

Mycorrhizal inoculation of pecan seedlings with some marketable truffles

GIAN MARIA NICCOLÒ BENUCCI¹, GREGORY BONITO², LEONARDO BACIARELLI
FALINI¹, MATTIA BENCIVENGA¹ and DOMIZIA DONNINI¹

¹Department of Applied Biology, University of Perugia, Borgo XX Giugno 74
I-06121 Perugia, gian.benucci@gmail.com

²Biology Department, Duke University, Durham, NC 27708, USA

Benucci G.M.N., Bonito G., Baciarelli Falini L., Bencivenga M., Donnini D.: *Mycorrhizal inoculation of pecan seedlings with some marketable truffles*. Acta Mycol. 47 (2): 179–184, 2012.

Pecan is the common name of *Carya illinoensis* (Wangenh.) K. Koch, an ectomycorrhizal tree native to North America, also frequently known as hickory. Mycorrhizal inoculations of pecan seedlings with: *Tuber aestivum* Vittad., *T. borchii* Vittad., *T. indicum* Cooke & Massee, and *T. lyonii* Butters are described and discussed.

Key words: *Carya illinoensis*, truffle spore-slurry, multi-cropping, *Tuber borchii*, *T. aestivum*, *T. indicum*, *T. lyonii*

INTRODUCTION

Pecan is the common name of *Carya illinoensis* (Wangenh.) K. Koch, also frequently known as hickory. It is a deciduous tree in the Juglandaceae family with pinnately compound leaves, which produces large and economically valuable nuts. This tree also establishes ectomycorrhizal (ECM) associations with hypogeous and epigeous fungi. Pecan is native to North America and grows naturally on the moist bottomland habitat along streams in the USA ranging from Indiana south to Kentucky and Alabama and from Iowa south to Texas, principally along the Mississippi river.

This species is cultivated largely in southeastern North America exclusively for the edible nuts, which are used for pecan pie, pecan pralines or food products including cereals, energy bars and candy bars. Pecan production of the United States is likely to be over 433 million dollar per year, and the industry is in a period of growth (USDA 2008). A large proportion of pecan harvests are being exported to China, a rapidly growing market. Large productive commercial pecan orchards have also been recently established in Brazil, Israel, and Australia (Wakeling et al. 2001).

In the late 1980's Hanlin and colleagues (1989) discovered truffles fruiting naturally in pecan orchards in Georgia, USA. Bonito and colleagues (2011) followed up on this study and showed that the ECM truffle genus *Tuber* (*Pezizaceae*) is naturally abundant in fungal communities of many pecan orchards in Georgia, including the species *Tuber lyonii* Butters, whose ascocarps are frequently found and collected. Although *T. lyonii* is sold and served in US restaurant with success, the market is still underdeveloped because of its low organoleptic qualities, low availability and low quality of truffles collected (Bonito et al. 2011; Benucci et al. 2012a). Truffles are harvested by raking, then mature and less aromatic immature truffles are marketed spreading a negative opinion on the product.

MATERIAL AND METHODS

To overcome these problems and evaluate whether pecan can associate with both native and non-native truffle species as first step of future potential multi-cropping of pecans and truffles, some investigation on ECM inoculations with *Tuber* spp. on pecan have been realized (Bonito et al. 2010; Bonito et al. 2011; Benucci et al. 2012a; Bonito et al. 2012). Spore inocula were prepared with appreciated and widely distributed truffle species: *Tuber aestivum* Vittad., *Tuber borchii* Vittad., *Tuber indicum* Cooke & Masee, *Tuber macrosporum* Vittad. and *Tuber lyonii* Butters.

After inoculations with spore slurries, pecan seedlings were grown in greenhouse conditions for several months (Fig. 1a), after which plants were harvested and mycorrhization levels quantified visually (Bencivenga et al. 1995). ECM were described morpho-anatomically and molecularly by species-specific polymerase chain reactions (PCRs) (Mello et al. 1999; Mello et al. 2002) and/or by sequencing and analyzing the internal transcribed spacer (ITS) (White 1990) and 28S large subunit (LSU) (Vilgalys, Hester 1990) nuclear ribosomal DNA (nrDNA) region.

RESULTS

Tuber borchii and *T. aestivum* formed abundant ECM on pecan (Fig. 1c, 1e), with $\approx 60\%$ and $\approx 40\%$ root-tip colonization, respectively (Benucci et al. 2012a). In contrast, no ECM of *T. macrosporum* were detected on pecan, which may indicate that this host is not compatible with this fungal species, although it is known that *T. macrosporum* ECM are difficult to obtain under laboratory and nursery conditions (Benucci et al. 2012b). Morphological and anatomical characteristics of *T. borchii* and *T. aestivum* on pecan have similar features to those that they display with other host species (e.g., oak, hazelnut), and their morphologies are consistent with previous reports (Giomaro et al. 2000; Granetti et al. 2005). In particular, *T. borchii* possessed straight needle-shaped cystidia and puzzle-like cell pattern (Fig. 1g, 1h), while *T. aestivum* possessed curly cystidia and angular cell pattern (Fig. 1f, 1i).

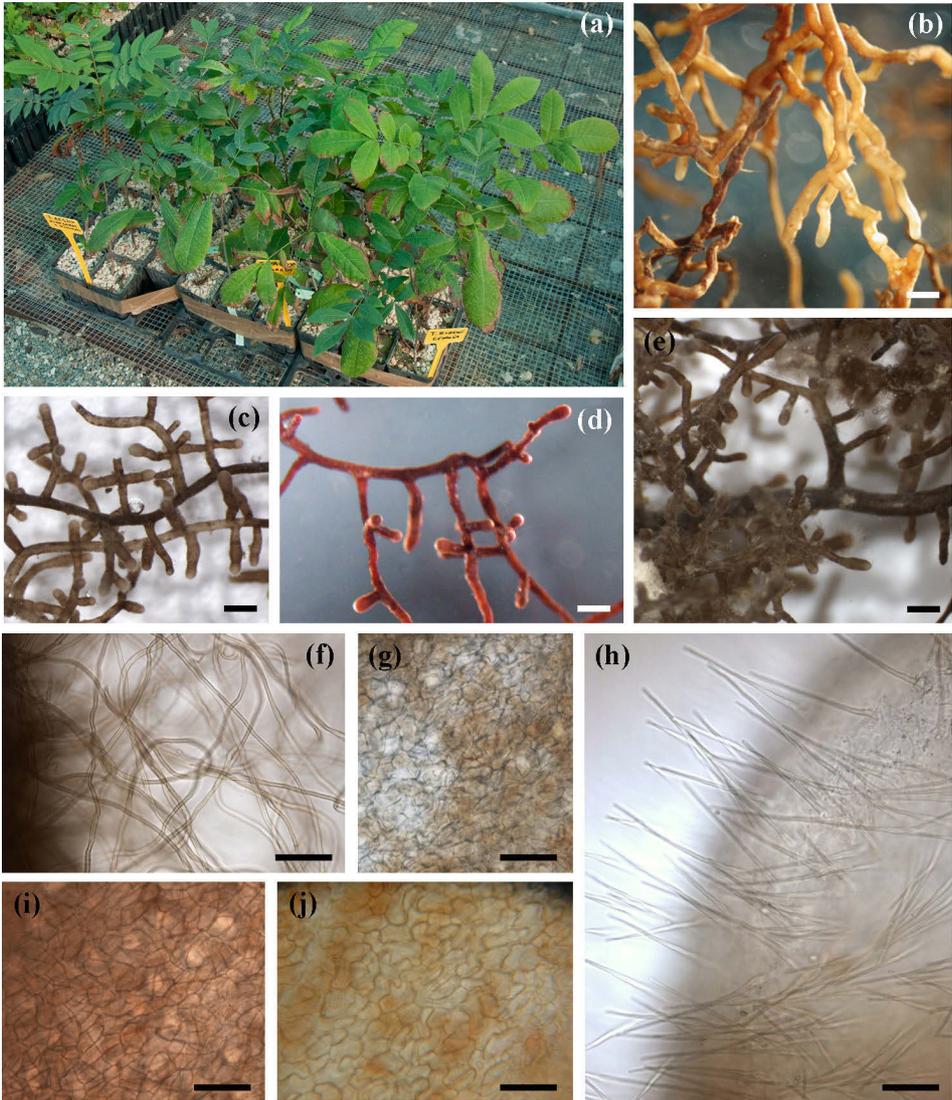


Fig. 1. Pecan seedlings and morphology of *T. aestivum*, *T. borchii*, *T. indicum* and *T. lyonii* ECM. (a) Pecan seedlings in the nursery; (b) *T. lyonii* ECM on pecan roots (scale bar=700 μm); (c) *T. borchii* ECM on pecan roots (scale bar=700 μm); (d) *T. indicum* ECM on pecan roots (scale bar=700 μm); (e) *T. aestivum* ECM on pecan roots (scale bar=700 μm); (f) curly cystidia of *T. aestivum* ECM (scale bar=30 μm); (g) outer mantle layer of *T. borchii* ECM (scale bar=30 μm); (h) needle-shape cystidia of *T. borchii* (scale bar=30 μm); (i) outer mantle layer of *T. aestivum* ECM (scale bar=30 μm); (j) outer mantle layer of *T. lyonii* ECM (scale bar=30 μm).

The molecular PCR assay confirmed that amplicons obtained from truffle ECM are consistent with our morphological and taxonomical assessment. Their ITS nrDNA sequences matched at 100% query coverage with 0.0 E-values and 99-100% identity to those present in GenBank for the same fungal species.

Bonito and colleagues (2010) were able to synthesize *T. indicum* ECM on *Carya illinoensis* seedlings using standard inoculation methods. The identification of ECM was confirmed by morphology (Fig. 1d) and ITS sequence data. ECM produced by *T. indicum* on pecan were unramified to irregularly pinnate, dark amber in colour and showed characteristic puzzle-like pseudoparenchyma cells in the outer mantle with emanating right-angle branching cystidia, as reported from other hosts (Geng et al. 2009).

Tuber lyonii ECM were also well formed on pecan seedlings 6 months after they had been inoculated with *T. lyonii* spores (Bonito et al. 2011). Ectomycorrhizas are characterized by a thin and smooth mantle with epidermoid puzzle-like cells (Fig. 1b, 1j). Molecular ITS sequences confirmed the identity of *T. lyonii* ECM.

DISCUSSION

Together these data demonstrate that European, Asian and American truffles are able to form healthy ECM on pecan, regardless of the fact that this plant is endemic to North America. These studies lead us to conclude that there is the potential to multi-crop various truffle species with pecan. To successfully cultivate pecan with the simultaneous production of truffles represents a unique opportunity to diversify agricultural outputs from a given tract of land, and to add extra income in an environmental and sustainable way (Benucci et al. 2013; Donnini et al. 2013).

Furthermore, given the attributes of pecan wood for lumber and flooring, opportunities exist to obtain additional revenue streams from the timber production at the end of the cultivation cycle or following stand thinning (Benucci et al. 2012a). In US, although some orchards were originally brought for lumber and furniture, most pecan trees taken out are chipped and used for smoking meats in fires and traditional barbecues. However, the wood is quite dense, making it hard on saw blades of modern mill equipment, and transport costs are high. Nonetheless, once CO₂ is regulated in the US, we expect more sustainable practices will develop and pecan wood and timber will be favored again for flooring and furniture.

CONCLUSIONS

In conclusion, given the ability of pecan to readily form ECM with economic truffle species from five different clades *Tuber* that includes North American, European, and Asian truffle species, pecan holds promise as a host plant that is compatible with the co-cropping of truffles and nuts. Whether or not *Tuber* ECM are maintained

when mycorrhized seedlings are planted out in an orchard setting, and whether these truffle species will fruit with pecan still need to be addressed however. Moreover, in humid climates pecan cultivation often requires heavy inputs of biocides to combat plant diseases and inputs of fertilizers to boost the production of nuts. Further studies are needed to address the potential accumulation of these harmful substances inside of truffles.

Acknowledgements. Suggestions and critical comments on the manuscript made by anonymous reviewers are greatly acknowledged. The authors are grateful to Andrea Vece and Andrea Gógán Csorbainé, who provided part of the truffles needed for mycorrhization trials.

REFERENCES

- Bencivenga M., Donnini D., Tanfulli M., Guiducci M. 1995. Tecnica di campionamento delle radici e degli apici radicali per la valutazione delle piante micorrizzate. *Micol Ital* 2: 35-47.
- Benucci G. M. N., Bonito G., Baciarelli Falini L., Bencivenga M. 2012a. Mycorrhization of pecan trees (*Carya illinoensis*) with commercial truffle species: *Tuber aestivum* Vittad. and *Tuber borchii* Vittad. *Mycorrhiza* 22: 383-392. DOI 10.1007/s00572-011-0413-z
- Benucci G. M. N., Bonito G., Baciarelli Falini L., Bencivenga M., Donnini D. 2013. Truffles, Timber, Food, and Fuel: Sustainable Approaches for Multi-cropping Truffles and Economically Important Plants. *Edible Ectomycorrhizal Mushrooms*, (Zambonelli A, Bonito G, eds), Springer-Verlag Berlin Heidelberg.
- Benucci G. M. N., Gogan Csorbai A., Baciarelli Falini L., Bencivenga M., Di Massimo G., Donnini D. 2012b. Mycorrhization of *Quercus robur* L., *Quercus cerris* L. and *Corylus avellana* L. seedlings with *Tuber macrosporium* Vittad. *Mycorrhiza* 22: 639-646. DOI 10.1007/s00572-012-0441-3
- Bonito G., Breneman T., Vilgalys R. 2011. Ectomycorrhizal fungal diversity in orchards of cultivated pecan (*Carya illinoensis*; Juglandaceae). *Mycorrhiza* 21: 601-612. DOI 10.1007/s00572-011-0368-0
- Bonito G., Trappe J. M., Donovan S., Vilgalys R. 2010. The Asian black truffle *Tuber indicum* can form ectomycorrhizas with North American host plants and complete its life cycle in non-native soils. *Fungal Ecology* 4: 83-93. DOI 10.1016/j.funeco.2010.08.003
- Bonito G., Smith M. E., Breneman T., Vilgalys R. 2012. Assessing ectomycorrhizal fungal spore banks of truffle producing soils with pecan seedling trap-plants. *Plant and Soil* 356: 357-366. DOI 10.1007/s11104-012-1127-5
- Donnini D., Gargano M. L., Perini C., et al. 2013. Wild versus cultivated mushrooms as a model of sustainable development. *Plant Biosystems*. DOI 10.1080/11263504.2012.754386
- Geng L. Y., Wang X. H., Yu F. Q., et al. 2009. Mycorrhizal synthesis of *Tuber indicum* with two indigenous hosts, *Castanea mollissima* and *Pinus armandii*. *Mycorrhiza* 19: 461-467.
- Giomaro G., Zambonelli A., Sisti D., Cecchini M., Evangelista V., Stocchi V. 2000. Anatomical and morphological characterization of mycorrhizas of five strains of *Tuber borchii* Vittad. *Mycorrhiza* 10: 107-114.
- Granetti B., De Angelis A., Materozzi G. 2005. Umbria terra di tartufi. Umbriagraf, Terni, Italy.
- Hanlin R. T., Wu M. L., Breneman T. B. 1989. The occurrence of *Tuber texense* in Georgia. *Mycotaxon* 34: 387-394.
- Mello A., Garnero L., Bonfante P. 1999. Specific PCR-primers as a reliable tool for the detection of white truffles in mycorrhizal roots. *New phytologist* 141: 511-516.
- Mello A., Cantisani A., Vizzini A., Bonfante P. 2002. Genetic variability of *Tuber uncinatum* and its relatedness to other black truffles. *Environ Microbiol* 4: 584-594.
- USDA 2008. National Agricultural Statistics Service. Noncitrus fruits and nuts: Fr Nt 1-3 (09). Unites States, <http://usda.mannlib.cornell.edu/MannUsda/viewDocumentInfo.do?documentID=1>
- Vilgalys R., Hester M. 1990. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* 172: 4238-4246.

- Wakeling L. T., Mason R. L., Bruce R., Caffin N. A. 2001. Composition of pecan cultivars Wichita and Western Schley [*Carya illinoensis* (Wangenh.) K. Koch] grown in Australia. *Journal of agricultural and food chemistry* 49: 1277-1281. DOI: 10.1021/jf000797d
- White T. J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *PCR protocols, a guide to methods and applications*, (Innis MA, Gelfand DH, Sninsky JJ, White TJ, eds), Academic Press Inc., New York , USA pp. 315-322.