

p53 Regulates Myogenesis by Triggering the Differentiation Activity of pRb

Alessandro Porrello, Maria Antonietta Cerone, Sabrina Coen, Aymone Gurtner, Giulia Fontemaggi, Letizia Cimino, Giulia Piaggio, Ada Sacchi, and Silvia Soddu

Molecular Oncogenesis Laboratory, Regina Elena Cancer Institute, Center for Experimental Research, 00158 Rome, Italy

Abstract. The p53 oncosuppressor protein regulates cell cycle checkpoints and apoptosis, but increasing evidence also indicates its involvement in differentiation and development. We had previously demonstrated that in the presence of differentiation-promoting stimuli, p53-defective myoblasts exit from the cell cycle but do not differentiate into myocytes and myotubes. To identify the pathways through which p53 contributes to skeletal muscle differentiation, we have analyzed the expression of a series of genes regulated during myogenesis in parental and dominant-negative p53 (dnp53)-expressing C2C12 myoblasts. We found that in dnp53-expressing C2C12 cells, as well as in p53^{-/-} pri-

mary myoblasts, pRb is hypophosphorylated and proliferation stops. However, these cells do not upregulate pRb and have reduced MyoD activity. The transduction of exogenous *TP53* or *Rb* genes in p53-defective myoblasts rescues MyoD activity and differentiation potential. Additionally, in vivo studies on the *Rb* promoter demonstrate that p53 regulates the *Rb* gene expression at transcriptional level through a p53-binding site. Therefore, here we show that p53 regulates myoblast differentiation by means of pRb without affecting its cell cycle-related functions.

Key words: p53 • Rb • MyoD • differentiation • muscle

Introduction

The differentiation of skeletal myoblasts is characterized by permanent withdrawal from the cell cycle, biochemical and morphological maturation into mononucleated myocytes, and fusion into multinucleated myotubes. This program depends on the expression and activation of myogenic basic helix-loop-helix (bHLH) regulatory factors belonging to the MyoD family (for review, see Weintraub et al., 1991) and on their cooperation with the myocyte-specific enhancer binding factors MEF2 (Melkentin and Olson, 1996; Yun and Wold, 1996). The activity of myogenic bHLH factors is regulated by the interaction with non-tissue-specific proteins. Members of the E2 family were shown to be necessary for binding of the myogenic bHLH factors to DNA, while this binding is inhibited by the presence of “inhibitor of differentiation” factors (for

review, see Wright, 1992). Other non-tissue-specific proteins contribute to the cell cycle withdrawal, as well as to the transcription of differentiation-associated genes. These proteins include members of the cyclin-dependent kinase inhibitors, such as p21^{Waf1/Cip1}, and cell-cycle regulators, such as pRb. p21^{Waf1/Cip1} is thought to inhibit pRb phosphorylation and thus to promote the cell cycle withdrawal that precedes terminal differentiation. This activity of p21^{Waf1/Cip1} was shown to be p53-independent (Halevy et al., 1995; Parker et al., 1995). The product of the *Rb* gene possesses two different, genetically dissociated functions in the differentiation process (Sellers et al., 1998). The hypophosphorylated form of pRb contributes to the cell-cycle withdrawal by acting as a transcriptional repressor of the E2F-regulated genes, relevant to the cell-cycle progression (Schneider et al., 1994; Weinberg, 1995, and references therein). In addition, pRb cooperates with MyoD to promote the expression of late markers of differentiation (Gu et al., 1993; Chen et al., 1996; Novitch et al., 1996) through the activation of the transcriptionally competent MEF2 factors (Novitch et al., 1999).

The product of the tumor suppressor gene *TP53* is a prominent regulator of cell-cycle checkpoints and apoptotic death. Furthermore, considerable experimental evidence has accumulated suggesting that a fine regulation of the p53 protein activity is required for optimal development and differentiation (for reviews, see Almog and Rot-

Drs. Porrello, Cerone, and Coen contributed equally to the work and should be considered co-first authors.

Address correspondence to S. Soddu, Molecular Oncogenesis Laboratory, Regina Elena Cancer Institute, Via delle Messi d'Oro 156, 00158 Rome, Italy. Tel.: 39 064985 2563. Fax: 39 064180 526. E-mail: soddu@ifo.it

Dr. Cerone's present address is Department of Pathology and Molecular Medicine, McMaster University Medical Center, Hamilton, Ontario, Canada L8N 3Z5.

Dr. Coen's present address is Lautemberg Center for General and Tumor Immunology, Hebrew University-Hadassa Medical School, Jerusalem, Israel 91120.

Dr. Cimino's present address is Laboratory of Genetic Expression, University of Rome La Sapienza, 00153 Rome, Italy.

ter, 1997; Choi and Donehower, 1999). p53 expression and/or activity increases during at least some differentiative processes such as hematopoiesis (Kastan et al., 1991), spermatogenesis (Almon et al., 1993), and myogenesis (Halevy et al., 1995; Soddu et al., 1996; Tamir and Bengal, 1998). Exogenous p53 expression can induce differentiation in a variety of tumor cells in vitro (reviewed in Almog and Rotter, 1997) and in vivo (Bossi et al., 2000), while alterations of the *TP53* gene in vivo frequently correlate with undifferentiated phenotypes of cancers (Feinstein et al., 1991; reviewed in Fagin, 1995). By in situ hybridization, it was observed that *TP53* mRNA is present at high levels in all mouse embryonic cells from embryonic day (E) 8.5 to E 10.5. At later stages of development, *TP53* expression becomes more pronounced during the differentiation of specific tissues and declines in mature tissues (Schmid et al., 1991). By using immortal and primary myoblasts and myeloblasts, we have previously demonstrated that the expression of *dnp53* proteins strongly reduces terminal differentiation into myotubes and granulocytes/monocytes. In contrast, *dnp53* expression does not affect the cell cycle withdrawal and the apoptotic death associated with these differentiation processes (Soddu et al., 1996; Mazzaro et al., 1999). Comparable results were obtained with MyoD-converted *p53*^{-/-} fibroblasts (Cerone et al., 2000). Interference with p53 mRNA or protein was shown to inhibit the development of *Xenopus laevis* (Wallingford et al., 1997). Despite the normal viability of the original *TP53* null mice (Donehower et al., 1992), it has become increasingly evident that a fine balance of p53 expression and activation is required for the normal development of total embryo or single organs (for review, see Choi and Donehower, 1999). The complete absence of p53 can result in reduced fertility and exencephaly in some mouse strains (Armstrong et al., 1995; Sah et al., 1995), or in mice receiving low folic acid concentration in the diet (Choi and Donehower, 1999). Not less deleterious is the overexpression or the aberrant activation of p53. This was shown by the early embryonic lethality of *TP53* transgenic or *mdm2* null mice, as well as by the rescue of the latter phenotype through the double knock out of the *mdm2* and *TP53* genes (Montes de Oca Luna et al., 1995). All these observations strongly support the involvement of the *TP53* gene in cell differentiation and development. Nevertheless, the pathways modulated by p53 in these processes are still unknown, as are the effectors of these biological events.

To identify the molecules that are downstream of p53 in differentiation, we took advantage of the C2C12 myoblast model we previously employed to demonstrate that *dnp53* expression interferes with terminal differentiation into skeletal myotubes (Soddu et al., 1996). Moreover, critical experiments were confirmed on *p53*^{+/+} and *p53*^{-/-} primary myoblasts. We compared the expression of myogenesis-regulated genes in parental and *dnp53*-expressing C2C12 myoblasts, in differentiation promoting conditions. The serial analysis of gene expression and the consequent biochemical and functional studies showed that p53 regulates, at the transcriptional level, the increased expression of pRb, which is involved in the execution of the muscle differentiation program together with MyoD. No modification was found in the pRb-dependent cell-cycle with-

drawal, indicating that during myogenesis p53 specifically acts on the differentiation functions of pRb.

Materials and Methods

Differentiation Assay

Mouse satellite cells (MSC)¹ were explanted from new-born mice and maintained in culture as described (Mazzaro et al., 1999). To avoid spontaneous differentiation, primary satellite and C2C12 cells were never allowed to reach confluence. Muscle differentiation was induced by serum withdrawal (DMEM plus 10 μ g/ml insulin and 5 μ g/ml transferrin, or plus 2% horse serum), as described (Soddu et al., 1996; Mazzaro et al., 1999). Differentiation was evaluated by indirect immunofluorescence for myosin heavy chain (MyHC) (Crescenzi et al., 1994). Differentiation index was calculated by counting at least 400 nuclei per dish, and applying the following formula: differentiation index = number of nuclei in differentiated cells/total number of nuclei.

Northern Blot Hybridization

Total cellular RNA was extracted by RNeasy Mini Kit (QIAGEN), following the manufacturer's instructions. Aliquots of 20 μ g per lane were electrophoresed through 1.5% agarose gel in the presence of formaldehyde. Gels were blotted onto Duralose nitrocellulose membrane (Stratagene) and hybridized according to standard protocols (Ausbel et al., 1987). Probes were gel-purified and labeled with [³²P]-dCTP (Dupont) by random primer extension (Ausbel et al., 1987).

Western Blotting

Cells were plated in 60-mm dishes (7.5 \times 10⁴/dish) and incubated in growth medium (GM) or in differentiation-promoting medium (DM) for different times. Approximately 30 min before harvesting the cells, Hoechst 33258 dye was added to the culture medium (GM or DM) to a final concentration of 1 μ g/ml, to stain nuclei. 20 different microscopic fields for each sample were analyzed by fluorescent microscope to estimate the total number of nuclei per dish. Equal numbers of nuclei were lysed in hot Laemmli buffer, loaded on 7 or 10% SDS-polyacrylamide gels, electrophoresed, and blotted onto nitrocellulose membranes (Bio-Rad Laboratories). Filters were immunoreacted with anti-pRb G3-245 mAb (PharMingen), anti-MyHC MF-20 mAb (Bader et al., 1982), or anti-Hsp70 N27F3-4 mAb (StressGen Biotechnologies). Immunoreactivity was determined using the ECL-chemiluminescence reaction (Amersham Corp.) following the manufacturer's instructions.

Plasmids, Viruses, and Cell Transduction

The following plasmids were used for transfections: pN53cG(Val135), carrying the temperature-sensitive p53Val¹³⁵ mutant gene (Michalovitz et al., 1990) and the neo gene; pRSVneo, carrying the selectable marker for G418 resistance; pLxSP, carrying the selectable marker for puromycin resistance; pMCK-luc, carrying 1,256 base pairs of the muscle creatine kinase (MCK) promoter upstream of the luciferase reporter gene (Novitch et al., 1999); pXRP1 and pd92, carrying the luciferase reporter gene driven by the human Rb promoter between -686 and -4 and between -686 and -92, respectively, relative to the translation start site (Osifchin et al., 1994); and pXRP1-mtp53, obtained from the pXRP1 vector by the QuikChange site-directed mutagenesis kit (Stratagene). In the last vector, the sequence of the p53 binding site (between -82 and -73) 5'-GGGCGTGCCC-3' was mutated in 5'-TGTCGTAAAC-3', and the mutation was confirmed by direct sequencing. C2C12 cells were transfected by CaPO₄-mediated DNA precipitation by using a 1:20 molar ratio between resistance and experimental vectors, when required. Selection was performed in the presence of 0.75 mg/ml G418, or 2 μ g/ml puromycin. *dnp53* protein expression was assessed by indirect immunofluorescence and Western blotting (Soddu et al., 1996). Integration of luciferase reporter vectors was assessed by direct PCR using two primers complementary to the luciferase gene.

¹Abbreviations used in this paper: Act D, actinomycin D; DM, differentiation-promoting medium; *dnp53*, dominant-negative p53; GM, growth medium; MCK, muscle creatine kinase; MSC, mouse satellite cell; MyHC, myosin heavy chain.

Recombinant adenoviruses Adp53 (Bacchetti and Graham, 1993), encoding a human wt-p53 protein and Ad-ΔRb (Chang et al., 1995), encoding a low phosphorylatable human Rb protein, and the replication defective, E1A-deleted dl312 control adenovirus were prepared and titrated by 293-cell infection. Recombinant retroviruses Ltsp53SN, encoding p53Val¹³⁵ mutant and the insertless LxSN, were produced by GP+E packaging cells.

Luciferase Activity

Luciferase activity was assessed as described (Osifchin et al., 1994) and normalized as reported for Western blotting.

In Vivo Footprinting and Chromatin Immunoprecipitation

For in vivo footprinting, proliferating and differentiated cells were treated with 0.06% dimethyl sulfate (Sigma-Aldrich) for 30 s. DNA was extracted and cleaved with piperidine (Sigma-Aldrich). As control, purified genomic DNA was methylated in vitro with 0.125% dimethyl sulfate for 2 min. In vivo footprintings were performed by ligation-mediated PCR (Martinez-Balbas et al., 1995) using the following primers: 5'-CCCGACTCCCGTTACAAAATAATG-3' (56.33°C), 5'-GAACGTC-CCCCGAGGAAAACCG-3' (61.65°C), and 5'-AACGTCCCCGAG-GAAAACCGGACGC-3' (65.8°C).

For chromatin cross-linked immunoprecipitation, proliferating and differentiated cells were cross-linked by adding formaldehyde directly to culture medium. DNA preparation, immunoprecipitation, and amplification were performed as described (Boyd et al., 1998). Affinity purified rabbit or sheep polyclonal antibodies were employed: anti-Sp1 sc-59-X (Santa Cruz Biotechnology, Inc.) and anti-p53 Ab-7 (Calbiochem). The following primers were employed to amplify the indicated promoters: the murine *Rb* promoter, 5'-GACGACGCGGGCGGAGACAGG-3', 5'-AACGTCCCCGAGGAAAACCGGACGC-3'; the human *Rb* promoter upstream the luciferase gene, 5'-GAGGGCGCGTCCGGTTTTTCTCA-3', 5'-AGCAATTGTTCAGGAACCAGGGCATAGC-3'; the murine *MyoD* promoter, 5'-CCTGGGGCTATTTATCCCCAGGGTAGCC-3', 5'-TAGACCACTGGAGAGGCTTGGGCAG-3'; the murine myosine light chain promoter, 5'-CTTCAGTCTCACCAGGGCTGTTCAC-3', 5'-CTCTCTCGGCTTCCTTTTATTCTGGGC-3', the murine desmin promoter, 5'-TGCTTCTAGCTGGGCCTTTC-3', 5'-CCGGACAG-GTCTCTACCATCCTTTC-3'.

Results

Myoblasts from p53^{-/-} Mice Have Reduced Differentiation Capacity

We have previously demonstrated that the expression of dnp53 proteins in primary and immortalized myoblasts inhibits their differentiation (Soddu et al., 1996; Mazzaro et al., 1999). These dnp53 proteins should form mixed, transcriptionally inactive oligomers with the endogenous wt-p53 protein (Milner and Medcalf, 1991). Recently, some weak but transcriptionally relevant heterotypic interactions have been described between some mutant p53 and p63 or p73 proteins (Davison et al., 1999; Di Como et al., 1999), which are members of the newly discovered p53 family (for reviews, see Oren, 1997; Kaelin, 1999). Since p63 and p73 knock-out mice present more evident developmental defects than p53^{-/-} mice (Mills et al., 1999; Yang et al., 1999, 2000), it is possible that the inhibition of myoblast differentiation induced by the dnp53 expression is due to the inactivation of other p53 family members. We have shown that MyoD-converted, p53^{-/-} fibroblasts have a significant reduction in their differentiation capacity, compared with the p53^{+/+} fibroblasts (Cerone et al., 2000). Still, to more directly demonstrate a p53-specific role in myogenesis, primary myoblasts were explanted from p53^{+/+} and p53^{-/-} mice to study their differentiation capacity.

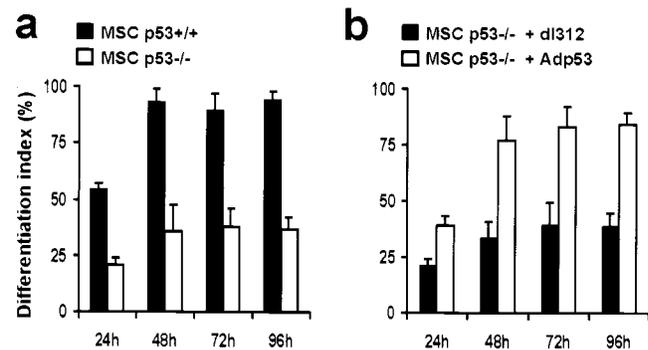


Figure 1. Effects of p53 impairment on muscle differentiation. (a) Time-course analysis of differentiation indices of mouse satellite cells (MSC) explanted from p53^{+/+} and p53^{-/-} mice. (b) p53^{-/-} MSC were induced to express wt-p53 protein by infection with the Adp53 recombinant adenovirus. The E1A-deleted dl312 adenovirus was used as negative control. Time course analysis of differentiation indices was performed as in a.

Time course analyses of differentiation showed a strong and persistent reduction of differentiation indices in p53^{-/-}, compared with p53^{+/+} myoblasts (Fig. 1 a). This reduction was completely rescued by the expression of an exogenous wt-p53 protein obtained by myoblast infection with a p53-carrying recombinant adenovirus (Bacchetti and Graham, 1993) (Fig. 1 b). These results demonstrate that the other members of the p53 family cannot substitute the p53-specific role in muscle differentiation.

p53-defective Myoblasts Do Not Upregulate Rb Expression

To identify the molecules downstream of p53 in the differentiation process, we evaluated the expression of a series of muscle differentiation-regulated genes in myoblasts derived from the C2C12 cell line. Total RNA was extracted from parental (C2C12) and dnp53-expressing (C2-dnp53) myoblasts (Soddu et al., 1996) maintained in GM, or incubated in the presence of DM for different times. As expected, C2-dnp53 cells had a strong reduction in the expression of muscle structural genes, like *MyHC*, compared with the parental cells (Fig. 2, a and b). A series of genes, including *MyoD*, *myogenin*, *p21^{Waf1/Cip1}*, *cyclin D1* and *D3*, and *Id-1*, were found to behave similarly in the two cell types (Fig. 2 a). In contrast, a strong difference in the *Rb* mRNA induction was present between parental and C2-dnp53 cells (Fig. 2, a and c), the latter being largely impaired in their ability to increase the *Rb* mRNA levels. This difference was even more marked at the protein level (Fig. 2 d) and was confirmed in p53^{-/-} primary myoblasts (e). Moreover, infection of the p53^{-/-} primary myoblasts with recombinant retroviruses carrying the dnp53 protein did not further inhibit MyHC and Rb expression (Fig. 2 e) and differentiation indices (15.3 and 16.7% in neo and dnp53-expressing cells, respectively), supporting the p53 specificity for this differentiation activity. Interestingly, Rb expression in proliferating conditions was similar in parental and p53-impaired cells, suggesting a specific p53 control on the differentiation-associated upregulation of the *Rb* gene.

To evaluate whether the differentiation-associated *Rb* upregulation was stably inhibited in the C2-dnp53 cells, or

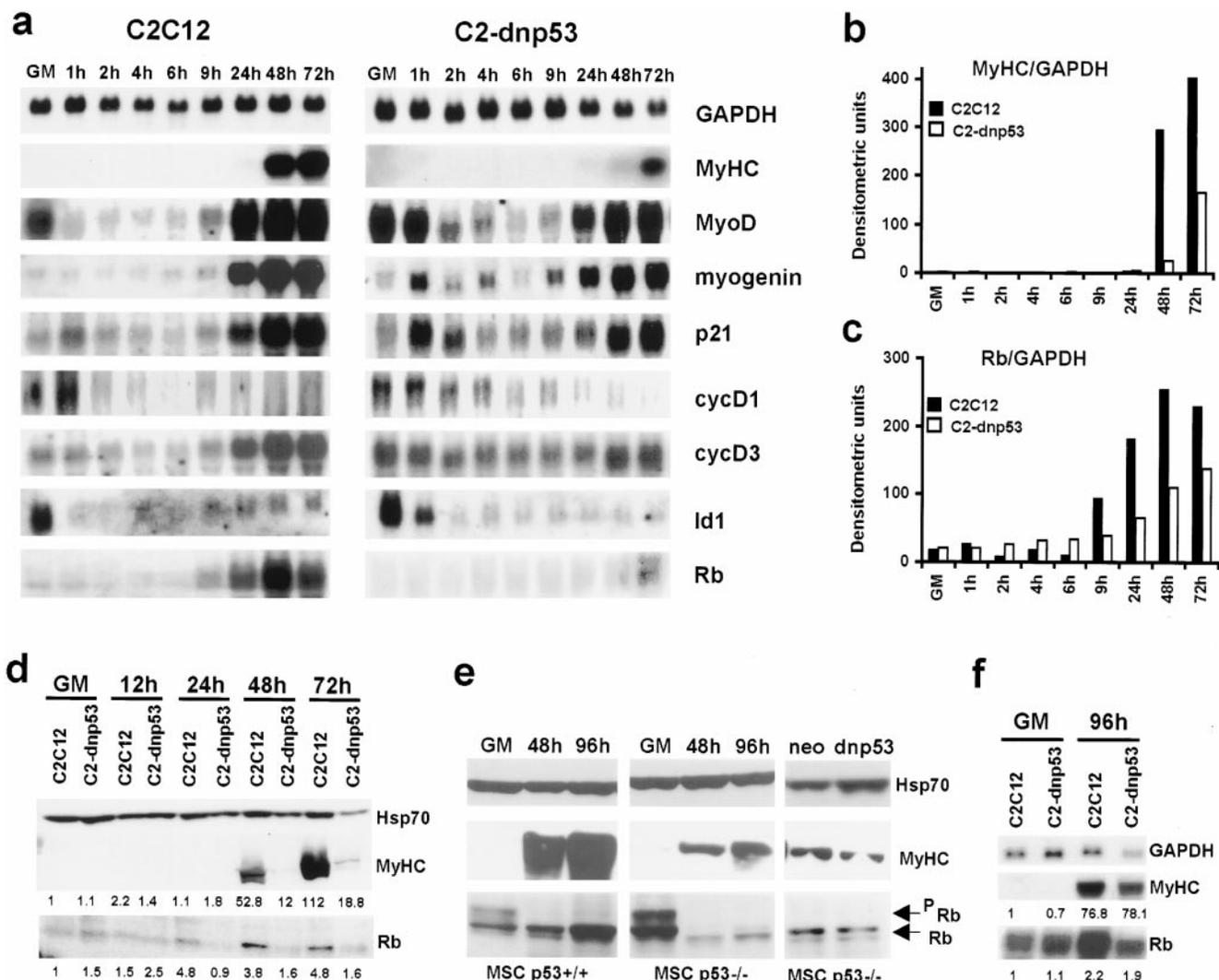


Figure 2. Serial analysis of muscle differentiation-regulated gene expression. (a) Northern blot hybridizations were performed on total RNA extracted from C2C12 and C2-dnp53 cells incubated in DM for the indicated times, or maintained in GM. (b and c) Quantitative analyses, relative to GAPDH, of the gene expression levels observed in a were evaluated for *MyHC* and *Rb* in the indicated cells. (d) C2C12 and C2-dnp53 cells, incubated as in a, were analyzed by Western blotting for the listed proteins. The numbers represent quantitative data relative to Hsp70. (e) p53^{+/+} and p53^{-/-} MSC were incubated as in a. p53^{-/-} MSC were also transduced with the indicated gene by retroviral infection and incubated for 96 h in DM. Total cell lysates from each of these cell lines were analyzed by Western blotting for the listed proteins. (f) C2C12 and C2-dnp53 cells were incubated in GM, or in DM for 96 h, and maintained at 32°C. Northern blot hybridization was performed for the indicated genes. The numbers represent quantitative data relative to GAPDH.

this inhibition was dependent on the sustained expression of exogenous dnp53 protein, we took advantage of this particular dnp53 protein being temperature sensitive (Michalovitz et al., 1990). Northern blot analysis was performed on parental and C2-dnp53 cells at the permissive temperature (32°C). The complete recovery of the differentiation ability of C2-dnp53 cells at 32°C (Soddu et al., 1996) was associated with that of *MyHC* and *Rb* expression levels (Fig. 2 f). These results indicate that inhibitions of differentiation and of *Rb* upregulation are reversible consequences of p53 inactivation.

p53 Does Not Interfere with the Cell Cycle-related Functions of pRb

Skeletal muscle cells belong to those cell types in which differentiation is associated with irreversible withdrawal

from the cell cycle (Crescenzi et al., 1995). In differentiating conditions, *Rb*^{-/-} myoblasts are defective in cell cycle withdrawal and expression of late markers of differentiation (Schneider et al., 1994; Novitch et al., 1996). Furthermore, those *Rb*^{-/-} myoblasts that stop proliferating and reach differentiation can re-enter the cell cycle after serum stimulation (Schneider et al., 1994). Since C2-dnp53 cells, which stop proliferating upon induction of differentiation (Soddu et al., 1996), are drastically inhibited in their differentiation capacity and do not upregulate *Rb* (as assessed at the population level), we evaluated whether these cells can re-enter the cell cycle upon serum stimulation. We found that most of the MyHC-negative, undifferentiated cells, which stop proliferating after serum starvation, reactivated DNA synthesis as well as parental cells (80% in the C2-dnp53 population and 70% in the parental one). By contrast, MyHC-positive C2-dnp53 cells were not able to

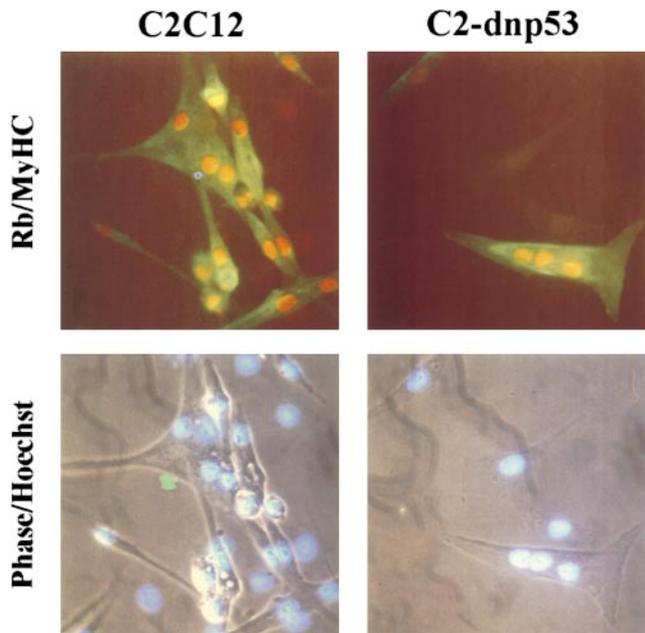


Figure 3. pRb expression in differentiated myotubes. (Top) Indirect immunofluorescence performed on C2C12 and C2-dnp53 cells 72 h after the incubation in DM. Anti-pRB mAb was revealed by TRITC-conjugated antiserum and anti-MyHC serum by FITC-conjugated antiserum. (Bottom) Hoechst-stained nuclei pictures superimposed on the relative phase contrast images of the same fields shown above.

reactivate the cell cycle any longer. Indeed, we could never find BrdUrd/MyHC double-positive cells in all the experiments performed. In addition, the expression of pRb protein was analyzed at the single-cell level by indirect immunofluorescence. As shown in Fig. 3, the C2-dnp53 cells that could differentiate had increased levels of pRb. Moreover, the hypophosphorylation of pRb, which is responsible for cell cycle withdrawal, was similarly present in parental and dnp53-expressing cells (data not shown), as well as in p53^{+/+} and p53^{-/-} myoblasts (Fig. 2 e). Taken together, these results indicate that p53 is not essential for the cell cycle-related functions of pRb.

p53 Controls the Differentiation Activity of pRb

A pRb function genetically distinct from cell cycle control affects the transcription of differentiation-related genes (Sellers et al., 1998; Novitch et al., 1999), pRb being required for the full function of the transcription factor MyoD during skeletal muscle differentiation (Gu et al., 1993; Novitch et al., 1996, 1999). Thus, MyoD transcriptional activity was measured in C2C12 and C2-dnp53 cells stably transfected with a reporter vector containing the MCK promoter. Upon induction of differentiation, a considerable accumulation of the luciferase reporter was found in parental C2C12 cells, while a strongly reduced accumulation was detectable in C2-dnp53 cells. This reduction was fully suppressed by infection with a recombinant adenovirus encoding an active pRb (Ad-ΔRb; Chang et al., 1995) (Fig. 4 a). Since MyoD activity was rescued by exogenous pRb overexpression, we evaluated the differentiation capacity of these cells. As shown in Fig. 4 b, Ad-ΔRb-infected C2-dnp53 cells differentiate as efficiently as

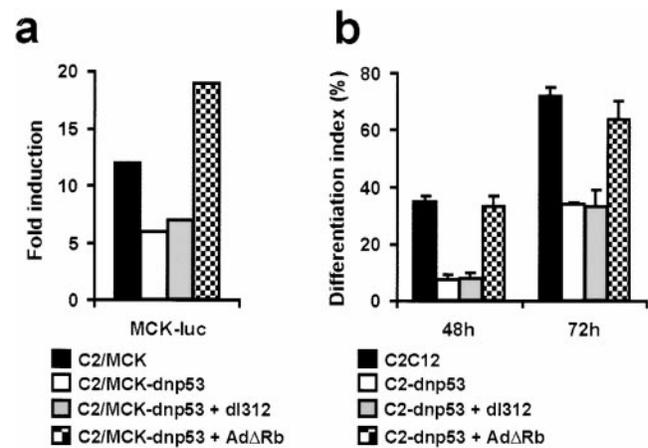


Figure 4. Effects of pRb overexpression in C2-dnp53 cells. (a) Activity of the MCK promoter in the cells indicated. C2C12 cells were stably transfected with the MCK reporter vector and subsequently infected with dnp53-carrying or insertless retrovirus. The C2-MCK-dnp53 cells were also infected with Ad-ΔRb virus to increase pRb expression or dl312 adenovirus as control. Fold induction values indicate the ratio between the values obtained in DM (48 h) and GM conditions. (b) Differentiation indices were evaluated at the indicated time points in C2C12 and C2-dnp53 cells before and after infection with adenovirus as in a.

the parental C2C12 cells, indicating that exogenous pRb overexpression is sufficient to recover the differentiation capacity of C2C12 cells lacking p53 activity. Altogether, these results strongly indicate that p53 controls the differentiation function of pRb.

Differentiation-associated Rb Upregulation Is Mediated at the Transcriptional and Posttranscriptional Levels

It has been suggested that, during myoblast differentiation, Rb mRNA upregulation is transcriptionally mediated (Martelli et al., 1994; Okuyama et al., 1996). Thus, the luciferase reporter vector pXRP1, which carries the -686 to -4 base-pair fragment of the human Rb promoter (Osifchin et al., 1994), was stably transfected into C2C12 cells. The kinetics of luciferase expression was evaluated in two different polyclonal populations upon induction of differentiation and compared with the levels of proliferating cells (time 0). A strong reduction in luciferase activity was present 24 h after differentiation induction. This initial downregulation was followed by the recovery of the promoter activity at the 48 h time point (Fig. 5 a), with no considerable difference between the two populations analyzed (b). Interestingly, in all the time courses performed, the differentiation-associated luciferase activity never reached the level measured in proliferating cells. This suggests the presence of a different type of Rb gene transcription between the proliferating and differentiating states. Thus, to detect possible differential occupations of cis-acting elements in the two conditions, in vivo footprintings were performed on the endogenous Rb promoter of C2C12 cells. The Rb promoter region responsible for the regulation during in vitro myogenesis (Okuyama et al., 1996) was analyzed. Proliferating and 48 h-differentiated cells showed different patterns of nucleotide protection (Fig. 5 c), supporting the idea of the presence of different transcriptional complexes in the two conditions compared.

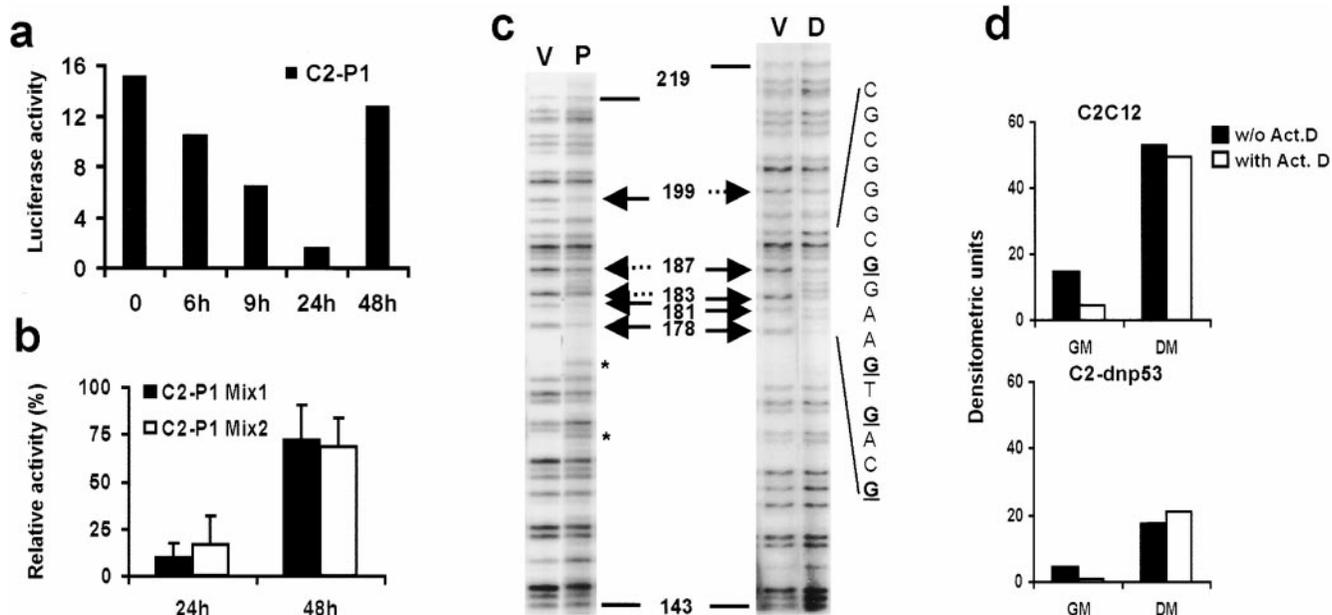


Figure 5. Rb promoter activity in muscle differentiation. (a) pRXP1-carrying C2C12 cells (C2-P1) were incubated in DM for different times. Time 0 indicates cells maintained in GM. Luciferase activity was normalized for number of nuclei. (b) Two different polyclonal populations of pRXP1-carrying C2C12 cells (C2-P1 Mix1 and C2-P1 Mix2) were incubated as in a. Columns report the luciferase activity of each cell line, relative to its own GM. (c) The region between -219 and -143 base-pairs upstream of the translation start site of the C2C12 endogenous *Rb* promoter was analyzed by in vivo footprinting in proliferating (P) and 48 h differentiated (D) cells; in vitro footprinting (V) was used as control. (Continuous arrows) Bands that are significantly protected; (dashed arrows) bands that are not protected; and (*) spurious bands. (d) Total RNA was extracted from C2C12 and C2-dnp53 cells maintained in GM or DM for 48 h, with or without Act D. Northern blottings were performed for the *Rb* and *GAPDH* genes. Histograms show quantitative analyses of the *Rb* gene expression levels relative to *GAPDH*.

Since the increment of *Rb* mRNA levels during C2C12 differentiation is not proportional to the increase in the *Rb* promoter activity in the same conditions, we asked whether posttranscriptional regulation of the *Rb* gene expression was also present. Northern blot analyses were performed on total RNA extracted from C2C12 and C2-dnp53 cells treated with the transcription inhibitor Actinomycin D (Act D). In proliferating conditions, reduction of *Rb* mRNA levels was found in both cell types compared with the relative untreated cells. At 48 h from differentiation induction, comparable amounts of *Rb* mRNA were present in cells treated and untreated with Act D, although, as expected, the absolute levels were different between parental and C2-dnp53 cells (Fig. 5 d).

Altogether, the results reported in this paragraph demonstrate that the differentiation-associated *Rb* upregulation is mediated at the transcriptional and posttranscriptional levels and that the latter type of control is p53 independent.

During Muscle Differentiation, p53 Contributes to the Transcriptional Regulation of the Rb Promoter through a p53-binding Site

Since the p53 transcriptional activity is required for the induction of differentiation (Soddu et al., 1996), we evaluated whether p53 contributes to the *Rb* gene transcription. pRXP1-carrying C2C12 cells were stably transduced with the dnp53 protein by retroviral infection and maintained as polyclonal populations. When these cells were incubated in DM, the *Rb* promoter showed the 24 h silencing that, however, was not followed by the normal 48 h recovery of transcriptional activity (Fig. 6 a). This finding indicates that p53

contributes to the increase of the *Rb* promoter activity that occurs during C2C12 differentiation after the initial silencing. The timing of this recovered activity correlates with the p53-specific transcriptional activity that we previously measured on the artificial p53-reporter vector PG₁₃-CAT (Soddu et al., 1996). Notably, no difference in *Rb* promoter activity was found between parental and dnp53-expressing C2C12 cells in proliferating conditions, as assessed by stable and transient transfections of C2C12 and C2-dnp53 cells with the pRXP1 (data not shown). These results further support the idea that p53 is involved only in the differentiation-associated upregulation of the *Rb* gene.

Since a sequence positively regulated by p53 had been mapped in the *Rb* promoter between -92 and -4 base-pairs upstream of the translation start site (Osifchin et al., 1994), we stably transfected into C2C12 cells a pRXP1 vector carrying a deletion of this region (pd92) (Osifchin et al., 1994). As shown in Fig. 6 b, after the expected promoter silencing at 24 h, no activation of pd92 transcription was present at 48 h, supporting the hypothesis that during C2C12 differentiation p53 modulates the *Rb* promoter through the p53-binding site present in this region. To confirm this hypothesis, the p53-binding site was mutated in the pRXP1 vector, creating the pRXP1-mtp53 that was stably transduced into C2C12 cells. Upon differentiation induction, the levels of luciferase activity of pd92 and pRXP1-mtp53 were comparable (Fig. 6 b), indicating that p53 regulates *Rb* transcription through this p53-binding site.

Because of the particularly high content in Guanine-Cytosine of the *Rb* promoter region containing the p53-binding site, we were not able to perform in vivo footprintings of this site. Thus, to verify the in vivo p53 binding to the

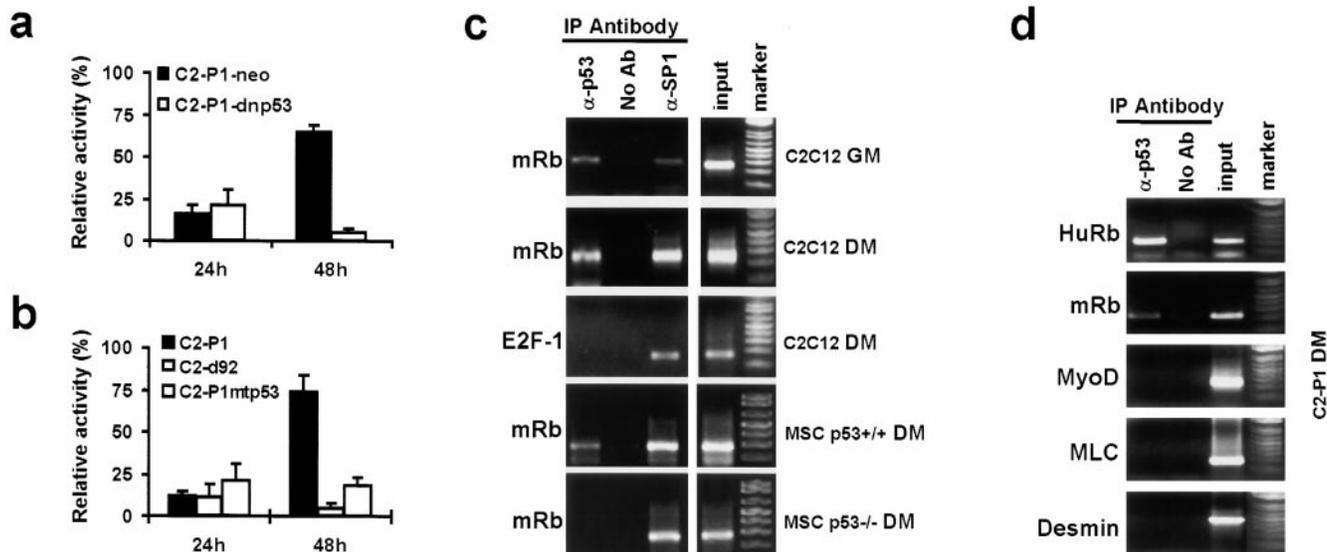


Figure 6. p53 activity on the Rb promoter. (a) C2-P1 cells were infected with a recombinant retrovirus carrying the *dnp53* gene or its insertless control. Relative luciferase activity was assessed on polyclonal populations as described in Fig. 5. (b) C2C12 cells were stably transduced with pXRp1 (C2-P1), pd92 (C2-d92), and pXRp1-mtp53 (C2-P1mtp53). Relative luciferase activity was assessed on polyclonal populations as in a. (c and d) Cross-linked chromatin from the indicated cells was immunoprecipitated with antibodies to p53, Sp1, or in the absence of antibodies, and analyzed by PCR with primers specific for the indicated promoters (see Materials and Methods). Sp1 was used as positive control in the promoters with an SP1-binding site; input corresponds to nonimmunoprecipitated cross-linked chromatin.

proximal region of the *Rb* promoter, a chromatin-cross-linked immunoprecipitation was performed (Boyd et al., 1998) in C2C12 cells maintained in the presence of GM or DM and in $p53^{+/+}$ and $p53^{-/-}$ MSC incubated in DM. Proteins were cross-linked to DNA by direct incubation of living cells with formaldehyde. DNA was extracted, immunoprecipitated with anti-p53 or anti-SP1 sera, and the *Rb* promoter sequence was amplified from the immunoprecipitate by PCR. We found the proximal region of the *Rb* promoter in the p53-bound cross-linked chromatin of C2C12 and $p53^{+/+}$ MSC (Fig. 6 c), indicating the presence of p53 in the complexes that bind the *Rb* promoter in vivo. The *Rb* promoter sequence could not be amplified from the anti-p53 immunoprecipitated chromatin obtained from $p53^{-/-}$ MSC, demonstrating the specificity of the anti-p53 immunoprecipitation. In the same experiment, the E2F promoter that contains the SP1-binding site, but not the p53-binding site, was used as control. To further confirm p53 binding to the *Rb* promoter, the C2-P1 cells carrying the exogenous human *Rb* promoter were in vivo cross-linked. The sequence of exogenous human and endogenous murine *Rb* promoters, as well as a series of muscle specific promoters that do not contain p53-binding sites were amplified from the same DNA immunoprecipitations (Wells et al., 2000). As shown in Fig. 6 d, both human and mouse *Rb* promoters could be amplified, while no amplification was detectable in the control promoters. These results support the conclusion that p53 binds the *Rb* promoter in muscle cells.

Discussion

The *TP53* gene has been largely characterized as a cell-cycle checkpoint controller and an apoptosis inducer. In addition, different studies performed in vivo and in vitro have indicated a p53 involvement in embryonic development and in cell differentiation. However, the mechanisms through which p53 influences differentiation and development are

substantially unknown. In the present report, we show that, during skeletal muscle differentiation, p53 is required to establish the increased level of pRb expression, which regulates the further steps of muscle differentiation (Fig. 7).

The pRb requirement in skeletal muscle differentiation has been extensively studied. Inactivation of the pRb function by proteins from DNA-tumor viruses, as well as studies performed with $Rb^{-/-}$ cells have shown that pRb is involved in cell-cycle withdrawal and in the expression of late markers of differentiation (Gu et al., 1993; Chen et al., 1996; Novich et al., 1996, 1999). These two pRb functions can be genetically and mechanistically dissociated, as it has been elegantly demonstrated by studying pRb mutants from high- and low-risk retinoblastomas (Sellers et al., 1998). pRb can inhibit DNA synthesis thanks to its ability to bind E2F and DNA, and to repress transcription through a specific domain. In contrast, the abilities to activate transcription and promote differentiation do not require E2F binding, but rather need cooperation with other types of transcription factors, such as MyoD. We found that, in the presence of differentiation-promoting stimuli, p53-impaired myoblasts maintain the capacity to hypophosphorylate pRb and to withdraw from the cell cycle, while losing their capacity to increase pRb expression and have strongly reduced MyoD activity and differentiation potential. These findings indicate that p53 discriminates between the cell cycle- and differentiation-related functions of pRb and provide a biological counterpart to the biochemical characterization of the pRb mutants described above (Sellers et al., 1998). Furthermore, our results are in good agreement with the observations indicating that the pRb hypophosphorylation responsible for the differentiation-associated cell cycle withdrawal involves a p53-independent $p21^{Waf1/Cip1}$ up-regulation (Halevy et al., 1995; Parker et al., 1995).

In physiological proliferating conditions, our p53-impaired myoblasts did not show any modification of the *Rb* gene expression, as shown by Northern and Western blot,

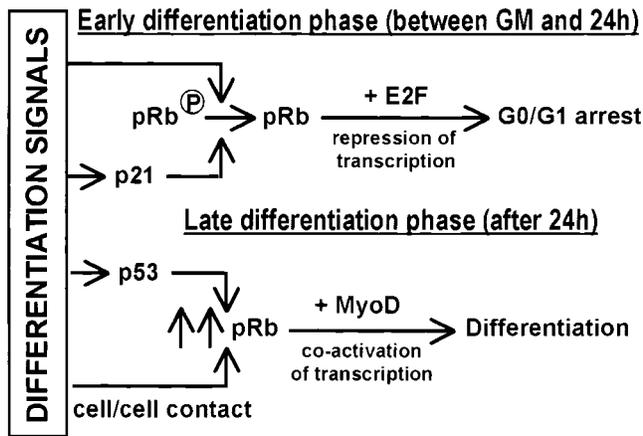


Figure 7. A model to explain p53 role in the induction of skeletal muscle differentiation. Early steps of differentiation include the activation of p21^{Waf1/Cip1} and the hypophosphorylation of pRb, which leads to the transcriptional repression of the E2F-activated genes. Findings (Halevy et al., 1995; Parker et al., 1995; Soddu et al., 1996; this report) indicate that these steps are p53 independent. After the cell-cycle withdrawal, p53 modifies the *Rb* gene transcription and contributes to the increase of pRb expression levels. Therefore, the upregulated pRb can cooperate with MyoD to transcribe later markers of differentiation. Cell-to-cell contacts can also increase pRb expression, overcoming the p53 requirement (Coen, S., and S. Soddu, unpublished results). Further and later functions of p53, such as the direct transcription of MCK promoter (Tamir and Bengal, 1998) are not included in this model since they are related to the activation of structural rather than regulatory genes.

and by promoter activation analyses. These observations are consistent with the notion that p53 is not involved in cell cycle control in normal proliferating conditions. In contrast, it is well known that different types of stressing stimuli promote p53 activation. In this type of situation, p53 is known to promote pRb hypophosphorylation and inhibition of DNA synthesis through the transcriptional induction of p21^{Waf1/Cip1} (Dulic et al., 1994). Indeed, compared with the parental cells, C2-dnp53 cells do not arrest in the G1 phase of the cell cycle in response to doxorubicin-induced DNA damage (Manni et al., manuscript submitted for publication). Together with the findings we obtained in differentiating conditions, these results indicate the presence of two different types of p53-dependent regulation of pRb. One operates through p21^{Waf1/Cip1} transcription, and the other through direct *Rb* transcription. These observations are consistent with the emerging idea that p53 regulates transcription of different genes, depending on the type of stimuli that provoked its activation (Zhao et al., 2000). Interestingly, the existence of a positively regulated p53-binding site on the *Rb* promoter has been known for several years (Osifchin et al., 1994), but no transcriptional induction of the *Rb* gene was found in apoptotic or growth-arresting situations, so far. Our results reveal the existence of a physiological condition in which p53 directly transactivates the *Rb* gene.

Recently, a family of p53-related proteins has been discovered. This family includes, besides p53, p73 and p63 proteins that share homology with the activation, the DNA-binding, and the oligomerization domains of p53. These proteins, when overexpressed in mammalian cells, can transactivate some of the p53 target genes and appear

to be involved in differentiation and development pathways (Oren, 1997; Kaelin, 1999; Mills et al., 1999; Yang et al., 1999, 2000). The genetic analysis we performed by comparing the differentiation abilities of primary myoblasts explanted from p53^{+/+} and p53^{-/-} mice allowed us to distinguish the function of p53 from those of its family members, demonstrating that the role played by p53 in muscle differentiation is specific of this protein. This result suggests that p53 and its family members have no overlapping activities on differentiation and thus might not be functionally redundant during ontogenesis. These considerations leave open the discrepancy between impaired differentiation of p53 null muscle cells in vitro and the apparently normal muscle development in p53 null mice. Temporary, slight defects in muscle development cannot be ruled out since, to our knowledge, detailed developmental analyses of skeletal muscles have not yet been performed in p53^{-/-} mice. On the other hand, an explanation for this discrepancy might be suggested by the observation that, in dnp53-expressing C2C12 cells, the inhibition of differentiation is present only when these cells are stimulated to differentiate by growth factor withdrawal (Soddu et al., 1996) (Fig. 7). It is known that skeletal muscle differentiation in vitro can be obtained by two different stimuli, growth factor deprivation and cell to cell contact (Martelli et al., 1994). We have found that cell confluence, even in the presence of serum, is able to increase pRb expression in a p53-independent manner (Coen, S., and S. Soddu, unpublished results). Differentiation-promoting stimuli due to cell to cell contact can be avoided in vitro, but not during in vivo development, suggesting that they can provide a functional replacement for p53. We do not know the molecular mediators of this effect nor whether it is relevant for development; nevertheless, the strong and frequent association between p53 alterations and the undifferentiated phenotypes described in many human cancers (Feinstein et al., 1991; Fagin, 1995) is in agreement with this hypothesis. Indeed, one peculiar characteristic of tumor transformation is the loss of contact inhibition. This would make tumor cells more sensitive than their normal counterparts to the absence of p53 in terms of differentiation capacity. If proved, this hypothesis might solve an apparent incongruity that has appeared between clinical and basic biological researches in the p53 field. While basic research has mostly studied the control of cell-cycle checkpoints and the apoptotic function of p53, a large number of clinical and epidemiological works have associated *TP53* gene alterations to loss of tumor differentiation (anaplasia). A commonly accepted model has it that genomic instability is the cause of anaplasia, through direct alterations of regulatory and structural differentiation genes. However, this hypothesis is in contrast with the differentiation-promoting effects of different compounds or of the exogenous expression of the wild-type p53 protein per se. Indeed, both approaches induce differentiation in tumor cells without correcting the genomic instability, suggesting a dual role for p53 in tumor development as gatekeeper (Levine, 1997) and differentiation controller. The finding that p53 regulates not only the cell-cycle progression function of pRb, but also its differentiation-promoting function, matches the *TP53* gene to the prototypic oncosuppressors, which are essential for embryonic development and play important roles in regulating cell cycle and differentiation.

We are particularly grateful to P.D. Robbins and M. Oren for generous gifts of *Rb* promoter vectors and p53-recombinant retroviruses, to C. Colussi for handling the p53^{-/-} mice, to G. Bossi for primary cell preparation, and to M. Crescenzi for constructive criticisms and helpful discussion.

This work was supported by Telethon-Italy and Associazione Italiana per la Ricerca sul Cancro. A. Gurtner is the recipient of a fellowship from Fondazione Italiana per la Ricerca sul Cancro.

Submitted: 31 May 2000

Revised: 16 October 2000

Accepted: 17 October 2000

References

- Almog, N., and V. Rotter. 1997. Involvement of p53 in cell differentiation and development. *Biochim. Biophys. Acta.* 1333:F1-F27.
- Almon, E., N. Goldfinger, A. Kapon, N. Schwartz, A.J. Levine, and V. Rotter. 1993. Testicular tissue-specific expression of the p53 suppressor gene. *Dev. Biol.* 156:107-116.
- Armstrong, J.F., M.H. Kaufman, D.J. Harrison, and A.R. Clarke. 1995. High-frequency developmental abnormalities in p53-deficient mice. *Curr. Biol.* 5:931-936.
- Ausbel, F.M., R. Brent, R.E. Kinston, D.D. Moore, J.G. Seidman, J.A. Smith, and K. Struhl. 1987. Current Protocols in Molecular Biology. Volume 1. Wiley Interscience, New York, NY. 3.5.9 and 4.9.1-4.9.7
- Bacchetti, S., and F. Graham. 1993. Inhibition of cell proliferation by an adenovirus vector expressing the human wild type p53 protein. *Int. J. Oncol.* 3:781-788.
- Bader, D., T. Masaki, and D.A. Fischman. 1982. Immunochemical analyses of myosin heavy chain during avian myogenesis in vivo and in vitro. *J. Cell Biol.* 95:763-770.
- Bossi, G., R. Scardigli, P. Musiani, R. Martinelli, M.P. Gentileschi, S. Soddu, and A. Sacchi. 2000. Development of a murine orthotopic model of leukemia: evaluation of TP53 gene therapy efficacy. *Cancer Gene Ther.* 7:135-143.
- Boyd, K.E., J. Wells, J. Graham, S.M. Bartley, and P.J. Farnham. 1998. c-Myc target gene specificity is determined by a post-DNA-binding mechanism. *Proc. Natl. Acad. Sci. USA.* 95:13887-13892.
- Cerone, M.A., A. Marchetti, G. Bossi, G. Blandino, A. Sacchi, and S. Soddu. 2000. p53 is involved in the differentiation but not in the differentiation-associated apoptosis of myoblasts. *Cell Death Diff.* 7:506-508.
- Chang, M.W., E. Barr, J. Seltzer, Y.Q. Jiang, G.J. Nabel, M.S. Parmacek, and J.M. Leiden. 1995. Cytostatic gene therapy for vascular proliferative disorders with a constitutively active form of the retinoblastoma gene product. *Science.* 27:518-522.
- Chen, P.L., D.J. Riley, S. Chen-Kiang, and W.-H. Lee. 1996. Retinoblastoma protein directly interacts with and activates the transcription factor NF-IL6. *Proc. Natl. Acad. Sci. USA.* 93:465-469.
- Choi, J., and L.A. Donehower. 1999. p53 in embryonic development: maintaining a fine balance. *Cell. Mol. Life Sci.* 55:38-47.
- Crescenzi, M., D.H. Crouch, and F. Tatò. 1994. Transformation by myc prevents fusion but not biochemical differentiation of C2C12 myoblasts: mechanisms of phenotypic correction in mixed culture with normal cells. *J. Cell Biol.* 125:1137-1145.
- Crescenzi, M., S. Soddu, and F. Tatò. 1995. Cell cycle reactivation in terminally differentiated cells by adenovirus infection. *J. Cell Physiol.* 162:26-35.
- Davison, T.S., C. Vagner, M. Kaghad, A. Ayed, D. Caput, and C.H. Arrow-smith. 1999. p73 and p63 are homotetramers capable of weak heterotypic interactions with each other but not with p53. *J. Biol. Chem.* 274:18709-18714.
- Di Como, C.J., C. Gaiddon, and C. Prives. 1999. p73 function is inhibited by tumor-derived p53 mutants in mammalian cells. *Mol. Cell Biol.* 19:1438-1449.
- Donehower, L.A., M. Harvey, B.L. Slagle, M.J. McArthur, C.A. Montgomery, Jr., J.S. Butel, and A. Bradley. 1992. Mice deficient for p53 are developmentally normal but susceptible to spontaneous tumors. *Nature.* 356:215-221.
- Dulic, V., W.K. Kaufmann, S.J. Wilson, T.D. Tlsty, E. Lees, J.W. Harper, S.J. Elledge, and S.I. Reed. 1994. p53 dependent inhibition of cyclin-dependent kinase activities in human fibroblasts during radiation-induced G1 arrest. *Cell.* 76:1013-1023.
- Fagin, J.A. 1995. Tumor suppressor genes in human thyroid neoplasms: p53 mutations are associated with undifferentiated thyroid cancers. *J. Endocrinol. Invest.* 18:140-142.
- Feinstein, E., R.P. Gale, J. Reed, and E. Canaani. 1991. p53 in chronic myelogenous leukemia in acute phase. *Proc. Natl. Acad. Sci. USA.* 88:6293-6297.
- Gu, W., J.W. Schneider, G. Condorelli, S. Kaushal, V. Mahdavi, and B. Nadal-Ginard. 1993. Interaction of myogenic factors and the retinoblastoma protein mediates muscle cell commitment and differentiation. *Cell.* 72:309-324.
- Halevy, O., B.G. Novitch, D.B. Spicer, S.X. Skapek, J. Rhee, G.J. Hannon, D. Beach, and A.B. Lassar. 1995. Correlation of terminal cell cycle arrest of skeletal muscle with induction of p21 by MyoD. *Science.* 267:1018-1021.
- Kastan, M.B., A.I. Radin, S.J. Kuerbitz, O. Onyekwere, C.A. Wolkow, C. Civin, K.D. Stone, T. Woo, Y. Ravindranath, and R.W. Craig. 1991. Levels of p53 protein increase with maturation in human hematopoietic cells. *Cancer Res.* 51:4279-4286.
- Kaelin, W.J., Jr. 1999. The emerging p53 gene family. *J. Nat. Cancer Inst.* 91: 594-598.
- Levine, A.J. 1997. p53, the cellular gatekeeper for growth and division. *Cell.* 88: 323-331.
- Martelli, F., C. Cenciarelli, G. Sartorelli, B. Polikar, A. Felsani, and M. Caruso. 1994. MyoD induces retinoblastoma gene expression during myogenic differentiation. *Oncogene.* 9:3579-3590.
- Martinez-Balbas, M.A., A. Dey, S. Rabindran, K. Ozato, and C. Wu. 1995. Displacement of sequence-specific transcription factors from mitotic chromatin. *Cell.* 83:29-38.
- Mazzaro, G., G. Bossi, S. Coen, A. Sacchi, and S. Soddu. 1999. The role of wild-type p53 in the differentiation of primary hemopoietic and muscle cells. *Oncogene.* 18:5831-5835.
- Melkintin, J.D., and E.N. Olson. 1996. Defining the regulatory networks for muscle development. *Curr. Opin. Genet. Dev.* 6:445-453.
- Michalovitz, D., O. Halevy, and M. Oren. 1990. Conditional inhibition of transformation and of cell proliferation by a temperature-sensitive mutant of p53. *Cell.* 62:671-680.
- Mills, A.A., B. Zheng, X.-J. Wang, H. Vogel, D.R. Roop, and A. Bradley. 1999. p63 is a p53 homologue required for limb and epidermal morphogenesis. *Nature.* 398:708-713.
- Milner, J., and E.A. Medcalf. 1991. Cotranslation of activated mutant p53 with wild type drives the wild-type p53 protein into the mutant conformation. *Cell.* 65:765-774.
- Montes de Oca Luna, R., D.S. Wagner, and G. Lozano. 1995. Rescue of early embryonic lethality in mdm2-deficient mice by deletion of p53. *Nature.* 378: 203-206.
- Novitch, B.G., G.J. Mulligan, T. Jacks, and A.B. Lassar. 1996. Skeletal muscle cells lacking the retinoblastoma protein display defects in muscle gene expression and accumulate in S and G2 phases of the cell cycle. *J. Cell Biol.* 135:441-456.
- Novitch, B.G., D.B. Spicer, P.S. Kim, W.L. Cheung, and A.B. Lassar. 1999. pRb is required for MEF2-dependent gene expression as well as cell-cycle arrest during skeletal muscle differentiation. *Curr. Biol.* 9:449-459.
- Okuyama, Y., Y. Sowa, T. Fujita, T. Mizumura, H. Nomura, T. Nikaido, T. Endo, and T. Sakai. 1996. ATF site of human Rb gene promoter is a responsive element of myogenic differentiation. *FEBS Lett.* 397:219-224.
- Oren, M. 1997. Lonely no more: p53 finds its kin in a tumor suppressor haven. *Cell.* 90:829-832.
- Osifchin, N.E., D. Jiang, N. Ohtani-Fujita, T. Fujita, M. Carrozza, S.-J. Kim, T. Sakai, and P.D. Robbins. 1994. Identification of a p53 binding site in the human retinoblastoma susceptibility gene promoter. *J. Biol. Chem.* 269:6383-6389.
- Parker, S.B., G. Eichele, P. Zhang, A.T. Sands, A. Bradley, E.N. Olson, J.W. Harper, and S.J. Elledge. 1995. p53-independent expression of p21^{Cip1} in muscle and other terminally differentiating cells. *Science.* 267:1024-1027.
- Sah, V.P., L.D. Attardi, G.J. Mulligan, B.O. Williams, R.T. Bronson, and T. Jacks. 1995. A subset of p53-deficient embryos exhibits exencephaly. *Nat. Genet.* 10:175-180.
- Schmid, P., A. Lorenz, H. Hameister, and M. Montenarh. 1991. Expression of p53 during mouse embryogenesis. *Development (Camb.)*. 113:857-865.
- Schneider, J.W., W. Gu, L. Zhu, V. Mahdavi, and B. Nadal-Ginard. 1994. Reversal of terminal differentiation mediated by p107 in Rb^{-/-} muscle cells. *Science.* 264:1467-1471.
- Sellers, W.K., B.G. Novitch, S. Miyake, A. Heith, G.A. Otterson, F.J. Kaje, A.B. Lassar, and W.J. Kaelin, Jr. 1998. Stable binding of E2F is not required for the retinoblastoma protein to activate transcription, promote differentiation, and suppress tumor cell growth. *Genes Dev.* 12:95-106.
- Soddu, S., G. Blandino, R. Scardigli, S. Coen, A. Marchetti, M.G. Rizzo, G. Bossi, L. Cimino, M. Crescenzi, and A. Sacchi. 1996. Interference with p53 protein inhibits hemopoietic and muscle differentiation. *J. Cell Biol.* 134:193-204.
- Tamir, Y., and E. Bengal. 1998. p53 protein is activated during muscle differentiation and participates with MyoD in the transcription of muscle creatine kinase gene. *Oncogene.* 17:347-356.
- Wallington, J.B., D.W. Seufert, V.C. Virta, and P.D. Vize. 1997. p53 activity is essential for normal development in *Xenopus*. *Curr. Biol.* 7:747-757.
- Weinberg, R.A. 1995. The retinoblastoma protein and the cell cycle control. *Cell.* 81:323-330.
- Weintraub, H., R. Davis, S. Tapscott, M. Thayer, M. Krause, R. Benezra, K.T. Blackwell, D. Turner, R. Rupp, S. Hollenberg, et al. 1991. The MyoD gene family: nodal point during specification of the muscle cell lineage. *Science.* 251:761-766.
- Wells, J., K.E. Boyd, C.J. Fry, S.M. Bartley, and P.J. Farnham. 2000. Target gene specificity of E2F and pocket protein family members in living cells. *Mol. Cell Biol.* 20:5797-5807.
- Wright, W.E. 1992. Muscle basic helix-loop-helix proteins and the regulation of myogenesis. *Curr. Opin. Genet. Dev.* 2:243-248.
- Yang, A., R. Schweitzer, D. Sun, M. Kaghad, N. Walker, R.T. Bronson, C. Tabin, A. Sharpe, D. Caput, C. Crum, and F. McKeon. 1999. p63 is essential for regenerative proliferation in limb, craniofacial and epithelial development. *Nature.* 398:714-718.
- Yang, A., N. Walker, R. Bronson, M. Kaghad, M. Oosterwegel, J. Bonnin, C. Vagner, H. Bonnet, P. Dikkes, A. Sharpe, et al. 2000. p73-deficient mice have neurological, pheromonal and inflammatory defects but lack spontaneous tumors. *Nature.* 404:99-103.
- Yun, K., and B. Wold. 1996. Skeletal muscle determination and differentiation: story of a core regulatory network and its context. *Curr. Opin. Cell Biol.* 8:877-889.
- Zhao, R., K. Gish, M. Murphy, Y. Yin, D. Notterman, W.H. Hoffman, E. Tom, D.H. Mack, and A.J. Levine. 2000. Analysis of p53-regulated gene expression patterns using oligonucleotide arrays. *Genes Dev.* 14:981-993