

Full Original Paper

Antifungal Activity Evaluation of the Constituents of *Haliclona baeri* and *Haliclona cymaeformis*, Collected from the Gulf of Thailand

Rawiwan Wattanadilok^{1,3}, Pichan Sawangwong², Cátia Rodrigues^{4,6}, Honorina Cidade^{4,6}, Madalena Pinto^{4,6}, Eugenia Pinto^{5,6}, Artur Silva⁷ and Anake Kijjoa^{3,8,*}

¹ Bangsaen Institute of Marine Science (BIMS), Burapha University, Bangsaen, Chonburi 20131, Thailand

² Department of Aquatic Science, Faculty of Sciences, Burapha University, Bangsaen, Chonburi 20131, Thailand

³ Centro Interdisciplinar de Investigação Marinha e Ambiental (CIIMAR), Universidade do Porto, Rua dos Bragas 289, 4050-123 Porto, Portugal

⁴ Laboratório de Química Orgânica, Faculdade de Farmácia, Universidade do Porto, Rua Aníbal Cunha 164, 4050-047 Porto, Portugal

⁵ Laboratório de Microbiologia, Faculdade de Farmácia, Universidade do Porto, Rua Aníbal Cunha 164, 4050-047 Porto, Portugal

⁶ Centro de Estudos de Química Orgânica, Fitoquímica e Farmacologia da Universidade do Porto (CEQOFFUP), Rua Aníbal Cunha 164, 4050-047 Porto, Portugal

⁷ Departamento de Química, Universidade de Aveiro, 4810-1933 Aveiro, Portugal

⁸ ICBAS-Instituto de Ciências Biomédicas Abel Salazar, Universidade do Porto, 4099-003 Porto, Portugal

* Author to whom correspondence should be addressed; Email: ankijjoa@icbas.up.pt

Received: 31 May 2007 / Accepted: 23 June 2007 / Published: 25 June 2007

Abstract: A new compound maleimide-5-oxime was isolated, together with 3,4-dihydroxybenzoic acid, tetillapyrone, from the ethyl acetate extract of the marine sponge *Haliclona baeri* while tetillapyrone, nortetillapyrone, *p*-hydroxybenzaldehyde and phenylacetic acid were isolated from the ethyl acetate extract of *Haliclona cymaeformis*,

collected from the Gulf of Thailand. The structures of tetillapyrone and nortetillapyrone were re-examined using HMBC correlations. Maleimide-5-oxime, tetillapyrone and nortetillapyrone were found to be inactive against three human tumor cell lines (the estrogen-dependent ER(+) MCF-7, the estrogen-independent ER(-) MDA-MB-231 and NCI-H460. Maleimide-5-oxime, *p*-hydroxybenzaldehyde, phenylacetic acid, tetillapyrone and nortetillapyrone were evaluated for their growth inhibitory effect against seven yeasts and eight filamentous fungi. Only nortetillapyrone showed antifungal activity, with a preponderance on the dermatophytic filamentous fungi.

Keywords: *Haliclona baeri*, *Haliclona cymaeformis*, tetillapyrone, nortetillapyrone, antifungal activity, dermatophytes

1. Introduction

The frequency of invasive fungal infection has risen substantially with the increasing numbers of immunocompromised patients, such as those infected with HIV, receiving cancer chemotherapy, immunosuppressive therapy, or treatment with broad-spectrum antibiotics [1]. The most frequently encountered infections are caused by the yeast *Candida albicans* and by species of filamentous fungus *Aspergillus*. Other fungal pathogens that have emerged in recent years include yeast species such as *Candida glabrata*, *C. krusei*, *C. tropicalis*, *Cryptococcus* and *Trichosporon* and filamentous fungi such as *Fusarium*, *Rhizopus* and *Rhizomucor* [2]. On the other hand, dermatophytes, which invade and multiply in keratinized tissues [3] and cause infection, have a worldwide distribution with geographical differences in the incidence and prevalence of different dermatophyte species [4] and the most common agents belong to three genera: *Epidermophyton*, *Trichophyton* and *Microsporum*. Majority of clinically used antifungals suffer from various drawbacks in terms of toxicity, efficacy and cost, and their frequent use has led to the emergence of resistant strains. Hence, there is a great demand for novel antifungal agents belonging to the wide range of structural classes, selectively acting on novel targets with fewer side effects [5].

There is mounting evidence that fungi display highly specific adaptations in the marine environment that include the production of unique secondary metabolites antibiotics [1]. The fact that marine organisms contain secondary metabolites different from their terrestrial counterparts in structure and biological activity has led to the hypothesis that marine organisms may contain efficient antifungal compounds with different modes of action and selective antifungal activity compared with human cells [6].

Sponges in the genus *Haliclona* are well-known for producing a variety of secondary metabolites, most commonly bioactive alkaloids [7] and many of which are cytotoxic [8]. Antifungal compounds isolated from *Haliclona* include haliclonadamine, an alkaloid isolated from *Haliclona* sp. from Palau [9] and alkyl amino alcohols, halaminols A, B and C from the tropical marine sponge *Haliclona* sp., collected on the Great Barrier Reef [10].

In the course of our investigation on bioactive compounds from the marine sponges from the Gulf of Thailand, we have investigated the chemical constituents of the two marine sponges *Haliclona baeri* and *Haliclona cymaeformis*, collected from the coastal area of Chonburi Province and from Koh Chang, Trad Province, Eastern Thailand, respectively. Isolated from *Haliclona baeri* were a new maleimide-5-oxime (**1**), 3,4-dihydroxybenzoic acid (**2**), tetillapyrone (**5** and **6**) while *Haliclona cymaeformis* furnished, besides tetillapyrone (**5** and **6**), *p*-hydroxybenzaldehyde (**3**), phenylacetic acid (**4**) and nortetillapyrone (**7** and **8**) (Figure 1). We have also re-examined the structures of tetillapyrone and nortetillapyrone, previously reported by our group from *Tetilla japonica* [11], as well as investigated the antifungal activity of the isolated compounds.

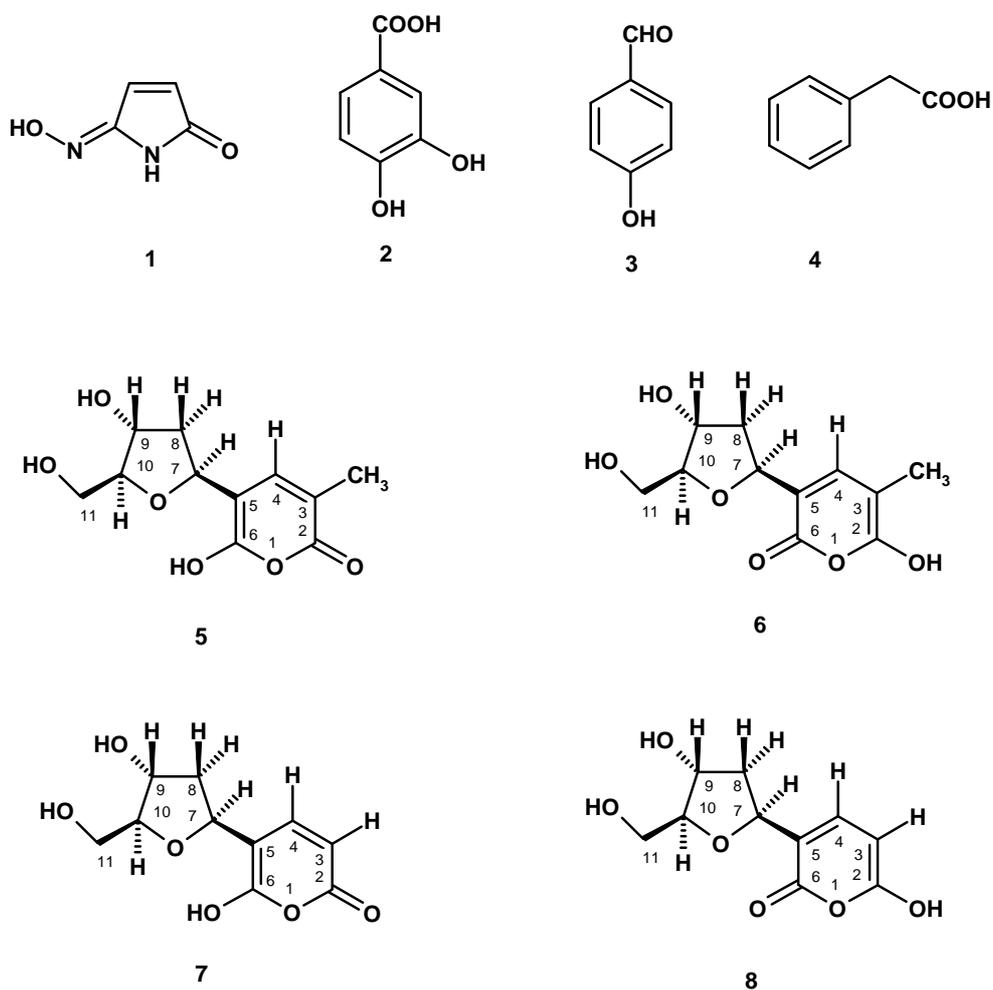


Figure 1. Compounds isolated from *Haliclona baeri* and *H. cymaeformis*.

2. Results and Discussion

We have previously reported isolation of tetillapyrone (**5**) and nortetillapyrone (**8**) from the marine sponge *Tetilla japonica*, collected from the Gulf of Thailand [11]. Though the structures of both compounds were established by ^1H , ^{13}C NMR, COSY, NOESY, HETCOR, HMBC and HRMS as well as by X-ray analysis in the case of tetillapyrone (**5**), there were still some uncertainties with respect to

the pyrone moiety. The correlations observed in the HMBC spectrum for both compounds raised some doubts about the correct position of the hydroxyl and carbonyl groups of the pyrone ring. In addition, the data obtained from the X-ray analysis did not provide any evidence if a hydrogen atom was attached to the oxygen atom on C-2 or on C-6. Besides, the positions of the hydroxyl and carbonyl groups of the pyrone ring in the structure of nortetillapyrone (**8**) were opposite to those observed in the pyrone ring of tetillapyrone (**5**). All of these evidences, combined with the new isolation of more quantities of tetillapyrone and nortetillapyrone from the marine sponges *H. cymaemormis* and *H. baeri*, have led us to re-examine their structures by careful analysis of the HMBC spectra of both compounds.

The structure of tetillapyrone (**5**), previously proposed by us [11], was based on the fact that one carbon signal was missing in the ^{13}C NMR spectrum and thus the carbon signal at $\delta 109.45$ was attributed to C-3 and C-5 of the pyrone ring. As the protons of the methyl group ($\delta 1.76\text{s}$) on C-3 of the pyrone ring and the hydroxyl proton at $\delta 11.28\text{s}$, showed crosspeaks with the carbon at $\delta 109.45$, we have concluded that the hydroxyl group was on C-6. In fact, it is improbable that C-3 and C-5 could have the same chemical shift value as both carbons have completely different environment. Carefully examining the HMBC at 500 MHz spectrum of tetillapyrone (Table 1), we have found that the methyl protons ($\delta 1.78\text{s}$) gave strong correlations to, besides C-4 ($\delta 136.21$), the carbons at $\delta 109.43$ (C-3), 150.54 (C-6) and 163.85 (C-2). This was very strange since the methyl protons were five bonds away from C-5. On the other hand, the hydroxyl proton at $\delta 11.30$ brs (OH-6) showed also correlation with the carbon at $\delta 109.43$ (C-3), which is also of five bonds distance. However, the only chance for this to happen is that tetillapyrone does not exist as only one compound (**5**) but instead as two compounds (**5** and **6**). Thus the correlations observed in the HMBC spectrum were due to the methyl protons ($\delta 1.78\text{s}$) with the carbons at $\delta 150.54$ (C-2 of compound **6**) and $\delta 163.85$ (C-2 of compound **5**), respectively. This also explains the correlation observed between the hydroxyl proton at $\delta 11.30$ brs (OH-2) and the carbon at $\delta 109.43$ (C-3) of compound **6**.

It is interesting to note that in the structure previously proposed for nortetillapyrone (**8**), the positions of the hydroxyl and carbonyl groups of the pyrone moiety were opposite to those found in the structure **5** of tetillapyrone. Though, the HMBC spectrum showed strong correlation between H-3 ($\delta 5.62$) and the carbon at $\delta 163.18$, like that in structure **5** of tetillapyrone, the correlation between H-3 and the hydroxyl proton at $\delta 11.23$ brs observed in the COSY spectrum, has led to the conclusion that nortetillapyrone should have the structure **8**. In this structure, the chemical shift values of the carbonyl carbon ($\delta 150.48$) and the enolic carbon ($\delta 163.18$) were opposite to those found in tetillapyrone (**5**). However, there is no satisfactory explanation for this incongruence. The only possibility is, like in the case of tetillapyrone, nortetillapyrone also exists, in solution, as two compounds (**7** and **8**). In this circumstance, the correlation observed in HMBC spectrum (Table 2) between H-4 ($\delta 5.52\text{d}$) and the carbon at $\delta 163.18$ (C-2) was due to structure **7**, while the correlation between H-4 ($\delta 5.52\text{d}$) and the hydroxyl group at $\delta 11.23$ brs (OH-2), observed in the COSY spectrum, was due to structure **8**.

Since structures **5** and **6** of tetillapyrone are tautomers, they can exist in equilibrium in the solution. In the same way, structures **7** and **8** are also tautomers of nortetillapyrone and thus can also exist in equilibrium in the solution (Figure 2).

Table 1. ^1H and ^{13}C NMR data (500 MHz, DMSO) of tetillapyrone (**5** and **6**).

Position	δH (Hz) ^a	δC mult ^b	COSY	HMBC
2	-----	150.54/163.85		
3	-----	109.43		
4	7.72d (1.0)	136.21	Me	Me, C-2/C-6, C-3
5	-----	?		
6	-----	150.54/163.85		
7	6.18t (6.4)	83.75	H-8	C-4, C-6
8	2.08m	39.49	H-7, H-9	C-7, C-9
9	4.25q (3.1)	70.45	H-8, H-10	
10	3.77dd (6.7, 3.8)	87.30	H-9, H-11	C-9, C-7
11	3.60dd (11.8, 3.8)	61.35	H-10	
	3.55dd (11.8, 3.8)			C-9
CH ₃	1.78d (1.0)	12.36	H-4	C-3, C-4, C-2/C-6
OH-2	11.30 brs	-----		C-3
OH-9	5.27d (4.1)	-----		C-8, C-9, C-10
OH-11	5.06t (5.1)	-----		C-10, C-11

^a*J* values in parenthesis. ^bMultiplicities deduced by DEPT.

Table 2. ^1H and ^{13}C NMR data (500 MHz, DMSO) of nortetillapyrone (**7** and **8**)

Position	δH (Hz) ^a	δC mult ^b	COSY	HMBC
2	-----	150.48/163.18		
3	6.52d (8.1)	101.78	H-4	C-4, C-2 (w)
4	7.85d (8.1)	140.56	H-3	C-7(w), C-3, C-2/C-6
5	-----	?		
6	-----	150.48/163.18		
7	6.13t (7.0)	84.11	H-8	C-4, C-2/C-6
8	2.10m	39.75	H-7, H-9	C-7, C-9
9	4.21m	70.40	H-8, H-10	
10	3.76dd (6.4, 3.2)	87.43	H-9, H-11	C-11
11	3.53m	61.27	H-10	
OH-2	11.23brs	-----		
OH-9	8.45brs	-----		
OH-11	-----	-----		

^a*J* values in parenthesis. ^bMultiplicities deduced by DEPT ; w = weak

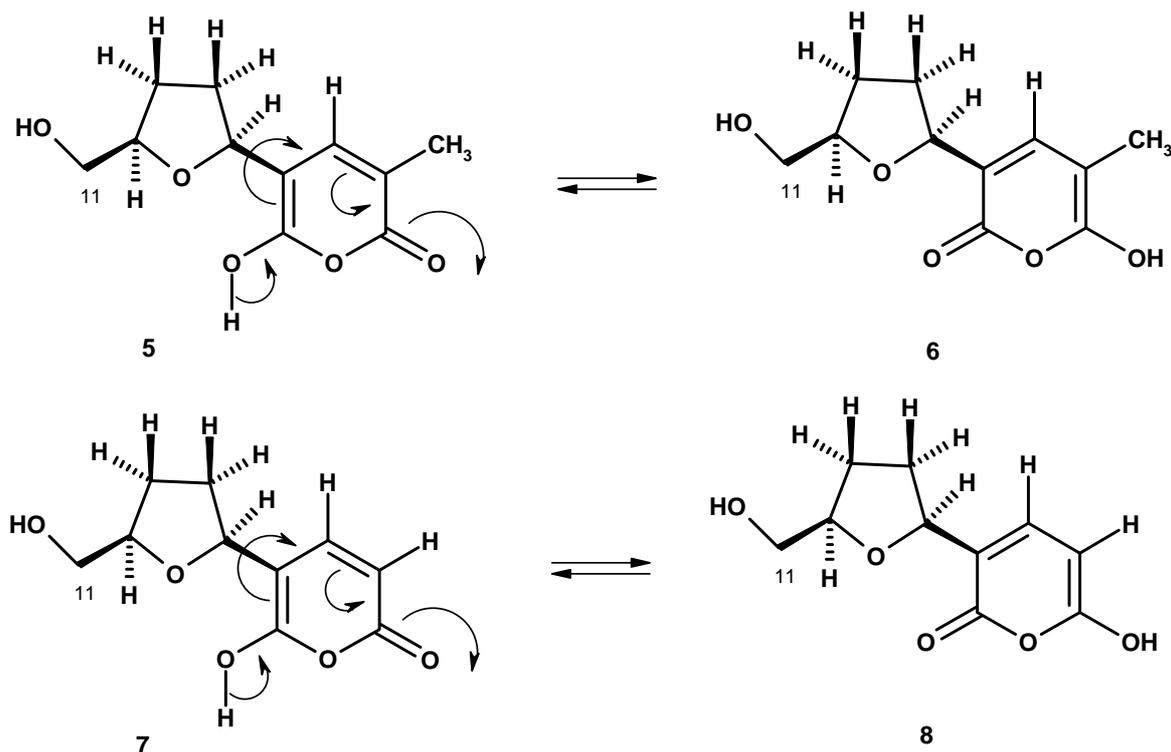


Figure 2. Tautomerization of tetillapyrone (**5**, **6**) and nortetillapyrone (**6**, **7**).

The structure of compound **1** was identified as maleimide-5-oxime by spectroscopic and HRMS data. EIHRMS of compound **1** gave the molecular ion peak at 122.02723, indicating its molecular formula as C₄H₄N₂O₂ (calcd.122.02728). The ¹H and ¹³C NMR spectra of compound **1** were very simple, showing the proton and chemical shift values very similar to those observed for 3-methylmaleimide-5-oxime, previously isolated from the marine sponge *Pseudoceratina purpurea* by our group [12]. However, the ¹HNMR of compound **1** showed two doublets of two coupled protons at δ 7.40d ($J=7.6$ Hz) and 5.45d ($J=7.6$ Hz), instead of the signals of the vinylic methyl protons (δ 1.73d, $J=0.9$ Hz) and a vinylic proton (δ 7.2.6d, $J=0.9$ Hz).

In order to investigate the biological activity of the constituents of the two marine sponges, maleimide-5-oxime (**1**), *p*-hydroxybenzaldehyde (**3**), phenylacetic acid (**4**), tetillapyrone (**5** and **6**), and nortetillapyrone (**7** and **8**) were evaluated for their growth inhibitory effect against seven yeasts (*Candida albicans*, *C. glabrata*, *C. dubliniensis*, *C. tropicalis*, *C. krusei*, *C. parapsilosis*, *Cryptococcus neoformans*), three non dermatophyte filamentous fungi (*Aspergillus fumigatus*, *A. flavus*, *A. niger*) and five dermatophyte filamentous fungi (*Microsporum gypseum*, *M. canis*, *Trichophyton mentagrophytes*, *T. rubrum* and *Epidermophyton floccosum*). Maleimide-5-oxime (**1**), *p*-hydroxybenzaldehyde (**3**), phenylacetic acid (**4**) and tetillapyrone (**5** and **6**) were found to be inactive against all the test organisms (MIC > 250 μ g/mL). Among the compounds tested, only nortetillapyrone (**7**, **8**) showed growth inhibitory effect against tested fungal strains. Interestingly, some selectivity was observed for this compound. It showed no activity against *Candida albicans*, *C. krusei* and *C. parapsilosis* even at maximum concentration tested (250 μ g/mL). However, nortetillapyrone showed some activity against *C. tropicalis* (250 μ g/mL) and good activity against *C. glabrata* (62.5-31.25

$\mu\text{g/mL}$), *C. dubliniensis* (62.5 $\mu\text{g/mL}$) and *Cryptococcus neoformans* (31.25 $\mu\text{g/mL}$). In Addition, this compound exhibited significant antifungal activity against dermatophytes with the MIC values ranging from 31.25 to 125 $\mu\text{g/mL}$. It was more active against *Microsporum canis* (31.25 $\mu\text{g/mL}$) and less active against *Trichophyton rubrum* (62.5-125 $\mu\text{g/mL}$). Nortetillapyrone showed no activity against *Aspergillus* species even at maximum concentration tested (250 $\mu\text{g/mL}$).

Table 3. Antifungal activity (MIC) of nortetillapyrone (**7**, **8**) for *Candida*, *Aspergillus* and dermatophyte strains.

Test strain	MIC ($\mu\text{g/ml}$)		
	nortetillapyrone (7 , 8)	Fluconazole	Amphotericin B
Yeasts			
<i>Candida albicans</i>	>250	1	NT
<i>C. glabata</i>	31.25-62.5	32	NT
<i>C. dubliniensis</i>	62.5	NT	NT
<i>C. tropicalis</i>	250	4	NT
<i>C. krusei</i>	>250	64-128	NT
<i>C. parapsilosis</i>	>250	<1	NT
<i>Cryptococcus neoformans</i>	31.25	NT	NT
Filamentous fungi, Aspergillus			
<i>Aspergillus fumigatus</i>	>250	NT	1
<i>A. flavus</i>	>250	NT	1-2
<i>A. niger</i>	>250	NT	2
Filamentous fungi, dermatophytes			
<i>Microsporum gypseum</i>	62.5	>128	NT
<i>M. canis</i>	31.25	128	NT
<i>Trichophyton mentagrophytes</i>	125	16-32	NT
<i>T. rubrum</i>	62.5-125	16-32	NT
<i>Epidermophyton floccosum</i>	62.5	16	NT

NT = not tested

Maleimide-5-oxime (**1**), tetillapyrone (**5** and **6**), and nortetillapyrone (**7** and **8**) were also evaluated for their capacity to inhibit the *in vitro* growth of three human tumor cell lines, namely the estrogen-dependent ER(+) MCF-7 (human breast adenocarcinoma), the estrogen-independent ER(-) MDA-MB-231 (human breast adenocarcinoma) and NCI-H460 (non-small cell lung cancer). Maleimide-5-oxime (**1**) and tetillapyrone (**5** and **6**) were found to be inactive against all the cell lines tested ($\text{GI}_{50} > 200 \mu\text{M}$) while nortetillapyrone (**7** and **8**) only weakly inhibited the growth of estrogen-independent ER (-) MDA-MB-231 cell line ($\text{GI}_{50} = 155.7 \mu\text{M}$) but was inactive against the estrogen-dependent ER (+) MCF-7 and NCI-H460 cell lines ($\text{GI}_{50} > 200 \mu\text{M}$).

3. Conclusion

As most antifungal compounds from marine origin are cytotoxic, consequently they have not generally been considered promising antifungal agents for clinical application [6]. Though many secondary metabolites from marine source were reported for their antifungal activity, their cytotoxic effect has not been measured. From this point of view, nortetillapyrone showed a very interesting biological activity profile. This compound not only selectively inhibits the growth of dermatophytes but also possesses no cytotoxicity against the tumor cell lines. Consequently, the structural feature of nortetillapyrone could serve as a model for development of a potential antifungal agent with therapeutic value.

4. Experimental

4.1. General

^1H and ^{13}C NMR spectra were recorded at ambient temperature in CDCl_3 and DMSO on a Bruker AMC instrument operating at 300.12 and 75.47 MHz, respectively or a Bruker DRX instrument operating at 500, respectively 125 MHz. EI mass spectra were measured on a Hitachi Perkin-Elmer RMV-6M instrument. HRMS spectra were measured on a Kratos concept II 2 sector mass spectrometer. The accelerating voltage was 8 kV. Si gel for chromatography was silica gel 60 (0.2-0.5 mm Merck) for analytical work and for preparative work TLC silica gel 60 GF 254 Merck.

4.2. Animal material

Haliclona cymaeformis (Esper, 1794) order Haplosclerida, family Chalinidae, was collected by Scuba diving in the Gulf of Thailand near Koh Chang (2 meter depth), Trad Province in March 2004. *Haliclona baeri* (Wilson, 1952) was also collected by Scuba diving in the in the coastal area of Chonburi Province (3 meter depth). The sponge materials were identified by Dr. Sumaitt Putchakarn and the vouchers of the specimens, registered as BIMS-I 953 and BIMS-I 1766, respectively were deposited at Bangsaen Institute of Marine Science, Burapha University, Bangsaen, Chonburi, Thailand. The materials were frozen immediately at -20°C prior to extraction.

4.3. Extraction, isolation and characterization of the constituents of *Haliclona baeri*

The sample (1.3 kg net weight) was thawed, homogenized with EtOH (3 L), allowed to stand for 24 h in a dark chamber and filtered. The residue on the filter paper was again extracted with EtOH (2x3L), the aqueous alcoholic extracts were combined, evaporated at reduced pressure to ca. 500 ml and extracted with EtOAc (3x500mL). The EtOAc extracts were combined and concentrated at reduced pressure to give the crude EtOAc extract. This (10 g) was chromatographed over a Si gel column (120g) and eluted with petrol- CHCl_3 and CHCl_3 - Me_2O , 350 ml fractions being eluted as follows. Frs 1-62 (petrol- CHCl_3 , 1:1), 63-78 (petrol- CHCl_3 , 3:7), 79-95 (petrol- CHCl_3 , 1:9), 96-182 (CHCl_3 - Me_2O , 4:1), 183-206 (CHCl_3 - Me_2O , 3:2). Frs 101-114 were combined (107.8 mg) and

purified by TLC (Si gel, CHCl₃-Me₂O-HCO₂H, 5:5:0.1) to give 8.6 mg of maleimide 5-oxime (**1**). Frs 125-198 were combined (185.6 mg) and purified by TLC (Si gel, CHCl₃-MeOH-HCO₂H, 93:7:1) to give 13.1 mg 3,4-dihydroxybenzoic acid (**2**) and 8.0 mg of tetillapyrone (**5** and **6**).

4.4. Extraction, isolation and characterization of the constituents of *Haliclona cymaeformis*

The sample of *Haliclona cymaeformis* (7.6 kg net weight) was thawed, homogenized with EtOH (10L), allowed to stand for 24 h in the dark chamber and filtered. The residue on the filter paper was again extracted with EtOH (2x10L), the aqueous alcoholic extracts were combined, evaporated at reduced pressure to give ca. 700 ml and extracted with EtOAc (3x1L). The EtOAc extracts were combined and concentrated at reduced pressure to give a crude EtOAc extract (21g). This extract was applied to flash chromatography column of Si Gel (150 g) and eluted with CHCl₃ (frs. 1-5; 5 x 200 ml), CHCl₃-MeOH, 49:1 (frs 6-15; 10 x 200 ml), CHCl₃-MeOH, 19:1 (frs16-20; 5x200 ml), CHCl₃-MeOH, 9:1 (frs 21-22; 2x500 ml), EtOAc (frs 23-24; 2 x500ml) and MeOH (frs 25-26; 2 x500ml). Frs 18-24 were combined (3.1g), applied on the flash chromatography column of ODS-A gel (35g) and eluted with mixture of MeOH-H₂O; 7:3 (sfr 1; 200ml), MeOH-H₂O; 4:1 (sfr 2; 200ml), MeOH-H₂O; 9:1 (sfr 3; 2x100ml), MeOH (sfr 4; 400ml), CHCl₃-MeOH-H₂O; 14:6:1 (sfr 5, 400ml). Sfr 1 (156 mg) was purified by TLC Si gel, CHCl₃-MeOH-HCO₂H, 19:1:0.1) to give 3.5 mg of *p*-hydroxybenzaldehyde (**3**) and 15 mg of tetillapyrone (**5** and **6**). Sfr 5 (103 mg) was purified by TLC Si gel, CHCl₃-MeOH-HCO₂H, 19:1:0.1) to give 7 mg of phenylacetic acid (**4**) and 17 mg of nortetillapyrone (**7** and **8**).

Maleimide-5-oxime (**1**): yellow viscous mass; EI HRMS 122.02723 (M⁺), cald for C₄H₄N₂O₂, 122.02728. ¹HNMR (300 MHz, DMSO): δ11.03 brs (-NH), 10.86 brs (=N-OH), 7.40d (*J*=7.6 Hz, H-4), 5.45d (*J*=7.6 Hz, H-3), ¹³CNMR (75.47 MHz, DMSO): δ164.37 (CO-2), 151.54 (C-5), 142.22 (CH-4), 100.24 (CH-3).

4.5. Biological Activity

5.5.1. Cell Growth Assay

The effect of compounds on the growth of human tumor cell lines were evaluated according to the procedure adopted by National Cancer Institute (NCI, USA) for the *in vitro* anticancer drug discovery screen which uses the protein-binding dye sulforhodamine B (SRB) to assess cell growth inhibition [13, 14] and according to the method described by Kijjoa *et al.* [15].

5.5.2. Antifungal Activity Determination

Antifungal activity of maleimide-5-oxime (**1**), *p*-hydroxybenzaldehyde (**3**), phenylacetic acid (**4**), tetillapyrone (**5** and **6**) and nortetillapyrone (**7** and **8**) was evaluated against *Candida*, *Aspergillus* and dermatophyte strains: three yeasts clinical strains, one of *Cryptococcus neoformans* PH1, one of *Candida dubliniensis* PH10 and one of *C. glabrata* D10R, as well as, four American Type Culture Collection (ATCC) type strains (*C. albicans* ATCC 10231, *C. tropicalis* ATCC 13803, *C. krusei*

ATCC 6258 and *C. parapsilosis* ATCC 90018); one *Aspergillus* clinical strain (*A. flavus* F44), as well as, two ATCC type strains (*A. niger* ATCC 16404, and *A. fumigatus* ATCC 46645); five dermatophyte clinical strains (*Microsporum canis* FF1, *M. gypseum* FF3, *Trichophyton rubrum* FF5, *T. mentagrophytes* FF7 and *Epidermophyton floccosum* FF9).

The fungal isolates were identified by standard microbiology methods and stored on Sabouraud Dextrose broth with glycerol at -70°C and subcultured in Sabouraud-Chloramphenicol agar. Minimal Inhibitory Concentration (MIC) was determined by a microdilution method according to the CLSI reference method M27-A2 for yeasts [16], and M38-A for filamentous fungi [17]. The test was performed in sterile 96 flat-bottom well microtitration plates and the medium used was MOPS-buffered RPMI 1640. Briefly, MICs were determined for all yeasts strains with the adjusted final inoculum of $1-4 \times 10^4$ CFU/ml, prepared from a 24-48 hours culture. With filamentous fungi (except for *Epidermophyton floccosum*), the final inoculum used was $4-8 \times 10^4$ CFU/mL, prepared from a sporulated culture. For *Epidermophyton floccosum*, the final inoculum was 8×10^4 to 1.6×10^5 CFU/mL. Two fold serial dilutions of compounds in DMSO, ranging from 250 to 15.125 µg/mL, were tested. Fungal growth controls, with and without DMSO, were included. The final DMSO concentration was 2%, and did not affect the fungal growth. In addition, reference antifungal compounds, fluconazole (Pfizer, United Kingdom) for yeasts and dermatophytes or amphotericin B (Sigma) for *Aspergillus*, were used as the standard antifungal drugs. Twofold serial dilutions ranging from 128 to 0.25 µg/mL for fluconazole and 16 to 0.063 µg/ml for amphotericin B were used. Plates were incubated at 30°C for 48 hours for *Candida* species, 72 hours for *Cryptococcus neoformans*, 4-5 days for *Aspergillus* species and 8 days for dermatophytes. Quality control determinations of the MIC of fluconazole were ensured by testing *C. parapsilosis* ATCC 90018. The results were within the recommended limits. All determinations were performed in duplicate and three independent experiments lead to concordant results.

Acknowledgements

This work was supported by FTC- Fundação para a Ciência e Tecnologia (Project POCI/MAR/58114/2004) and Unidade de ID 226/94; FEDER and POCI, and GRICES. Work in Thailand was supported by the Thailand Research Fund (No. TRG 4780003). We thank Dr. Sumaitt Puchakarn, BIMS, Burapha University, Thailand, for collection and identification of the sponge materials and Dr. Graham Eaton, Department of Chemistry, University of Leicester, UK for HRMS.

References

1. Li, H.Y.; Matsunaga, S.; Fusetani, N. Antifungal metabolites from marine sponges. *Curr. Org. Chem.* **1998**, *2*, 649-682.
2. Richardson, M.D. Changing patterns and trends in systematic fungal infections. *J. Antimicrobial Chemother.* **2005**, *56*, i5-i11.

3. Gupta, A.K and Tu, L.Q. Dermatophytes: Diagnosis and treatment. *J. Am. Acad. Dermatol.* **2006**, *54*, 1050-1055.
4. Johnson, L. Dermatophytes-the skin eaters. *Mycologist* **2003**, *17*, 147-149.
5. Sundriyal, S.; Sharma, R. K.; Jain, R. Current advances in antifungal targets and drug development. *Curr. Med. Chem.* **2006**, *13*, 1321-1335.
6. M. Donia and M. Hartmann. Marine natural products and their potential applications as anti-infective agents. *THE LANCET Infectious Disease* **2003**, *3*, 338-348.
7. Faulkner, D.J. Marine Natural Products. *Nat. Prod. Rep.* **1993**, *10*, 497-539.
8. Rashid, M.A.; Gustafson, K.R.; Boswell, J. L.; and Boyd M.R. Haligramides A and B, two new cytotoxic hexapeptides from the marine sponge *Haliclona nigra*. *J. Nat. Prod.* **2000**, *63*, 956-959.
9. Fahy, E.; Molinski, T.F.; Harper, M.K.; Sulliwán, B.W.; Faulkner, D. J. Haliclonadiamine: an antimicrobial alkaloid from the sponge *Haliclona* sp. *Tetrahedron Lett.* **1988**, *29*, 3427-3428.
10. Clark R. J.; Garson, M.J, Hooper, J.N.A. Halaminols A-C, three new alkyl amino alcohol from the sponge *Haliclona* sp. *J. Nat. Prod.* **2001**, *64*, 1568-1571.
11. Watanadilok, R.; Sonchaeng, P.; Kijjoa, A.; Damas, A.M.; Gales, L.; Silva, A.M.S.; Herz, W. Tetillapyrone and nortetillapyrone, two unusual hydroxypyran-2-ones from the marine sponge *Tetilla japonica*. *J. Nat. Prod.* **2001**, *64*, 1056-1058.
12. Kijjoa, A.; Bessa, J.; Wattanadilok, R.; Sawangwong, P.; Nascimento, M.S.J.; Pedro, M.; Silva, A.M.S.; Eaton, G.; van Soest, R.; Herz, W. Dibromotyrosine derivatives, a maleimide, aplysamine -2 and other constituents of the marine sponge *Pseudoceratina purpurea*. *Zeitschrift fur Naturforschung* **2005**, *60b*, 904-908.
13. Monks, A.; Scudiero, D.; Skehan, P.; Shoemaker, R.; Paul, K.; Vistica, D.; Hose, C.; Langley, J.; Cronise, P.; Vaigro-Wolff, A.; Gray-Goodrich, M.; Campbell, H.; Mayo, J. Boyd. M. Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines. *J. Natl. Cancer Inst.* **1991**, *83*, 757-776.
14. Skehan, P.; Storeng, R.; Scudiero, D.; Monks, A.; McMahon, J. Vistica, D.; Warren, J.T.; Bokesch, H.; Kenny, S.; Boyd, M.R. New calorimetric cytotoxic assay for anticancer drug screening. *J. Natl. Cancer Inst.* **1990**, *82*, 1107-1112.
15. Kijjoa, A.; Wattanadilok, R.; Campos, N.; Nascimento, M.S.J.; Pinto, M.; Herz W. Anticancer activity Evaluation of Kuanoniamines A and C isolated from the marine sponge *Oceanapia sagittaria*, collected from the Gulf of Thailand. *Mar. Drugs* **2007**, *5*, 6-22.
16. National Committee for Clinical Laboratory Standards 2002. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Yeasts: approved standard. Document M27-A2. Wayne, PA: National Committee for Clinical Laboratory Standards.

17. National Committee for Clinical Laboratory Standards 2002. Reference Method for Broth Dilution Antifungal Susceptibility Testing of Filamentous Fungi: approved standard. Document M38-A. Wayne, PA: National Committee for Clinical Laboratory Standards.

Sample Availability: Available from the author.

© 2007 by MDPI (<http://www.mdpi.org>). Reproduction is permitted for noncommercial purposes.