

Original Research Article

A Co-relative Study of ADA and CYFRA 21-1 in Serum and Pleural Effusion Secondary to Tuberculosis and Cancer

Sumeru Samanta^{*1}, Ashish Sharma², Biswajit Das³, Ayaz K Mallick⁴, Amit Kumar⁵¹Assistant Professor and Research Fellow, Department of Biochemistry, Rohilkhand Medical College and Hospital, Bareilly, Uttar Pradesh, India.²Associate Professor and Guide, Department of Biochemistry, Geetanjali Medical College and Hospital, Geetanjali University, Udaipur, Rajasthan, India³Professor, Co-Guide and HOD, Department of Biochemistry, Rohilkhand Medical College and Hospital, Bareilly, Uttar Pradesh, India.⁴Associate Professor, Department of Biochemistry, Rohilkhand Medical College and Hospital, Bareilly, Uttar Pradesh, India.⁵Associate Professor, Department of Pulmonary Medicine, Rohilkhand Medical College and Hospital, Bareilly, Uttar Pradesh, India.

*Corresponding author

Sumeru Samanta

Email: sumeru_samanta@yahoo.co.in

Abstract: Lung cancer and Pulmonary Tuberculosis are two major public health problems associated with significant morbidity and mortality in India. Wrong diagnosis of lung cancer cases as pulmonary tuberculosis delays the onset of anti-cancer chemotherapy and initiation of DOTS thus increases complication in malignancy patients. In this context easy, cost effective diagnostic tool at primary level must be the priority and need of hour. This study was done to evaluate any significance of ADA, CYFRA 21-1 in serum and pleural effusion secondary to tuberculosis and lung cancer. Case control study was carried out on 100 cases of tuberculous effusion, 50 cases of malignant effusion and 100 age and gender matched apparently healthy controls. Correlation between ADA and CYFRA 21-1 was evaluated to find any significance between three groups. Blood and pleural fluid samples were collected and analyzed by using Erba Mannheim Chem 5 plus V2 semi autoanalyzer and LISA SCANII Elisa reader. Statistical analysis was done by using ANOVA and student's 't' test. P value <0.05 was considered significant. ADA levels in serum and pleural fluid was significantly higher in pulmonary TB group than lung cancer group ($p < 0.001$) and both are higher than control group ($p < 0.001$). CYFRA 21-1 in serum and pleural fluid was significantly higher in lung cancer group than TB group ($p < 0.001$) but both were higher than control group ($p < 0.001$). The results suggests early quantization of these parameters can differentiate pulmonary tuberculosis from lung cancer and thus can decrease the mortality rate of lung cancer cases though more extensive study with increased sample size may provide more insights.

Keywords: Differential diagnosis, malignant pleural effusion, tuberculous effusion, ADA, CYFRA 21-1.

INTRODUCTION

The etiological diagnosis of exudative pleural effusion poses a significant dilemma in clinical practice, especially in terms of the differentiation between malignant and benign pleural effusion as there is significant difference in treatment and prognosis [1].

Statistics for India revealed 63,000 new lung cancer cases are reported each year [2] which was once considered to be rare [3] and now responsible for 1.38 million deaths worldwide out of which significant contributor is also India [4].

Tuberculosis is another cause of exudative pleuritis and is most major health problems associated with significant morbidity and mortality in India along with lung cancer. In 2012, India declared tuberculosis to be notifiable disease [5] and it is the highest tuberculosis burden country with WHO statistics revealed 2.2 million cases out of global 8.7 million cases so approximately 40% of Indian population is infected with *Mycobacterium tuberculosis* bacteria, the vast majority of whom have latent than active tuberculosis [6,7]. So, any patient arrives with

predominantly lymphocytic exudative pleural effusion, the suspicion is targeted towards diagnosis of malignant or tuberculous pleuritis primarily. Diagnostic dilemma happens as traditional methods of diagnosis tuberculosis fail to recognise it [8] moreover, microbiological results reported so late that decisions regarding management of the symptoms are already taken and initiated which causes wrong diagnosis and management of lung cancer as pulmonary tuberculosis. Most often to deal with this diagnostic dilemma combination of pleural biopsy culture and histology is done which increases diagnostic chances up to 90% in tuberculous pleuritis [9] but it is an invasive approach and frequently requires more than six samples [10]. Lung cancer accounts to 68% of all cases of malignant pleural effusion [11], the diagnosis is mostly done by cytopathological study of pleural fluid but sensitivity is only 50%. It can be increased to 80% if needle biopsy of pleura is performed [12]. Due to such low sensitivity sometimes the results are inconclusive and thus thoracoscopy is done to identify the type and loci of malignancy [13].

Because diagnosis of these two common causes of pleural effusion which is often having similar biochemical profiles and predominance of lymphocytes with other diagnostic difficulties already discussed can delay or misdiagnose a case of lung cancer as sputum negative pulmonary tuberculosis is very high and often these patients presumptively treated with anti-tubercular drugs not only delays the diagnosis of lung cancer but also causes progression of the disease to stage IIIB or IV by that time they are beyond the scope of curative resection [14,15]. In this context, the objective of our present study were to describe the characteristics and laboratory performances of ADA and CYFRA 21-1 in serum and pleural effusion of patients suffering from tuberculosis and lung cancer as these are non invasive as well as in expensive test and can be performed in primary health care setup.

Adenosine deaminase (ADA; EC 3.5.4.4) is an enzyme required for converting adenosine to inosine, a stage in purine catabolism. Since 1978, when ADA activity was found high in tuberculous pleural exudates as well as in serum [16], since then activity of total ADA has been used to diagnose tuberculous pleural effusion but degrees of sensitivity and specificity varies in different study [17 – 21].

CYFRA 21-1 is a cytokeratin-19 fragment; an acid type of cytoplasmic protein having molecular weight of 40 KD is a major component of cytoskeleton intermediate filaments of simple epithelial cells and is over expressed in various carcinomas. Following cell death it is released in the serum in the form of soluble fragments [22 – 26]. CYFRA 21-1 is a potential marker for malignant pleural effusion and is not only found in serum but also present in pleural fluid [27]. So we have

included these two parameters to find out its efficacy for differential diagnosis between malignant pleural effusion and tuberculous pleural effusion.

MATERIALS AND METHODS

This case control study was conducted in the Department of Biochemistry from October 2014 till January 2016 in collaboration with Department of Pulmonary Medicine, Rohilkhand medical college and hospital, Bareilly, Uttar Pradesh. Ethical clearance was procured from Institutional Ethical Committee with vide reference no. IEC/64/2014.

We have taken 100 cases of diagnosed tuberculous effusion, 50 cases of lung cancer who were earlier considered as smear negative pulmonary tuberculosis cases and administered DOTS in primary health centre later referred to Department of Pulmonary Medicine as complication started and 100 cases of age and sex matched apparently healthy controls (who appeared for general health check up in study age and sex group).

We have only considered exudative pleural effusion cases as per Light's criteria i.e. (a) Pleural fluid/Serum total protein ratio > 0.5, (b) Pleural fluid/serum LDH ratio > 0.6, (c) Pleural Fluid LDH > 200.0 IU/L[28] and Roth *et al.* i.e. Serum-Pleural Effusion Albumin Gradient of ≤ 1.2 gm/dL suggests exudates and > 1.2 gm/dL suggests transudates [29].

Standardised Diagnostic Criteria for Tuberculosis were:

- Pleural biopsy demonstrating a granulomatous process.
- Detection of Mycobacterium tuberculosis in pleural fluid or tissue by Z-N staining.
- A compatible clinical history and radiological examination, in patients with a lymphocytic exudates and ADA levels higher than 24 IU/L as well as favourable clinical evaluation after specific treatment.

Standardised Diagnostic Criteria for Lung Cancer:

- Finding of neoplastic cells in pleural fluid or tissues obtained by needle biopsy.
- CT scan of thorax.
- In inconclusive cases, diagnosis was established by thoracoscopy guided biopsy or surgery.

The following patients were excluded from our study:

- Pleural exudates other than tuberculosis and lung cancer.
- Other cases of cancer.
- Chronic diseases like DM, hypertension etc.
- Any liver, renal and muscular disorders.
- Known HIV positive cases.

The following parameters were evaluated i.e. Adenosine deaminase (ADA) and Cytokeratin fragment CYFRA 21-1 in serum and pleural effusion secondary to tuberculosis and lung cancer. Evaluation of ADA was done in Erba Chem 5 plus V2 semi auto analyzer by enzymatic kinetic ADAZYME method procured from Tulip Diagnostics and CYFRA 21-1 was done by sandwich ELISA method procured from Elabscience Biotechnology and reading was taken by LISA SCANII ELISA reader and automated ELISA washer by Erba Mannheim.

After taking informed consent from patients pleural fluid was collected by thoracocentesis done by Department of pulmonary medicine and 4 mL blood was collected in serum separation tube (SST) by venipuncture under aseptic condition. Serum was separated after allowing the blood to stand for 30 min at room temperature and then centrifuged at 2000 rpm for 5 min. Fresh samples were used for our study.

Data was presented as mean ± SD, comparison between cases (TB and Lung cancer group) was done by Unpaired student's 't' test. Significance between three groups (TB, Lung cancer and Control groups) was calculated by using one way ANOVA. P value <0.05 was considered as statistically significant. Statistical analysis and ROC curve analysis was done by using licensed SPSS 16.0 software (SPSS Inc., Chicago, IL, USA) for windows. We have used licensed MS Excel software to present and prepare charts and graphs for our present study data.

RESULT

The demographic distribution of our study population for the pulmonary tuberculosis and lung cancer group is shown in Table-1. Patients with pulmonary tuberculosis were significantly of lower age group (mean ±SD for males 45.46±13.525; for females 40.918±13.781) than lung cancer group (for males 58.186±8.764; for females 62.428±7.656) with 'p' value <0.0001.

The biochemical analysis of serum and pleural fluid ADA in pulmonary tuberculosis and lung cancer is shown in Table-2 which shows concentration of serum and pleural fluid ADA was significantly higher in TB group (mean ± SD of serum ADA 33.731±7.355 IU/L; pleural fluid ADA 107.08±23.09 IU/L) than that of lung cancer group (mean ±SD of serum ADA 16.206±6.356 IU/L; pleural fluid ADA 31.828±11.913 IU/L) when compared by unpaired student's 't' test 'p' value were <0.001 for both serum as well as pleural fluid values but both were significantly higher than control group (mean±SD of serum ADA 5.512±1.862 IU/L) with 'p' value < 0.001 (for both TB and Lung cancer group when compared with control group). ANOVA of serum ADA in TB group, Lung cancer group and Control group was 'p' value 0.00 and F value 647.8327. When plotted in ROC curve in TB vs. Lung cancer the best cut off values for serum ADA was 20.5 IU/L (sensitivity 98%, specificity 86%) shown in Fig-1. For pleural effusion the best cut off values was 59.7 IU/L (sensitivity 99%, specificity 98%) presented as Fig-2.

The biochemical analysis of serum and pleural fluid CYFRA 21-1 is shown in Table-3 which shows serum CYFRA 21-1 was significantly higher in Lung cancer group (mean±SD; 14.004± 10.578 ng/mL) than pulmonary tuberculosis group (mean±SD 1.6951±0.553 ng/mL) with 'p' value < 0.001. But both were higher than serum values in control group (0.976±0.421 ng/mL). ANOVA of serum CYFRA 21-1 in TB group, Lung cancer group and control group was 'p' value 0.00 and F value 143.9277. When plotted in ROC curve, the most probable cut off value of serum CYFRA 21-1 was 2.99 ng/mL (sensitivity 100%, specificity 100%) which is presented as Fig-3..

Pleural fluid CYFRA 21-1 was also significantly higher in lung cancer group (mean±SD; 79.918±34.973) than TB group (11.486±4.798 ng/mL) with 'p' value < 0.001. When plotted in ROC curve the most probable cut off value was 23.15 ng/mL (sensitivity 100% and specificity 100%) which is shown in Fig-4.

Table-1: Demographic Distribution of study population

	Pulmonary Tuberculosis group	Lung Cancer Group	Control Group
Male	63	43	63
Female	37	7	37
Mean Female age (yrs)	40.918 ±13.781	62.428±7.656	40.287±13.757
Mean Male age (yrs)	45.46±13.525	58.186 ±8.764	45.662±13.635

Table-2: Values of ADA (IU/L) in TB, Lung cancer and Control Group

Tuberculosis Group		Lung Cancer Group		Control Group
Serum	Pleural Fluid	Serum	Pleural Fluid	Serum
33.731±7.355 IU/L *vs. Lung cancer 'p' <0.001 *vs. Control 'p' <0.001	107.08±23.09 IU/L *vs. Lung cancer 'p' <0.001	16.206±6.356 IU/L *vs. TB group 'p' <0.001 *vs. Control group 'p' <0.001	31.828±11.913 IU/L *vs. TB group 'p' <0.001	5.512±1.862 IU/L
ANOVA analysis of serum ADA in TB, Lung Cancer & Control Groups				
'p' Value = 0.00, F value= 647.8327, Fcrit= 3.032361 Between groups; SS= 40282.22366, df=2, MS= 20141.11 Within groups; SS=7679.2277, df=247, MS=31.08999				

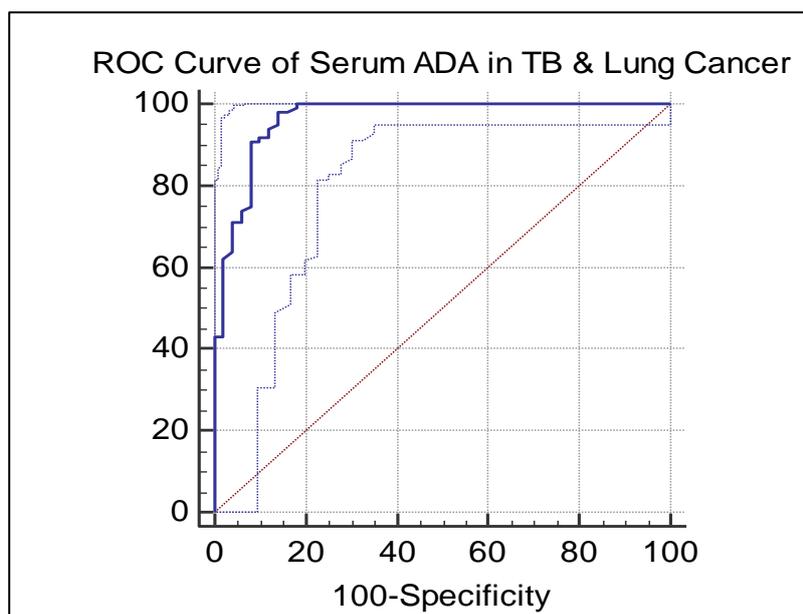


Fig-1:ROC Curve of Serum ADA in TB & Lung Cancer

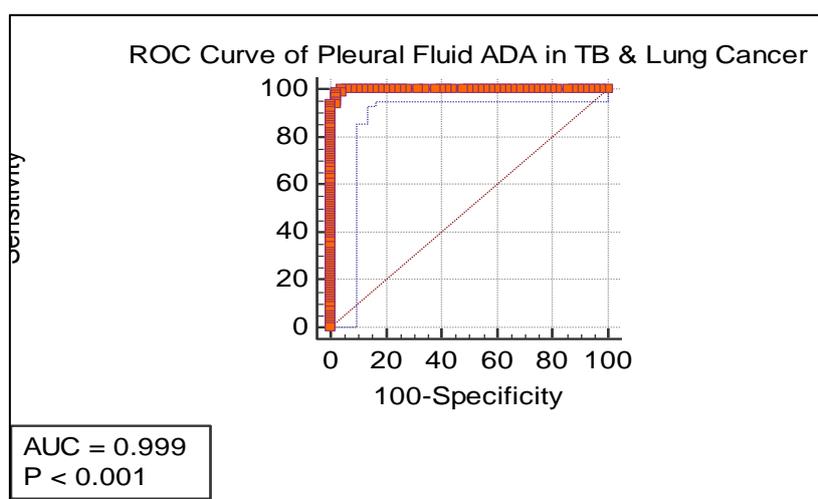


Fig-2:ROC Curve of Pleural fluid ADA in TB & Lung Cancer

Table-3: Values of CYFRA 21-1 (ng/mL) in TB, Lung Cancer and Control Groups

Tuberculosis Group		Lung Cancer Group		Control Group
Serum	Pleural Fluid	Serum	Pleural Fluid	Serum
1.6951±0.553 ng/mL *vs. Lung cancer 'p' <0.001 *vs. Control 'p' <0.001	11.486±4.798 ng/mL *vs. Lung cancer 'p' <0.001	14.004±10.578 ng/mL *vs. TB group 'p' <0.001 *vs. Control group 'p' <0.001	79.918±34.973 ng/mL *vs. TB group 'p' <0.001	0.976±0.421 ng/mL
ANOVA analysis of serum CYFRA 21-1 in TB, Lung Cancer & Control Groups				
'p' Value = 0.00, F value= 143.9277, Fcrit= 3.032361 Between groups; SS= 6445.846, df=2, MS= 3222.923 Within groups; SS=5530.987, df=247, MS=22.39266				

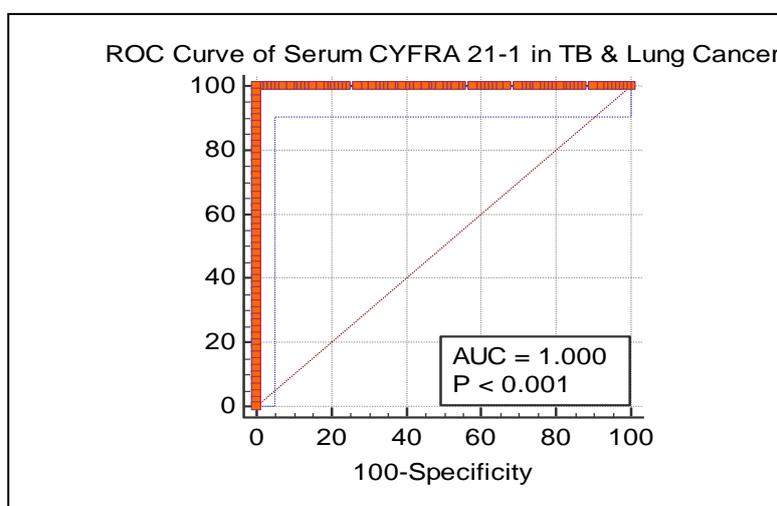


Fig-3: ROC Curve of Serum CYFRA 21-1 in TB & Lung Cancer

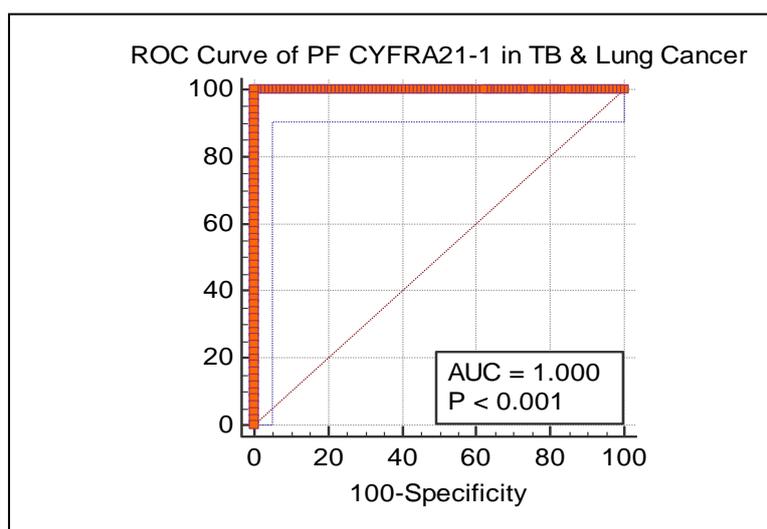


Fig-4: ROC Curve of Pleural fluid CYFRA 21-1 in TB & Lung Cancer

DISCUSSION

Pleural effusions are common complications observed in wide variety of diseases including pulmonary tuberculosis and lung cancer. Thoracoscopy

is considered as gold diagnostic standard and analysis of removed fluid is the fastest and easiest way of assessment of causes [30]. Cytological, biochemical and microbiological analysis of pleural fluid is fundamental

for adequate screening although 20% of the cases remain inconclusive [31] to differentiate benign pleural effusion from malignant pleural effusion thus more expensive examination is required which may not be bearded by our low economic group of population in both rural as well as urban areas thus increases the morbidity and mortality of the patients.

In our study with respect to gender we have found men are more predisposed to both tuberculosis and lung cancer (63% of TB cases and 86% of lung cancer cases). Tuberculosis usually predominates among younger age group (mean \pm SD; males 45.46 \pm 13.525; females 40.918 \pm 13.781) than lung cancer group (mean \pm SD; males 58.186 \pm 8.764; females 62.428 \pm 7.656) which is very similar to other global studies [32-34] although incidence of lung cancer is increasing alarmingly in females who were never smokers [35]. In our study we have got 7 such cases who were never smokers in which 4 cases are having adenocarcinoma, 2 cases of squamous cell carcinoma and 1 case of large cell carcinoma which is following similar pattern according to incidence rate as studied by Noronha V *et al.*[36].In the present study as a rule first we have analyzed and segregated exudates arising from pulmonary tuberculosis and lung cancer by Light's criteria and Roth *et al.* followed by confirmation by our diagnostic criteria and then subjected to further analysis. Serum and pleural fluid activity has been proved to be a valuable biochemical marker that has high sensitivity and specificity for TB diagnosis [37].Serum and pleural fluid ADA activity were significantly higher in TB group than lung cancer group ('p' value <0.001) but both were significantly higher than control group (mean \pm SD 5.512 \pm 1.862). The cut off value for serum ADA was 20.5 IU/L for TB group with 98% sensitivity and 86% specificity with area under ROC curve 0.965 and standard error 0.0158; for pleural fluid ADA the best cut off value was 59.7 IU/L with 99% sensitivity and 98% specificity with area under ROC curve 0.999 and standard error 0.00143 was a significant predictor for pulmonary tuberculosis cases and can differentiate TB cases from malignancy cases significantly which is in accordance to study conducted by Mo-Lung Chen *et al.* [38].Our findings contradict the study conducted by Light *et al.* [30] and Sharma *et al.*[39] who has suggested that lower ADA levels with lesser sensitivities and specificities among Asians in comparison to their European and Caucasian compatriots might compromise its usefulness in TB detection in these population. Our sensitivity and specificity were remarkably high due to improvement in diagnostic methodology as well as quality of the reagent as we have used enzymatic kinetic method instead of Giusti's method or NADH linked method. Exclusion of other increased cell mediated immune response to pathological causes like empyema; liver diseases etc. may also causes significant improvement in achieving

greater diagnostic accuracy which was reflected in the area under ROC curve and increase in sensitivity and specificity of the method.

We have evaluated the diagnostic performance of CYFRA 21-1 in serum and pleural fluid in tuberculosis and malignancies in differentiation between these two. There are wide ranges of markers for the detection of malignant pleural effusion but it lacks sufficient diagnostic accuracy in discriminating lung cancer cases from TB cases in early stage. One of the promising tumour markers is CYFRA 21-1 and it can be detected by sandwich ELISA method from both pleural fluid as well as serum [40].

Serum and pleural fluid CYFRA 21-1 is significantly higher in lung cancer group than tuberculosis group ('p' value < 0.001) but both were significantly higher than control group (mean \pm SD, CYFRA 21-1 0.976 \pm 0.421). When ROC curve analysis was done it shows cut off values of CYFRA 21-1 in serum is 2.99 ng/mL with 100% sensitivity and 100% specificity, area under ROC curve was 1.00 and standard error was 0.00. Up on ROC curve analysis of pleural fluid CYFRA 21-1 the most probable cut off value is 23.15 ng/mL with 100% sensitivity and 100% specificity. The values are in accordance with study conducted by David *et al.*[41], Li *et al.*[42], Liang *et al.*[43], Huang *et al.*[44] and Dalia H Farag *et al.*[45]. This finding could be attributed to increased Cytokeratin fragment solubility due to modification at the amino and carboxyl terminals of keratin by phosphorylation, glycosylation and transglutamination which mainly occurs during transformation of normal cells to malignant cells. Higher values with high sensitivity and specificity of CYFRA 21-1 in lung cancer cases is further caused due to proteolytic degradation of keratin during cell lysis, abnormal mitosis and tumour necrosis[44].Thus quantisation of serum and pleural fluid CYFRA 21-1 is an excellent discriminator between pulmonary tuberculous effusion and malignant pleural effusion.

CONCLUSION:

Biochemical analytes like Adenosine deaminase and Cytokeratin fragment CYFRA 21-1 levels in serum and pleural fluid is a useful and efficient tool to differentiate between two common exudative causes i.e. tuberculosis and lung cancer along with other specific test as they possess higher sensitivity and specificity. As these tests are easy, inexpensive and thus can help us in early segregation of lung cancer cases from pulmonary tuberculosis in primary health care setup and decrease mortality and morbidity significantly. The limitation of our study is limited sample size and study was conducted in a single region. Larger sample size and multi-centric studies could be done to obtain wider insights.

REFERENCE

1. Light RW. Pleural effusion related to metastatic malignancies. In: Light RW ed. *Pleural Diseases*. 4th Ed. Philadelphia: Lippincott; 2001. Pg 108-134.
2. Ganesh B, Sushama S, Monika S, Suvarna P. A case control study risk factor for lung cancer in Mumbai; *Indian Asian Pac J Cancer, Prev* 2011; 12:357-362.
3. Parkin DM, Muir CS. Cancer incidence in five continents: comparability and quality of data. IARC Sci Publication 1992; 120: 45-173.
4. Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. Estimates of worldwide burden of cancer in 2008. *GLOBOCAN 2008, Int. J of Cancer* (2010); 127: 2893-2917.
5. Sinha K. "Finally, tuberculosis declared as notifiable disease." *Times of India*, May 9th 2012.
6. *Global Tuberculosis Control 2012*. World Health Organization, Geneva, 2012.
7. Revised National Tuberculosis Control Programme (RNTCP), Govt. Of India, 2012.
8. Ferrer J. Tuberculous Pleural Effusion and Tuberculous Empyema. *Semin Respir Crit Care Med*. 2001; 22:637-646.
9. Valdes L, Alvarez D, San Jose E, Panella P, Valle JM. Tuberculous Pleurisy. A study of 254 patients. *Archives of Internal Medicine*, 1998;158:2017-2021.
10. Kirsch CM, Kroe DM, Azzi RL, Jensen WA, Kagawa FT, Wehner JH. The optimal number of pleural biopsy specimens for diagnosis of tuberculous pleurisy. *Chest*, 1997;112:702-706.
11. American Thoracic Society. Management of malignant pleural effusion. *American J Respiratory Critical Care Medicine*. 2000;162(5): 1987-2001.
12. Romero S, Fernandez C, Arriero JM, Espasa A, Candela A, Martin C. CEA, CA 15-3 and CYFRA 21-1 in serum and pleural fluid of patients with pleural effusion. *European Respiratory Journal*, 1996;9(1):17-23.
13. Lodden Kemper R. Thoracoscopy – state of art. *European Respiratory Journal*. 1998;11(1):213-221.
14. Singh VK, Chandra S, Kumar S, Pangtey G, Mohan A, Guleria R. A common medical error: Lung cancer misdiagnosed as sputum negative tuberculosis. *Asian Pac J Cancer Prev* 2009;10:335-338.
15. Mountain CF. Revision in International system for staging lung cancer. *Chest*, 1997;111:1710-1717.
16. Geicia Mouco JC. CNS tuberculosis. *Neurologic Clinics*. Marra CM Ed. 1999;17(4) : 737-760.
17. Piras MA, Gakis C, Budroni M, Andreoni G. Adenosine deaminase activity in pleural effusion: an aid to differential diagnosis. *British Medical Journal* 1978;2:1751-1752.
18. Ocana I, Martinez Vasquez JM, Ribera E. Adenosine deaminase in serum and pleural fluids: tests for diagnosis of tuberculous pleural effusion. *Chest*, 1983;84:51-53.
19. Petterson J, Ojala K, Weber TH. Adenosine deaminase in the diagnosis of pleural effusion. *Acta Med Scand*, 1984;215:299-304.
20. Fontan J, Vereza Hernando H, Garcia-Buela JP, Dominiguez Juncal L, Martin Egana MT, Montero Martinez MC. Diagnostic value of simultaneous determination of pleural adenosine deaminase and pleural lysozyme/serum lysozyme ratio in pleural effusion. *Chest* 1988; 93:303-307.
21. Lamsal M, Gautam N, Bhatta N, Majhi S, Baral N, Bhattacharya SK. Diagnostic utility of Adenosine deaminase activity in pleural fluid and serum of tuberculosis and non-tuberculosis respiratory disease patients. *South East Asian J Tropical Medicine and Public Health*; Vol. 38(2) 2007; 363-369.
22. Boden Muller H. Technical evaluation of a new automated tumor marker assay: the enzyme test CYFRA 21-1. In: Tumor associated antigen, oncogenes, receptors, cytokeratins in tumor diagnostics and therapy at the beginning of nineties. Klapdor R (Ed.) W Zuckshwerds: Munich 1992. Page 137-138.
23. Moll R, Franke WW, Schiller DL, Geiger B, Krepler R. The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. *Cell*(31)1982; 11-24.
24. Feng Yang Chun, Feng Min, Zhang Di, Huang Yan Chun. Diagnostic value of CYFRA 21-1; SCC and CEA for differentiating of early stages NSCLC from benign lung diseases. *International J Clinical and Experimental Medicine*, 2015;8(7):11295-11300.
25. Schneider J. Tumour markers in detection of lung cancer. *Adv Clin Chem*. 2006;42:1-41.
26. Lai RS, Hsu HK, Lu JY, Ger LP, Lai NS. CYFRA 21-1 enzyme linked immunosorbent assay. Evaluation as a tumor marker in NSCLC. *Chest* 1996; 109:995-1000.
27. Wagner IC, Guimaraes MJ, Da Silva LK, De Melo FM, Muniz MT. Evaluation of serum and pleural levels of tumor markers CEA, CYFRA 21-1 and CA 15-3 in patients with pleural effusion. *J Bras Pneumol*. 2007; 33(2):185-191.
28. Light RW, Mac Gregor MI, Luchsinger PC. Pleural effusion, the diagnostic separation of transudates and exudates. *Ann intern meet*, 1972;77:507.
29. Roth BJ, O Meara TF, Gragem WH. The serum – effusion albumin gradient in the evaluation of pleural effusions. *Chest* 1990; 98:546.
30. Light RW. Clinical manifestations and useful tests. In: *Pleural Diseases*. 4th Ed. Philadelphia, Lippincott-Williams and Wilkins, 2001. Pg 42-86.
31. Villena V, Lopez Encuentraz A, Echave Sustaeta J, Alvarez Martinez C, Martin Escribano P. Prospective study of 1000 consecutive patients

- with pleural effusion. Etiology of the effusion and characteristics of the patients. Archives Bronchopneumology, 2002; 38:21-26.
32. World Health Organization. URL: www.who.int/gtb/policyrd/gender&tb.htm
33. Parkin DM, Bray F. Global Cancer Statistics. 2002. CA Cancer J Clin 2005;55:74-108.
34. Antunes G, Neville E. BTS guidelines for the management of malignant pleural effusions. Thorax 2003; 58:29-38.
35. Toh CK, Gao F, Lim WT, Leong SS, Fong KW, Yap SP. Never-Smokers with lung cancer: Epidemiologic evidence of a distinct disease entity. J Clin Oncol 2006; 24: 2245-2251.
36. Noronha V, Dikshit R, Raut N, Joshi A, Pramesh CS, George K. Epidemiology of lung cancer in India: Focus on differences between non-smoker and smokers; a single centre experience. Indian J of Cancer; Jan-March 2012; Vol.49, pg 74-81.
37. Perez-rodriguez E, Walton IJP, Hernandez JJS, Pallaris E, Rubi J, Castro DJ, Diaz Nuevo G. ADA₁/ADA_p ratio in pleural tuberculosis: an excellent diagnostic parameter in pleural fluid. Respir Med 1999; 93(11):816-821.
38. Chen ML, Yu WC, Lam CW, Au KM, Kong FY. Diagnostic value of pleural adenosine deaminase activity in tuberculous pleurisy. Clin Chim Acta, 2004;341:101-107.
39. Sharma SK, Suresh V, Mohan A, Kaur P, Saha P, Kumar A. A prospective study of sensitivity and specificity of adenosine deaminase estimation in the diagnosis of tuberculous pleural effusion. Indian J Chest Dis Allied Science, 2001; 43:149-55.
40. Blok BK. Thoracocentesis. In: Roberts JR & Hedges JR (Eds.) Roberts: Clinical Procedure in Emergency medicine; 5th Ed. Saunders Elsevier, Philadelphia.2009:160-174.
41. David S, Boris Z, Ariella B, Dekel S. Diagnostic value of CYFRA 21-1, CEA, CA 19-9, CA 15-3, and CA 125 assays in pleural effusions: analysis of 116 cases and review of the literature. The Oncologists; 2005, 10(7):501-507.
42. Li C, Cheng B, Ge W, Gao F. Clinical values of CYFRA 21-1, NSE, CA 15-3, CA 19-9, and CA 125 assays in the elderly patients with pleural effusion. Int. J Clin. Practice, 2007; 61(3):444-448.
43. Liang QL, Shi HZ, Qin XJ, Liang XD, Jiang J, Yang HB. Diagnostic accuracy of tumor markers for malignant pleural effusion: a meta analysis. Thorax; 2008; 63: 35-41.
44. Huang, Wen W, Tsao, Lai, Su, Cheng C, Tseng and Chih-E. Diagnostic value of Her-2/neu, CYFRA 21-1 and CEA levels in malignant pleural effusions. Pathology, 2010; 42(3):224-228.
45. Dalia H Farag, Eman EL, Haididi, Mohammed O, EL Maraghy, Maha M Huseein. Pleural CYFRA 21-1 and CA 15-3 in differentiation of malignant from benign pleural effusion. Life Science Journal 2012; 9(3):499-505.