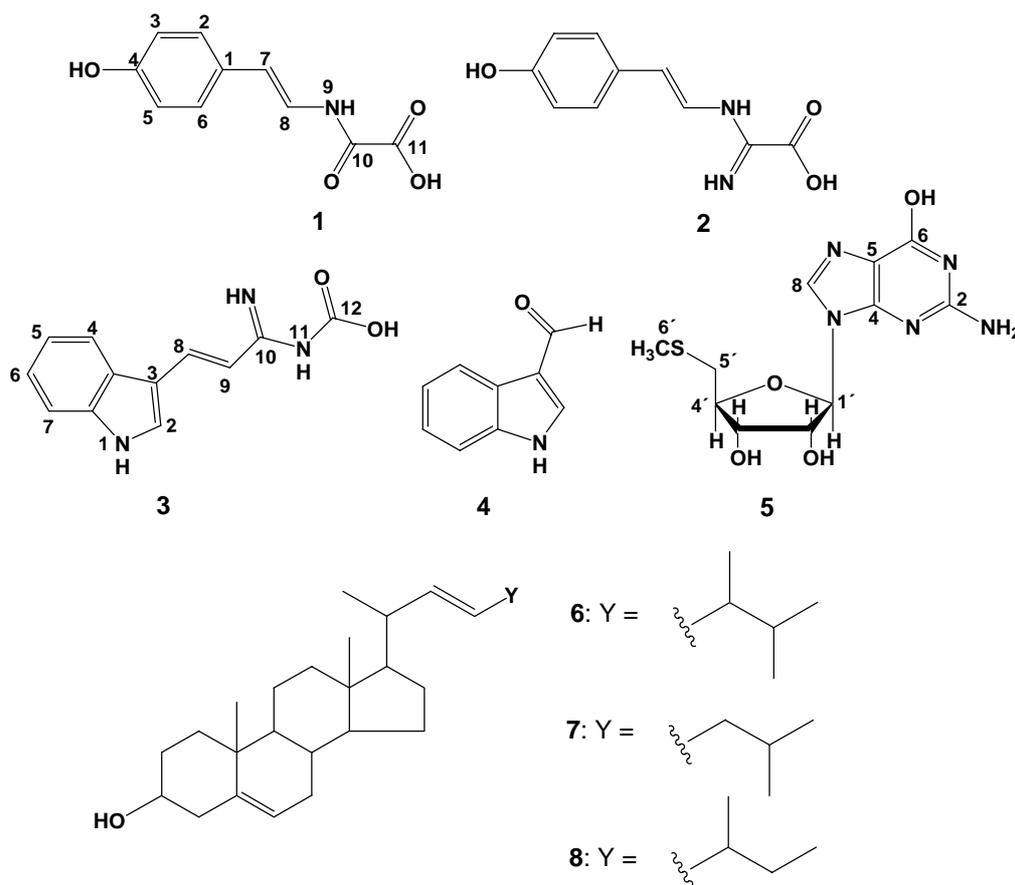
**Abstract:** The Mediterranean sponge *Hamigera hamigera* (family Anchinoideae) was studied since its total extract showed deterrent activity in a fish feeding assay. Eight compounds were isolated from the biologically active fractions and four of these proved to be new natural products, hamigeroxalamic acid (**1**), hamigeramine (**2**), hamigeramide (**3**) and hamiguanosinol (**5**). The structures of the new compounds were elucidated by 1D and 2D NMR spectroscopy and mass spectrometry .

**Keywords:** *Hamigera hamigera*, hamigeroxalamic acid, hamigeramime, hamigeramide, hamiguanosinol.

---

### Introduction

Marine sponges are known as a prolific source of biologically active and structurally unique metabolites. In the search for biologically active substances from marine sponges, the family Anchinoidae was shown to be a rich source for secondary metabolites which include tripeptide and dimeric peptide alkaloids, sulfur-containing compounds [1, 2, 3] chlorine-containing phenolic compounds [4] and antifungal and cytostatic macrolides [5, 6]. The genus *Hamigera* belongs to the same family. Biologically active brominated compounds have previously been isolated from *Hamigera tarangensis* [7, 8].



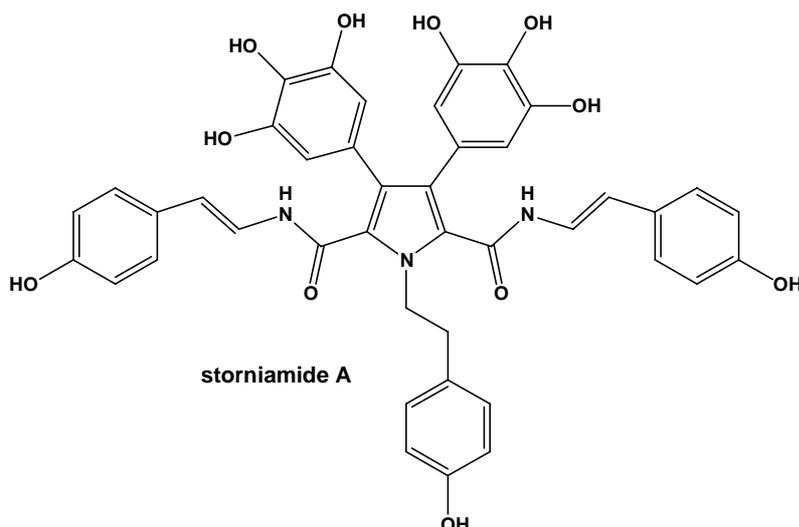
**Figure 1.** Compounds isolated from the fish feeding deterrent extracts of the Mediterranean sponge *Hamigera hamigera*.

The Mediterranean sponge *Hamigera hamigera* was studied because the total extract showed biological activity in a fish-feeding deterrence assay. This study led to the isolation of four new compounds including hamigeroxalamic acid (1), hamigeramine (2), hamigeramide (3) and a sulfated nucleoside, hamiguanosinol (5) together with the known compounds, 1*H*-indole-3-carbaldehyde (4), and three steroid derivatives 6, 7, and 8.

## Results and Discussion

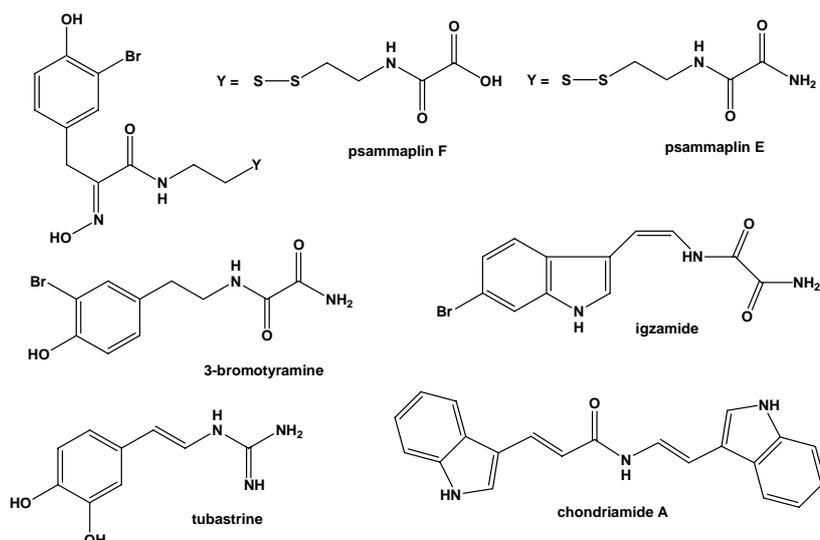
The conspicuous dark red colored sponge *Hamigera hamigera* was collected at a depth of 5 to 7 m off the island of Elba (Mediterranean Sea). The biologically active EtOAc and aqueous extracts were subjected to a series of chromatographic separation steps which yielded four new compounds **1**, **2**, **3**, and **5**, while four known compounds (**4**, **6**, **7**, and **8**) were also isolated. The structures of the known compounds, indole-3-carboxaldehyde (**4**) [9, 10], 22-dehydrocampasterol (**6**), 27-nor-22-dehydrocampasterol (**7**), and 24-methylcholesta-5,22-dien-3 $\beta$ -ol (**8**) [11, 12, 13] were identified from comparison of their  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data with those found in literature. The structures of the new compounds were identified unambiguously by 1D and 2D NMR spectroscopy. Through-bond homonuclear ( $^1\text{H}$ - $^1\text{H}$  COSY) and heteronuclear (long range  $^{13}\text{C}$ - $^1\text{H}$ ) correlations were used to establish assignments and atom connectivities. Chemical shifts were compared with literature data for compounds containing similar structural subunits.

Hamigeroxalamic acid (**1**) was isolated as a pale yellowish white amorphous powder. Positive and negative ion ESIMS established the molecular weight of the compound which is compatible with a molecular formula of  $\text{C}_{10}\text{H}_9\text{NO}_4$ . The  $^1\text{H}$ -NMR spectra (see Table 1) revealed an AA'BB' system for 2H doublets at  $\delta$  7.09 and 6.67 which indicated a *para*-disubstituted benzene ring. The broad proton signal at  $\delta$  9.31 (OH) suggested **1** was phenolic as it gave a  $^2J$  correlation with the  $^{13}\text{C}$  NMR signal at  $\delta$  156.4 and  $^3J$  correlations to the C-3 and C-5 at  $\delta$  116.5 of the AA'BB' system. The presence of a 1,2-disubstituted double bond adjacent to the NH group or *trans*-enamide substructure were observed as implied by the coupling constants and respective proton signals at  $\delta$  6.35 (H-7, d,  $J = 14.5$  Hz) and  $\delta$  7.10 (H-8, dd,  $J = 14.5$  and 10.7 Hz) which coupled further with the broad NH doublet proton at  $\delta$  10.07 (d,  $J = 10.7$  Hz) [14, 15]. A similar substructure was found in storniamides [14], peptide alkaloids isolated from a Patagonian sponge *Cliona* sp. The carbon signals of the olefins at  $\delta$  114.0 and 122.1 were also compatible to those found in the latter compounds (*ca.*  $\delta$  114 and 120). From the  $^{13}\text{C}$ -NMR spectrum of hamigeroxalamic acid, signals at  $\delta_{\text{C}}$  163.0 and 158.3 were observed indicating the presence of two carbonyl carbons which were comparable to those of the terminal oxalamic (NHCOCOOH) functionality in psammaplin F with chemical shifts of 161.9 and 159.2 ppm [17]. The ESI-MS/MS performed on the negative pseudo molecular ion  $m/z$  at 206  $[\text{M}-\text{H}]^-$  gave a daughter ion at  $m/z$  134.2 indicating the loss of an oxoacetic acid [COCOOH] as a result of fragmentation of the amide linkage of the oxalamic acid functionality. A similar fragmentation pattern was also observed with the storniamides [14]. This confirms that **1** is *N*-[2-(4-hydroxy-phenyl)-vinyl]-oxalamic acid which was assigned the trivial name hamigeroxalamic acid.

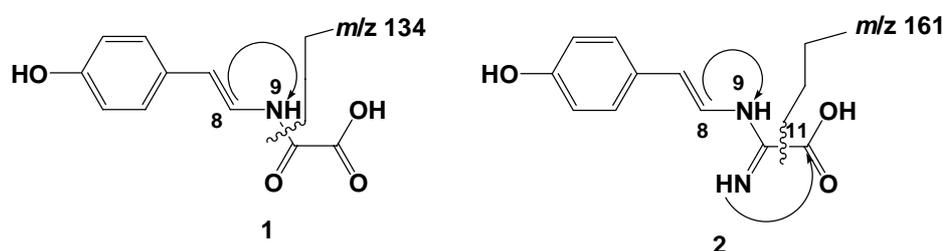


Hamigeramine (**2**) was isolated as a yellowish white powder. The ESIMS of **2** showed a peak at  $m/z$  207  $[M+H]^+$  in accordance with the molecular formula  $C_{10}H_{10}N_2O_3$ .  $^1H$ - and  $^{13}C$  NMR data were comparable to those of hamigeroxalamic acid which also revealed the presence of a 4-hydroxystyryl unit, as found in **1**. Its  $^1H$  NMR spectrum also showed both the AA'BB' spin system and *trans* vinyl amine pattern of resonances. The signals for the NH at  $\delta$  10.97 and the corresponding olefinic proton at  $\delta$  7.29 appeared very broad such that the expected doublet for a *trans* vinyl amine coupling was not resolved. However, the COSY spectrum confirmed the presence of the *trans* vinyl amine function through the correlation observed between  $\delta_H$  10.97 and  $\delta_H$  7.29. The  $^{13}C$  NMR spectrum of hamigeramine (**2**) was very similar to that of hamigeroxalamic acid (**1**) except for a significant change in chemical shifts for the carbonyl functionalities ( $\delta$  155.7 and  $\delta$  155.1) which were to higher field, while the  $^1H$  signals of the *trans* olefinic protons were to lower field. There were also additional broad singlets at  $\delta$  9.29 and  $\delta$  9.10 that were first thought to indicate the presence of a terminal  $NH_2$  as in oxalamide. However, significant discrepancies were observed for the carbonyl signals in the  $^{13}C$  NMR spectrum of **2** when compared to those of psammaplins E [17] (in  $CD_3OD$ ), 3-bromotyramine amide [18] (in  $CD_3OD$ ), and igzamide [19] (in DMSO), all of which possess an oxalamide terminal functional group. The carbonyl shifts of the oxalamide unit for these known compounds were reported at *ca.* 162 and 165 ppm in  $CD_3OD$  while in DMSO, the chemical shifts were at *ca.* 161 and 157 ppm. The carbon chemical shifts at *ca.* 155 could also suggest the presence of an imine group as found in tubastrine [16, 20, 21]. The  $^1H$  and  $^{13}C$  chemical shifts ( $\delta_H$  6.81 and 7.29,  $\delta_C$  121.3 and 119.5) of the olefinic protons of hamigeramine (**2**) were comparable to those of tubastrine (*ca.*  $\delta_H$  6.70 and 7.20,  $\delta_C$  120 and 119). The amino imino acetic functionality in hamigeramine (**2**) was detected through the HMBC correlations of the olefinic H-8 ( $\delta$  7.29) with C-10 ( $\delta$  155.1) and of the imino NH ( $\delta$  9.29) with C-11 ( $\delta$  155.7).

ESI-MS/MS fragmentation of the negative pseudo molecular ion  $[M-H]^-$  at  $m/z$  205 gave a



daughter ion at  $m/z$  161 instead of at  $m/z$  134 as in hamigeroxalamic acid (**1**). This indicated the loss of only a COOH unit and absence of the amide linkage found in **1**, again implying the presence of a different terminal functionality for hamigeramine (**2**), and confirmed that hamigeramine **2** was [2-(4-hydroxy-phenyl)-vinylamino]-imino-acetic acid.



**Figure 2.** ESI-MS/MS fragmentation of hamigeroxalamic acid and hamigeramine in the negative mode. The vinyl amine function in both compounds was confirmed by the COSY correlation between H-8 and NH-9 while the amino imino acetic functionality in hamigeramine was detected through the HMBC correlation between the imino NH and C-11.

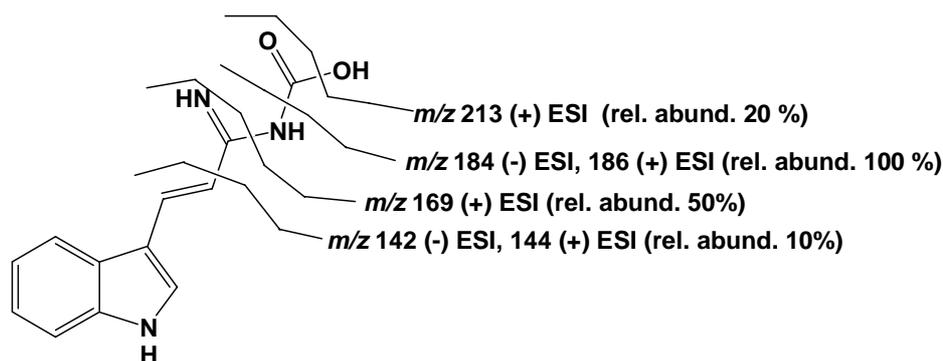
Hamigeramide (**3**) was obtained as a yellowish white powder from the biologically active EtOAc extract. It has the molecular formula of  $C_{12}H_{11}N_3O_2$  as suggested by EIMS. Its  $^1H$  NMR spectrum (Table 2) was very similar to those of 3-indoleacrylamide and its acid congener [22] except for the difference in signals observed for the 1,2-*trans*-disubstituted olefinic protons. The  $^{13}C$  data for **3** were obtained from a HMBC spectrum and significant differences compared to the known compounds were observed for the olefinic carbons. The  $^1H$  signals of the olefinic system of indoleacrylamide and its acid congener were observed at *ca.* 6.80 and 8.00 ppm while their carbon shifts were detected at *ca.* 115 and 138 ppm. The same chemical shifts were also observed for the

bis-indole alkaloids, chondriamides, isolated from the red alga *Chondria atropurpurea* [10, 23]. The functionality common to both the chondriamides and indoleacrylamide is the presence of a carbonyl moiety adjacent to the *trans* olefinic bond. The  $^1\text{H}$  signals of the olefinic system of hamigeramide (**3**) were observed at 7.18 and 7.10 ppm while their carbon shifts were detected at 118.2 and 117.3 ppm which suggested that **3** had a unique terminal functionality. Since the  $^1\text{H}$  and  $^{13}\text{C}$  chemical shifts of the olefinic protons were comparable to those of hamigeramine (**2**), this could suggest the presence of an adjacent imino amino moiety. Due to the very small amount of hamigeramide isolated, it was not possible to obtain a  $^{13}\text{C}$  NMR spectrum to prove the presence of the imino group. The HMBC spectra also did not show any discernible correlation between any of the olefinic protons to the expected imino carbon chemical shift at *ca.* 155 ppm. Ion trap ESI MS/MS showed the presence of the imino amino moiety and its connectivity were confirmed (Figure 3). ESIMS fragmentation positive mode of the  $[\text{M}+\text{H}]^+$  ion at  $m/z$  230.21 gave a major daughter ion peak at  $m/z$  186.1  $[\text{M}-\text{CO}_2\text{H}]^+$  which indicated the loss of the terminal acid unit, followed by subsequent loss of an  $\text{NH}_3$  unit ( $m/z$  169.3) and  $\text{C}=\text{NH}$  ( $m/z$  144.1). This was confirmed by the (-) ESIMS fragmentation of the  $[\text{M}-\text{H}]^-$  ion at  $m/z$  228.62 which gave a major daughter ion peak at  $m/z$  184.1  $[\text{M}-\text{CO}_2\text{H}]^-$ . The daughter ion peak at  $m/z$  169.3 in the (+) ESIMS fragmentation also indicated the absence of an enamide unit as in hamigeroxalamic acid (**1**) and hamigeramine (**2**) which further suggested that the imino unit was adjacent to the 1,2-*trans* disubstituted double bond. This accounted for the relatively sharp doublet of both the olefinic protons in the  $^1\text{H}$  NMR spectrum of hamigeramide (**3**) compared to hamigeramine (**2**). The fragmentation pattern suggested an imino carbamic acid terminal moiety and thus hamigeramide was elucidated as 1-imino-3-(1H-indol-3-yl)-allyl]-carbamic acid.

**Table 1.**  $^1\text{H}$ ,  $^{13}\text{C}$ -NMR, and HMBC data of compounds **1** and **2** in DMSO-  $d_6$ .

No	1			2		
	$\delta_{\text{H}}$ (m, <i>J</i> in Hz)	$\delta_{\text{C}}$	HMBC ( $\delta_{\text{H}}$ to $\delta_{\text{C}}$ )	$\delta_{\text{H}}$ (m, <i>J</i> in Hz)	$\delta_{\text{C}}$	HMBC ( $\delta_{\text{H}}$ to $\delta_{\text{C}}$ )
1		128.2 s			125.9 s	
2	7.09 (d, 8.5)	127.9 d	C-1, C-4, C-7	7.25 (d, 8.4)	127.5 d	C-1, C-6, C-7
3	6.67 (d, 8.5)	116.5 d	C-1, C-4, C-5	6.75 (d, 8.4)	115.6 d	C-1, C-4, C-5
4	9.31 (OH, br s)	156.4 s	C-3, C-4, C-5	9.62 (OH, br s)	157.3 s	C-3, C-4, C-5
5	6.67 (d, 8.5)	116.5 d	C-1, C-3, C-4	6.75 (d, 8.4)	115.6 d	C-1, C-3, C-4
6	7.09 (d, 8.5)	127.9 d	C-1, C-4, C-7	7.25 (d, 8.4)	127.5 d	C-1, C-2, C-7
7	6.35 (d, 14.5)	114.0 d	C-2, C-6, C-8	6.81 (d, 13.9)	121.3 d	C-2, C-6, C-8
8	7.10 (dd, 14.5, 10.7)	122.1 d	C-1, C-7	7.29 (br d, 13.9)	119.5 d	C-7, C-9, C-10
9	10.07 (NH, d, 10.7)			10.97 (NH, br s)		
10		163.0 s		9.29 (NH, br s) <sup>a</sup>	155.1 s	C-11
11		158.3 s		9.10 (OH, br s) <sup>a</sup>	155.7 s	

<sup>a</sup> These assignments are interchangeable but the signal at 9.29 ppm shows a correlation to C-11.



**Figure 3.** MS/MS fragmentation of the pseudomolecular ion peak at  $m/z$  230.21  $[M+H]^+$  and  $m/z$  228.62  $[M-H]^-$  of hamigeramide (**3**).

**Table 2.**  $^1H$ ,  $^{13}C$ -NMR, and HMBC data of compound **3** in DMSO- $d_6$ .

No.	$\delta_H$ (m, $J$ in Hz)	$\delta_C^a$	HMBC ( $\delta_H$ to $\delta_C$ )
1	11.20 (NH, br s)		
2	7.50 (s)	127.3 d	C-3, C-3a, C-7a, C-8
3		111.2 s	
3a		124.5 s	
4	7.90 (d, 7.8)	120.1 d	C-6, C-3a, C-7a
5	7.08 (t, 7.8)	119.0 d	C-7, C-3a
6	7.15 (t, 7.8)	121.5 d	C-7a
7	7.40 (d, 7.8)	112.3 d	C-5, C-3a
7a		138.4 s	
8	7.10 (d, 14.0)	117.3 d	C-2, C-3a
9	7.18 (d, 14.0)	118.2 d	C-3
10			
11	8.91 (NH, br s)		
12		b	

<sup>a</sup> Carbon assignments were determined indirectly from the HMBC.

<sup>b</sup> There are two further very broad exchangeable signals at *ca.* 9.3 and 10.8 ppm. The carboxylic acid  $^{13}C$  signal was not detected.

Hamiguanosinol (**5**) was isolated as a pale yellowish white amorphous solid from the biologically active aqueous phase. The ESI-MS showed the molecular ion peak at  $m/z$  314  $[M+H]^+$ , corresponding to the molecular formula  $C_{11}H_{15}N_5O_4S$ , as confirmed by HREIMS. The  $^1H$ - and  $^{13}C$ -NMR spectra of hamiguanosinol (**5**) were similar to those of 6-hydroxy guanosine [24]. The  $^1H$ -NMR signals at  $\delta$  5.60,  $\delta$  4.50, 3.95, 4.05, 2.80, and 2.70, together with the  $^{13}C$ -NMR signals at  $\delta$  87.0, 84.0, 74.0, 71.5 and 35.0, indicated the presence of a  $\beta$ -ribose moiety. The remaining  $^1H$ - and  $^{13}C$ -NMR signals at  $\delta_H$  10.57 (1H, s), 7.90 (1H, s) and 6.40 (2H, br s) and  $\delta_C$  at 135.0 (d), 118.0 (d), 157.2 (s) and 153.5 (s), respectively were assigned to the 2-amino purine nucleus. The occurrence of the  $\beta$ -ribose moiety was confirmed by the HMBC spectrum which showed correlations between exchangeable hydroxyl protons that resonated at  $\delta_H$  5.40 and 5.20 with  $\delta_{C-3'}$  71.5/  $\delta_{C-1'}$  87.0 and  $\delta_{C-2'}$

74.0 /  $\delta_{C-4}$  84.0, respectively. The only difference from known nucleoside, 6-hydroxy guanosine, was the presence of a methyl singlet at  $\delta_H$  2.05 which showed a direct correlation with the quaternary carbon at  $\delta_C$  16.0 suggesting the presence of a SCH<sub>3</sub> group. This was confirmed from the ESI-MS/MS experiment which showed daughter ions at  $m/z$  163 for the methylthioribosyl moiety while a subsequent fragment ion at  $m/z$  97 indicated the presence of the thiomethyl (SCH<sub>3</sub>) moiety. For the guanine nucleus, the presence of a hydroxyl group at C-6 instead of a keto function was confirmed by signals at  $\delta_H$  10.57 and  $\delta_C$  157.2 instead of a <sup>13</sup>C signal at *ca.*170 ppm for a keto amide function. The hydroxyl substituent at C-6 was also evident from the four bond HMBC correlations of  $\delta_H$  7.90 (H-8) with  $\delta_C$  157.2 (C-6). The attachment of the ribosyl moiety with the purine unit at N-9 was confirmed by HMBC correlations of the anomeric proton (H-1') with C-4 and C-8. Hamiguanosinol (**5**) was then concluded to be 5'-deoxy-5'-methylthio-6-hydroxy-guanosine. The fish deterrent activity of the aqueous phase was accounted for by the presence of hamiguanosinol. Nucleoside compounds are known for their antiviral activity and the first notable discovery of biologically active marine metabolites was actually unusual nucleosides. They were first isolated from a Caribbean sponge in the 1950s, and later served as lead structures for the development of the important anti-viral drugs such as Ara-A known commercially as acyclovir [25].

The fish deterrent activity of the isolated compounds was not quantified due to the very small amounts isolated. However, the biological activity of related compounds should be mentioned. Hamigeroxalamic acid (**1**) is among the few examples of natural products to date that carry the oxalamic functionality. Another important natural product having the same oxalamic moiety is psammaplin F which was found to selectively inhibit histone deacetylase, which is an epigenic modifier in the silencing of tumour suppression genes. It is also important to note that among the 10 known psammaplin derivatives, only psammaplin F, the oxalamic congener, was found to inhibit HDAC. Hamigeramide (**3**) is basically an indole alkaloid. Related compounds are the bis-indole alkaloids chondramides that were reported to exhibit anthelmintic and antiviral activity against HSV II [10].

## Material and Methods

### *General experimental procedures*

Optical rotation data were recorded on a Perkin-Elmer Model 341 LC polarimeter. 1D and 2D NMR spectra (chemical shifts in ppm) were recorded on Bruker AMX 300, ARX 400, or DRX 500 NMR spectrometers using standard Bruker software. Mass spectra (ESIMS) were recorded on a ThermoFinnigan Deca LCQ ion trap mass spectrometer. High resolution EIMS were recorded on a Finnigan MAT 8430 and on an Intectra AMD 402. For HPLC analysis, samples were injected into a HPLC system equipped with a photodiode-array detector (Gynkotek, München, Germany). Routine detection was at 254, 280, and 340 nm. The separation column (125 x 4 mm, i.d.) was prefilled with Eurospher 100-C<sub>18</sub>, 5  $\mu$ m (Knauer, Berlin, Germany). Separation was achieved by applying a

linear gradient from 100% H<sub>2</sub>O (with 0.2% TFA) to 100% MeOH over 40 min. Compounds were purified by semipreparative HPLC which was conducted on a Merck Hitachi LaChrome L-7100 pump and Merck Hitachi LaChrome L-7400 UV detector. Chromatograms were recorded on a Merck Hitachi D-2000 Chromato-Integrator. Separation columns (300 x 8 mm, i.d.) were prefilled with Eurospher 100-C<sub>18</sub>, 7 μm (Knauer, Berlin, Germany). TLC was performed on precoated TLC plates with Si gel 60 F<sub>254</sub> and Si gel RP-18 F<sub>254</sub> (Merck, Darmstadt, Germany). Compounds were detected by UV absorbance or by fluorescence at 254 and 366 nm. Solvents were distilled prior to use and spectral grade solvents were used for spectroscopic measurements.

**Table 3.** <sup>1</sup>H, <sup>13</sup>C-NMR, and HMBC data of compound **5** in DMSO-*d*<sub>6</sub>.

No.	δ <sub>H</sub> (m, J in Hz)	δ <sub>C</sub>	<sup>1</sup> H- <sup>1</sup> H COSY	HMBC (δ <sub>H</sub> to δ <sub>C</sub> )
2	(NH <sub>2</sub> ) 6.40 (br s)	153.5 s		
4		153.0 s		
5		118.0 s		
6	(OH) 10.57 (br s)	157.2 s		
8	7.90 s	135.0 d		C-5, C-4, C-6
1'	5.60 (d, 5.7)	87.0 d	H-2'	C-8, C-4, C-3'
2'	4.50 (q, 5.7)	74.0 d	H-1', H-3'	C-1', C-4'
2'-OH	5.40 (d, 6.3)			C-1', C-3'
3'	3.95 (m)	71.5 d	H-2', H-4'	C-1'
3'-OH	5.20 (d, 5.1)			C-2', C-4'
4'	4.05 (t, 6.9)	84.0 d	H-3', H-5'A, H-5'B	C-3'
5'A	2.80 (dd, 13.9, 6.9)	35.0 t	H-5' B, H-4'	C-6', C-3', C-4'
5'B	2.70 (dd, 13.9, 6.9)			
6'	2.05 (s)	16.0 q	H-5'A, H-4'	C-5'

### Animal Material

The sponge *Hamigera hamigera* was collected in April 2000 from the Mediterranean Sea near Elba, Italy, at a depth of 5 to 7 m. The sponge was frozen immediately at -20° until work up. The taxonomic identification was carried out by Dr. R. Van Soest (Instituut voor Systematiek en populatie Biologie, Zoological Museum, Amsterdam). A voucher specimen (ZMA.POR 14397) was deposited at the Department of Coelenterates and Porifera, Zoologisch Museum, University of Amsterdam.

### Extraction and isolation

The freeze-dried sponge was extracted with CH<sub>2</sub>Cl<sub>2</sub> and subsequently with MeOH, the resulting total crude extract obtained with both solvents was evaporated to dryness *in vacuo*. The total crude extract (5.3 g) was partitioned between H<sub>2</sub>O and cyclohexane, then H<sub>2</sub>O and EtOAc and finally between H<sub>2</sub>O and n-BuOH. The EtOAc and H<sub>2</sub>O extracts were found to be active in the fish-feeding assay. The biologically active extracts were chromatographed separately over a Sephadex LH-20

column using MeOH as an eluent. Each fraction was collected to a volume of 5 mL and monitored by TLC on a pre-coated silica gel plates (Merck, Darmstadt, Germany) which were developed under the appropriate solvent system. The pooled non-polar fractions from the EtOAc extract were subjected to normal-phased silica gel column chromatography which was eluted with hexane:EtOAc (90:10) resulting in the isolation of steroids **6** (3.1 mg), **7** (2.1 mg), and **8** (1.8 mg) while the polar fraction was chromatographed over a reversed-phase C<sub>18</sub> column and eluted with MeOH:H<sub>2</sub>O (1:1) which yielded the new compound **2**. The polar fractions obtained from the EtOAc extract afforded the isolation of new compounds **1** (7.5 mg), **2** (2.5 mg), **3** (3.5 mg), and the known compound indole-3-carbaldehyde (**4**, 2.0 mg), after the extract was chromatographed over a reversed-phase C<sub>18</sub> column which was also eluted with MeOH:H<sub>2</sub>O (1:1). The compounds were purified by semi-preparative reversed phase HPLC using a gradient of H<sub>2</sub>O and MeOH. The aqueous extract was subjected to Sephadex LH-20 column chromatography using MeOH as eluent and yielded the new nucleoside (**5**, 2.3 mg) which was also further purified by semi-preparative reversed phase HPLC using a gradient of H<sub>2</sub>O and MeOH.

### *Spectroscopic Data*

**Hamigeroxalamic acid (N-[2-(4-hydroxy-phenyl)-vinyl]-oxalamic acid) (1):** yellowish white powder; UV  $\lambda_{\max}$  (MeOH) nm 248.5, 320.1; <sup>1</sup>H NMR and <sup>13</sup>C NMR (DMSO-d<sub>6</sub>): Table 1; (+) ESIMS (ion trap)  $m/z$  246.2 [M+K]<sup>+</sup>; (−) ESIMS  $m/z$  206.2 [M−H]<sup>−</sup>, 252.0 [M+HCOOH−H]<sup>−</sup>, 413.1 [2M−H]<sup>−</sup>. The results of the MS/MS fragmentation of the (+) and (−) ESIMS (ion trap) are shown in Figure 2.

**Hamigeramine ([2-(4-Hydroxy-phenyl)-vinylamino]-imino-acetic acid) (2):** yellowish white powder; UV  $\lambda_{\max}$  (MeOH) nm 226.7, 324.0; <sup>1</sup>H NMR and <sup>13</sup>C NMR (DMSO-d<sub>6</sub>): Table 1; (+) FABMS  $m/z$  207.1 [M+H]<sup>+</sup>, 229 [M+Na]<sup>+</sup>; (+) ESIMS (ion trap)  $m/z$  207.1 [M+H]<sup>+</sup>, 412.9 [2M+H]<sup>+</sup>; (−) ESIMS (ion trap) 205.4 [M−H]<sup>−</sup>, 411.5 [2M−H]<sup>−</sup>, 617.1 [3M−H]<sup>−</sup>. The results of the MS/MS fragmentation of the (+) and (−) ESIMS (ion trap) are shown in Figure 2.

**Hamigeramide (3):** yellowish white powder; UV  $\lambda_{\max}$  (MeOH) nm 228.0, 346.0; <sup>1</sup>H NMR and <sup>13</sup>C NMR (DMSO-d<sub>6</sub>): Table 2; (+) ESIMS (ion trap)  $m/z$  230.2 [M+H]<sup>+</sup>, 252.1, 459.0 [2M+H]<sup>+</sup>; (−) ESIMS (ion trap) 228.6 [M−H]<sup>−</sup>, 457.6 [2M−H]<sup>−</sup>, 686.3 [3M−H]<sup>−</sup>. The results of the MS/MS fragmentation of the (+) and (−) ESIMS (ion trap) are shown in Figure 3.

**Hamiguanosinol (5):** pale yellowish white amorphous powder,  $[\alpha]_D^{20} +15.8^\circ$  (MeOH, *c* 0.10); UV  $\lambda_{\max}$  (MeOH) nm 254.6, 275.0 (sh); <sup>1</sup>H NMR and <sup>13</sup>C NMR (DMSO-d<sub>6</sub>): Table 3; (+)ESIMS  $m/z$  314.5 [M+H]<sup>+</sup>, 627.3 [2M+H]<sup>+</sup>; (−)ESIMS  $m/z$  312.1[M−H]<sup>−</sup>, 625.8 [2M−H]<sup>−</sup>, 938.9 [3M−H]<sup>−</sup>; EIMS (70eV)  $m/z$  [M]<sup>+</sup> 313 (13), 236 (12), 163 (12), 151 (16), 138 (44), 121 (93), 84 (100); HRESIMS  $m/z$  312.0763 [M−H]<sup>−</sup> (calcd for C<sub>11</sub>H<sub>14</sub>N<sub>5</sub>O<sub>4</sub>S, 312.0767).

### *Feeding experiments with Blennius sphinx*

For the feeding experiments, crude sponge extracts and/or fractions were incorporated into the artificial fish diet. The artificial fish diet was treated with physiological concentrations of secondary sponge metabolites as determined in 10 mL ethanol preserved sponge material to match the natural concentrations of the metabolites found in the sponge. All metabolites used for the feeding experiments were dissolved in MeOH and incorporated into 1.053 g of ground commercial fish food granules (TetraMarin Granulat, TetraWerke, Melle, Germany). This mixture was then added to 9 mL of 2% agar at 48° C to give a volume of 10 mL. Control diet was likewise prepared by mixing the food granules in MeOH without the sponge extracts. Freshly prepared treated diet and control diet were then separately poured into rectangular molds (2 x 25 x 250 mm) placed on a meshed screen. The food diet was allowed to set on the meshed screen which were then cut to six equivalent food strips. Each of the food strips contained a treated diet and a control diet, each covering an area of 10 x 15 meshed screen squares.

The feeding experiments were then performed in an aquarium (size: 170 x 80 x 40 cm) using 70 individuals of the polyphagous Mediterranean fish *Blennius sphinx*. The food strips were placed at the bottom of the aquarium. The food strips were checked at regular time intervals and were removed when approximately one-third of the total food mass was consumed. To determine the amount of control and treated food eaten, the number of empty squares in the meshed screen were counted [25, 27]. The results obtained from several replicate experiments were pooled and analyzed with a paired t-test.

## Acknowledgments

W.H. would like to thank the government of Egypt for financial support. The authors are grateful to Dr. R. Van Soest for identification of the sponge and to C. Kakoschke and B. Jaschok-Kentner for recording NMR data (GBF, Braunschweig). Financial support by a grant of the BMBF (“BIOTECMARIN”) is gratefully acknowledged.

## References and Notes

1. Casapullo, A.; Finamore, E.; Minale, L.; Zollo F. A dimeric peptide alkaloid of a completely new type, Anchinopeptolide A, from the marine sponge *Anchinoe tenacior*. *Tetrahedron Lett.* **1993**, *34*, 6297-6300.
2. Casapullo, A.; Minale, L.; Zollo, F. The unique 6-(p-hydroxyphenyl)-2-H-3,4-dihydro-1,1-dioxo-1,4-thiazine and the new tripeptide L-glu-gly-4-hydroxystirylamine from the marine sponge *Anchinoe tenacior*. *Tetrahedron Lett.* **1994**, *35*, 2421-2422.

3. Casapullo, A.; Minale, L.; Zollo, F. Four new dimeric peptide alkaloids, anchinopeptolides B-D, and cycloanchinopeptolide C, congeners of anchinopeptolide A, from the Mediterranean marine sponge *Anchinoe tenacior* *J. Nat. Prod.* **1994**, *57*, 1227-1233.
4. Rudi, A.; Stein, Z.; Green, S.; Goldberg, I.; Kashman, Y. Phorbazoles A-D, novel chlorinated phenylpyrrolyloxazoles from the marine sponge *Phorbas* aff. *clathrata*. *Tetrahedron Lett.* **1994**, *35*, 2589-2492.
5. Searle, A. S.; Molinski, T. F. Absolute configuration of Phorboxazole A and B from the marine sponge *Phorbas* sp. 1. Macrolide and hemiketal rings. *J. Am. Chem. Soc.* **1996**, *118*, 9422-9423.
6. Molinski, T. F. Absolute configuration of Phorboxazole A and B from the marine sponge *Phorbas* sp. 2. C-43 and complete stereochemistry. *Tetrahedron Lett.* **1996**, *37*, 7879-7880.
7. Wellington, K. D.; Cambie, R. C.; Rutledge, P. S.; Bergquist, R. Chemistry of sponges. 19. Novel bioactive metabolites from *Hamigera tarangensis*. *J. Nat. Prod.* **2000**, *63*, 79-85.
8. Cambie, R. C.; Lai, A. R.; Kernan, M. R.; Bergquist, P. R. Chemistry of sponges. A novel brominated benzocyclooctane derivative from *Hamigera hamigera*. *J. Nat. Prod.* **1995**, *58*, 940-942.
10. Guella, G.; Mancini, I.; Zibrowius, H.; Pietra, F. 160. Aplysinopsin-type alkaloids from *Dendrophyllia* sp., a Scleractinian coral of the family Dendrophylliidae of the Philippines. Facile photochemical (Z/E) *Helv. Chim. Acta* **1989**, *72*, 1444-1450.
11. Anjaneyulu, V.; Rao, K. N. R.; Kobayashi M. (24S)-24-Methylcholesta-4-ene-3 $\beta$ ,6 $\beta$ -diol from a gorgonian (*Rumphella aggregata*) of the Andaman and Nicobar Islands *Ind. J. Chem.* **1995**, *34B*, 78-80.
12. Wright, J. McInnes, A. Shmizu, S. Smith, D. Walter, J. Identification of C-24 alkylepimers of marine sterols by <sup>13</sup>C nuclear magnetic resonance spectroscopy. *Can. J. Chem.* **1978**, *56*, 1898-1903.
13. Itoh, T.; Sica, D.; Djerassi, C. Minor and trace sterols in marine invertebrate. Part 1. 35. Isolation and structure elucidation of seventy four sterols from the sponge *Axinella cannabina*. *J. Chem. Soc. Perkin Trans 1*, **1983**, 147-153.
14. Palermo, J. A.; Brasco, M. F. R.; Seldes, A. M. Stroniamides A-D: Alkaloids from a Patagonian sponge *Cliona* sp. *Tetrahedron* **1996**, *52*, 2727-2734.
15. Bokesch, H. R.; Pannell, L. K. McKee, T. C.; Boyd, M. R. Coscinamides A, B, and C, three new indole alkaloids from the marine sponge *Coscinoderma* sp. *Tetrahedron Lett.*, **2000**, *41*, 6305-6308.
17. Piña, I. C.; Gautschi, J. T.; Wang, G.-Y.-S.; Sanders, M. L.; Schmitz, F. J.; France, D.; Cornell-Kennon, S.; Sambucetti, L. C.; Remiszewski, S. W.; Perez, L. B.; Bair, K. W.; Crews, P. Psammaplins from the sponge *Pseudoceratina purpurea*: Inhibition of both histone deacetylase and DNA methyltransferase. *J. Org. Chem.* **2003**, *68*, 3866-3873.

18. Pettit, G. R.; Butler, M. S.; Williams, M. D.; Filiatrault, M. J.; Pettit, R. K. Isolation and structure of hemibastadinols 1-3 from the Papua New Guinea marine sponge *Ianthella basta*. *J. Nat. Prod.* **1996**, *59*, 327-934.
19. Dumdei, E.; Andersen, R. J. Igzamide, a metabolite of the marine sponge *Plocamissma igzo*. *J. Nat. Prod.* **1993**, *56*, 792-794.
20. Sakai, R.; Higa, T. Tubastrine, a new guanidostyrene from the coral *Tubastrea aurea*. *Chem. Lett.* **1987**, 122-128.
21. Sperry, S.; Crews, P. Dihydrotubastrines: Phenethylguanidine analogues from the Indo-Pacific marine sponge *Petrosia* cf. *contignata*. *J. Nat. Prod.* **1998**, *61*, 859-861.
22. Davyt, D.; Entz, W.; Fernandez, R.; Mariezcurrena, R.; Mombrú, A. W.; Saldaña, J.; Domínguez, L.; Coll, J.; Manta, E. A new indole derivative from the red alga *Chondria atropurpurea*. Isolation, structure determination, and anthelmintic activity. *J. Nat. Prod.* **1998**, *61*, 1560 – 1563.
24. Pretsch, E.; Seibl, J.; Simon, W.; Clerc, T. Tabellen zur Strukturaufklärung organischer Verbindungen mit spektroskopischen Methoden. **1990**, Heidelberg, Springer Verlag, p.C212.
25. Balzarini, J.; Haller-Meier, F., De Clercq, E., Meier, C. Antiviral activity of cycloaligenyl prodrugs of acyclovir, carbovir and abacavir. *Antivir. Chem. Chemother.* **2001**, *12*, 301-306.
27. Hay, M. E.; Stachowicz, J. J.; Cruz-Rivera, E.; Bullard, S.; Deal, M. S.; Linquist, N. Bioassays with marine and freshwater macroorganisms. In: Haynes KF, Millar J. G. (eds) *Methods in Chemical Ecology*. **1998**, Vol. 2. Bioassay methods. Chapman and Hall, New York, pp 39-141.

*Sample Availability:* Samples are available from the authors.