




Review

Toxicity of Heavy Metals and Recent Advances in Their Removal: A Review

Manar K. Abd Elnabi ^{1,2}, Nehal E. Elkaliny ¹, Maha M. Elyazied ¹, Shimaa H. Azab ¹, Shawky A. Elkhalfifa ¹, Sohaila Elmasry ³, Moustafa S. Mouhamed ¹, Ebrahim M. Shalamesh ¹, Naira A. Alhoriény ¹, Abeer E. Abd Elaty ¹, Ibrahim M. Elgendy ¹, Alaa E. Etman ¹, Kholod E. Saad ¹, Konstantina Tsigkou ⁴ , Sameh S. Ali ^{1,5,*} , Michael Kornaros ^{4,*}  and Yehia A.-G. Mahmoud ¹

- ¹ Botany Department, Faculty of Science, Tanta University, Tanta 31527, Egypt; manar.abdelnabi@ejust.edu.eg (M.K.A.E.); nehal31012429@science.tanta.edu.eg (N.E.E.); maha31012879@science.tanta.edu.eg (M.M.E.); shaimaa31012609@science.tanta.edu.eg (S.H.A.); shawqe31012093@science.tanta.edu.eg (S.A.E.); mostafa30974505@science.tanta.edu.eg (M.S.M.); ibrahem231058915@science.tanta.edu.eg (E.M.S.); nira231058550@science.tanta.edu.eg (N.A.A.); abeer31012615@science.tanta.edu.eg (A.E.A.E.); ibrahim31012107@science.tanta.edu.eg (I.M.E.); alaa231058671@science.tanta.edu.eg (A.E.E.); khaloud31012947@science.tanta.edu.eg (K.E.S.); yehiamahmoud@science.tanta.edu.eg (Y.A.-G.M.)
- ² Biotechnology Program, Institute of Basic and Applied Science (BAS), Egypt-Japan University of Science and Technology, New Borg El-Arab City 21934, Egypt
- ³ Microbiology Department, Faculty of science, Damanhour University, Behaira 22514, Egypt; s.abdelazez00804@sci.dmu.edu.eg
- ⁴ Department of Chemical Engineering, University of Patras, 1 Karatheodori str, 26504 Patras, Greece; ktsigkou@chemeng.upatras.gr
- ⁵ Biofuels Institute, School of the Environment and Safety Engineering, Jiangsu University, Zhenjiang 212013, China
- * Correspondence: samh@ujs.edu.cn or samh_samir@science.tanta.edu.eg (S.S.A.); kornaros@chemeng.upatras.gr (M.K.)



Citation: Abd Elnabi, M.K.; Elkaliny, N.E.; Elyazied, M.M.; Azab, S.H.; Elkhalfifa, S.A.; Elmasry, S.; Mouhamed, M.S.; Shalamesh, E.M.; Alhoriény, N.A.; Abd Elaty, A.E.; et al. Toxicity of Heavy Metals and Recent Advances in Their Removal: A Review. *Toxics* **2023**, *11*, 580. <https://doi.org/10.3390/toxics11070580>

Academic Editors: Junhao Qin, Peidong Su, Feng Zhu and Lin Ding

Received: 28 May 2023

Revised: 14 June 2023

Accepted: 24 June 2023

Published: 3 July 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

Abstract: Natural and anthropogenic sources of metals in the ecosystem are perpetually increasing; consequently, heavy metal (HM) accumulation has become a major environmental concern. Human exposure to HMs has increased dramatically due to the industrial activities of the 20th century. Mercury, arsenic lead, chrome, and cadmium have been the most prevalent HMs that have caused human toxicity. Poisonings can be acute or chronic following exposure via water, air, or food. The bioaccumulation of these HMs results in a variety of toxic effects on various tissues and organs. Comparing the mechanisms of action reveals that these metals induce toxicity via similar pathways, including the production of reactive oxygen species, the inactivation of enzymes, and oxidative stress. The conventional techniques employed for the elimination of HMs are deemed inadequate when the HM concentration is less than 100 mg/L. In addition, these methods exhibit certain limitations, including the production of secondary pollutants, a high demand for energy and chemicals, and reduced cost-effectiveness. As a result, the employment of microbial bioremediation for the purpose of HM detoxification has emerged as a viable solution, given that microorganisms, including fungi and bacteria, exhibit superior biosorption and bio-accumulation capabilities. This review deals with HM uptake and toxicity mechanisms associated with HMs, and will increase our knowledge on their toxic effects on the body organs, leading to better management of metal poisoning. This review aims to enhance comprehension and offer sources for the judicious selection of microbial remediation technology for the detoxification of HMs. Microbial-based solutions that are sustainable could potentially offer crucial and cost-effective methods for reducing the toxicity of HMs.

Keywords: heavy metals; bioremediation; heavy metals toxicity; biosorption; biomineralization; biotransformation

1. Introduction

Heavy metals (HMs) are defined as metallic elements with a greater density than water [1]. HMs are naturally occurring elements with a large atomic weight and at least five times the density of water [2]. Assuming that heaviness and toxicity are intertwined, HMs also include metalloids, such as arsenic (As), that can induce toxicity at low exposure levels [3]. Various heavy metals, such as chromium (Cr), cadmium (Cd), nickel (Ni), copper (Cu), zinc (Zn), lead (Pb), mercury (Hg), and As, are recognized as biologically dispensable and detrimental to the aquatic ecosystem [4]. Anthropogenic sources, including but not limited to the nonferrous metallurgical industry, mining, mineral processing, electroplating, leather tanning, and chemical industries, are commonly attributed to the discharge of hazardous metals such as HMs [5]. The level of toxicity exhibited by these hazardous materials is contingent upon the amount and length of time that living organisms are exposed to them. Figure 1 depicts the origins and impacts of HMs on humans through the food chain. Prolonged exposure to HMs through dermal contact, ingestion of contaminated food, and inhalation can result in the development of diverse disorders in both humans and other organisms [6]. In addition, the issue of soils contaminated with HM has become an increasingly significant environmental concern worldwide [7].

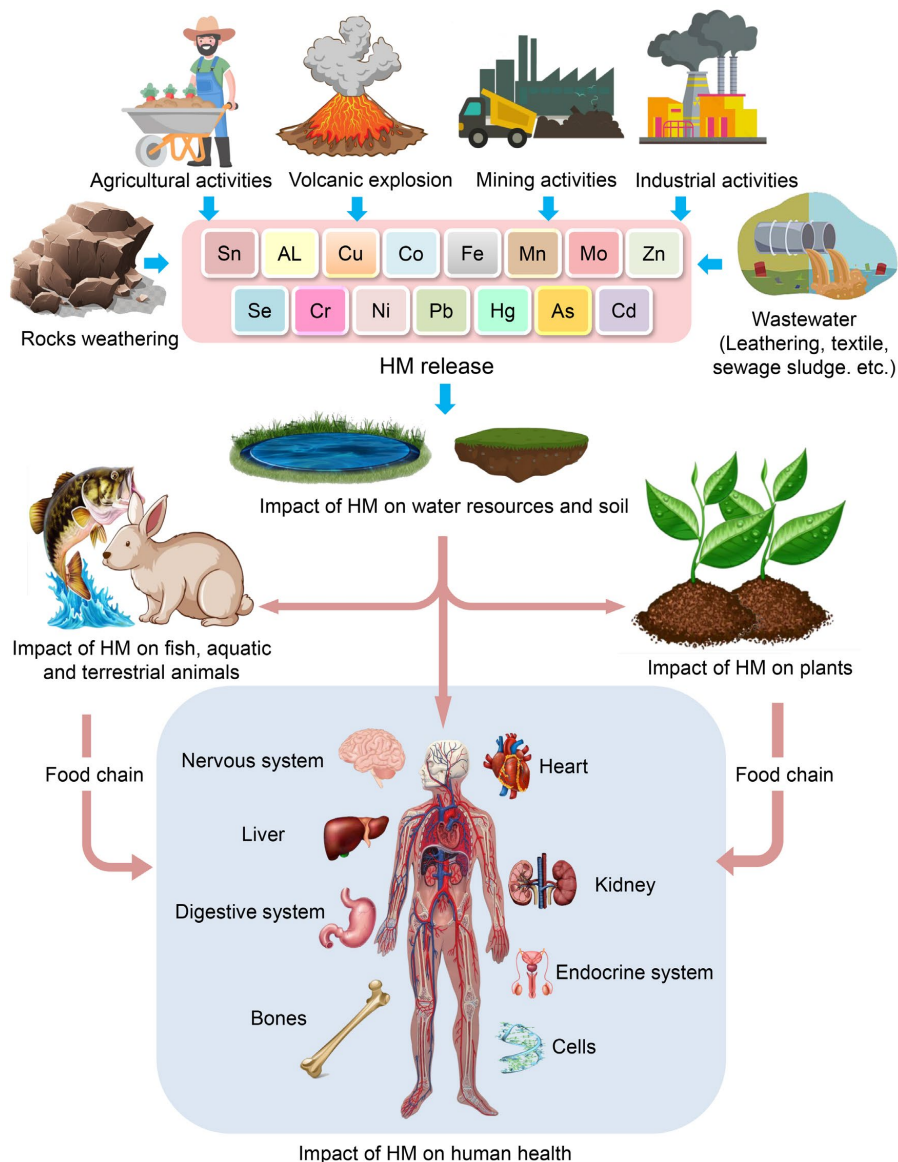


Figure 1. Origins and impacts of HMs on humans through the food chain.

The presence of HMs in composting materials poses a considerable obstacle to their utilization. Consequently, there has been a significant focus on mitigating the toxicity of HMs during the composting process. The composting of HMs primarily impacts microorganisms, and the toxicity levels of different HMs exhibit variability [8]. Various techniques, including chemical, physical, or a combination of both, can be employed to degrade HMs. A considerable number of these techniques are neither ecologically sustainable nor financially viable. Microbial remediation is a viable alternative technique due to the efficient adaptability of microorganisms to their environment [9–14]. The remediation of HMs through microbial bioremediation involves various mechanisms, including bioaugmentation, biosorption, biomineralization, and biotransformation [15]. The implementation of various techniques such as remediation, microbial fuel cells, biofilm, nanomaterials, cell immobilization, and genetic engineering have shown great potential in the removal of HMs. The aim of this review is to provide updated knowledge regarding the abundance of HMs and their potential toxicity on living organisms and plants, as well as their negative impact on human health. It additionally offers mechanistic insights and highlights research uncertainties pertaining to HM remediation via biological and green remediation approaches.

2. Impact of HMs on Human Health

The exposure of humans to HMs is a result of industrial activities and can manifest through various means, such as ingestion, inhalation, and dermal absorption [16]. HMs are categorized based on their function in biological systems as either essential or non-essential HMs. The presence of excessive amounts of essential or non-essential metals can lead to physiological or morphological disorders or genetic mutations [17]. Table 1 categorizes HMs according to their adverse impacts on the human body.

Table 1. Classification of HMs based on their impact on the human body.

HM Compounds	Impact HM on Human
Lead (Pb), mercury (Hg), arsenic (As), and cadmium (Cd)	High degree of toxicity, carcinogenic, and induce damage to multiple organs
Tin (Sn) and aluminum (Al)	Less toxic to human body
Barium (Ba) and lithium (Li)	Non-essential for physiological and biochemical functions
Copper (Cu), zinc (Zn), nickel (Ni), manganese (Mn), molybdenum (Mo), cobalt (Co), selenium (Se), chromium (Cr), and iron (Fe)	Essential for physiological and biochemical functions

The presence of HMs in the ecosystem is attributed to anthropogenic activities, which are the primary sources of human exposure to these substances [18]. HMs are found in the Earth's crust in the form of ores, which are extracted as minerals during mining operations. The majority of ores contain HMs, including but not limited to As, Fe, Pb, Zn, gold (Au), Ni, silver (Ag), and Co, in the form of sulfides. Conversely, some ores contain metals such as Mn, Al, and Se in the form of oxides. In the course of mining operations, the extraction process results in the discharge of HMs from the ore, which are subsequently dispersed into the surrounding environment. These metals remain in the soil and may be transported to other locations via air and water currents. Moreover, the utilization of HMs in industrial applications results in the discharge of certain elements into the atmosphere through combustion or into soil and water systems as effluents. Additionally, industrial commodities such as paints, cosmetics, pesticides, and herbicides can also function as origins of heavy metals (Figure 1).

HMs have the potential to be conveyed to various locations on soil and water bodies via erosion, run-off, or acid rain. The diagram presented in Figure 2 illustrates the trajectory of HM origins and the subsequent impact on human exposure. The toxicity of HMs to biological systems is caused by their ability to form bonds with sulfhydryl groups and generate reactive oxygen species (ROS). The aforementioned phenomenon results in the deactivation of crucial macromolecules, leading to the manifestation of oxidative stress and

depletion of glutathione. Upon exposure to toxic metals and subsequent entry into the body, a multitude of processes occur, including the potential interaction or inhibition of certain metabolic pathways [19]. Consequently, a plethora of deleterious impacts on both human and animal populations are observed. The aforementioned conditions encompass a range of medical issues such as organ dysfunction, metabolic irregularities, hormonal imbalances, congenital anomalies, compromised immune system function, and malignancy [20,21]. Consequently, numerous international organizations establish regulations pertaining to the existence of metals in the environment, foods, and drinking water. The subsequent section delineates the origin and toxicity of distinct HMs on human health.

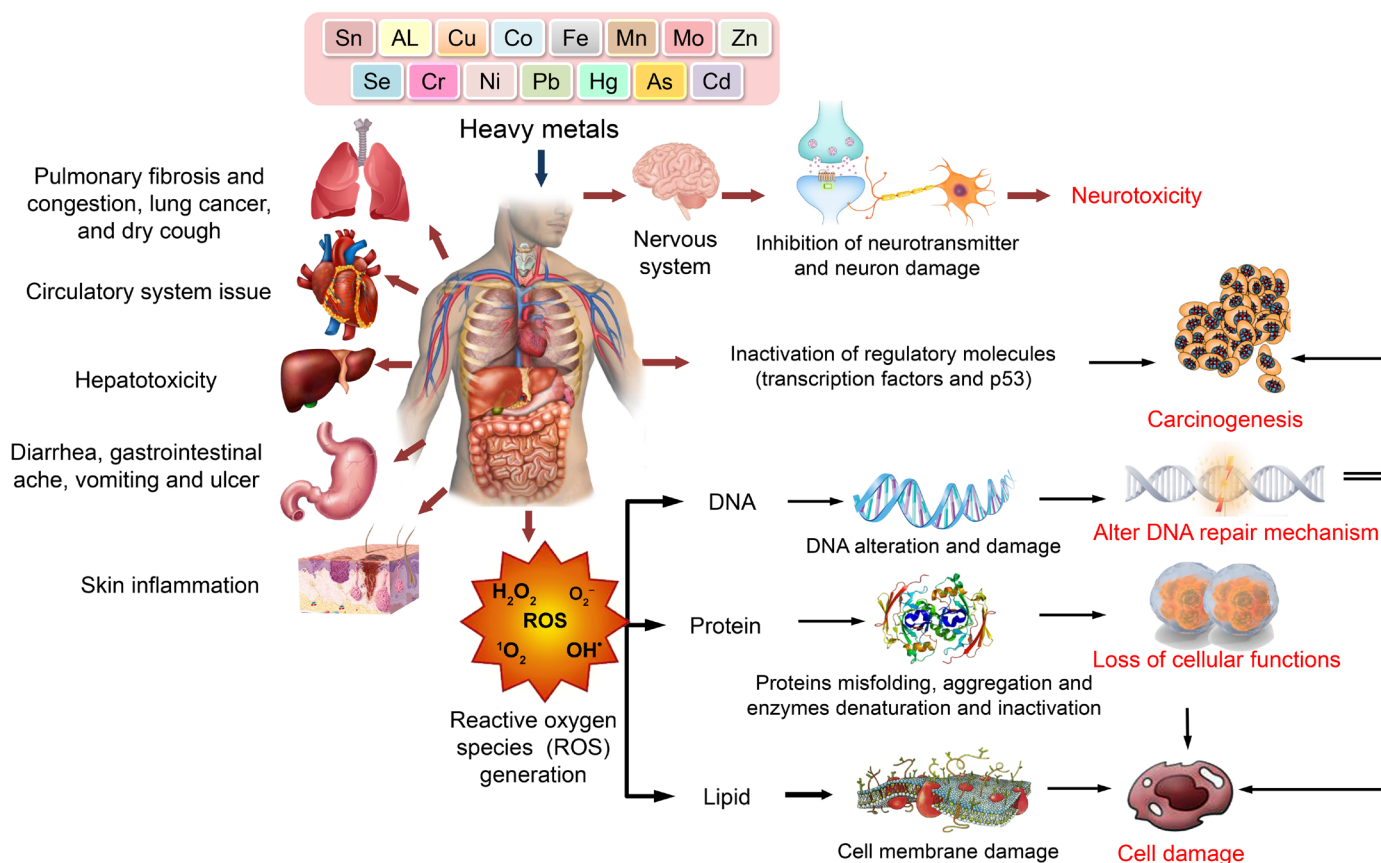


Figure 2. Oxidative stress and human organ toxicity following exposure to HMs.

2.1. Toxicity in Humans following Exposure to HMs

2.1.1. Effect of Arsenic

Arsenic (As) is a hazardous metallic element that can be found in various geological formations, as well as in water and air. As exhibits carcinogenic properties in humans and is associated with various detrimental health outcomes in both the short and prolonged periods [22]. The process of As biotransformation within the human body involves the methylation of various As compounds, ultimately resulting in the production of detrimental end metabolites such as mono-methylmalonic acid (MMA) and di-methyl arsenic acid (DMA). The thiol group of cells is impacted, leading to impaired functionality in cell enzymes, respiration, and mitotic division [8]. A correlation has been observed between exposure to As and the incidence of cardiovascular disease, including hypertension. The impact of exposure to As and its metabolites, namely MMA and DMA, on hypertension during pregnancy has been reported [23]. The findings revealed that pregnant women with low concentrations of DMA experienced an increase in systolic, diastolic, and main arterial pressure, which could potentially result in adverse cardiovascular health outcomes for both the mother and the child [23].

2.1.2. Effect of Mercury

Mercury (Hg) is a hazardous HM that is ubiquitous in the environment. It has the potential to undergo methylation, leading to the formation of methylmercury (MeHg), which can accumulate in the food web. The consumption of seafood has been linked to human exposure to Hg [24]. Lipoperoxidation and nitrite concentration were found to be elevated while total antioxidant capacity was reduced as a result of the presence of Hg in the forms of HgII and MeHg [25]. Multiple studies have demonstrated that exposure to Hg can have significant adverse effects on children. Specifically, prenatal exposure to Hg has been found to negatively impact child growth, which may be attributed to a decrease in parasympathetic modulation of cardiac autonomic function in children [26]. A positive correlation has been reported between exposure to Hg and blood pressure measures during childhood [27]. There is also a positive association between high levels of Hg and dyslipidemia in adults [28]. Several studies on the effect of Hg on liver function found that liver enzymes increased significantly with Hg exposure [29].

2.1.3. Effect of Lead

Lead (Pb) is a toxic HM that has the tendency to accumulate in various tissues including blood, bones, and most body organs [30]. Exposure to Pb has been associated with various neurological disorders such as Alzheimer's disease, Parkinson's disease, amyotrophic lateral sclerosis, and attention-deficit/hyperactivity disorder [31]. There is an observed association between Pb and cardiovascular diseases, and it is regarded as a risk factor for vascular complications in individuals with diabetes [32]. The impact of Pb exposure on liver and kidney functions, as well as on white blood cell count, serum urea, creatinine, aspartate transaminase, alanine transaminase, hemoglobin, and hematocrit concentrations, was observed to be more pronounced in the high blood lead level group as compared to the low blood lead level group [33]. Numerous studies have extensively examined the impact of Pb on children. The findings of these studies have demonstrated that exposure to Pb has adverse effects on the physical growth of children, particularly boys [34]. Exposure to Pb has the potential to alter the concentration of sex hormones, thereby impacting the functioning of the reproductive system [35].

2.1.4. Effect of Chromium

The accumulation of chromium (Cr) in body organs has been found to have potential implications for human health [36]. Cr has a deleterious impact on bronchial epithelial cells, potentially through the aberrant modulation of apoptosis-related proteins, cytoskeletal proteins, and energy metabolism-associated proteins [37]. The impact of Cr on fetal growth during pregnancy has been investigated [38]. The findings of the study suggest that Cr may have a toxic effect on fetal growth. Cr is regarded as a carcinogen that has been linked to the development of lung cancer [39]. The manifestation of hyperpigmented skin is a consequence of exposure to elevated levels of Cr [40].

2.1.5. Effect of Copper

Copper (Cu) is a vital micronutrient for humans. However, excessive levels of Cu can lead to toxic and harmful effects [41]. The accumulation of Cu has been observed to induce mutations and aggregation of mitochondrial protein, leading to a decrease in the activity of the primary antioxidant enzyme and an increase in the production of toxic ROS [42]. A correlation between the elevation of Cu and fibrosis in renal tissues has been established [43]. The presence of Cu has a detrimental impact on male reproductive function, leading to a decrease in sperm count and mobility [44]. There exists a correlation between exposure to copper and obesity [45]. The presence of Cu has an impact on the maintenance of homeostasis of crucial elements such as Ca, Fe, and Mn, which are linked to the induction of oxidative stress. This, in turn, can result in the development of neurodegenerative disorders [46].

2.1.6. Effect of Nickel

Nickel (Ni) is ubiquitously distributed in various environmental compartments such as air, water, and soil [47]. The adverse impacts of Ni on humans have been explored, particularly in the context of pregnancy. The findings revealed a positive association between Ni exposure in pregnant women and preterm delivery [48]. Individuals diagnosed with diabetes exhibited elevated concentrations of Ni in their urine [49]. Offspring may experience congenital heart defects that are associated with maternal exposure to Ni [50]. Ni has been found to have a correlation with immunological disorders, as well as the occurrence of Type I hypersensitivity and Type IV immune reactions in individuals experiencing chronic systemic symptoms associated with Ni [51].

2.1.7. Effect of Uranium

Recent ecological research has examined the potential correlation between prolonged consumption of uranium (U) through drinking water and an elevated risk of leukemia as well as kidney, lung, and colorectal cancer in both genders [52]. It has been evidenced that the absorption of U leads to an increase in ROS, DNA strand breaks, and alterations in gene expression, all of which have adverse clinical consequences [53]. The hexavalent uranyl ion (UO_2^{2+}) has been observed to accumulate in kidney and bone tissues. This accumulation has been found to cause acute and chronic damage to the kidneys, while also increasing the likelihood of osteosarcoma and ontogenesis [54]. Exposure to U either accidentally or through contamination of food or water can lead to a decline in hematopoietic function or bone marrow illness, which can subsequently result in a range of systemic effects [55].

2.1.8. Effect of Cadmium

Cadmium (Cd) is a toxic HM that has detrimental effects on human health [47]. The impact of Cd on the vascular endothelium has been observed to stimulate the release of antithrombotic substances, leading to the production of various inflammatory indicators [56]. Vijayakumar et al. [57] have conducted a study to ascertain the biological effects of Cd on the proliferation and metastasis of prostate cancer and basal breast tumors. Breast tumor cells may experience a reduction in their anti-oxidative defenses, leading to the initiation of ROS formation upon exposure to Cd [58]. Furthermore, the proximal tubule is the site of Cd accumulation and nephron destruction [59].

2.1.9. Effect of Iron

Iron (Fe) is an essential element for various biological processes, including but not limited to DNA replication, mitochondrial respiration, and oxygen transport, that are crucial for the survival of almost all living organisms. Fe is a redox-active element that plays a role in the generation of ROS, which can cause damage to cellular membranes, DNA, and proteins [60]. Fe is an indispensable constituent of human physiology and participates in numerous cellular metabolic pathways, such as the conveyance of oxygen [61]. Fe deficiency anemia is the most commonly occurring form of anemia worldwide [62]. Fe deficiency can have adverse effects on the development and functioning of the immune system [63]. Fe deficiency during pregnancy can pose risks to both the mother and fetus [64]. In contrast, an overabundance of iron has been associated with heightened susceptibility to cardiovascular disease, gestational diabetes, and fetal complications, as well as the generation of oxidative stress and cellular harm [65].

2.1.10. Effect of Vanadium

Vanadium (V) is present in various environmental compartments such as soil, water, and air. Various tissues and organs, such as the kidney, central nervous system, lung, lymphoid organs, and immune system, undergo histological and physiological changes as a result of this HM [66]. Exposure to V has been associated with various adverse health effects, including respiratory dysfunction, kidney toxicity, biochemical and hematologic changes, toxicity for both reproduction and development, mutagenicity, neurotoxicity, and

immunotoxicity [67]. V has an impact on the digestive, respiratory, and cardiovascular systems [68]. The condition known as olfactory dysfunction is the result of exposure to low concentrations of V through the intranasal route. This condition is marked by a decrease in the volume of the olfactory bulb and a decline in dopaminergic neurotransmission to the olfactory bulb [69].

2.1.11. Effect of Cobalt

Cobalt (Co) is a mineral that occurs naturally and can be detected in over a hundred organic and inorganic compounds [70]. The exposure to Co has been associated with inflammations of the upper respiratory tract, including rhinitis and bronchitis, as well as respiratory ailments affecting the lower respiratory system. Simultaneous exposure to certain substances can result in the development of fibrotic alterations in pulmonary tissue, which can subsequently lead to the onset of asthma [71]. Exposure to CO may result in various adverse effects, including lung fibrosis, hepatotoxicity, and carcinogenesis [72]. Exposure to Co can result in various neural system disorders such as memory loss, neuropathies, optic atrophy, and bilateral nerve deafness [73]. The impact of Co on the heart can result in instances of single cardiomyopathy, hypertension, and reversible electrocardiographic alterations [74].

2.1.12. Effect of Thallium

Thallium (Tl) exhibits greater toxicity than Hg, Cd, and Pb [75]. Chronic Tl poisoning, characterized by symptoms such as anorexia and headaches, can result from prolonged exposure to low levels of Tl [76]. Respiratory muscle paralysis may lead to a state of coma in severe instances [77]. One of the characteristic indications of Tl poisoning is the occurrence of hair loss subsequent to the contraction of hair follicles. Additional symptoms encompass issues related to digestion, discomfort, mental health, and cardiovascular system complications [78]. The incidence of fetal mortality, preterm birth or low birth weight has been found to be associated with Tl toxicity during pregnancy [79].

2.2. Mechanism of Intoxication following Exposure to HMs

Upon ingestion via food or water, HMs undergo acidification within the stomach's acidic environment. Under acidic conditions, the aforementioned elements undergo oxidation and attain their respective oxidative states (Zn^{2+} , As^{3+} , Cd^{2+} , As^{2+} , and Pb^{2+}). These states can effectively interact with biological molecules, including proteins and enzymes, to form robust and enduring bonds [80]. The aggregation of proteins may be induced by HM, as evidenced by the observed As-induced protein aggregation, which was found to be dependent on concentration. Furthermore, the aggregates exhibited a diverse array of proteins that were notably enriched in functionalities pertaining to metabolic processes, protein folding, protein synthesis, and stabilization [81]. The ability of these agents to induce protein aggregation in vivo is likely contingent upon their cellular uptake/export efficiency and their unique biological mechanisms. Figure 3 depicts the diverse mechanisms involved in HM intoxication.

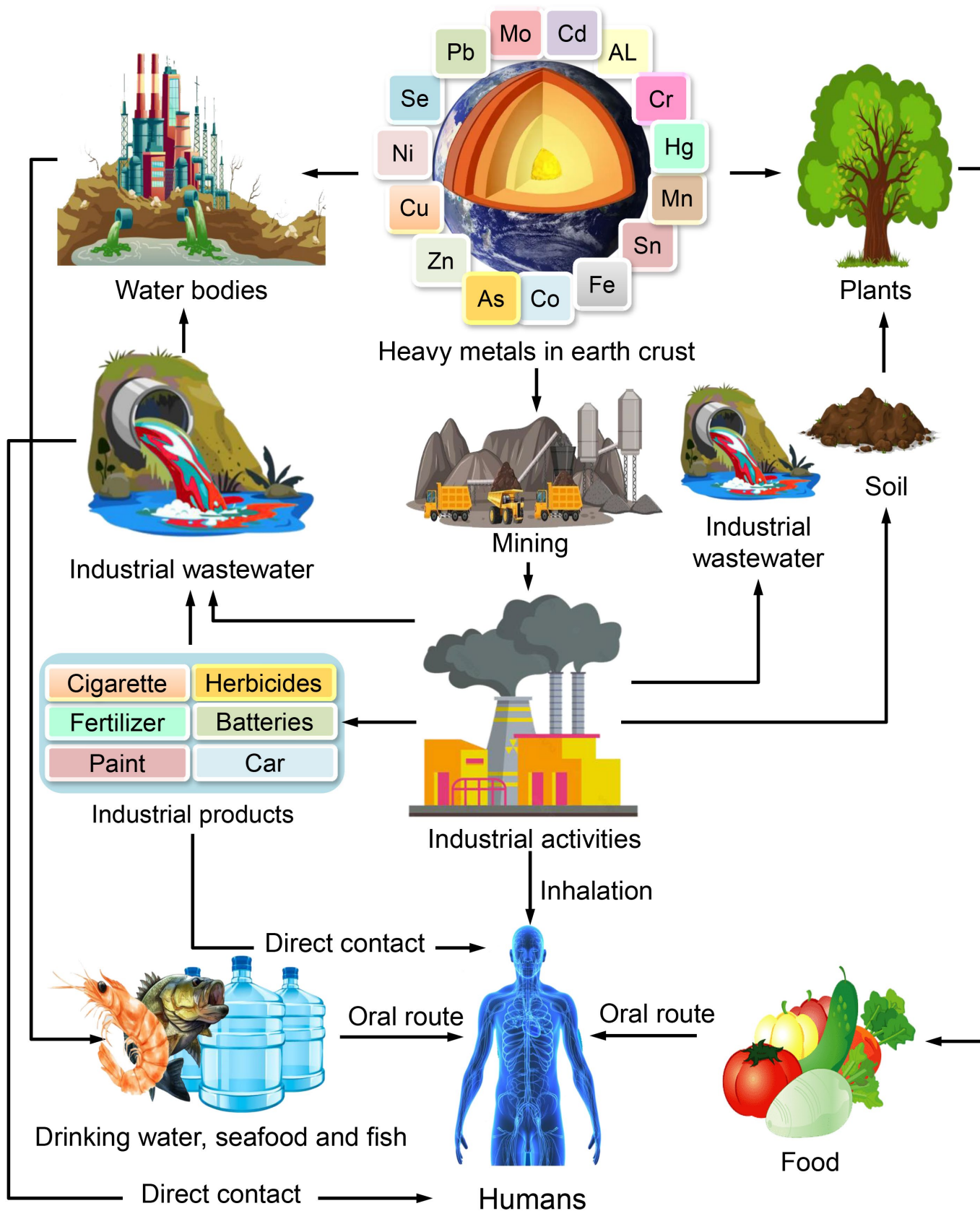


Figure 3. Intoxication mechanisms in humans following exposure to HMs.

3. Impact of HMs on Aquatic Animals

HMs are naturally occurring elements that are also generated through human activities, such as industrialization and agriculture. These HMs can enter aquatic ecosystems through direct and indirect means. The transportation of HMs occurs in tandem with the water cycle, whereby they are conveyed from upstream to downstream along river courses, ultimately depositing in vast water bodies such as large lakes, rivers, or oceans [82]. Whilst HMs have the capability to enter water directly, a considerable quantity of HMs are assimilated into the ground and sediment, which function as a conveyance or prospective origin of HMs in aquatic environments [83]. It has been observed that sediment-bound HMs have the potential to migrate into water, and the rate of their release is influenced by various factors such as the type of HMs, as well as the physico-chemical properties of both the water and sediment, including pH, salinity, specific surface area, and cation exchange capacity [83]. Aquatic invertebrates can ingest and accumulate HMs in lakes, rivers, and marine environments through either water or food pathways. HMs have the potential to undergo transfer from aquatic invertebrates to higher trophic levels, ultimately leading to biomagnification. This process can result in harmful consequences for all organisms, including humans. The ingestion of various HMs by aquatic invertebrates has the potential to impact aquatic ecosystems, as depicted in Figure 4.

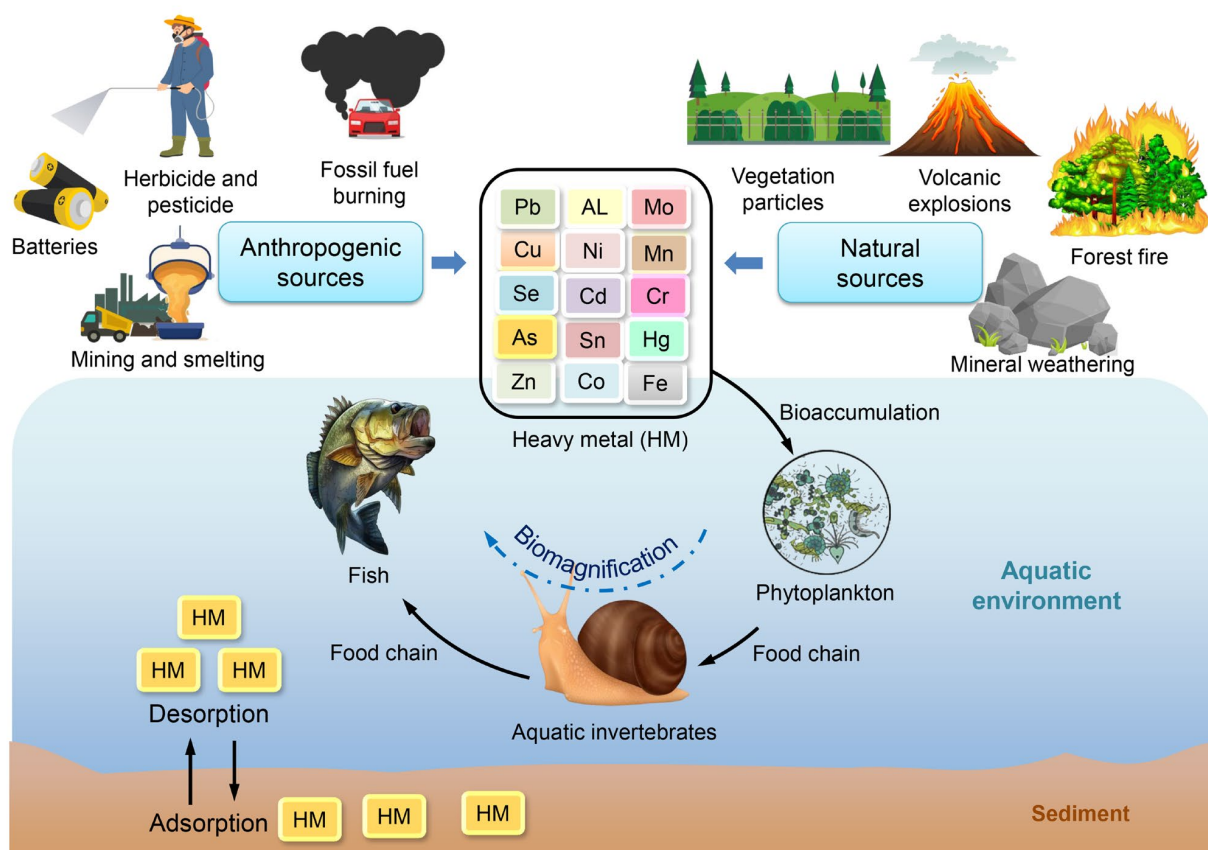


Figure 4. Distribution of HMs in aquatic environments.

The biological properties of pure elements and their compounds may exhibit differences in toxicity when considering metals, owing to variations in solubility, oxidation state, and bioavailability. The presence of Cr has been found to impede the growth and reproductive capabilities of aquatic invertebrates, induce oxidative stress and DNA damage, and potentially result in teratogenic and carcinogenic effects [84]. Cu has been found to have various detrimental impacts on aquatic organisms, including but not limited to inducing oxidative stress, modifying enzyme activity, impeding growth or reproduction, interfering

with the endocrine system, and diminishing energy acquisition [85]. The exposure of aquatic invertebrates to Zn has the potential to induce oxidative stress and alter various physiological processes, including immune responses, metabolism, and detoxification [86]. As has the potential to cause various physiological and biochemical toxic effects, including but not limited to growth inhibition, oxidative stress, fecundity reduction, apoptosis, and immunotoxicity [87]. Even at low concentrations, Cd demonstrates deleterious impacts on aquatic invertebrates. The impact of Cd resulted in a reduction in the growth rate and reproductive capacity in freshwater fleas. Additionally, the substance caused neuronal toxic effects in the oyster [88]. The presence of Pb in aquatic ecosystems can result in toxicity, which may manifest as the inhibition of crucial enzymatic activity and the occurrence of oxidative stress-induced harm to cell membranes [89]. Hg exhibits a notable attraction towards the sulfur (-SH) group and can readily attach to the cysteine component found in proteins. This phenomenon leads to functional impairment and toxicity in aquatic invertebrates [90].

4. Impact of HMs on Soil and Plants

The stress induced by HMs is a significant factor that adversely impacts the agricultural yield of plants. HMs are widely acknowledged as a constituent of soil. However, their concentration in soil and plants can have detrimental effects on the environment [91]. HMs have the ability to transfer through the food chain after being absorbed by plants, ultimately accumulating in the bodies of animals and humans and posing potential health risks [92]. HMs have the potential to infiltrate groundwater or surface waters and subsequently undergo absorption by plants or turn into emissions as gases that are sent into the atmosphere. Alternatively, they may form semi-permanent associations with soil constituents such as clay or organic matter, ultimately impacting human health at a later time. It is imperative to keep soil pollution levels below the prescribed toxicity thresholds to ensure sustainability [93]. The utilization of chemical fertilizers and pesticides has been found to elevate the likelihood of soil contamination by HMs, which can subsequently accumulate in the tissue of crops cultivated in such contaminated soil [94]. The mobility of HMs in soil is significantly influenced by the presence of total organic matter and pH levels. Upon introduction of HMs to soil, their downward movement is limited by the soil's capacity to retain the metals or by the interaction of the metals with the adjacent waste matrix, which may enhance their mobility. High concentrations of HMs in soil can have adverse effects on the development, growth, and nitrogen fixation of legumes [95]. Microbial remediation refers to the biological process by which microorganisms adsorb or transform heavy metals into less toxic products. Microbial remediation is a process that primarily involves various mechanisms such as biosorption, bioaccumulation, bioleaching, biovolatilization, and biomineralization [96], as illustrated in Figure 5, for the purpose of addressing HM contamination in soil.

There was a decrease in the concentration of HMs in the soil as the depth increased [97]. The transport of HMs ions is influenced by both their properties and the texture of the soil. The duration required for the infiltration of HMs in sandy and compound soil is comparatively shorter than that in loess. The migration resistance of loess to all metals is significantly higher when compared to sandy and compound soil. HMs are immobilized in the soil through the formation of complexes with ligands, which can be either organic or inorganic in nature. When subjected to leaching conditions, HMs present in soil have the potential to contaminate surface water and groundwater, thereby exacerbating water pollution and adversely impacting aquatic organisms [98].

The detection of Hg in soil solutions poses a challenge owing to its robust affinity with soil particles and organic substances. Inorganic Hg can cause toxicity to humans and disrupt soil biota across all trophic levels [99]. The conversion of Hg^{2+} in soil into methylmercury, a highly toxic substance that accumulates in plants and is subsequently enriched in the food chain, poses a significant risk to human health through chronic poisoning. This process occurs through various mechanisms, including absorption, migration, and

transformation [100]. The presence of Hg in soil has a notable effect on both the nitrogen cycle of the soil and the prevalence of microorganisms inhabiting the soil [99].

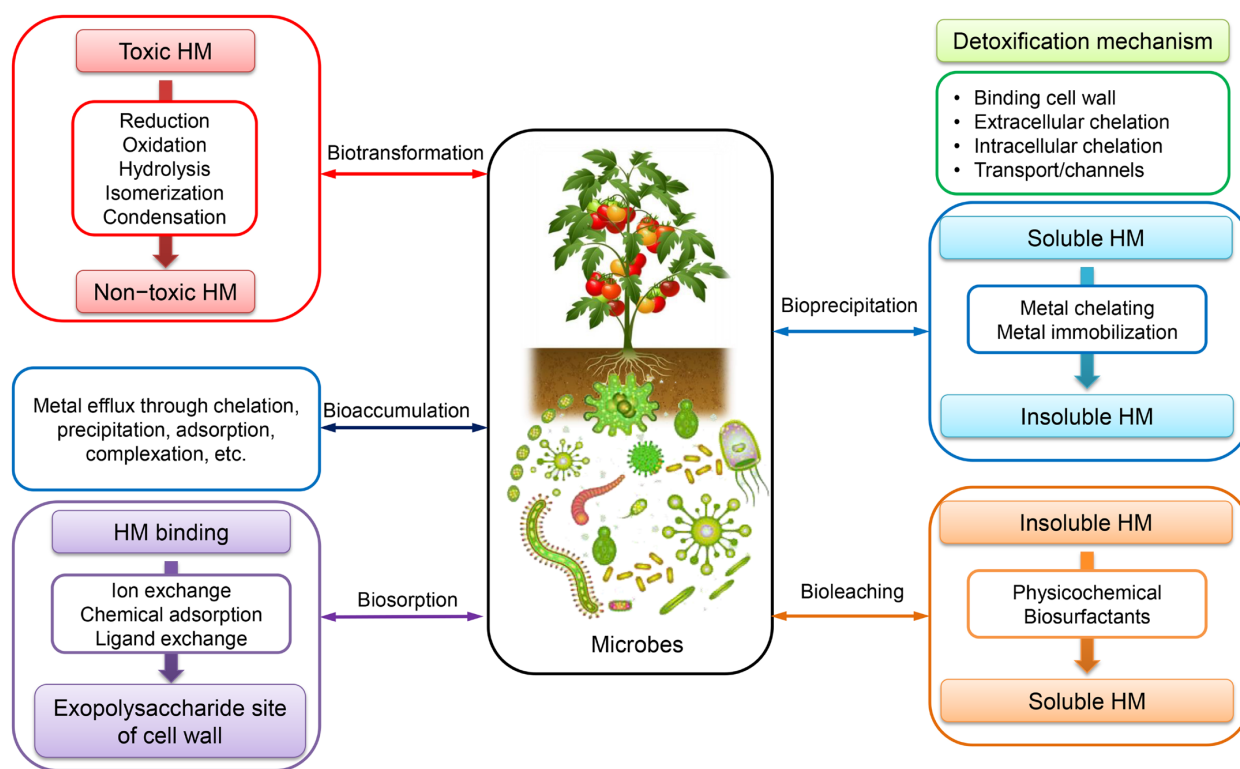


Figure 5. Microbial remediation mechanisms of soils contaminated with HMs.

Elevated concentrations of Cu have deleterious effects on various metabolic pathways in plants, including but not limited to photosynthesis, respiration, and enzymatic activity. Cu has been found to enhance the production of ROS in plants and upregulate the expression of genes associated with oxidative stress response [101]. Elevated concentrations of Cd have a significant impact on the development of soil microorganisms and the activities of enzymes. In contrast to the majority of other metals, it has been observed that Cd exhibits a higher degree of plant absorption and can translocate into the edible portions prior to the onset of phytotoxicity [91].

Cr is a noxious element for plants. Cr is a naturally occurring element that is ubiquitously distributed in the environment, encompassing the soil, air, and water compartments [102]. Cr is present in two oxidation states, namely trivalent and hexavalent, in soil. However, hexavalent Cr is comparatively more toxic and mobile than its trivalent counterpart [103]. The presence of Cr in soil has a negative impact on both soil fertility and microbial activity, ultimately resulting in decreased plant yield [104]. The presence of Cr compounds can have detrimental effects on the growth and development of plants. This can manifest in altered germination processes and hindered development of roots, stems, and leaves, ultimately leading to reduced yield and dry matter production [105]. The elevated concentration of soil Pb has been associated with certain chemical fertilizer production enterprises. Pb has a tendency to accumulate in the roots and lower above-ground portions of plants. While plant roots are not highly receptive to Pb absorption, the presence of this element can negatively impact various metabolic processes in plants, including root development, photosynthesis, and water uptake [93]. The enzymatic activity of soil biota is hindered by Pb, leading to the buildup of partially digested organic matter [106].

Phytoremediation techniques represent an environmentally friendly technology that facilitates the detoxification of polluted metal-lands, water, and groundwater through the implementation of various processes such as phytoextraction, phytostabilization, phyto-

volatilization, rhizofiltration, and phytofiltration [107]. The utilization of plants for the remediation of contaminated sites, known as phytoremediation, is a sustainable approach that has been extensively investigated in both field and laboratory settings. HMs exhibit resistance to remediation through both microbial and physiological mechanisms, and they persist in soil for extended periods, thereby posing a potential threat to the ecosystem [108]. It is imperative to implement remedial measures to mitigate the consumption of metals by the atmospheric, terrestrial, and aquatic ecosystems, and to curtail the contamination of groundwater [109]. Numerous phytoremediation techniques entail the utilization of rhizodegradation and phytodegradation mechanisms to facilitate the decomposition of both organic and inorganic pollutants. The diagram presented in Figure 6 illustrates the implementation of phytoremediation as a means of addressing the issue of HMs in plants.

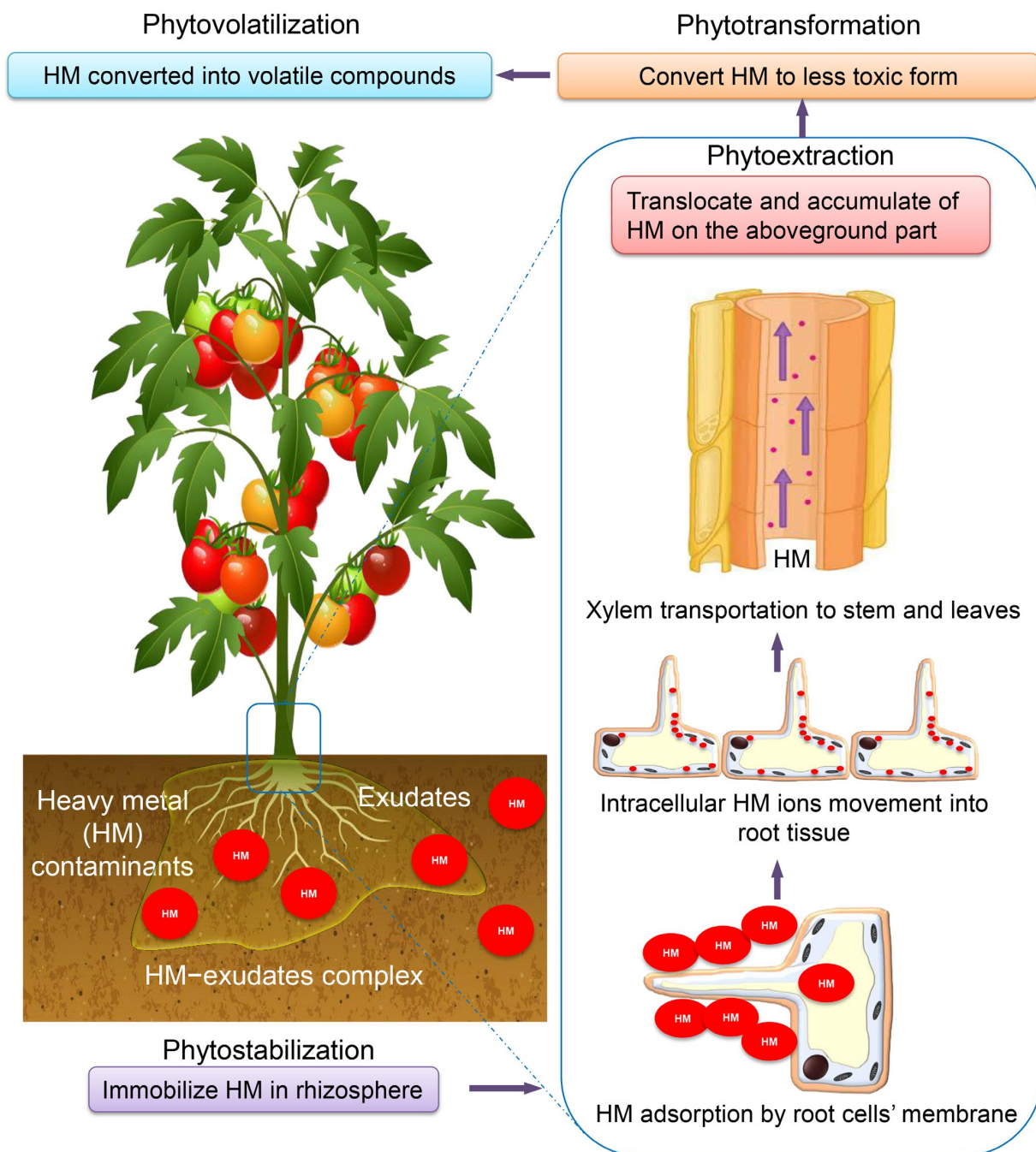


Figure 6. Transfer route and phytoremediation mechanism of HMs into plant cells and tissues.

5. Toxic Effect of HMs on Microbial Cells

The toxicity of HMs is attributed to multiple mechanisms, which include degradation of enzymatic functions leading to lethality, functioning as redox catalysts in the generation of ROS, disruption of ion regulation, and direct effects on the composition of DNA and proteins. Cu has the ability to catalyze the production of ROS, thereby serving as soluble electron carriers. The aforementioned phenomenon has the potential to induce significant harm to cytoplasmic molecules, DNA, lipids, and other proteins [110]. HMs have an impact on the rate of enzyme activity, which is influenced by various soil factors such as pH, organic content, and chemical composition. The negative effects of toxic metal-contaminated soil can be attributed to the decrease in urease, catalase, and lipolytic activity. There exists an inverse correlation between the concentration of HMs and soil enzyme activity. A decrease in bioavailability of Cd, Cu, and Pb in treated soil results in an increase in soil enzyme activity [111]. Figure 7 illustrates the adverse impacts of HMs on microbial cells. Cr has the potential to induce oxidative harm and denaturation of microorganisms, thereby impeding their bioremediation efficacy. The electric interaction between the negatively charged phosphate groups of DNA and intracellular cationic Cr complexes could potentially affect transcription and replication processes, leading to mutagenesis [110]. Both Cd and Pb exhibit deleterious impacts on microorganisms, causing harm to their cell membranes and disrupting the integrity of their DNA structure. The aforementioned impairment arises due to the dislodgment of metallic elements from their initial binding locations or bonding interactions [112]. The manipulation of DNA structure can have an impact on microbial morphology, metabolism, and growth by inducing functional disruption, cell membrane disruption, inhibition of enzyme activity, and oxidative phosphorylation [113].

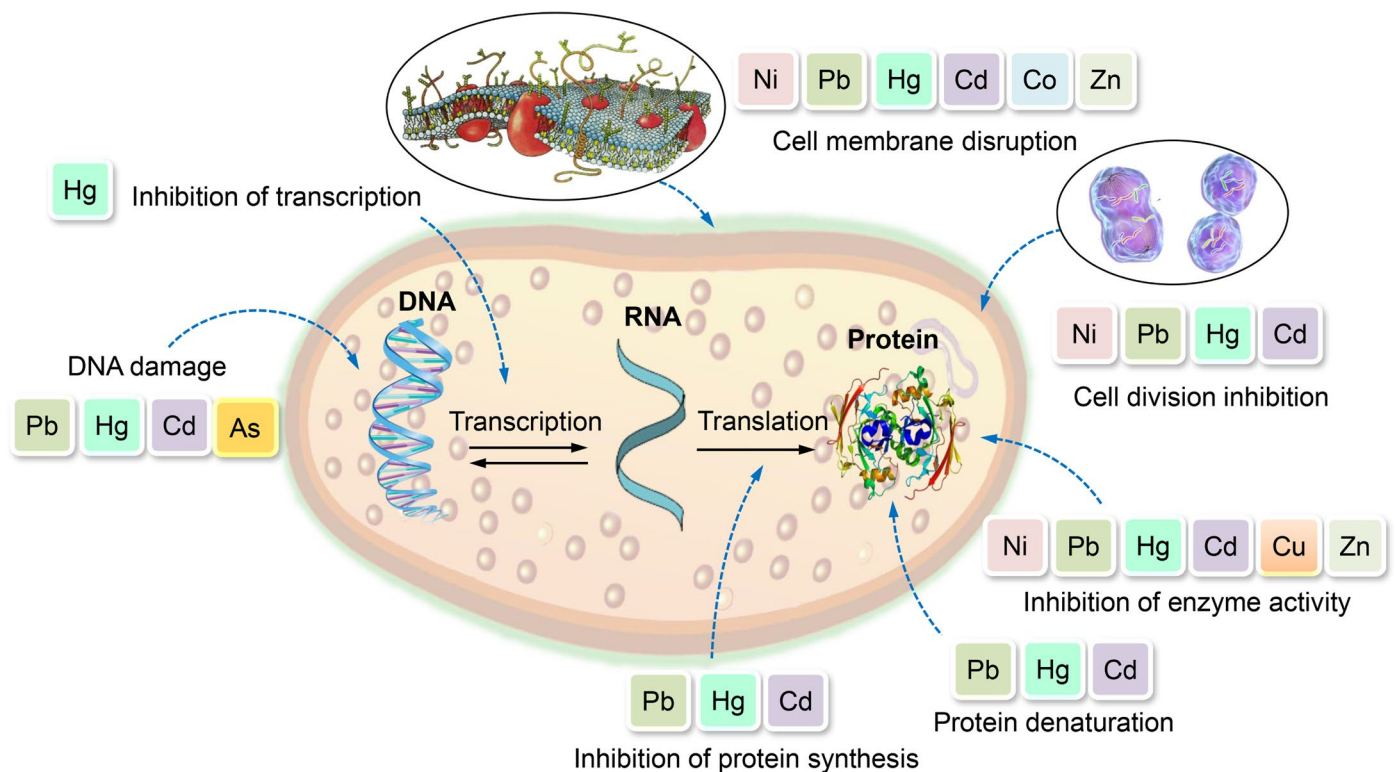


Figure 7. Toxicity of HMs on microorganisms.

6. Methods for the Remediation of HMs

Removal of HMs is typically accomplished by physical remediation, chemical remediation, and bioremediation [114–119]. The advantages and disadvantages of remediation technologies are summarized in Table 2.

Table 2. Remediation approaches of HMs.

Remediation Type	Approach	HM	Advantages	Disadvantages	References
Physical	Replacement	Ni, Cd, Cr, Zn, Cu, Pb	Appropriate for smaller contaminated sites	Expensive	[114]
	Thermal desorption	Hg	<ul style="list-style-type: none"> • Effective • Safe • Lack of secondary pollutants • Short pretreatment period 	<ul style="list-style-type: none"> • Expensive devices • Long desorption time 	[115]
Chemical	Oxidation/reduction	Cu, As, Sb, Cr, Pb, Se	Suitable for soils that are heavily polluted with HMs	<ul style="list-style-type: none"> • Expensive • Need large quantity of chemicals 	[116]
	Leaching	Cr, Zn, Cu, Pb, Ni, Cd	Suitable for soils that are heavily polluted with HMs	Formation of secondary pollutants	[117]
	Electrokinetic remediation-permeable reactive barrier	Cd, Pb, Cr, Ni	Suitable for soils that are heavily polluted with HMs	Time-consuming	[118]
	Stabilization	Pb, Cd, Cu, Mn, Zn, As, Fe, Ni	<ul style="list-style-type: none"> • Effective • Low cost • Convenient 	Pollutants cannot be removed thoroughly	[118]
Bioremediation	Microbial remediation	Cr, Pb, Ni, Zn, As,	<ul style="list-style-type: none"> • Effective • Low cost • Lack of secondary pollutants 	<ul style="list-style-type: none"> • Concentration limit • Time-consuming 	[119]
	Phytoremediation	Se, Cd, Cu, Pb, Zn	<ul style="list-style-type: none"> • Convenient • Eco-friendly • Low cost 	<ul style="list-style-type: none"> • Time-consuming • Non-specific • Low efficiency 	[119]

7. Bioremediation: Interaction of Microorganisms with HMs

Bioremediation is an innovative technology that employs a variety of biological agents, including bacteria, fungi, algae, yeasts, moulds, and plants, to effectively eliminate, detoxify, transform, or neutralise the adverse effects of HMs [120]. In contrast to several physico-chemical techniques commonly employed for mitigating HM pollution, bioremediation presents a variety of economically viable advantages owing to its notable efficacy in HM removal, cost-efficiency, ease of management, and widespread availability in both contaminated soil and water [121]. Microorganisms are a significant factor in the bioremediation of HMs among biological agents. These organisms are not only proficient in the dissolution of HMs, but also actively participate in the oxidation and reduction of transition HMs. The utilisation of microorganisms' metabolic abilities for the eradication of HM pollution is a form of green technology. Overall, most HMs have been classified as toxic. However, various biological organisms have developed distinct resistance mechanisms and intricate intracellular pathways to facilitate the utilization, interaction, adaptation, and detoxification of HMs for the purpose of cellular regeneration [122]. Figure 8 illustrates the various mechanisms employed by microorganisms for the elimination of HMs, including detoxification, biosorption, degradation, mineralization, and transformation from highly toxic to low toxic forms [122,123]. Furthermore, it has been suggested that the utilization of fungi and bacteria may have a potential role in the bioremediation of HM, as depicted in Table 3 [124–140].

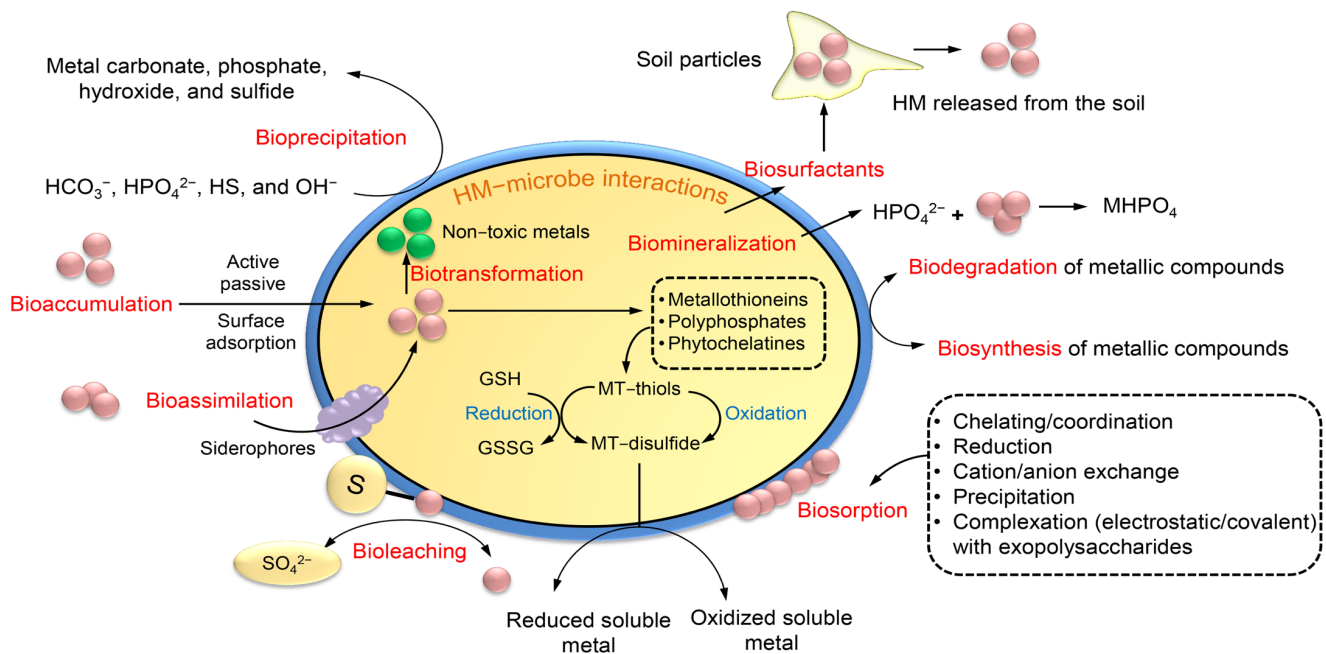


Figure 8. Mechanisms of microbial bioremediation of heavy metals. GSH, glutathione; GSSG, glutathione disulfide; MT, Metallothionein.

Table 3. Efficient bacterial and fungal species used in HM removal.

Microorganisms	HM	Efficiency of Removal (%)	References
<i>Penicillium</i> spp. XK10	Cd	32.2	[124]
<i>Cladosporium</i> sp. XM01	Mn	91.5	[125]
<i>Penicillium janthinillum</i> GXCR	Pb, Fe, Cu	85–99	[126]
<i>Mucor rouxii</i>	Pb, Zn,	11–35	[127]
<i>Aspergillus niger</i>	Cd, Cr	43–98	[128]
<i>Aspergillus flavus</i>	As, Pb, Cr, Ni	46–97	[129]
<i>Trichoderma brevicompactum</i>	Cu, Cr, Cd, Zn	4–64	[130]
<i>Aspergillus fumigatus</i>	Cd, Cr	69–79	[131]
<i>Penicillium simplicissimum</i>	Cr, Pb, Cu, Cd, Zn	28–88	[131]
<i>Trichoderma virens</i>	Cu, Cr	63–70	[132]
<i>Pseudomonas putida</i>	Cu	25.4	[133]
<i>Bacillus subtilis</i>	Cu	37.8	[133]
<i>Pseudomonas aeruginosa</i>	Hg	60	[134]
<i>Sporosarcina pastaurii</i>	Cd, Zn, Pb	94–98	[135]
<i>Variovorax boronicumulans</i>	Cd, Zn, Pb	73–95	[135]
<i>Stenotrophomonas rhizophila</i>	Cd, Zn, Pb	63–96	[135]
<i>Citrobacter freundii</i>	Cd, Cu, Fe, Mn, Zn	40–80	[136]
<i>Pseudomonas aeruginosa</i>	Cu, Zn	18–65	[137]
<i>Acinetobacter</i> sp. B9	Ni	69	[138]
<i>Bacillus thuringiensis</i>	Hg	62	[139]
<i>Streptomyces</i> sp.	Pb	83	[140]

7.1. Mechanisms of HM Removal by Fungi

The most common mechanisms of HM mycoremediation are bioaugmentation, bioaccumulation, biosorption, biomineralization, and biotransformation [41] (Figure 9). The implementation of fungi in bioaugmentation exhibits the most superior performance in terms of both metal elimination and enzymatic activity [141]. Bioaccumulation is a biological phenomenon wherein microorganisms synthesize proteins to recycle HMs for utilization in various cellular processes such as enzyme catalysis, signaling, and stabilization of charges in biomolecules [142]. Fungi have the ability to mitigate the harmful

impact of HMs by generating inorganic acids and organic acid complexes that facilitate the leaching of HMs. This characteristic makes fungi a promising candidate for the sustainable management of wastewater. Certain types of fungi have the ability to synthesize metal transport proteins that facilitate the efflux of heavy metals from the cell or their sequestration into vacuoles [143]. The elevated levels of proteins resulting from the heightened concentration of iron facilitate the synthesis of crucial proteins that participate in the detoxification processes of *Aspergillus flavus*, thereby mitigating the deleterious effects of iron [144].

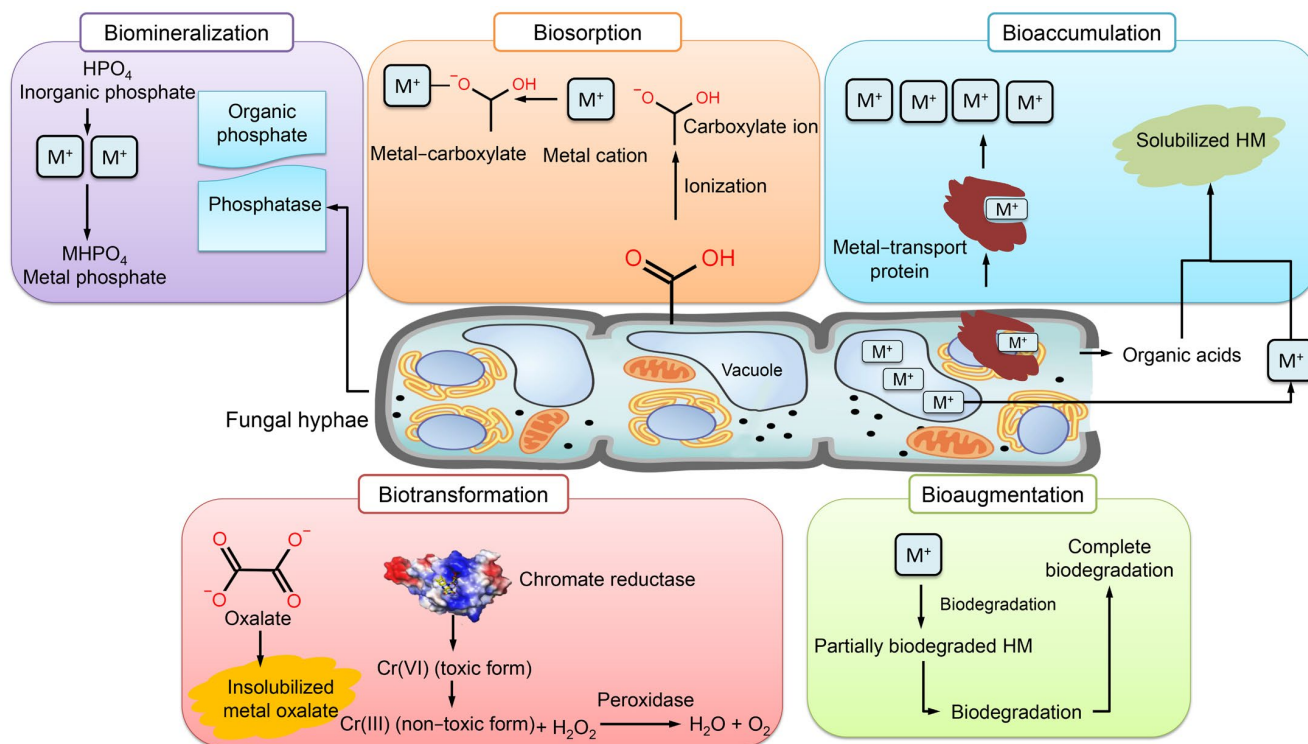


Figure 9. Mechanisms of HM removal by fungi.

Biosorption has emerged as a viable alternative for HM remediation. Fungi are known to primarily employ this technique through the ion exchange mechanism that occurs between their sorption materials or various polymeric substances, such as EPS [145]. The present approach involves the utilization of adsorbents as biological agents that facilitate the binding of HMs to non-living biomass of microorganisms, particularly through the functional groups of cell wall sorption components. The binding of metal ions is contingent upon the pH level, which influences the electrostatic interaction between metal ions and a functional group. At low pH levels, two distinct phenomena take place: firstly, the functional groups carried by the mycelium undergo ionization, resulting in the generation of a negative charge. Secondly, the occurrence of protonation in a solution can be attributed to the presence of free hydrogen ions resulting from an acidic pH. As a result of these two mechanisms, a significant quantity of unbound protons present in the solution adhere to the negatively charged sites on the cellular wall of the fungus [145]. Under optimal pH conditions, the fungus exhibits an elevated binding affinity and establishes electrostatic interactions with positively charged metal ions. The influx of metal complexes into the fungal cell results in a decrease in the quantity of metal ions absorbed from the solution, thereby affecting the fungus’s retention capacity and ultimately leading to a reduction in its biosorption potential [146]. The augmentation of the incubation period results in an elevation of both biomass production and functional groups, leading to a corresponding increase in the quantity of biosorbent metal attached to said groups.

The metal that is biosorbed in the fungal cell wall is bound by metal-chelating proteins, and subsequently either restored in vacuoles or precipitated in the fungal inner cell surface through the formation of intracellular organic acid [41]. The process of metal binding to cell surface binding sites is contingent upon a variety of factors, including the intricate structure of microorganisms, extracellular precipitation, intracellular accumulation, oxidation and reduction, and methylation/demethylation [147]. Fungi possess inner walls with multiple layers on their adsorbent surface, which are accountable for the sorption of metals. As a result of metal adsorption, the surface of fungi appears rough. The fungal surface is comprised of a capsule layer that is characterized by the presence of carboxyl groups. These groups are instrumental in facilitating the mechanism of metal biosorption [148]. Metal cations can effectively bind to the fungal surface with the aid of anionic ligands, including carboxyl and amino groups. Metal ions exhibit an affinity for functional groups containing atoms with electron-donating properties. The carboxyl group of galacturonic acid, which is the primary constituent of fungal capsules, facilitates cation binding by attracting charges. The presence of oxygen in carboxylate ions facilitates anionic binding and enhances the attraction toward metal cations [148].

The process of biomineralization pertains to the capacity of living organisms to form minerals as a means of mitigating the harmful effects of metal toxicity. The biomineralization of metal phosphate is initiated by the production of acid phosphatase by fungi in a medium containing phosphate, leading to the formation of phosphate binders [149]. Certain fungi exhibit uneven diffusion of metals such as Pb^{+2} in their mycelia, resulting in the manifestation of green fluorescence with granular distribution. This phenomenon is attributed to the biomineralization of metals [150]. The concentration of precipitated calcium carbonate increases concomitantly with the rise in calcium oxide levels within the fungal culture inhabiting the contaminated concrete cube. The aforementioned process induces ion activity, thereby facilitating the mineralization of the solution, leading to the substantial precipitation of calcium carbonate. The entrapment of Cr by *Candida orthopsilosis* resulting in the formation of calcium carbonate precipitate has been observed to enhance the removal of HM and facilitate the mineralization of the surfaces of concrete cubes [151].

Biotransformation involves the conversion of toxic metal forms into less toxic forms through oxidation–reduction or mineralization–demineralization reactions [152]. The capacity of fungi to generate oxalate, which combines with HMs to form coordination compounds that function as chelators, enables them to convert soluble metals into insoluble particles, specifically insoluble metal oxalates [145]. A range of fungi have the ability to undergo biotransformation of chromium, arsenic, and other HMs into non-toxic forms. Additionally, these fungi are capable of accumulating the transformed metals in their hyphae, thereby preventing the entry of toxic metals into the roots of plants [153].

7.2. Mechanisms of HM Removal by Bacteria

Biosorption is a biological physicochemical process that is employed for the removal of recalcitrant compounds, including metal ions [112]. The utilization of this approach is characterized by its cost-effectiveness, high efficiency in eliminating toxicity, absence of the requirement for supplementary nutrients, and minimal production of chemical and/or biological sludge [112]. The analysis of the supernatant derived from suspension cultures of bacteria revealed that the primary factor responsible for metal sequestration was the soluble exopolysaccharides (EPSs). EPS is composed of carbohydrates and proteins, which jointly facilitate the biosorption of Zn and Pb [154]. The secretion of EPS has been observed in various bacterial strains, including *Paenibacillus jamilae*, *Bacillus firmus*, *Bacillus licheniformis* KX657843, *Herbaspirillum* sp., and *Paenibacillus peoriae* TS7. The biosorption capacity of the substance in question exhibited notable efficacy in the elimination of various metals, including Zn^{2+} , Pb^{2+} , Ni^{2+} , Cu^{2+} , Cd^{2+} , Co^{2+} , and Hg. This can be attributed to the electrostatic interactions that occur between the negatively charged functional groups of EPS and the positively charged metal ion (Figure 10A) [155]. The biosorption process is primarily influenced by the quantity of biosorbent utilized. Bacterial immobilization

is a biological process that relies on the utilization of bacteria's inherent characteristics to eliminate various pollutants. The fundamental principle of the mechanism involves the interplay between bacteria and HMs, facilitated by the immobilization of the biocatalyst onto diverse materials that serve as carriers, providing safeguarding against external factors. This immobilization enables the microbial cells to be readily available for degradation purposes [156]. Two mechanisms, namely the attachment method and the encapsulation method, can be utilized to elucidate bacterial immobilization mechanisms, as depicted in Figure 10B.

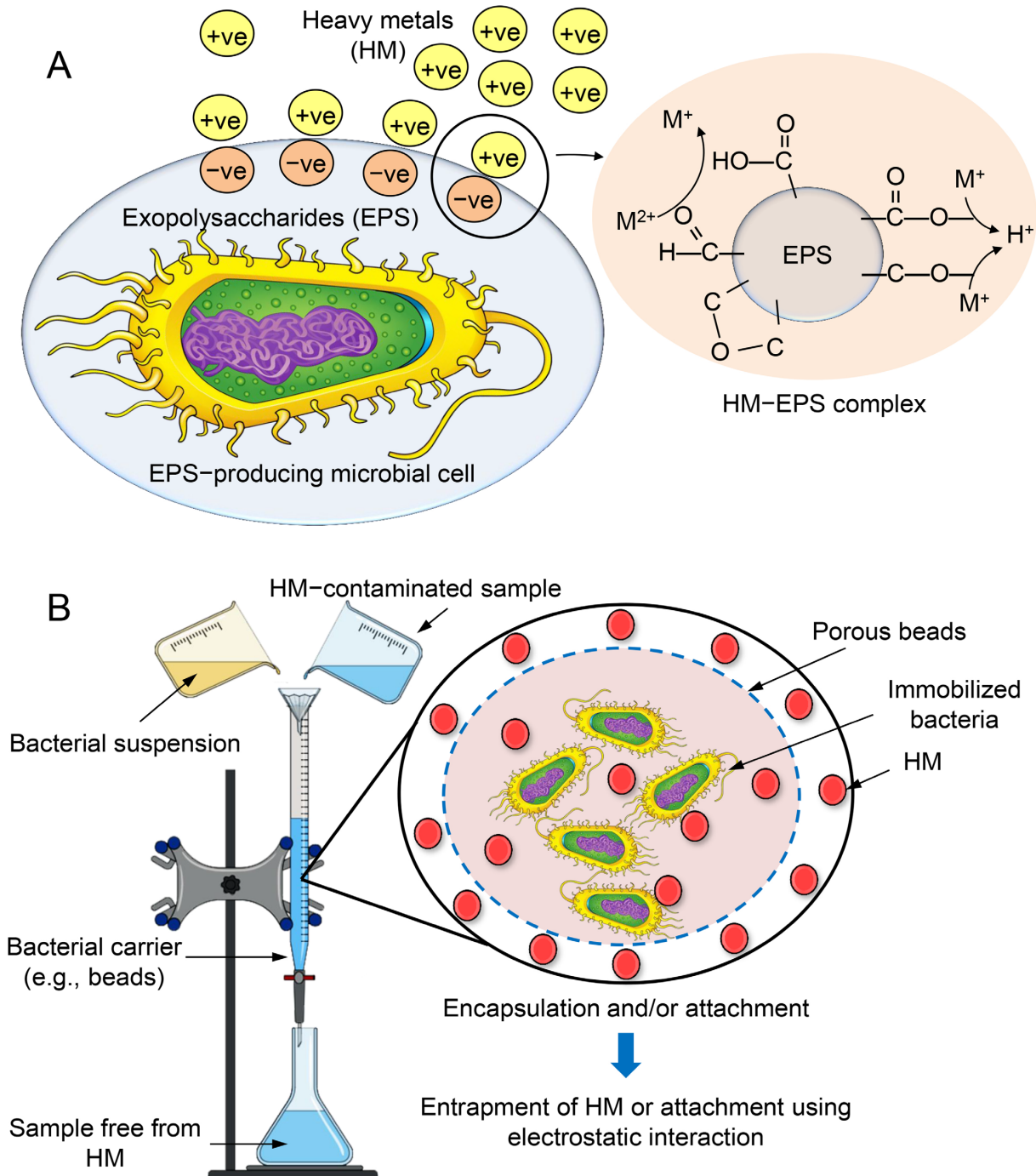


Figure 10. Mechanisms of bacterial exopolysaccharide (A) and encapsulation (B) for HM removal.

Biobleaching is a microbial-mediated process employed for the purpose of solubilizing minerals from waste solids. The process of biobleaching has been found to be both cost-effective and efficient in the removal of a diverse range of HMs, including but not limited to Cd, Cu, Cr, Fe, Ni, Pb, and Zn. The involvement of mesophilic bacteria in metal leaching is attributed to their ability to generate acids such as sulphuric acid, which facilitate the mobilization of metals. The reduction in pH and increase in oxidation–reduction potential (ORP) can effectively create an optimal environment for the removal of heavy metals (HMs) from sludge [157]. Sulfur-oxidizing and iron-oxidizing bacteria are the predominant bacterial strains employed in biobleaching [8]. The process of biobleaching can occur either through the direct metabolic activity of leaching bacteria or indirectly through the by-products of bacterial metabolism [8]. The direct mechanism involves the binding of bacteria with HMs, leading to the oxidation of the latter to their soluble metal forms. Bacteria have been observed to convert sulphur to sulfuric acid through an indirect mechanism. This process results in a reduction in the pH of the medium and an increase in the solubility of HMs, as demonstrated in Figure 11 [157]. Biomineralization refers to the process by which various solid minerals, such as carbonates, phosphates, silicates, and sulphates are formed from metal ions that are subsequently precipitated by microorganisms that are actively involved in the process. Microbial-induced precipitation is the governing mechanism of biomineralization, which encompasses microbial-induced carbonate perception that is contingent upon urea hydrolysis, pH, and temperature [135]. The process of biomineralization involves the participation of various microorganisms, including photosynthetic microorganisms and sulfate-reducing bacteria, which contribute significantly to mineral precipitation through autotrophic and heterotrophic pathways [158].

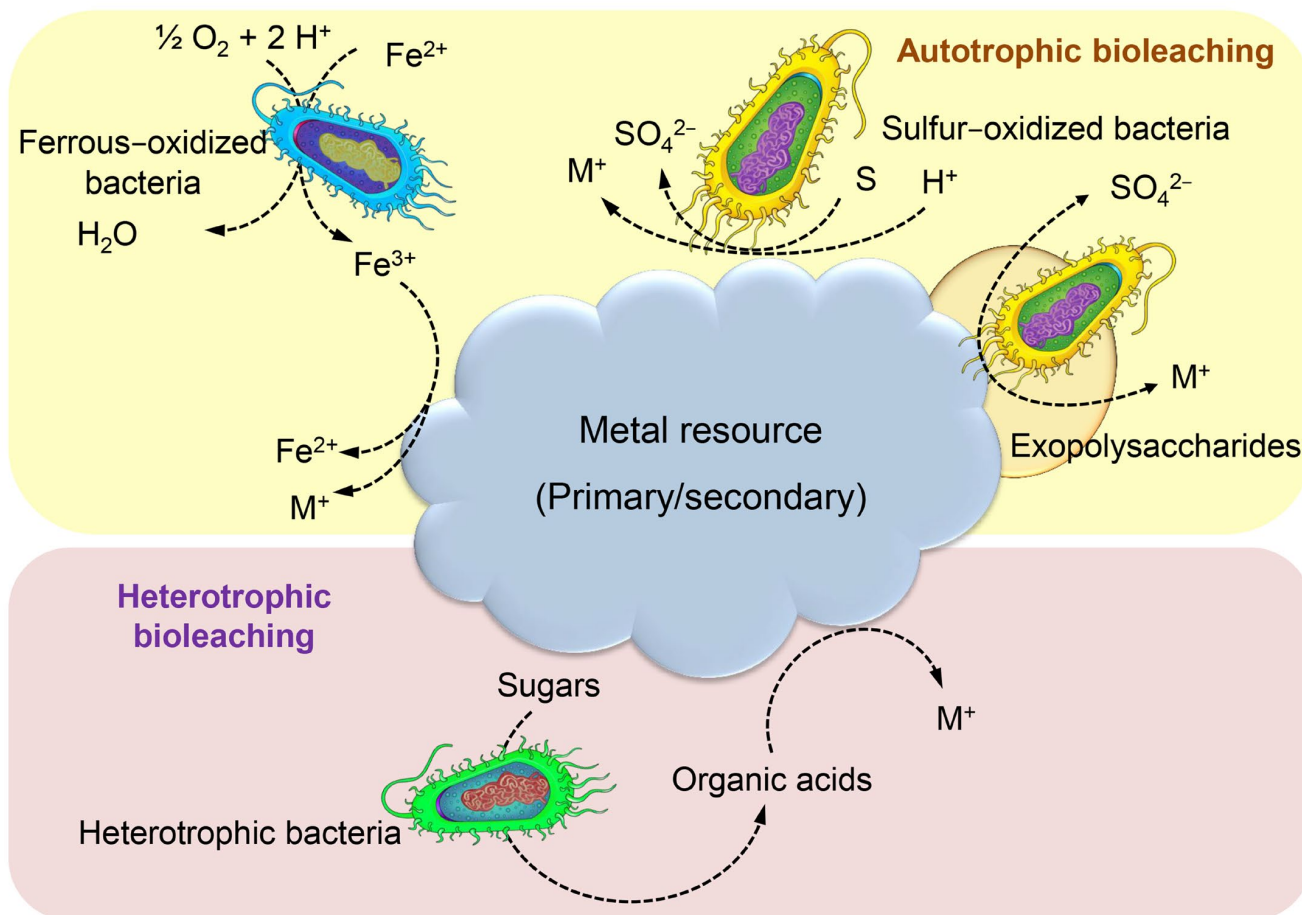


Figure 11. Bacterial biobleaching mechanism for HM removal.

7.3. Advances in Microbial Technologies for HM Removal

Extensive endeavors have been undertaken over a prolonged period to tackle the protracted persistence of HMs in the environment. Notwithstanding, the issue persists as a result of the increasing utilization of HM and the absence of feasible eradication remedies [159]. In order to mitigate the potential ecological ramifications of HM pollution, it is imperative to develop innovative and robust ecological technologies that can effectively extract and reclaim HMs from contaminated environments. The utilization of microbial bioremediation has been identified as a cost-effective approach to address the issue of HM contamination in environmental sites. Several techniques that hold promise for the removal of HMs from soil, including microbial fuel cells, biofilm, nanotechnology, and genetic engineering have been identified [160].

7.3.1. Microbial Fuel Cells

Microbial fuel cells (MFCs) are bio-electrochemical devices that facilitate the decomposition of organic waste into smaller molecules, thereby liberating protons (H^+) and electrons (e^-), which subsequently culminates in the production of energy. The diagram presented in Figure 12A illustrates the general structure and functioning of the MFC. Microorganisms generate electrons and protons at the anode through the process of oxidizing organic materials. Additionally, these microorganisms assimilate metal ions into their biomass. The transportation of electrons occurs through an external circuit, while the diffusion of protons toward the cathode takes place through the solution [161]. In principle, it is possible for any chemical possessing a high redox potential to function as the cathodic electron acceptor within the context of an MFC. Certain heme molecules exhibit significantly elevated redox potentials. Hence, they can serve as electron acceptors, wherein metallic ions amalgamate with protons and undergo reduction, thereby facilitating their elimination from the HMs and subsequent retrieval [162].

MFCs have been identified as a feasible approach to mitigate environmental contamination. The proposed approach presents a viable remedy for the treatment of wastewater and soil, along with the removal and recovery of HMs [163]. MFCs have the capability to convert chemical energy into electrical energy via oxidation mechanisms facilitated by microorganisms or enzymatic catalysis. MFCs provide a number of advantages in comparison to conventional fuel cells. The production of fuel can be achieved through the utilization of diverse organic or inorganic materials, including but not limited to soil sediments and organic waste [164]. The utilization of this technology has the potential to offer significant benefits in regions lacking access to electrical power and to improve soil characteristics. Notwithstanding its potential benefits, the technology presents a number of obstacles, including suboptimal rates of electrical generation. Consequently, various research endeavors have been undertaken to enhance electrical efficiency [165]. The removal of HMs from wastewater and soil has been the subject of extensive research utilizing MFC-based techniques [164–166]. Given the limited power output of MFCs, it is advisable to employ them primarily as sensors for detecting the toxicity of HMs [167].

7.3.2. Biofilm Technology

The microbial communities comprising bacteria, fungi, microalgae, and yeast are crucial in mitigating the deleterious impact of inorganic metallic salts, HMs, and organic waste on soil and water quality [168]. The replacement of physicochemical procedures with microbial metabolism for efficient utilization is a more environmentally friendly, secure, and effective strategy [169]. The microbial population generates EPSs, which enhance the efficacy of HM elimination. EPS substances are a class of biogenic macromolecules composed of proteins, uronic acid, and polysaccharides. These compounds are synthesized by microorganisms as a mechanism of self-preservation in response to various environmental stressors, such as extreme temperatures, acidic or alkaline conditions, and other physiological challenges. The aforementioned stressors enhance the efficacy of metal sequestration in the bioremediation of heavy metals [169]. The formation of biofilms is a consequence

of the association of one or more species, and the composition and configuration of EPS can be influenced by environmental factors [170]. EPS contains a significant amount of anionic charge. The sequestration of metallic ions occurs in substantial quantities [171]. The utilization of microbial biofilm has demonstrated a high degree of efficacy in the removal of HMs from both wastewater and soil.

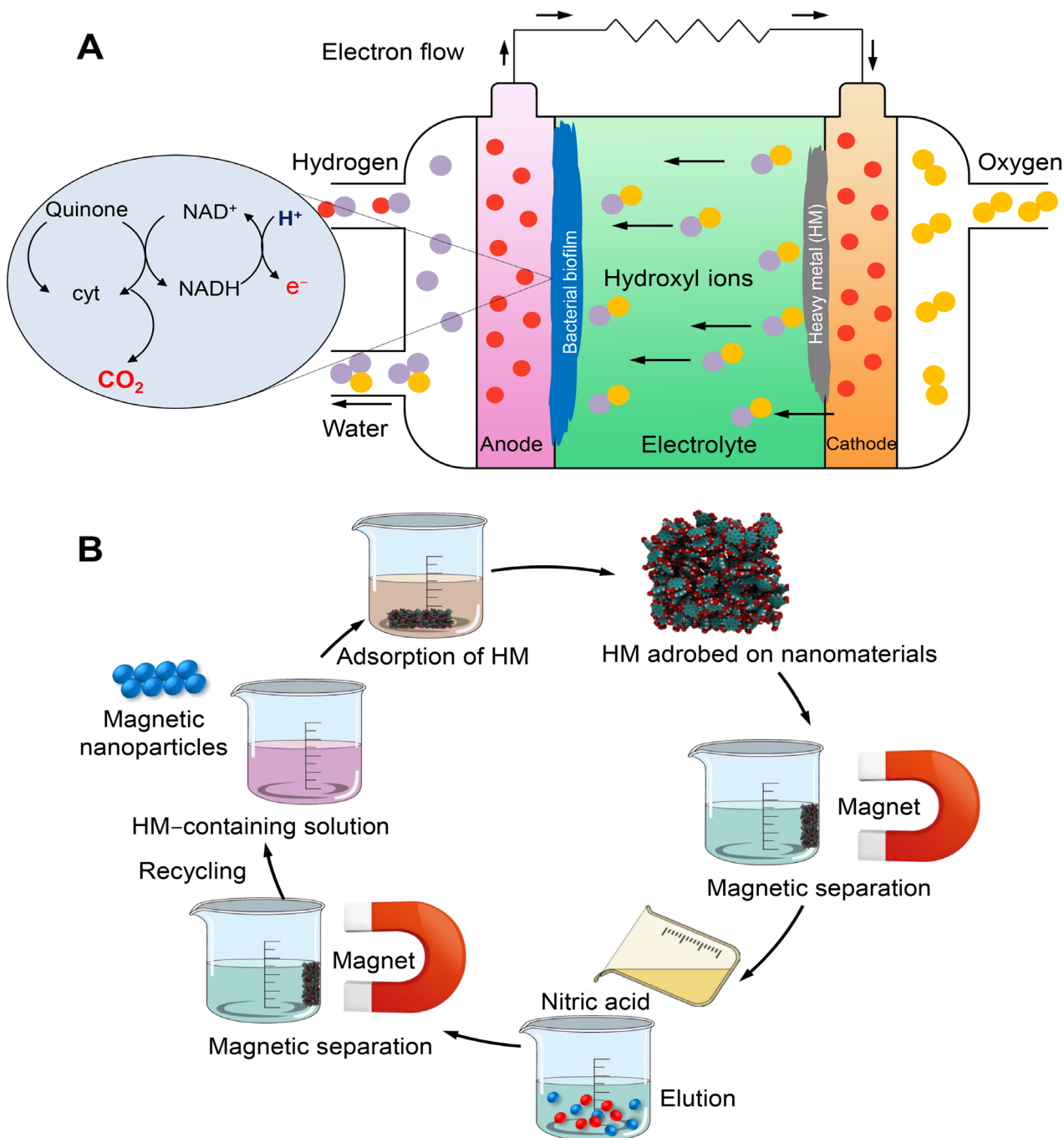


Figure 12. Mechanisms of HM removal via microbial fuel cells (A) and adsorption of metal ions across mesoporous magnetite nanoparticles (B).

7.3.3. Nanomaterial for HM Removal

Carbon nanotubes (CNTs) have demonstrated exceptional adsorption capabilities toward various HMs such as Cu, Pb, and Cr, owing to their potential adsorption active sites. The utilization of CNTs for the purpose of removing HMs from wastewater is

impeded by the considerable expenses associated with their production, as well as the difficulty of isolating CNTs from the wastewater stream [172]. The utilization of silica-based nanomaterials has been investigated as a means of extracting HMs from polluted sites, owing to their exceptional surface properties [173]. The promotion of HM adsorption can be achieved through the utilization of nano-silica as the foundation for nanocomposites or by modifying its surface with functional groups such as -NH₂ and -SH. Silica-based nanomaterials possess remarkable surface properties and are non-toxic, which makes them advantageous [174]. The employment of nanotechnology exhibits considerable potential in the realm of environmental remediation, particularly in the removal of HMs. The manipulation of material properties has been utilized for the purpose of removing HMs from the environment. The material exhibits exceptional surface properties and quantum effects due to its reduction to the nanoscale [172]. The principal modality by which nanomaterials eliminate HMs is through their elevated adsorption capacity and discernment for HMs (Figure 12B).

7.3.4. Biotechnological Tools

The application of contemporary biotechnological techniques such as genetic engineering and recombinant DNA technology has expedited the process of biodegradation and remediation of HMs [173]. Metagenomics and metabolic investigations offer insights into microbial diversity, population, and functional composition with respect to metal resistance genes. These findings can be leveraged to improve the efficacy of microbial strains in the removal or degradation of heavy metals. The field of genetic engineering has enabled the transfer of advantageous traits from one species to another, resulting in the development of specific strains for the purpose of bioremediating soil, sludge, or polluted water [175].

8. Conclusions

The issue of toxicity caused by HMs has emerged as a significant environmental challenge, with detrimental impacts on agricultural productivity and food security. The toxic effects of HMs result in their accumulation in the environment, which has adverse effects on human health, plant growth, and marine ecosystems. HMs can gain entry into the human body through various routes, such as ingestion via drinking water or food, inhalation through air, or through dermal exposure. After being absorbed, HMs are retained and accumulate within the human body. The accumulation of noxious metals in biological systems results in a range of deleterious consequences on diverse body tissues and organs. Various physical and chemical approaches have been suggested as potential remedies for the pollution caused by HMs. Nonetheless, the effectiveness of these techniques has been limited, and progress in this area has been slow. Therefore, alternative methodologies utilizing bioremediation have been explored. The employment of microbial bioremediation has emerged as an effective, reliable, and eco-friendly alternative for HM detoxification. However, HMs can interfere with the cellular activities of microorganisms depending on the existing concentration. The metal-tolerant microbes can enhance HM bioremediation. Microbes exhibit varying cellular structures, and their mechanisms for dealing with HM toxicity are typically dependent on the affinity of cellular biomolecules for metal ions and the stability of the specific metal in question. Microorganisms employ enzymatic systems or develop resistance through various mechanisms as a means of self-protection against HMs. However, the development of large-scale applications of combined remediation technology is recommended. In the context of practical application, one can acquire knowledge from both domestic and international experiences and enhance the judicious utilization of cutting-edge technologies. The mitigation of harmful effects caused by HMs necessitates the implementation of both scientific and community-based remediation technologies. Furthermore, forthcoming studies will derive advantages from assessing novel targets as protective procedures against organ toxicity triggered by HMs.

Author Contributions: Writing—original draft, M.K.A.E., N.E.E., M.M.E., S.H.A., S.A.E., S.E., M.S.M., E.M.S., N.A.A., A.E.A.E., I.M.E., A.E.E., and K.E.S., K.T.; Software, S.S.A.; Writing—review and editing, Validation, Visualization, S.S.A., M.K., and Y.A.-G.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data will be given upon request.

Conflicts of Interest: The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

1. Fergusson, J.E. *The Heavy Elements: Chemistry, Environmental Impact and Health Effects*; Pergamon Press: Oxford, UK, 1990; Volume 614.
2. Tchounwou, P.B.; Yedjou, C.G.; Patlolla, A.K.; Sutton, D.J. Heavy Metal Toxicity and the Environment. *Exp. Suppl.* **2012**, *101*, 133–164.
3. Duffus, J.H. “Heavy Metals” a Meaningless Term? (IUPAC Technical Report). *Pure Appl. Chem.* **2002**, *74*, 793–807. [[CrossRef](#)]
4. Jacob, J.M.; Karthik, C.; Saratale, R.G.; Kumar, S.S.; Prabakar, D.; Kadirvelu, K.; Pugazhendhi, A. Biological Approaches to Tackle Heavy Metal Pollution: A Survey of Literature. *J. Environ. Manag.* **2018**, *217*, 56–70. [[CrossRef](#)]
5. Sarker, A.; Al Masud, M.A.; Deepo, D.M.; Das, K.; Nandi, R.; Ansary, M.W.R.; Islam, A.R.M.T.; Islam, T. Biological and Green Remediation of Heavy Metal Contaminated Water and Soils: A State-of-the-Art Review. *Chemosphere* **2023**, *332*, 138861. [[CrossRef](#)] [[PubMed](#)]
6. Anyanwu, B.; Ezeiofor, A.; Igweze, Z.; Orisakwe, O. Heavy Metal Mixture Exposure and Effects in Developing Nations: An Update. *Toxics* **2018**, *6*, 65. [[CrossRef](#)] [[PubMed](#)]
7. Wu, C.; Li, F.; Yi, S.; Ge, F. Genetically Engineered Microbial Remediation of Soils Co-Contaminated by Heavy Metals and Polycyclic Aromatic Hydrocarbons: Advances and Ecological Risk Assessment. *J. Environ. Manag.* **2021**, *296*, 113185. [[CrossRef](#)]
8. Chen, S.-Y.; Wu, J.-Q.; Sung, S. Effects of Sulfur Dosage on Continuous Bioleaching of Heavy Metals from Contaminated Sediment. *J. Hazard. Mater.* **2022**, *424*, 127257. [[CrossRef](#)] [[PubMed](#)]
9. Ali, S.S.; Abdelkarim, E.A.; Elsamahy, T.; Al-Tohamy, R.; Li, F.; Kornaros, M.; Zuurro, A.; Zhu, D.; Sun, J. Bioplastic Production in Terms of Life Cycle Assessment: A State-of-the-Art Review. *Environ. Sci. Ecotechnol.* **2023**, *15*, 100254. [[CrossRef](#)] [[PubMed](#)]
10. Koutra, E.; Mastropetros, S.G.; Ali, S.S.; Tsigkou, K.; Kornaros, M. Assessing the Potential of *Chlorella Vulgaris* for Valorization of Liquid Digestates from Agro-Industrial and Municipal Organic Wastes in a Biorefinery Approach. *J. Clean. Prod.* **2021**, *280*, 124352. [[CrossRef](#)]
11. Ali, S.S.; Elsamahy, T.; Abdelkarim, E.A.; Al-Tohamy, R.; Kornaros, M.; Ruiz, H.A.; Zhao, T.; Li, F.; Sun, J. Biowastes for Biodegradable Bioplastics Production and End-of-Life Scenarios in Circular Bioeconomy and Biorefinery Concept. *Bioresour. Technol.* **2022**, *363*, 127869. [[CrossRef](#)]
12. Abdelfattah, A.; Ali, S.S.; Ramadan, H.; El-Aswar, E.I.; Eltawab, R.; Ho, S.-H.; Elsamahy, T.; Li, S.; El-Sheekh, M.M.; Schagerl, M.; et al. Microalgae-Based Wastewater Treatment: Mechanisms, Challenges, Recent Advances, and Future Prospects. *Environ. Sci. Ecotechnol.* **2022**, *13*, 100205. [[CrossRef](#)] [[PubMed](#)]
13. Ali, S.S.; El-Sheekh, M.; Manni, A.; Ruiz, H.A.; Elsamahy, T.; Sun, J.; Schagerl, M. Microalgae-Mediated Wastewater Treatment for Biofuels Production: A Comprehensive Review. *Microbiol. Res.* **2022**, *265*, 127187. [[CrossRef](#)] [[PubMed](#)]
14. Al-Tohamy, R.; Ali, S.S.; Li, F.; Okasha, K.M.; Mahmoud, Y.A.-G.; Elsamahy, T.; Jiao, H.; Fu, Y.; Sun, J. A Critical Review on the Treatment of Dye-Containing Wastewater: Ecotoxicological and Health Concerns of Textile Dyes and Possible Remediation Approaches for Environmental Safety. *Ecotoxicol. Environ. Saf.* **2022**, *231*, 113160. [[CrossRef](#)] [[PubMed](#)]
15. Kumar, L.; Bharadvaja, N. Microbial Remediation of Heavy Metals. In *Microbial Bioremediation & Biodegradation*; Springer: Singapore, 2020.
16. Ali, H.; Khan, E.; Ilahi, I. Environmental Chemistry and Ecotoxicology of Hazardous Heavy Metals: Environmental Persistence, Toxicity, and Bioaccumulation. *J. Chem.* **2019**, *2019*, 1–14. [[CrossRef](#)]
17. Mitra, S.; Chakraborty, A.J.; Tareq, A.M.; Emran, T.B.; Nainu, F.; Khushro, A.; Idris, A.M.; Khandaker, M.U.; Osman, H.; Alhumaydhi, F.A.; et al. Impact of Heavy Metals on the Environment and Human Health: Novel Therapeutic Insights to Counter the Toxicity. *J. King Saud Univ.-Sci.* **2022**, *34*, 101865. [[CrossRef](#)]
18. Verma, S.; Bhatt, P.; Verma, A.; Mudila, H.; Prasher, P.; Rene, E.R. Microbial Technologies for Heavy Metal Remediation: Effect of Process Conditions and Current Practices. *Clean Technol. Environ. Policy* **2021**. [[CrossRef](#)]
19. Rehman, A.U.; Nazir, S.; Irshad, R.; Tahir, K.; ur Rehman, K.; Islam, R.U.; Wahab, Z. Toxicity of Heavy Metals in Plants and Animals and Their Uptake by Magnetic Iron Oxide Nanoparticles. *J. Mol. Liq.* **2021**, *321*, 114455. [[CrossRef](#)]

20. Li, R.; Wu, H.; Ding, J.; Fu, W.; Gan, L.; Li, Y. Mercury Pollution in Vegetables, Grains and Soils from Areas Surrounding Coal-Fired Power Plants. *Sci. Rep.* **2017**, *7*, 46545. [[CrossRef](#)]
21. Deng, Y.; Wang, M.; Tian, T.; Lin, S.; Xu, P.; Zhou, L.; Dai, C.; Hao, Q.; Wu, Y.; Zhai, Z.; et al. The Effect of Hexavalent Chromium on the Incidence and Mortality of Human Cancers: A Meta-Analysis Based on Published Epidemiological Cohort Studies. *Front. Oncol.* **2019**, *9*, 24. [[CrossRef](#)]
22. Negahdari, S.; Sabaghan, M.; Pirhadi, M.; Alikord, M.; Sadighara, P.; Darvishi, M.; Nazer, M. Potential Harmful Effects of Heavy Metals as a Toxic and Carcinogenic Agent in Marine Food-An Overview. *Egypt. J. Vet. Sci.* **2021**, *52*, 379–385. [[CrossRef](#)]
23. Wang, X.; Wu, Y.; Sun, X.; Guo, Q.; Xia, W.; Wu, Y.; Li, J.; Xu, S.; Li, Y. Arsenic Exposure and Metabolism in Relation to Blood Pressure Changes in Pregnant Women. *Ecotoxicol. Environ. Saf.* **2021**, *222*, 112527. [[CrossRef](#)] [[PubMed](#)]
24. Sundseth, K.; Pacyna, J.; Pacyna, E.; Pirrone, N.; Thorne, R. Global Sources and Pathways of Mercury in the Context of Human Health. *Int. J. Environ. Res. Public Health* **2017**, *14*, 105. [[CrossRef](#)] [[PubMed](#)]
25. Teixeira, F.B.; de Oliveira, A.C.A.; Leão, L.K.R.; Fagundes, N.C.F.; Fernandes, R.M.; Fernandes, L.M.P.; da Silva, M.C.F.; Amado, L.L.; Sagica, F.E.S.; de Oliveira, E.H.C.; et al. Exposure to Inorganic Mercury Causes Oxidative Stress, Cell Death, and Functional Deficits in the Motor Cortex. *Front. Mol. Neurosci.* **2018**, *11*, 125. [[CrossRef](#)] [[PubMed](#)]
26. Papadopoulou, E.; Botton, J.; Caspersen, I.H.; Alexander, J.; Eggesbø, M.; Haugen, M.; Iszatt, N.; Jacobsson, B.; Knutsen, H.K.; Meltzer, H.M.; et al. Maternal Seafood Intake during Pregnancy, Prenatal Mercury Exposure and Child Body Mass Index Trajectories up to 8 Years. *Int. J. Epidemiol.* **2021**, *50*, 1134–1146. [[CrossRef](#)]
27. Farzan, S.F.; Howe, C.G.; Chen, Y.; Gilbert-Diamond, D.; Korricks, S.; Jackson, B.P.; Weinstein, A.R.; Karagas, M.R. Prenatal and Postnatal Mercury Exposure and Blood Pressure in Childhood. *Environ. Int.* **2021**, *146*, 106201. [[CrossRef](#)]
28. Kang, P.; Shin, H.Y.; Kim, K.Y. Association between Dyslipidemia and Mercury Exposure in Adults. *Int. J. Environ. Res. Public Health* **2021**, *18*, 775. [[CrossRef](#)]
29. Lee, S.; Cho, S.-R.; Jeong, I.; Park, J.B.; Shin, M.-Y.; Kim, S.; Kim, J.H. Mercury Exposure and Associations with Hyperlipidemia and Elevated Liver Enzymes: A Nationwide Cross-Sectional Survey. *Toxics* **2020**, *8*, 47. [[CrossRef](#)]
30. Charkiewicz, A.E.; Backstrand, J.R. Lead Toxicity and Pollution in Poland. *Int. J. Environ. Res. Public Health* **2020**, *17*, 4385. [[CrossRef](#)]
31. Wang, T.; Zhang, J.; Xu, Y. Epigenetic Basis of Lead-Induced Neurological Disorders. *Int. J. Environ. Res. Public Health* **2020**, *17*, 4878. [[CrossRef](#)]
32. Obeng-Gyasi, E.; Ferguson, A.C.; Stamatakis, K.A.; Province, M.A. Combined Effect of Lead Exposure and Allostatic Load on Cardiovascular Disease Mortality—A Preliminary Study. *Int. J. Environ. Res. Public Health* **2021**, *18*, 6879. [[CrossRef](#)]
33. Nakhaee, S.; Amirabadizadeh, A.; Brent, J.; Mehrpour, O. Impact of Chronic Lead Exposure on Liver and Kidney Function and Haematologic Parameters. *Basic Clin. Pharmacol. Toxicol.* **2018**, *124*, 621–628. [[CrossRef](#)]
34. Zhou, C.-C.; He, Y.-Q.; Gao, Z.-Y.; Wu, M.-Q.; Yan, C.-H. Sex Differences in the Effects of Lead Exposure on Growth and Development in Young Children. *Chemosphere* **2020**, *250*, 126294. [[CrossRef](#)] [[PubMed](#)]
35. Dehghan, S.F.; Mehrifar, Y.; Ardalan, A. The Relationship between Exposure to Lead-Containing Welding Fumes and the Levels of Reproductive Hormones. *Ann. Glob. Health* **2019**, *85*, 125. [[CrossRef](#)] [[PubMed](#)]
36. Chakraborty, R.; Renu, K.; Eladl, M.A.; El-Sherbiny, M.; Elsherbiny, D.M.A.; Mirza, A.K.; Vellingiri, B.; Iyer, M.; Dey, A.; Valsala Gopalakrishnan, A. Mechanism of Chromium-Induced Toxicity in Lungs, Liver, and Kidney and Their Ameliorative Agents. *Biomed. Pharmacother.* **2022**, *151*, 113119. [[CrossRef](#)] [[PubMed](#)]
37. Xia, B.; Yuan, J.; Pang, L.; He, K. Chromium [Cr(VI)] Exposure Causes Cytotoxicity of Human Bronchial Epithelial Cells (16-HBE) and Proteomic Alterations. *Int. J. Toxicol.* **2022**, *41*, 225–233. [[CrossRef](#)]
38. Peng, Y.; Hu, J.; Li, Y.; Zhang, B.; Liu, W.; Li, H.; Zhang, H.; Hu, C.; Chen, X.; Xia, W.; et al. Exposure to Chromium during Pregnancy and Longitudinally Assessed Fetal Growth: Findings from a Prospective Cohort. *Environ. Int.* **2018**, *121*, 375–382. [[CrossRef](#)]
39. Baszuk, P.; Janasik, B.; Pietrzak, S.; Marciniak, W.; Reszka, E.; Białkowska, K.; Jabłońska, E.; Muszyńska, M.; Lesicka, M.; Derkacz, R.; et al. Lung Cancer Occurrence-Correlation with Serum Chromium Levels and Genotypes. *Biol. Trace Elem. Res.* **2021**, *199*, 1228–1236. [[CrossRef](#)]
40. Al Hossain, M.M.A.; Yajima, I.; Tazaki, A.; Xu, H.; Saheduzzaman, M.; Ohgami, N.; Ahsan, N.; Akhand, A.A.; Kato, M. Chromium-Mediated Hyperpigmentation of Skin in Male Tannery Workers in Bangladesh. *Chemosphere* **2019**, *229*, 611–617. [[CrossRef](#)]
41. Kumar, V.; Dwivedi, S.K. Bioremediation Mechanism and Potential of Copper by Actively Growing Fungus *Trichoderma* Lixii CR700 Isolated from Electroplating Wastewater. *J. Environ. Manag.* **2021**, *277*, 111370. [[CrossRef](#)]
42. Kahlson, M.A.; Dixon, S.J. Copper-Induced Cell Death. *Science* **2022**, *375*, 1231–1232. [[CrossRef](#)]
43. Niu, Y.; Zhang, Y.; Zhu, Z.; Zhang, X.; Liu, X.; Zhu, S.; Song, Y.; Jin, X.; Lindholm, B.; Yu, C. Elevated Intracellular Copper Contributes a Unique Role to Kidney Fibrosis by Lysyl Oxidase Mediated Matrix Crosslinking. *Cell Death Dis.* **2020**, *11*, 211. [[CrossRef](#)] [[PubMed](#)]
44. Tvrdá, E.; Peer, R.; Sikka, S.C.; Agarwal, A. Iron and Copper in Male Reproduction: A Double-Edged Sword. *J. Assist. Reprod. Genet.* **2015**, *32*, 3–16. [[CrossRef](#)] [[PubMed](#)]
45. Yang, H.; Liu, C.-N.; Wolf, R.M.; Ralle, M.; Dev, S.; Pierson, H.; Askin, F.; Steele, K.E.; Magnuson, T.H.; Schweitzer, M.A.; et al. Obesity Is Associated with Copper Elevation in Serum and Tissues. *Metallomics* **2019**, *11*, 1363–1371. [[CrossRef](#)]

46. Witt, B.; Stiboller, M.; Raschke, S.; Friese, S.; Ebert, F.; Schwerdtle, T. Characterizing Effects of Excess Copper Levels in a Human Astrocytic Cell Line with Focus on Oxidative Stress Markers. *J. Trace Elem. Med. Biol.* **2021**, *65*, 126711. [[CrossRef](#)] [[PubMed](#)]
47. Genchi, G.; Carocci, A.; Lauria, G.; Sinicropi, M.S.; Catalano, A. Nickel: Human Health and Environmental Toxicology. *Int. J. Environ. Res. Public Health* **2020**, *17*, 679. [[CrossRef](#)] [[PubMed](#)]
48. Chen, X.; Li, Y.; Zhang, B.; Zhou, A.; Zheng, T.; Huang, Z.; Pan, X.; Liu, W.; Liu, H.; Jiang, Y.; et al. Maternal Exposure to Nickel in Relation to Preterm Delivery. *Chemosphere* **2018**, *193*, 1157–1163. [[CrossRef](#)] [[PubMed](#)]
49. Titcomb, T.J.; Liu, B.; Lehmler, H.; Snetselaar, L.G.; Bao, W. Environmental Nickel Exposure and Diabetes in a Nationally Representative Sample of US Adults. *Expo. Health* **2021**, *13*, 697–704. [[CrossRef](#)]
50. Yu, M.; Zhang, J. Serum and Hair Nickel Levels and Breast Cancer: Systematic Review and Meta-Analysis. *Biol. Trace Elem. Res.* **2017**, *179*, 32–37. [[CrossRef](#)]
51. Büyüköztürk, S.; Gelincik, A.; Ünal, D.; Demirtürk, M.; Çelik, D.D.; Erden, S.; Çolakoğlu, B.; Erdem Kuruca, S. Oral Nickel Exposure May Induce Type I Hypersensitivity Reaction in Nickel-Sensitized Subjects. *Int. Immunopharmacol.* **2015**, *26*, 92–96. [[CrossRef](#)]
52. Redvers, N.; Chischilly, A.M.; Warne, D.; Pino, M.; Lyon-Colbert, A. Uranium Exposure in American Indian Communities: Health, Policy, and the Way Forward. *Environ. Health Perspect.* **2021**, 129. [[CrossRef](#)]
53. Surdyk, S.; Itani, M.; Al-Lobaidy, M.; Kahale, L.A.; Farha, A.; Dewachi, O.; Akl, E.A.; Habib, R.R. Weaponised Uranium and Adverse Health Outcomes in Iraq: A Systematic Review. *BMJ Glob. Health* **2021**, *6*, e004166. [[CrossRef](#)] [[PubMed](#)]
54. Wang, X.; Dai, X.; Shi, C.; Wan, J.; Silver, M.A.; Zhang, L.; Chen, L.; Yi, X.; Chen, B.; Zhang, D.; et al. A 3,2-Hydroxypyridinone-Based Decorporation Agent That Removes Uranium from Bones In Vivo. *Nat. Commun.* **2019**, *10*, 2570. [[CrossRef](#)] [[PubMed](#)]
55. Joseph, S.J.; Arunachalam, K.D.; Murthy, P.B.; Ramalingam, R.; Musthafa, M.S. Uranium Induces Genomic Instability and Slows Cell Cycle Progression in Human Lymphocytes in Acute Toxicity Study. *Toxicol. Vitro* **2021**, *73*, 105149. [[CrossRef](#)] [[PubMed](#)]
56. Sears, C.G.; Poulsen, A.H.; Eliot, M.; Howe, C.J.; James, K.A.; Harrington, J.M.; Roswall, N.; Overvad, K.; Tjønneland, A.; Raaschou-Nielsen, O.; et al. Urine Cadmium and Acute Myocardial Infarction among Never Smokers in the Danish Diet, Cancer and Health Cohort. *Environ. Int.* **2021**, *150*, 106428. [[CrossRef](#)] [[PubMed](#)]
57. Vijayakumar, V.; Abern, M.R.; Jagai, J.S.; Kajdacsy-Balla, A. Observational Study of the Association between Air Cadmium Exposure and Prostate Cancer Aggressiveness at Diagnosis among a Nationwide Retrospective Cohort of 230,540 Patients in the United States. *Int. J. Environ. Res. Public Health* **2021**, *18*, 8333. [[CrossRef](#)] [[PubMed](#)]
58. Amadou, A.; Praud, D.; Coudon, T.; Danjou, A.M.N.; Faure, E.; Deygas, F.; Grassot, L.; Leffondré, K.; Severi, G.; Salizzoni, P.; et al. Exposure to Airborne Cadmium and Breast Cancer Stage, Grade and Histology at Diagnosis: Findings from the E3N Cohort Study. *Sci. Rep.* **2021**, *11*, 23088. [[CrossRef](#)]
59. Yan, L.-J.; Allen, D.C. Cadmium-Induced Kidney Injury: Oxidative Damage as a Unifying Mechanism. *Biomolecules* **2021**, *11*, 1575. [[CrossRef](#)]
60. Rajan, M.; Anderson, C.P.; Rindler, P.M.; Romney, S.J.; Ferreira dos Santos, M.C.; Gertz, J.; Leibold, E.A. NHR-14 Loss of Function Couples Intestinal Iron Uptake with Innate Immunity in *C. Elegans* through PQM-1 Signaling. *Elife* **2019**, *8*. [[CrossRef](#)]
61. Verna, G.; Sila, A.; Liso, M.; Mastronardi, M.; Chieppa, M.; Cena, H.; Campiglia, P. Iron-Enriched Nutritional Supplements for the 2030 Pharmacy Shelves. *Nutrients* **2021**, *13*, 378. [[CrossRef](#)]
62. Killip, S.; Bennett, J.M.; Chambers, M.D. Iron Deficiency Anemia. *Am. Fam. Physician* **2007**, *75*, 671–678.
63. Weyh, C.; Krüger, K.; Peeling, P.; Castell, L. The Role of Minerals in the Optimal Functioning of the Immune System. *Nutrients* **2022**, *14*, 644. [[CrossRef](#)]
64. Means, R.T. Iron Deficiency and Iron Deficiency Anemia: Implications and Impact in Pregnancy, Fetal Development, and Early Childhood Parameters. *Nutrients* **2020**, *12*, 447. [[CrossRef](#)]
65. Quezada-Pinedo, H.G.; Cassel, F.; Duijts, L.; Muckenthaler, M.U.; Gassmann, M.; Jaddoe, V.W.; Reiss, I.K.; Vermeulen, M.J. Maternal Iron Status in Pregnancy and Child Health Outcomes after Birth: A Systematic Review and Meta-Analysis. *Nutrients* **2021**, *13*, 2221. [[CrossRef](#)]
66. Rojas-Lemus, M.; López-Valdez, N.; Bizarro-Nevarés, P.; González-Villalva, A.; Ustarroz-Cano, M.; Zepeda-Rodríguez, A.; Pasos-Nájera, F.; García-Peláez, I.; Rivera-Fernández, N.; Fortoul, T.I. Toxic Effects of Inhaled Vanadium Attached to Particulate Matter: A Literature Review. *Int. J. Environ. Res. Public Health* **2021**, *18*, 8457. [[CrossRef](#)]
67. Imtiaz, M.; Rizwan, M.S.; Xiong, S.; Li, H.; Ashraf, M.; Shahzad, S.M.; Shahzad, M.; Rizwan, M.; Tu, S. Vanadium, Recent Advancements and Research Prospects: A Review. *Environ. Int.* **2015**, *80*, 79–88. [[CrossRef](#)] [[PubMed](#)]
68. Filler, G.; Kobrzynski, M.; Sidhu, H.K.; Belostotsky, V.; Huang, S.-H.S.; McIntyre, C.; Yang, L. A Cross-Sectional Study Measuring Vanadium and Chromium Levels in Paediatric Patients with CKD. *BMJ Open* **2017**, *7*, e014821. [[CrossRef](#)] [[PubMed](#)]
69. Ngwa, H.A.; Kanthasamy, A.; Jin, H.; Anantharam, V.; Kanthasamy, A.G. Vanadium Exposure Induces Olfactory Dysfunction in an Animal Model of Metal Neurotoxicity. *Neurotoxicology* **2014**, *43*, 73–81. [[CrossRef](#)]
70. Mahey, S.; Kumar, R.; Sharma, M.; Kumar, V.; Bhardwaj, R. A Critical Review on Toxicity of Cobalt and Its Bioremediation Strategies. *SN Appl. Sci.* **2020**, *2*, 1–12. [[CrossRef](#)]
71. Wahlqvist, F.; Bryngelsson, I.-L.; Westberg, H.; Vihlborg, P.; Andersson, L. Dermal and Inhalable Cobalt Exposure—Uptake of Cobalt for Workers at Swedish Hard Metal Plants. *PLoS ONE* **2020**, *15*, e0237100. [[CrossRef](#)] [[PubMed](#)]
72. Magaye, R.; Zhao, J.; Bowman, L.; Ding, M. Genotoxicity and Carcinogenicity of Cobalt-, Nickel- and Copper-Based Nanoparticles. *Exp. Ther. Med.* **2012**, *4*, 551–561. [[CrossRef](#)] [[PubMed](#)]

73. Zhang, N.; Yang, S.; Yang, J.; Deng, Y.; Li, S.; Li, N.; Chen, X.; Yu, P.; Liu, Z.; Zhu, J. Association between Metal Cobalt Exposure and the Risk of Congenital Heart Defect Occurrence in Offspring: A Multi-Hospital Case-Control Study. *Environ. Health Prev. Med.* **2020**, *25*, 38. [[CrossRef](#)] [[PubMed](#)]
74. Linna, A.; Uitti, J.; Oksa, P.; Toivio, P.; Virtanen, V.; Lindholm, H.; Halkosaari, M.; Sauni, R. Effects of Occupational Cobalt Exposure on the Heart in the Production of Cobalt and Cobalt Compounds: A 6-Year Follow-Up. *Int. Arch. Occup. Environ. Health* **2020**, *93*, 365–374. [[CrossRef](#)] [[PubMed](#)]
75. Karbowska, B. Presence of Thallium in the Environment: Sources of Contaminations, Distribution and Monitoring Methods. *Environ. Monit. Assess.* **2016**, *188*, 1–19. [[CrossRef](#)]
76. Kremer, D.; Riemersma, N.L.; Groothof, D.; Sotomayor, C.G.; Eisenga, M.F.; Post, A.; Knobbe, T.J.; Touw, D.J.; Bakker, S.J.L. Plasma Thallium Concentration, Kidney Function, Nephrotoxicity and Graft Failure in Kidney Transplant Recipients. *J. Clin. Med.* **2022**, *11*, 1970. [[CrossRef](#)]
77. Lin, G.; Yuan, L.; Bai, L.; Liu, Y.; Wang, Y.; Qiu, Z. Successful Treatment of a Patient with Severe Thallium Poisoning in a Coma Using Prussian Blue and Plasma Exchange: A Case Report. *Medicine* **2019**, *98*, e14629–e14629. [[CrossRef](#)] [[PubMed](#)]
78. Ahmed, M.J.; Mia, M.L. A New Simple, Highly Sensitive and Selective Spectrofluorimetric Method for the Speciation of Thallium at Pico-Trace Levels in Various Complex Matrices Using N-(Pyridin-2-Yl)-Quinoline-2-Carbothioamide. *RSC Adv.* **2021**, *11*, 32312–32328. [[CrossRef](#)]
79. Xia, W.; Du, X.; Zheng, T.; Zhang, B.; Li, Y.; Bassig, B.A.; Zhou, A.; Wang, Y.; Xiong, C.; Li, Z.; et al. A Case-Control Study of Prenatal Thallium Exposure and Low Birth Weight in China. *Environ. Health Perspect.* **2016**, *124*, 164–169. [[CrossRef](#)]
80. Zwolak, I. Epigallocatechin Gallate for Management of Heavy Metal-Induced Oxidative Stress: Mechanisms of Action, Efficacy, and Concerns. *Int. J. Mol. Sci.* **2021**, *22*, 4027. [[CrossRef](#)]
81. Tamás, M.J.; Sharma, S.K.; Ibstedt, S.; Jacobson, T.; Christen, P. Heavy Metals and Metalloids as a Cause for Protein Misfolding and Aggregation. *Biomolecules* **2014**, *4*, 252–267. [[CrossRef](#)]
82. Patel, P.; Raju, N.J.; Reddy, B.C.S.R.; Suresh, U.; Sankar, D.B.; Reddy, T.V.K. Heavy Metal Contamination in River Water and Sediments of the Swarnamukhi River Basin, India: Risk Assessment and Environmental Implications. *Environ. Geochem. Health* **2017**, *40*, 609–623. [[CrossRef](#)]
83. Miranda, L.S.; Wijesiri, B.; Ayoko, G.A.; Egodawatta, P.; Goonetilleke, A. Water-Sediment Interactions and Mobility of Heavy Metals in Aquatic Environments. *Water Res.* **2021**, *202*, 117386. [[CrossRef](#)]
84. Kim, B.-M.; Kim, B.; Nam, S.-E.; Eom, H.-J.; Lee, S.; Kim, K.; Rhee, J.-S. Reductive Transformation of Hexavalent Chromium in Ice Decreases Chromium Toxicity in Aquatic Animals. *Environ. Sci. Technol.* **2022**, *56*, 3503–3513. [[CrossRef](#)] [[PubMed](#)]
85. Morroni, L.; Gaion, A.; Broccoli, A.; Ferrari, S.; Pellegrini, D.; Sartori, D. Influence of Salinity on Copper Toxicity in Paracentrotus Lividus and Arbacia Lixula Embryos. *Water* **2022**, *15*, 65. [[CrossRef](#)]
86. Wang, N.; Kunz, J.L.; Cleveland, D.M.; Stevens, J.A.; Hammer, E.J.; Van Genderen, E.; Ryan, A.C.; Schlekot, C.E. Evaluation of Acute and Chronic Toxicity of Nickel and Zinc to 2 Sensitive Freshwater Benthic Invertebrates Using Refined Testing Methods. *Environ. Toxicol. Chem.* **2020**, *39*, 2256–2268. [[CrossRef](#)] [[PubMed](#)]
87. Byeon, E.; Kang, H.-M.; Yoon, C.; Lee, J.-S. Toxicity Mechanisms of Arsenic Compounds in Aquatic Organisms. *Aquat. Toxicol.* **2021**, *237*, 105901. [[CrossRef](#)] [[PubMed](#)]
88. Perić, L.; Stinga Perusco, V.; Nerlović, V. Differential Response of Biomarkers in the Native European Flat Oyster *Ostrea Edulis* and the Non-Indigenous Pacific Oyster *Crassostrea Gigas* Co-Exposed to Cadmium and Copper. *J. Exp. Mar. Bio. Ecol.* **2020**, *523*, 151271. [[CrossRef](#)]
89. Meng, J.; Wang, W.-X.; Li, L.; Zhang, G. Tissue-Specific Molecular and Cellular Toxicity of Pb in the Oyster (*Crassostrea Gigas*): mRNA Expression and Physiological Studies. *Aquat. Toxicol.* **2018**, *198*, 257–268. [[CrossRef](#)]
90. Xu, X.; Shi, L.; Wang, M. Comparative Quantitative Proteomics Unveils Putative Mechanisms Involved into Mercury Toxicity and Tolerance in *Tigriopus Japonicus* under Multigenerational Exposure Scenario. *Environ. Pollut.* **2016**, *218*, 1287–1297. [[CrossRef](#)]
91. Alengebawy, A.; Abdelkhalek, S.T.; Qureshi, S.R.; Wang, M.-Q. Heavy Metals and Pesticides Toxicity in Agricultural Soil and Plants: Ecological Risks and Human Health Implications. *Toxics* **2021**, *9*, 42. [[CrossRef](#)]
92. Chen, D.; Liu, X.; Bian, R.; Cheng, K.; Zhang, X.; Zheng, J.; Joseph, S.; Crowley, D.; Pan, G.; Li, L. Effects of Biochar on Availability and Plant Uptake of Heavy Metals – A Meta-Analysis. *J. Environ. Manag.* **2018**, *222*, 76–85. [[CrossRef](#)]
93. Sabra, M.; Aboulnasr, A.; Franken, P.; Perreca, E.; Wright, L.P.; Camehl, I. Beneficial Root Endophytic Fungi Increase Growth and Quality Parameters of Sweet Basil in Heavy Metal Contaminated Soil. *Front. Plant Sci.* **2018**, *9*, 1726. [[CrossRef](#)] [[PubMed](#)]
94. Kladsomboon, S.; Jaiyen, C.; Choprathumma, C.; Tusai, T.; Apilux, A. Heavy Metals Contamination in Soil, Surface Water, Crops, and Resident Blood in Uthai District, Phra Nakhon Si Ayutthaya, Thailand. *Environ. Geochem. Health* **2019**, *42*, 545–561. [[CrossRef](#)] [[PubMed](#)]
95. Mondal, N.K.; Das, C.; Datta, J.K. Effect of Mercury on Seedling Growth, Nodulation and Ultrastructural Deformation of *Vigna radiata* (L) Wilczek. *Environ. Monit. Assess.* **2015**, *187*, 241. [[CrossRef](#)]
96. Verma, S.; Kuila, A. Bioremediation of Heavy Metals by Microbial Process. *Environ. Technol. Innov.* **2019**, *14*, 100369. [[CrossRef](#)]
97. Wu, Q.; Leung, J.Y.S.; Geng, X.; Chen, S.; Huang, X.; Li, H.; Huang, Z.; Zhu, L.; Chen, J.; Lu, Y. Heavy Metal Contamination of Soil and Water in the Vicinity of an Abandoned E-Waste Recycling Site: Implications for Dissemination of Heavy Metals. *Sci. Total Environ.* **2015**, *506*, 217–225.

98. Wang, Z.; Lei, G. Study on Penetration Effect of Heavy Metal Migration in Different Soil Types. *IOP Conf. Ser. Mater. Sci. Eng.* **2018**, *394*, 52033. [[CrossRef](#)]
99. Mahbub, K.R.; King, W.L.; Siboni, N.; Nguyen, V.K.; Rahman, M.M.; Megharaj, M.; Seymour, J.R.; Franks, A.E.; Labbate, M. Long-Lasting Effect of Mercury Contamination on the Soil Microbiota and Its Co-Selection of Antibiotic Resistance. *Environ. Pollut.* **2020**, *265*, 115057. [[CrossRef](#)]
100. Xu, J.; Zhang, J.; Lv, Y.; Xu, K.; Lu, S.; Liu, X.; Yang, Y. Effect of Soil Mercury Pollution on Ginger (*Zingiber Officinale* Roscoe): Growth, Product Quality, Health Risks and Silicon Mitigation. *Ecotoxicol. Environ. Saf.* **2020**, *195*, 110472. [[CrossRef](#)]
101. Htwe, T.; Onthong, J.; Duangpan, S.; Techato, K.; Chotikarn, P.; Sinutok, S. Effect of Copper Contamination on Plant Growth and Metal Contents in Rice Plant (*Oryza sativa* L.). *Commun. Soil Sci. Plant Anal.* **2020**, *51*, 2349–2360. [[CrossRef](#)]
102. Sharma, A.; Kapoor, D.; Wang, J.; Shahzad, B.; Kumar, V.; Bali, A.S.; Jasrotia, S.; Zheng, B.; Yuan, H.; Yan, D. Chromium Bioaccumulation and Its Impacts on Plants: An Overview. *Plants* **2020**, *9*, 100. [[CrossRef](#)]
103. Nasiri, A.; Jamshidi-Zanjani, A.; Khodadadi Darban, A. Application of Enhanced Electrokinetic Approach to Remediate Cr-Contaminated Soil: Effect of Chelating Agents and Permeable Reactive Barrier. *Environ. Pollut.* **2020**, *266*, 115197. [[CrossRef](#)]
104. Abbas, A.; Azeem, M.; Naveed, M.; Latif, A.; Bashir, S.; Ali, A.; Bilal, M.; Ali, L. Synergistic Use of Biochar and Acidified Manure for Improving Growth of Maize in Chromium Contaminated Soil. *Int. J. Phytoremediation* **2019**, *22*, 52–61. [[CrossRef](#)] [[PubMed](#)]
105. Singh, D.; Sharma, N.L.; Singh, C.K.; Yerramilli, V.; Narayan, R.; Sarkar, S.K.; Singh, I. Chromium (VI)-Induced Alterations in Physio-Chemical Parameters, Yield, and Yield Characteristics in Two Cultivars of Mungbean (*Vigna radiata* L.). *Front. Plant Sci.* **2021**, *12*, 735129. [[CrossRef](#)] [[PubMed](#)]
106. Liu, J.; Liu, F.; Ding, C.; Ma, F.; Yu, H.; Shi, Y.; Zhang, X. Response of *Trametes Hirsuta* to Hexavalent Chromium Promotes Laccase-Mediated Decolorization of Reactive Black 5. *Ecotoxicol. Environ. Saf.* **2020**, *205*, 111134. [[CrossRef](#)] [[PubMed](#)]
107. Wang, L.; Hou, D.; Shen, Z.; Zhu, J.; Jia, X.; Ok, Y.S.; Tack, F.M.G.; Rinklebe, J. Field Trials of Phytomining and Phytoremediation: A Critical Review of Influencing Factors and Effects of Additives. *Crit. Rev. Environ. Sci. Technol.* **2019**, *50*, 2724–2774. [[CrossRef](#)]
108. Suman, J.; Uhlik, O.; Viktorova, J.; Macek, T. Phytoextraction of Heavy Metals: A Promising Tool for Clean-Up of Polluted Environment? *Front. Plant Sci.* **2018**, *9*, 1476. [[CrossRef](#)]
109. Yan, A.; Wang, Y.; Tan, S.N.; Mohd Yusof, M.L.; Ghosh, S.; Chen, Z. Phytoremediation: A Promising Approach for Revegetation of Heavy Metal-Polluted Land. *Front. Plant Sci.* **2020**, *11*, 359. [[CrossRef](#)]
110. Igiri, B.E.; Okoduwa, S.I.R.; Idoko, G.O.; Akabuogu, E.P.; Adeyi, A.O.; Ejiogu, I.K. Toxicity and Bioremediation of Heavy Metals Contaminated Ecosystem from Tannery Wastewater: A Review. *J. Toxicol.* **2018**, *2018*, 1–16. [[CrossRef](#)]
111. Matinian, N.N.; Gusareva, A.L.; Bakhmatova, K.A.; Sheshukova, A.A. Microbiological Indicators and Heavy Metal Concentration in Ecological Assessment of Urban Soils of Saint Petersburg, Russia. *Geogr. Environ. Sustain.* **2020**, *13*, 214–223. [[CrossRef](#)]
112. Priyadarshane, M.; Das, S. Biosorption and Removal of Toxic Heavy Metals by Metal Tolerating Bacteria for Bioremediation of Metal Contamination: A Comprehensive Review. *J. Environ. Chem. Eng.* **2021**, *9*, 104686. [[CrossRef](#)]
113. Ayangbenro, A.; Babalola, O. A New Strategy for Heavy Metal Polluted Environments: A Review of Microbial Biosorbents. *Int. J. Environ. Res. Public Health* **2017**, *14*, 94. [[CrossRef](#)]
114. Gong, Y.; Zhao, D.; Wang, Q. An Overview of Field-Scale Studies on Remediation of Soil Contaminated with Heavy Metals and Metalloids: Technical Progress over the Last Decade. *Water Res.* **2018**, *147*, 440–460. [[CrossRef](#)] [[PubMed](#)]
115. Zhao, C.; Dong, Y.; Feng, Y.; Li, Y.; Dong, Y. Thermal Desorption for Remediation of Contaminated Soil: A Review. *Chemosphere* **2019**, *221*, 841–855. [[CrossRef](#)] [[PubMed](#)]
116. Liang, W.; Wang, G.; Peng, C.; Tan, J.; Wan, J.; Sun, P.; Li, Q.; Ji, X.; Zhang, Q.; Wu, Y.; et al. Recent Advances of Carbon-Based Nano Zero Valent Iron for Heavy Metals Remediation in Soil and Water: A Critical Review. *J. Hazard. Mater.* **2022**, *426*, 127993. [[CrossRef](#)] [[PubMed](#)]
117. Hu, W.; Niu, Y.; Zhu, H.; Dong, K.; Wang, D.; Liu, F. Remediation of Zinc-Contaminated Soils by Using the Two-Step Washing with Citric Acid and Water-Soluble Chitosan. *Chemosphere* **2021**, *282*, 131092. [[CrossRef](#)]
118. Wang, D.; Li, G.; Qin, S.; Tao, W.; Gong, S.; Wang, J. Remediation of Cr(VI)-Contaminated Soil Using Combined Chemical Leaching and Reduction Techniques Based on Hexavalent Chromium Speciation. *Ecotoxicol. Environ. Saf.* **2021**, *208*, 111734. [[CrossRef](#)]
119. Wang, J.; Hou, L.; Yao, Z.; Jiang, Y.; Xi, B.; Ni, S.; Zhang, L. Aminated Electrospun Nanofiber Membrane as Permeable Reactive Barrier Material for Effective In-Situ Cr(VI) Contaminated Soil Remediation. *Chem. Eng. J.* **2021**, *406*, 126822. [[CrossRef](#)]
120. Jeyakumar, P.; Debnath, C.; Vijayaraghavan, R.; Muthuraj, M. Trends in Bioremediation of Heavy Metal Contaminations. *Environ. Eng. Res.* **2022**, *28*, 220630–220631. [[CrossRef](#)]
121. Medfu Tarekegn, M.; Zewdu Salilih, F.; Ishetu, A.I. Microbes Used as a Tool for Bioremediation of Heavy Metal from the Environment. *Cogent Food Agric.* **2020**, *6*, 1783174. [[CrossRef](#)]
122. Sreedevi, P.R.; Suresh, K.; Jiang, G. Bacterial Bioremediation of Heavy Metals in Wastewater: A Review of Processes and Applications. *J. Water Process Eng.* **2022**, *48*, 102884. [[CrossRef](#)]
123. Jasrotia, S.; Kansal, A.; Mehra, A. Performance of Aquatic Plant Species for Phytoremediation of Arsenic-Contaminated Water. *Appl. Water Sci.* **2015**, *7*, 889–896. [[CrossRef](#)]
124. He, Y.; Li, C.; Sun, Z.; Zhang, W.; He, J.; Zhao, Y.; Xu, Z.; Zhao, W. *Penicillium* Spp. XK10, Fungi with Potential to Repair Cadmium and Antimony Pollution. *Appl. Sci.* **2023**, *13*, 1228. [[CrossRef](#)]

125. Wang, M.; Xu, Z.; Huang, Y.; Dong, B. Static Magnetic Field Enhances *Cladosporium* Sp. XM01 Growth and Fungal Mn(II) Oxidation. *J. Hazard. Mater.* **2022**, *437*, 129332. [[CrossRef](#)] [[PubMed](#)]
126. Wang, R.; Fan, X.-W.; Li, Y.-Z. Efficient Removal of a Low Concentration of Pb(II), Fe(III) and Cu(II) from Simulated Drinking Water by Co-Immobilization between Low-Dosages of Metal-Resistant/Adapted Fungus *Penicillium Janthinillum* and Graphene Oxide and Activated Carbon. *Chemosphere* **2022**, *286*, 131591. [[CrossRef](#)]
127. Rusu, L.; Grigoraş, C.-G.; Simion, A.-I.; Suceveanu, E.-M.; Istrate, B.; Harja, M. Biosorption Potential of Microbial and Residual Biomass of *Saccharomyces Pastorianus* Immobilized in Calcium Alginate Matrix for Pharmaceuticals Removal from Aqueous Solutions. *Polymers* **2022**, *14*, 2855. [[CrossRef](#)] [[PubMed](#)]
128. Khan, I.; Aftab, M.; Shakir, S.; Ali, M.; Qayyum, S.; Rehman, M.U.; Haleem, K.S.; Touseef, I. Mycoremediation of Heavy Metal (Cd and Cr)–Polluted Soil through Indigenous Metallotolerant Fungal Isolates. *Environ. Monit. Assess.* **2019**, *191*. [[CrossRef](#)]
129. Kumar, V.; Pandita, S.; Singh Sidhu, G.P.; Sharma, A.; Khanna, K.; Kaur, P.; Bali, A.S.; Setia, R. Copper Bioavailability, Uptake, Toxicity and Tolerance in Plants: A Comprehensive Review. *Chemosphere* **2021**, *262*, 127810. [[CrossRef](#)] [[PubMed](#)]
130. Zhang, D.; Yin, C.; Abbas, N.; Mao, Z.; Zhang, Y. Multiple Heavy Metal Tolerance and Removal by an Earthworm Gut Fungus *Trichoderma Brevicompactum* QYCD-6. *Sci. Rep.* **2020**, *10*, 6940. [[CrossRef](#)]
131. Chen, S.H.; Cheow, Y.L.; Ng, S.L.; Ting, A.S.Y. Mechanisms for Metal Removal Established via Electron Microscopy and Spectroscopy: A Case Study on Metal Tolerant Fungi *Penicillium Simplicissimum*. *J. Hazard. Mater.* **2019**, *362*, 394–402. [[CrossRef](#)]
132. Tansengco, M.; Tejano, J.; Coronado, F.; Gacho, C.; Barcelo, J. Heavy Metal Tolerance and Removal Capacity of *Trichoderma* Species Isolated from Mine Tailings in Itogon, Benguet. *Environ. Nat. Resour. J.* **2018**, *16*, 39–57.
133. Melo, A.; Quintelas, C.; Ferreira, E.C.; Mesquita, D.P. The Role of Extracellular Polymeric Substances in Micropollutant Removal. *Front. Chem. Eng.* **2022**, *4*, 778469. [[CrossRef](#)]
134. Imron, M.F.; Kurniawan, S.B.; Abdullah, S.R.S. Resistance of Bacteria Isolated from Leachate to Heavy Metals and the Removal of Hg by *Pseudomonas Aeruginosa* Strain FZ-2 at Different Salinity Levels in a Batch Biosorption System. *Sustain. Environ. Res.* **2021**, *31*, 14. [[CrossRef](#)]
135. Jalilvand, N.; Akhgar, A.; Alikhani, H.A.; Rahmani, H.A.; Rejali, F. Removal of Heavy Metals Zinc, Lead, and Cadmium by Biomineralization of Urease-Producing Bacteria Isolated from Iranian Mine Calcareous Soils. *J. Soil Sci. Plant Nutr.* **2019**, *20*, 206–219. [[CrossRef](#)]
136. Goma, E.Z.; El-Meihy, R.M. Bacterial Biosurfactant from *Citrobacter Freundii* MG812314.1 as a Bioremoval Tool of Heavy Metals from Wastewater. *Bull. Natl. Res. Cent.* **2019**, *43*, 69.
137. Mulligan, C.N.; Yong, R.N.; Gibbs, B.F. Heavy Metal Removal from Sediments by Biosurfactants. *J. Hazard. Mater.* **2001**, *85*, 111–125. [[CrossRef](#)]
138. Bhattacharya, A.; Gupta, A. Evaluation of *Acinetobacter* Sp. B9 for Cr (VI) Resistance and Detoxification with Potential Application in Bioremediation of Heavy-Metals-Rich Industrial Wastewater. *Environ. Sci. Pollut. Res.* **2013**, *20*, 6628–6637. [[CrossRef](#)]
139. Saranya, K.; Sundaramanickam, A.; Shekhar, S.; Swaminathan, S.; Balasubramanian, T. Bioremediation of Mercury by *Vibrio Fluvialis* Screened from Industrial Effluents. *Biomed Res. Int.* **2017**, *2017*, 6509648. [[CrossRef](#)]
140. Hamdan, A.M.; Abd-El-Mageed, H.; Ghanem, N. Biological Treatment of Hazardous Heavy Metals by *Streptomyces Rochei* ANH for Sustainable Water Management in Agriculture. *Sci. Rep.* **2021**, *11*, 9314. [[CrossRef](#)]
141. Hassan, A.; Pariatamby, A.; Ossai, I.C.; Hamid, F.S. Bioaugmentation Assisted Mycoremediation of Heavy Metal and/Metalloid Landfill Contaminated Soil Using Consortia of Filamentous Fungi. *Biochem. Eng. J.* **2020**, *157*, 107550. [[CrossRef](#)]
142. Diep, P.; Mahadevan, R.; Yakunin, A.F. Heavy Metal Removal by Bioaccumulation Using Genetically Engineered Microorganisms. *Front. Bioeng. Biotechnol.* **2018**, *6*, 157. [[CrossRef](#)]
143. Khilji, S.A.; Aqeel, M.; Maqsood, M.F.; Khalid, N.; Tufail, A.; Sajid, Z.A.; Al-Surhane, A.A.; Hashem, M.; Alamri, S.; Al-Mutairi, K.A.; et al. *Hemarthria Compressa*—*Aspergillus Niger*—*Trichoderma Pseudokoningii* Mediated Trilateral Perspective for Bioremediation and Detoxification of Industrial Paper Sludge. *Sustainability* **2021**, *13*, 12266. [[CrossRef](#)]
144. El-Sayed, M.T.; Ezzat, S.M.; Taha, A.S.; Ismaiel, A.A. Iron Stress Response and Bioaccumulation Potential of Three Fungal Strains Isolated from Sewage-Irrigated Soil. *J. Appl. Microbiol.* **2022**, *132*, 1936–1953. [[CrossRef](#)]
145. Geetha, N.; Bhavya, G.; Abhijith, P.; Shekhar, R.; Dayananda, K.; Jogaiah, S. Insights into Nanomycoremediation: Secretomics and Mycogenic Biopolymer Nanocomposites for Heavy Metal Detoxification. *J. Hazard. Mater.* **2021**, *409*, 124541. [[CrossRef](#)] [[PubMed](#)]
146. Sharma, R.; Jasrotia, T.; Sharma, S.; Sharma, M.; Kumar, R.; Vats, R.; Kumar, R.; Umar, A.; Akhtar, M.S. Sustainable Removal of Ni(II) from Waste Water by Freshly Isolated Fungal Strains. *Chemosphere* **2021**, *282*, 130871. [[CrossRef](#)] [[PubMed](#)]
147. Kumar, V.; Dwivedi, S.K. Mycoremediation of Heavy Metals: Processes, Mechanisms, and Affecting Factors. *Environ. Sci. Pollut. Res.* **2021**, *28*, 10375–10412. [[CrossRef](#)]
148. Sundararaju, S.; Manjula, A.; Kumaravel, V.; Muneeswaran, T.; Vennila, T. Biosorption of Nickel Ions Using Fungal Biomass *Penicillium* Sp. MRF1 for the Treatment of Nickel Electroplating Industrial Effluent. *Biomass Convers. Biorefinery* **2020**, *12*, 1059–1068. [[CrossRef](#)]
149. Coelho, E.; Reis, T.A.; Cotrim, M.; Mullan, T.K.; Renshaw, J.; Rizzutto, M.; Corrêa, B. *Talaromyces Amestolkiae* Uses Organic Phosphate Sources for the Treatment of Uranium-Contaminated Water. *BioMetals* **2022**, *35*, 335–348. [[CrossRef](#)] [[PubMed](#)]
150. Luo, D.; Qiang, S.; Geng, R.; Shi, L.; Song, J.; Fan, Q. Mechanistic Study for Mutual Interactions of Pb²⁺ and *Trichoderma Viride*. *Ecotoxicol. Environ. Saf.* **2022**, *233*, 113310. [[CrossRef](#)]

151. Schaefer, S.; Steudtner, R.; Hübner, R.; Krawczyk-Bärsch, E.; Merroun, M.L. Effect of Temperature and Cell Viability on Uranium Biomineralization by the Uranium Mine Isolate *Penicillium simplicissimum*. *Front. Microbiol.* **2021**, *12*, 802926. [[CrossRef](#)]
152. Saeid, A.; Cepoi, L.; Jastrzębska, M.; Nomngongo, P.N. Bioremediation of Heavy Metal Ions Contaminated Soil. *Soil Bioremediation* **2021**, 87–114.
153. Husna; Hussain, A.; Shah, M.; Hamayun, M.; Qadir, M.; Iqbal, A. Heavy Metal Tolerant Endophytic Fungi *Aspergillus Welwitschiae* Improves Growth, Ceasing Metal Uptake and Strengthening Antioxidant System in *Glycine max* L. *Environ. Sci. Pollut. Res.* **2021**, *29*, 15501–15515. [[CrossRef](#)]
154. Pagliaccia, B.; Carretti, E.; Severi, M.; Berti, D.; Lubello, C.; Lotti, T. Heavy Metal Biosorption by Extracellular Polymeric Substances (EPS) Recovered from Anammox Granular Sludge. *J. Hazard. Mater.* **2022**, *424*, 126661. [[CrossRef](#)] [[PubMed](#)]
155. Sivashankar, R.; Sathya, A.B.; Kanimozhi, J.; Deepanraj, B. Characterization of the Biosorption Process. In *Biosorption for Wastewater Contaminants*; John Wiley & Sons: Hoboken, NJ, USA, 2021; pp. 102–116.
156. Bouabidi, Z.B.; El-Naas, M.H.; Zhang, Z. Immobilization of Microbial Cells for the Biotreatment of Wastewater: A Review. *Environ. Chem. Lett.* **2018**, *17*, 241–257. [[CrossRef](#)]
157. Kamizela, T.; Worwag, M. Processing of Water Treatment Sludge by Bioleaching. *Energies* **2020**, *13*, 6539. [[CrossRef](#)]
158. Maity, J.P.; Chen, G.-S.; Huang, Y.-H.; Sun, A.-C.; Chen, C.-Y. Ecofriendly Heavy Metal Stabilization: Microbial Induced Mineral Precipitation (MIMP) and Biomineralization for Heavy Metals within the Contaminated Soil by Indigenous Bacteria. *Geomicrobiol. J.* **2019**, *36*, 612–623. [[CrossRef](#)]
159. Kumbhar, P.; Savla, N.; Banerjee, S.; Mathuriya, A.S.; Sarkar, A.; Khilari, S.; Jadhav, D.A.; Pandit, S. Microbial Electrochemical Heavy Metal Removal: Fundamental to the Recent Development. *Wastewater Treat.* **2021**, 521–542. [[CrossRef](#)]
160. Tian, Y.; Zhu, Q.; Yuan, J.; Kneepkens, R.; Yue, Y.; Zhang, C. Direct Embryotoxicity of Chromium (III) Exposure during Preimplantation Development. *J. Reprod. Dev.* **2021**, *67*, 283–290. [[CrossRef](#)]
161. Lim, S.S.; Fontmorin, J.-M.; Pham, H.T.; Milner, E.; Abdul, P.M.; Scott, K.; Head, I.; Yu, E.H. Zinc Removal and Recovery from Industrial Wastewater with a Microbial Fuel Cell: Experimental Investigation and Theoretical Prediction. *Sci. Total Environ.* **2021**, *776*, 145934. [[CrossRef](#)]
162. Ezziat, L.; Elabed, A.; Ibensouda, S.; El Abed, S. Challenges of Microbial Fuel Cell Architecture on Heavy Metal Recovery and Removal From Wastewater. *Front. Energy Res.* **2019**, *7*, 1. [[CrossRef](#)]
163. Gustave, W.; Yuan, Z.-F.; Li, X.; Ren, Y.-X.; Feng, W.-J.; Shen, H.; Chen, Z. Mitigation Effects of the Microbial Fuel Cells on Heavy Metal Accumulation in Rice (*Oryza sativa* L.). *Environ. Pollut.* **2020**, *260*, 113989. [[CrossRef](#)]
164. Wang, L.; Xu, D.; Zhang, Q.; Liu, T.; Tao, Z. Simultaneous Removal of Heavy Metals and Bioelectricity Generation in Microbial Fuel Cell Coupled with Constructed Wetland: An Optimization Study on Substrate and Plant Types. *Environ. Sci. Pollut. Res.* **2021**, *29*, 768–778. [[CrossRef](#)]
165. Zhang, J.; Cao, X.; Wang, H.; Long, X.; Li, X. Simultaneous Enhancement of Heavy Metal Removal and Electricity Generation in Soil Microbial Fuel Cell. *Ecotoxicol. Environ. Saf.* **2020**, *192*, 110314. [[CrossRef](#)] [[PubMed](#)]
166. Kabutey, F.T.; Antwi, P.; Ding, J.; Zhao, Q.; Quashie, F.K. Enhanced Bioremediation of Heavy Metals and Bioelectricity Generation in a Macrophyte-Integrated Cathode Sediment Microbial Fuel Cell (MSMFC). *Environ. Sci. Pollut. Res.* **2019**, *26*, 26829–26843. [[CrossRef](#)] [[PubMed](#)]
167. Adekunle, A.; Gomez Vidales, A.; Woodward, L.; Tartakovsky, B. Microbial Fuel Cell Soft Sensor for Real-Time Toxicity Detection and Monitoring. *Environ. Sci. Pollut. Res.* **2020**, *28*, 12792–12802. [[CrossRef](#)] [[PubMed](#)]
168. Wei, Z.; Tang, M.; Huang, Z.; Jiao, H. Mercury Removal from Flue Gas Using Nitrate as an Electron Acceptor in a Membrane Biofilm Reactor. *Front. Environ. Sci. Eng.* **2021**, *16*, 20. [[CrossRef](#)]
169. Khosravi, A.; Javdan, M.; Yazdanpanah, G.; Malakootian, M. Removal of Heavy Metals by *Escherichia coli* (*E. coli*) Biofilm Placed on Zeolite from Aqueous Solutions (Case Study: The Wastewater of Kerman Bahonar Copper Complex). *Appl. Water Sci.* **2020**, *10*, 167. [[CrossRef](#)]
170. Henagamage, A.P.; Peries, C.M.; Seneviratne, G. Fungal-Bacterial Biofilm Mediated Heavy Metal Rhizo-Remediation. *World J. Microbiol. Biotechnol.* **2022**, *38*, 85. [[CrossRef](#)]
171. Grujić, S.; Vasić, S.; Radojević, I.; Čomić, L.; Ostojić, A. Comparison of the *Rhodotorula mucilaginosa* Biofilm and Planktonic Culture on Heavy Metal Susceptibility and Removal Potential. *Water Air Soil Pollut.* **2017**, *228*, 73. [[CrossRef](#)]
172. Rikta, S.Y. Application of Nanoparticles for Disinfection and Microbial Control of Water and Wastewater. *Nanotechnol. Water Wastewater Treat.* **2019**, 159–176.
173. Shahpiri, A.; Mohammadzadeh, A. Mercury Removal by Engineered *Escherichia coli* Cells Expressing Different Rice Metallothionein Isoforms. *Ann. Microbiol.* **2018**, *68*, 145–152. [[CrossRef](#)]
174. Yang, J.; Hou, B.; Wang, J.; Tian, B.; Bi, J.; Wang, N.; Li, X.; Huang, X. Nanomaterials for the Removal of Heavy Metals from Wastewater. *Nanomaterials* **2019**, *9*, 424. [[CrossRef](#)]
175. Shehata, M.; Yamazaki, K. *ichi* Using Recombinant *E. coli* Displaying Surface Heavy Metal Binding Proteins for Removal of Pb²⁺ from Contaminated Water. *J. Bioremediation Biodegrad.* **2018**, *9*. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.