

RESEARCH ARTICLE

# Delta Opioid Receptors: The Link between Exercise and Cardioprotection

Juliana P. Borges<sup>1</sup>, Karine S. Verdoorn<sup>2</sup>, Anissa Daliry<sup>1</sup>, Scott K. Powers<sup>3</sup>, Victor H. Ortenzi<sup>4</sup>, Rodrigo S. Fortunato<sup>4</sup>, Eduardo Tibiriçá<sup>1</sup>, Marcos Adriano Lessa<sup>1\*</sup>

1. Laboratory of Cardiovascular Investigation, Oswaldo Cruz Institute, FIOCRUZ, Rio de Janeiro, RJ, Brazil, 2. Federal University of Rio de Janeiro, Macaé, RJ, Brazil, 3. Department of Applied Physiology and Kinesiology, University of Florida, Gainesville, FL, United States of America, 4. Carlos Chagas Filho Biophysics Institute, Federal University of Rio de Janeiro, Rio de Janeiro, RJ, Brazil

\*[malessa@ioc.fiocruz.br](mailto:malessa@ioc.fiocruz.br)



CrossMark  
click for updates

 OPEN ACCESS

**Citation:** Borges JP, Verdoorn KS, Daliry A, Powers SK, Ortenzi VH, et al. (2014) Delta Opioid Receptors: The Link between Exercise and Cardioprotection. PLoS ONE 9(11): e113541. doi:10.1371/journal.pone.0113541

**Editor:** John Calvert, Emory University, United States of America

**Received:** August 4, 2014

**Accepted:** October 27, 2014

**Published:** November 21, 2014

**Copyright:** © 2014 Borges et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Data Availability:** The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper and its Supporting Information files.

**Funding:** JPB, MAL and ET received funding from Oswaldo Cruz Institute, Conselho Nacional de Desenvolvimento Científico e Tecnológico and Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

## Abstract

This study investigated the role of opioid receptor (OR) subtypes as a mechanism by which endurance exercise promotes cardioprotection against myocardial ischemia-reperfusion (IR) injury. Wistar rats were randomly divided into one of seven experimental groups: 1) control; 2) exercise-trained; 3) exercise-trained plus a non-selective OR antagonist; 4) control sham; 5) exercise-trained plus a kappa OR antagonist; 6) exercise-trained plus a delta OR antagonist; and 7) exercise-trained plus a mu OR antagonist. The exercised animals underwent 4 consecutive days of treadmill training (60 min/day at ~70% of maximal oxygen consumption). All groups except the sham group were exposed to an in vivo myocardial IR insult, and the myocardial infarct size (IS) was determined histologically. Myocardial capillary density, OR subtype expression, heat shock protein 72 (HSP72) expression, and antioxidant enzyme activity were measured in the hearts of both the exercised and control groups. Exercise training significantly reduced the myocardial IS by approximately 34%. Pharmacological blockade of the kappa or mu OR subtypes did not blunt exercise-induced cardioprotection against IR-mediated infarction, whereas treatment of animals with a non-selective OR antagonist or a delta OR antagonist abolished exercise-induced cardioprotection. Exercise training enhanced the activities of myocardial superoxide dismutase (SOD) and catalase but did not increase the left ventricular capillary density or the mRNA levels of HSP72, SOD, and catalase. In addition, exercise significantly reduced the protein expression of kappa and delta ORs in the heart by 44% and 37%, respectively. Together, these results indicate that ORs contribute to the cardioprotection conferred by endurance exercise, with the delta OR subtype playing a key role in this response.

## Introduction

The concept that regular exercise is cardioprotective against ischemia-reperfusion (IR) cardiac injury is well established in animal models [1], [2], and human epidemiological studies also suggest that physically active individuals are protected against IR-induced myocardial injury [3–5]. Indeed, physically active individuals have a lower incidence of myocardial infarction and a greater survival rate following a heart attack compared with their less active counterparts [1], [3].

In reference to animal investigations of cardioprotection, well-controlled animal studies beginning in the late 1970s have provided convincing evidence that regular bouts of endurance exercise provide cardioprotection against IR-induced injury [1], [2]. Although it is clear that endurance exercise produces a cardioprotective phenotype, the mechanisms responsible for this phenomenon remain unclear. Some studies suggest that exercise promotes cardioprotection, at least in part, by direct effects on the myocardium [1], [2], [6]. Specific mechanisms that may be responsible for exercise-induced cardioprotection include the following: (a) increased nitric oxide production and cardiac antioxidant capacity [2], [7–11]; (b) expansion of the coronary capillary network and enlargement of the coronary artery diameter [2], [7]; (c) increased production of heat shock proteins (HSP) [2], [7], [8]; (d) reduced production of reactive oxygen species (ROS) in myocardial mitochondria during IR [7], [12]; and (e) improved function of sarcolemmal and/or mitochondrial ATP-sensitive potassium channels (sarco/mito K<sup>+</sup> ATP channels) [7], [8].

Additionally, recent evidence suggests that endogenous opioids may contribute to exercise-induced cardioprotection [13], [14]. Dickson et al. [13] demonstrated that the exercise-induced reduction in infarct size (IS) was abolished following the blockade of opioid receptors (OR) with naloxone. Similarly, Michelsen et al. [14] showed that naloxone treatment blocked the exercise-induced cardioprotection transferred from a remotely preconditioned human donor to an isolated perfused rabbit heart using a dialysate of plasma.

Identical to exercise-induced cardioprotection, ischemic preconditioning (IPC), which consists of performing intermittent brief exposures of the myocardium to ischemia, protects the heart against a subsequent severe episode of ischemia [15]. Strong evidence also indicates that endogenous opioids are involved in cardioprotection induced by IPC [14], [16–20]. Intriguingly, despite the putative role of the opioid system as a mechanism for IPC, the involvement of different subtypes of ORs in the cardioprotective effect of exercise is not completely understood. To date, few studies have investigated the role that OR activation plays in exercise-induced cardioprotection [13], [21]. Therefore, the current experiments were designed to identify the specific OR subtype responsible for exercise-induced cardioprotection. Based upon our preliminary experiments, we hypothesized that the delta OR subtype plays an essential role in exercise-induced cardioprotection.

## Materials and Methods

### Animals and ethics statement

Male Wistar rats (250–300 g) were housed under controlled light (12:12 h light-dark cycle) and temperature ( $22 \pm 1^\circ\text{C}$ ) conditions with free access to water and standard rat chow.

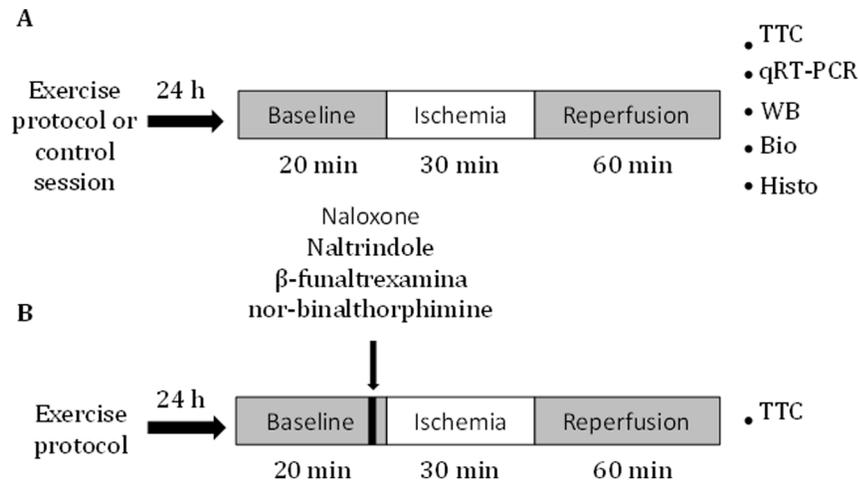
This study was carried out in strict accordance with the Guidelines for the Care and Use of Laboratory Animals of the National Institutes of Health. The Oswaldo Cruz Foundation Animal Welfare Committee approved all protocols (permit number: LW-4/11). All surgeries were performed under sodium pentobarbital anesthesia, and all efforts were made to minimize animal suffering.

### Experimental design

These experiments evaluated whether the delta OR subtype plays an essential role in exercise-induced cardioprotection. The experimental groups were subjected to endurance exercise training and treated with both non-selective and selective OR antagonists prior to exposure to a myocardial IR insult. The rats were randomly divided into one of seven experimental groups: 1) control (CT;  $n=20$ ); 2) exercised (exe;  $n=18$ ); 3) exercised plus treatment with a non-selective OR antagonist (Exe+NAL;  $n=10$ ); 4) control sham (Sham;  $n=10$ ); 5) exercised plus treatment with the kappa OR (KOR) antagonist (Exe+KOR;  $n=5$ ); 6) exercised plus treatment with the delta OR (DOR) antagonist (Exe+DOR;  $n=5$ ); and 7) exercised plus treatment with a mu OR (MOR) antagonist (Exe+MOR;  $n=5$ ). All of the groups were exposed to IR injury surgery (see below), with the exception of the sham group, which was submitted to sham injury surgery without exposure to IR. Following reperfusion, the left ventricles were excised to histologically quantify the IS and assess a variety of biochemical measurements ([Figure 1](#)).

### Opioid receptor (OR) blockade

To identify which of the three OR subtypes are involved in exercise-induced cardioprotection, 10 min prior to the induction of myocardial ischemia, select groups of animals received an intravenous injection of a selective OR antagonist. Specifically, the group of animals assigned to the DOR antagonist received 3 mg/kg of naltrindole. Similarly, animals assigned to MOR blockade were treated with 3 mg/kg of  $\beta$ -funaltrexamine, and animals assigned to KOR blockade were injected with 3 mg/kg of nor-binalthorphan. Finally, animals assigned to the non-selective OR group received 3 mg/kg of naloxone to block all OR receptor subtypes. The 3 mg/kg dose of OR antagonists was chosen based on previous results from Schultz et al. [[17](#)], [[44](#)] and on preliminary experiments from our lab.



**Figure 1. Diagrammatic representation of the experimental design of the study.** (A) Left ventricles obtained from the exercised and control groups were submitted to qRT-PCR, Western blotting (WB), histochemical (Histo), and biochemical (Bio) analyses and TTC staining. (B) Left ventricles obtained from the non-selective (naloxone) and selective OR antagonist (naltrindole,  $\beta$ -funaltrexamine and nor-binalthorphan) groups were submitted to TTC staining.

doi:10.1371/journal.pone.0113541.g001

## Endurance exercise training

The rats were familiarized to treadmill running using a low-speed, motor-driven rodent treadmill (HT 2.0, Hectron Fitness Equipment, RJ, Brazil). The animals began by walking at 12 m/min (0% grade) for 15 min/day on three consecutive days. Following this brief period of familiarization, maximal oxygen consumption ( $VO_{2max}$ ) was measured during the last stage of a graded treadmill exercise test. The graded exercise protocol began with the rats running at 10 m/min (0% grade) with the treadmill speed increasing by 3 m/min every 3 min until the animals could no longer maintain the desired running speed.  $VO_{2max}$  was measured by assessing the total airflow through the treadmill chamber and assessing the oxygen content of the expired gas using an electronic oxygen analyzer (AVS Projetos, SP, Brazil). The measurement of  $VO_{2max}$  was then used to establish the appropriate treadmill speed to elicit a relative exercise intensity of  $\sim 70\% VO_{2max}$ .

Following the  $VO_{2max}$  test, the animals completed an additional four consecutive days of exercise training (60 min/day, 0% grade, at  $\sim 70\%$  of  $VO_{2max}$ ). This exercise training protocol was selected because this training program has been shown to promote cardioprotection against IR injury [21]. The exercise training occurred between 8:00 and 10:00 am each day. To control for handling/environmental stress between experimental groups, the animals assigned to the sedentary groups were placed on a non-moving treadmill for 60 minutes. Twenty-four hours after the last exercise or control session, the animals were exposed to IR injury surgery.

## Surgical procedures and infarct area assessment

Anesthesia was induced and maintained with sodium pentobarbital (70 mg/kg intraperitoneally (ip) and 5 mg/kg intravenously (iv), respectively), and the animals were tracheotomized and artificially ventilated (Rodent Ventilator 7025, Ugo Basile, Varese, Italy) with a respiratory frequency of 50 breaths/min. Polyethylene-tipped cannulas filled with heparinized saline solution were inserted into the left femoral artery and the right jugular vein for direct measurements of arterial pressure and drug administration, respectively. The arterial cannula was connected to a strain-gauge transducer (TSD104A, BIOPAC Systems, CA, USA), and pressure signals were channeled through a signal amplifier (DA100C, BIOPAC Systems, CA, USA). Data were acquired at a rate of 1,000 samples/s through the MP150 system (BIOPAC Systems, CA, USA) and recorded using AcqKnowledge 3.7.3 for Windows (BIOPAC Systems, CA, USA).

The animals were submitted to left thoracotomy between the fourth and fifth ribs, and after 20 min of baseline, myocardial IR injury was induced by the occlusion of the left anterior descending coronary artery for 30 min, followed by reperfusion for 60 min. The sham group underwent the same surgical procedure without ischemia. To distinguish necrotic from viable myocardium, the left ventricles were submitted to a gross histological double-staining technique using the Evan's blue/triphenyl tetrazolium chloride (TTC) method as previously described [13]. The myocardial infarct area was assessed in each heart by a blinded observer using planimetry (Image J, NIH Image, USA), and the infarct area was expressed as a percentage of the area at risk (% AAR).

## Analysis of cardiac capillary density

To measure the left ventricular structural capillary density, the samples were dehydrated in a graded series of ethanol (70%, 95%, and 100%) and cut using the "orthrip" method [22]. This method generates isotropic, uniform, and random sections of biological specimens for the quantitative assessment of three-dimensional anisotropic structures (such as the left ventricle) from two-dimensional sections. Therefore, the organ was cut three times consecutively. The first section was random, the second section was orthogonal to the first, and the third section was orthogonal to the second [23]. After the samples were embedded in paraffin blocks, they were cut into 5- $\mu$ m-thick sections and stained with FITC-conjugated *G. simplicifolia lectin* (1:150 dilutions) in a dark chamber at room temperature for 30 min. For each rat, at least five randomly selected microscopic fields were examined by microscopy (Olympus BX51/WI; Olympus, PA, USA) and analyzed with Archimed 3.7.0 software (Microvision, Lisses, France). The left ventricular structural capillary density was calculated by dividing the capillary volume density by the fiber volume density ( $V_{V[\text{cap}]} / V_{V[\text{fib}]}$ ).

## Determination of antioxidant enzyme activities

Samples of frozen left ventricles were homogenized with an Ultra-Turrax (Janke & Kunkel IKA, Labortechnik, Germany) in nine volumes of 10 mM Tris-HCl pH 7.4 containing 0.9% NaCl, 1 mM PMSF and 0.5 g/ml aprotinin. The homogenate was centrifuged at  $720 \times g$  at 4°C for 10 min, and the supernatant was utilized for enzymatic measurements [24], [25]. Protein content was determined by the Bradford method using bovine serum albumin as the standard [26]. All enzyme activity assays were performed at 37°C with at least four biological replicates for the Exe and CT groups. Readings were obtained using a GBC 920 UV-Vis spectrophotometer (GBC Scientific Equipment, Australia), and the results were expressed as units (mol/min) per mg of protein. The total superoxide dismutase (SOD) activity was determined according to a method described elsewhere [27]. Briefly, the total SOD activity was assessed by the rate of cytochrome c reduction at 550 nm before and after adding the heart homogenate (15 µg protein) in a reaction medium containing 50 mM phosphate buffer (pH 8.0), 0.1 mM EDTA, 0.01 mM potassium cyanide, 0.02 mM cytochrome c, 0.05 mM xanthine, and 8 mU xanthine oxidase. A 50% decrease in the rate of cytochrome c reduction was considered as one international unit (IU) of total SOD activity and was expressed as IUs per mg of protein. Catalase activity was measured by the rate of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) decomposition to water and oxygen, according to a previous method reported in the literature [28]. The difference between the rate of absorbance decrease at 240 nm in the presence or absence of heart homogenates (30 µg protein) was measured in a reaction medium composed of 50 mM phosphate buffer (pH 7.0), 0.002% Triton X-100, 0.1 mM EDTA, and 15 mM H<sub>2</sub>O<sub>2</sub> in a final volume of 1 ml. The amount of H<sub>2</sub>O<sub>2</sub> was calculated using the molar extinction coefficient (43.6 M/cm), and catalase activity was expressed in units (µmol of H<sub>2</sub>O<sub>2</sub> consumed per min) by mg protein.

## Quantitative RT-PCR

Total RNA was extracted from 30 mg of at least four samples of frozen left ventricles obtained from the CT and Exe groups using the RNeasy Fibrous Tissue Mini kit (Qiagen, Hilden, Germany), following the manufacturer's instructions. The RNA yield and purity were determined using a NanoDrop spectrometer (Thermo Fisher Scientific, MA, USA). Next, 1 µg of total RNA was reverse-transcribed into cDNA using the High Capacity cDNA Reverse Transcriptase kit (Applied Biosystems, CA, USA) according to the manufacturer's instructions. The amplification reactions were performed in 96-well plates (Applied Biosystems, CA, USA) at a final volume of 25 µl and contained 1 µl of  $10 \times$  diluted cDNA, 12.5 µl of  $2 \times$  Power SYBR Green PCR Master Mix (Applied Biosystems, CA, USA) and 150 nM of each forward and reverse primer (Thermo Fisher Scientific, MA, USA) (Table 1). The levels of mRNA were measured using the ViiA 7 Real Time PCR System (Thermo Fisher Scientific, MA, USA). The amplification program consisted of an initial cycle at 50°C for 2 min, followed by 95°C for 10 min, and 40 cycles of 95°C for 15 s and 60°C for 1 min. Each cDNA was

**Table 1.** Primer sequences.

Target	Primer Sequence	
EcSOD	Forward	GGCCCAGCTCCAGACTGA
	Reverse	CTCAGGTCCCCGAATCATG
CuZnSOD	Forward	CGGCTTCTGTCGTCTCCT
	Reverse	GTTACCGCTTGCCTTCT
MnSOD	Forward	TTAACGCGCAGATCATGCA
	Reverse	CCTCGGTGACGTTTCAGATTGT
Catalase	Forward	ACTCAGGTGCGGACATTC
	Reverse	GGAGTTGACTGGTCCAGAAGAGCC
HSP72	Forward	GCTCATCAAGCGCAACTCCAC
	Reverse	TCGTACACCTGGATCAGCACCC
β-Actin	Forward	CCACCCGCGAGTACAACCTTCTT
	Reverse	GAAGCCGGCCTTGACATGCC

EcSOD, endothelial cell superoxide dismutase (SOD); CuZnSOD, copper zinc SOD; MnSOD, manganese SOD; HSP72, heat shock protein 72.

doi:10.1371/journal.pone.0113541.t001

amplified in duplicate, and a corresponding sample without reverse transcriptase (no-RT sample) was included as a negative control. The specificity of the RT-PCR products was confirmed by melting curve analysis and by omission of the reverse transcriptase. All of the samples were normalized to β-actin mRNA using the  $\Delta\Delta C_t$  method.

### Western blotting

Samples of frozen left ventricles from the Exe and CT groups were minced on ice and homogenized with a RW20 tissue processor (Digital IKA, Staufen, Germany) for 2 min at 1,745 rpm in homogenization buffer containing 250 mM sucrose, 1 mM imidazole, pH 7.6 (adjusted with Tris), 1 mM EDTA and 0.05 g protease inhibitor. The homogenates were centrifuged at 3,500 rpm for 15 min (SS34 rotor, SORVALL RC-5B centrifuge, Thermo Fisher Scientific, MA, USA), and the supernatant was collected, centrifuged again at 32,900 rpm for 60 min (70 Ti rotor, Beckman Coulter centrifuge, Beckman Coulter, CA, USA) (modified from Dostanic et al., 2004) [29], and kept on ice. Then, a 10-μl aliquot (in triplicate) was used for the protein concentration assays, which were performed using the Folin phenol method described by Lowry et al. [30] with 5% SDS added to the samples and bovine serum albumin used as the standard. Sixty micrograms of protein were separated by 10% polyacrylamide gel electrophoresis and subsequently electroblotted onto nitrocellulose membranes (Amersham GE Healthcare, Buckinghamshire, United Kingdom). The resulting membranes were then stained with Ponceau S and analyzed to verify equal loading and transfer. The membranes were blocked overnight with 5% skim milk at 4°C in phosphate-buffered saline solution containing Tris-buffered saline (TBS). The blots were then incubated in blocking buffer with rabbit polyclonal IgG antibodies directed

against OR subtypes mu (1:1,000, Santa Cruz Biotechnology, SC-15310, CA, USA), kappa (1:1,000, Santa Cruz Biotechnology, SC-9112, CA, USA) and delta (1:1,500, Santa Cruz Biotechnology, SC- 9111, CA, USA) for 1 h at room temperature. After washing with TBS-T (TBS with 0.1% Tween 20), the blots were incubated at room temperature for 1 h with the appropriate secondary polyclonal antibody (anti-rabbit 1:5,000, Amersham GE Healthcare, Buckinghamshire, United Kingdom) coupled to peroxidase and washed again with TBS-T. The membranes were then treated with chemiluminescent reagents (luminol and enhancer; Amersham GE Healthcare, Buckinghamshire, United Kingdom) and exposed to light-sensitive film. Images of these films were analyzed using Scion software (Scion Co, MD, USA). The results were expressed in arbitrary units (a.u.).

### Reagent sources

All drugs and reagents were purchased from Sigma Chemical (MO, USA), with the exception of naloxone, naltrindole,  $\beta$ -funaltrexamine, and nor-binalthorphimine (Tocris Bioscience, MN, USA).

### Statistical analysis

Statistical comparisons between groups were performed using a one-way analysis of variance. Hemodynamic parameters were evaluated using a two-way analysis of variance. The Bonferroni post-test was used to localize the significant differences. P values  $<0.05$  were considered statistically significant. The results are expressed as the mean  $\pm$  SD.

## Results

### Hemodynamic parameters

The baseline hemodynamic parameters were not significantly different between experimental groups. Not surprisingly, ischemia produced a significant decrease in systolic, mean and diastolic arterial pressure 5 min after coronary ligation in all groups compared with the sham group ([Table 2](#)).

### Infarct size (IS)

As expected, the sham group experienced no myocardial infarction in response to the sham surgery. Importantly, the Exe group showed a significantly reduced IS compared with the CT group ( $27.6 \pm 3.5$  vs.  $42.0 \pm 3.0\%$ ;  $P < 0.05$ ). This cardioprotection was abolished when ORs were blocked with naloxone, maintaining an infarct area similar to that of the CT group (Exe+Nal:  $39.5 \pm 2.9\%$ ;  $P < 0.05$  - [Figure 2A](#)).

To determine which OR mediated this cardioprotective effect, specific receptor subtype antagonists were used. MOR and KOR blockade did not interfere in the

**Table 2.** Hemodynamic parameters in response to IR injury.

		Baseline	5 min after ischemia	30 min after ischemia	60 min after reperfusion
CT	SAP	126 ± 12	84 ± 17	96 ± 16	103 ± 25
	MAP	102 ± 11	67 ± 18	83 ± 18	83 ± 24
	DAP	82 ± 12	54 ± 16	68 ± 18	66 ± 21
	HR	428 ± 59	451 ± 24	456 ± 36	416 ± 73
Exe	SAP	122 ± 14	84 ± 19	124 ± 29	119 ± 11
	MAP	105 ± 12	69 ± 18	105 ± 29	97 ± 11
	DAP	89 ± 12	58 ± 16	89 ± 27	77 ± 10
	HR	449 ± 34	454 ± 27	473 ± 26	462 ± 31
Exe + Nal	SAP	124 ± 19	90 ± 29	109 ± 27	124 ± 23
	MAP	106 ± 16	76 ± 27	90 ± 27	101 ± 21
	DAP	88 ± 13	65 ± 26	74 ± 26	83 ± 18
	HR	452 ± 32	465 ± 43	471 ± 34	463 ± 37
Sham	SAP	118 ± 7	119 ± 9*	121 ± 13	128 ± 7
	MAP	101 ± 7	101 ± 9*	100 ± 13	103 ± 9
	DAP	87 ± 7	85 ± 9*	81 ± 12	81 ± 10
	HR	452 ± 39	455 ± 36	445 ± 33	441 ± 45

The values represent the means ± SD, n=10 for all groups. CT, control group; Exe, exercised group; Exe+Nal, exercised + naloxone group; SAP, systolic arterial pressure (mmHg); MAP, mean arterial pressure (mmHg); DAP, diastolic arterial pressure (mmHg); HR, heart rate (bpm). \*P<0.05 vs. all groups.

doi:10.1371/journal.pone.0113541.t002

exercise-induced cardioprotection, these groups presented IS similar to Exe group (Exe+MOR:  $28.1 \pm 2.7\%$ ; Exe+KOR:  $28.2 \pm 4.8\%$  - [Figure 2B](#)). Nevertheless, DOR blockade completely prevented the cardioprotection seen in the Exe group, the IS was similar to that observed in the CT group (Exe+DOR:  $39.6 \pm 2.0\%$  - [Figure 2B](#)).

### Left ventricle structural capillary density

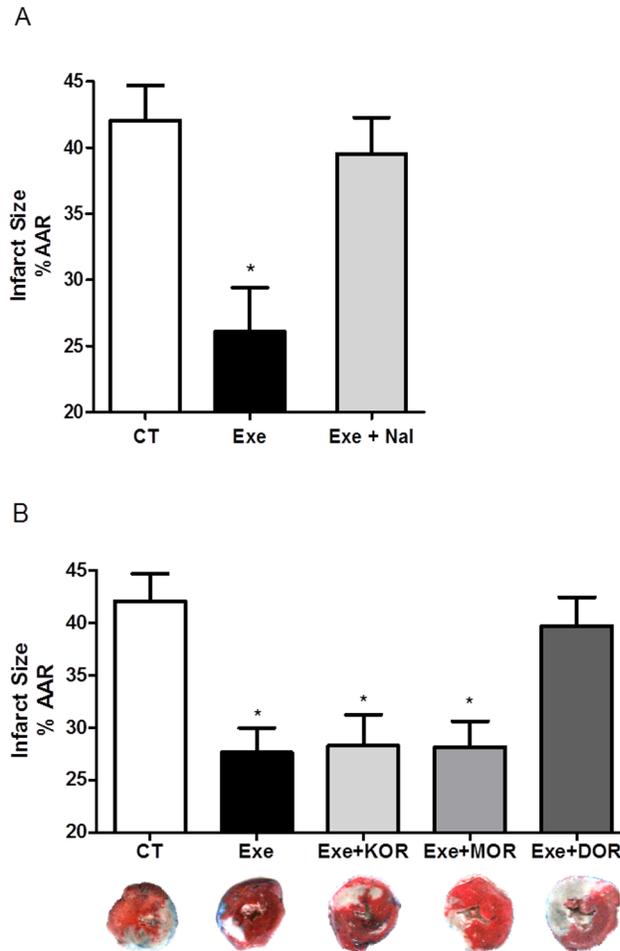
The left ventricle structural capillary density was similar in both the Exe ( $0.22 \pm 0.01 V_{v[cap]}/V_{v[fib]}$ ) and CT groups ( $0.21 \pm 0.01 V_{v[cap]}/V_{v[fib]}$ ) - [Figure 3](#).

### Antioxidant enzyme activities

Exercise caused a significant increase in the total SOD (CT:  $52.2 \pm 2.2$  vs. Exe:  $57.8 \pm 1.2$  U/mg; P<0.05) and catalase activities (CT:  $0.49 \pm 0.00$  vs. exe:  $0.62 \pm 0.05$  U/mg; P<0.05 - [Figure 4](#)).

### HSP72 and antioxidant enzyme mRNA expression

In the mRNA expression measurements of antioxidant enzymes, the endothelial cell SOD (EcSOD), copper zinc SOD (CuZnSOD), manganese SOD (MnSOD) and catalase expression levels revealed no differences between the Exe and CT



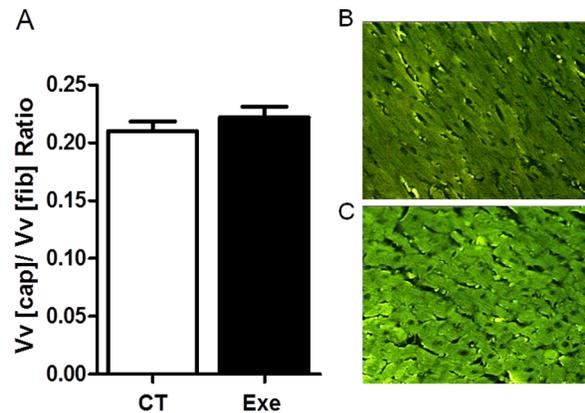
**Figure 2. IS after ischemia-reperfusion injury following non-selective (A) and selective (B) opioid receptor antagonists.** CT (control group, n=10); Exe (exercised group, n=10); Exe+Nal (non-specific antagonist group, n=10); Exe+KOR (kappa opioid receptor antagonist group, n=5); Exe+MOR (mu opioid receptor antagonist group, n=5); and Exe+DOR (delta opioid receptor antagonist group, n=5). \* P <0.05 vs. CT.

doi:10.1371/journal.pone.0113541.g002

groups. For each enzyme, the fold change in expression relative to the control was  $1.07 \pm 0.86$ ,  $0.94 \pm 0.18$ ,  $0.93 \pm 0.19$  and  $1.13 \pm 0.38$ , respectively (Figure 5). Whereas exercise induced no adaptations in any antioxidant enzyme expression, HSP72 mRNA expression in the Exe group was significantly lower compared with the CT group ( $0.53 \pm 0.18$ -fold control;  $P < 0.05$  - Figure 5).

### OR protein expression

The Exe group presented significantly reduced protein expression levels for KOR compared with the CT group ( $0.82 \pm 0.09$  vs.  $0.46 \pm 0.08$  a.u.;  $P < 0.05$ ), whereas DOR expression was not different between the groups ( $0.61 \pm 0.12$  vs.  $0.38 \pm 0.08$  a.u. - Figure 6). The immunoblot bands for DOR and KOR can be



**Figure 3. Left ventricle structural capillary density.** The bars represent capillary density in the control and exercised groups (A). The representative images used to determine the structural capillary density with capillaries stained with FITC-conjugated *G. simplicifolia* lectin. Magnification,  $\times 200$ ; scale bar, 100  $\mu\text{m}$  in the CT (B) and Exe groups (C). Exe, exercised group (n=5); CT, control group (n=5).

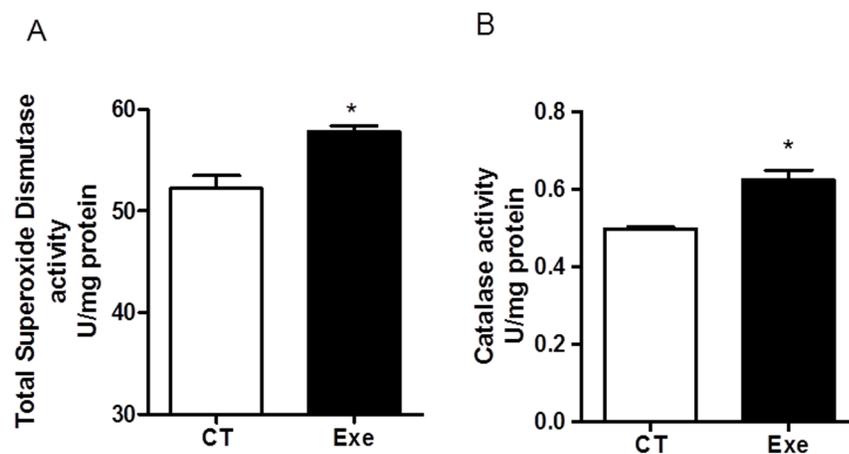
doi:10.1371/journal.pone.0113541.g003

found in Figures S1 and S2, respectively. MOR was not detected by Western blotting in the left ventricle in the studied animals.

## Discussion

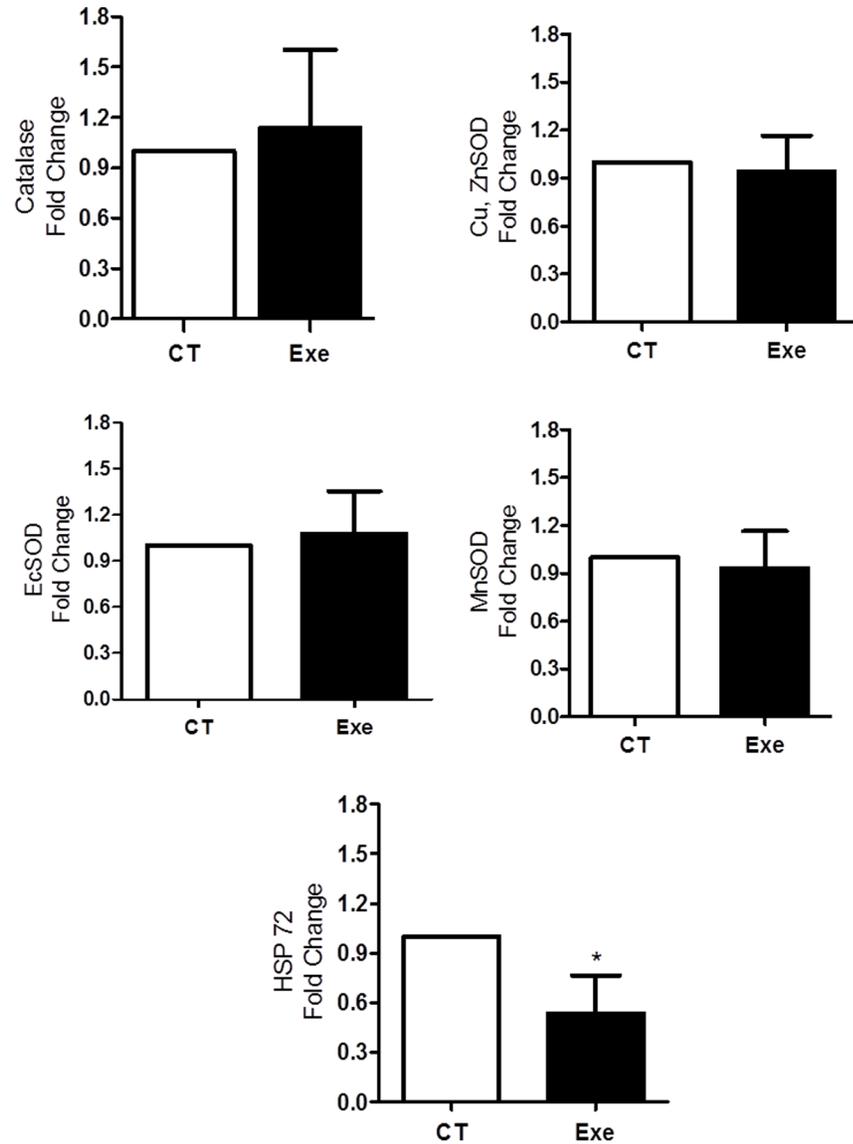
### Overview of major findings

These experiments provide new and important information regarding the mechanism responsible for exercise-induced cardioprotection. Specifically, our experiments reveal for the first time that the cardioprotective effect of endurance exercise is mediated, at least in part, through activation of the DOR subtype. Our



**Figure 4. Total superoxide dismutase (A) and catalase (B) activities in the control and exercised groups.** Exe, exercised group (n=5); CT, control group (n=5). \*  $P < 0.05$  vs. CT.

doi:10.1371/journal.pone.0113541.g004



**Figure 5. mRNA expression of key antioxidant enzymes and HSP72.** Copper zinc superoxide dismutase (SOD) (CuZnSOD), endothelial cell SOD (EcSOD), manganese SOD (MnSOD) and heat shock protein 72 (HSP72). The values represent control fold changes, normalized by  $\beta$ -actin. Exe (exercised group, n=3); and CT (control group, n=5). \*  $P < 0.05$  vs. CT.

doi:10.1371/journal.pone.0113541.g005

exercise training protocol also increased SOD and catalase activity in the heart but did not increase the left ventricle capillary density, mRNA levels of key antioxidant enzymes or mRNA levels of HSP72. A brief discussion of each of these findings follows.

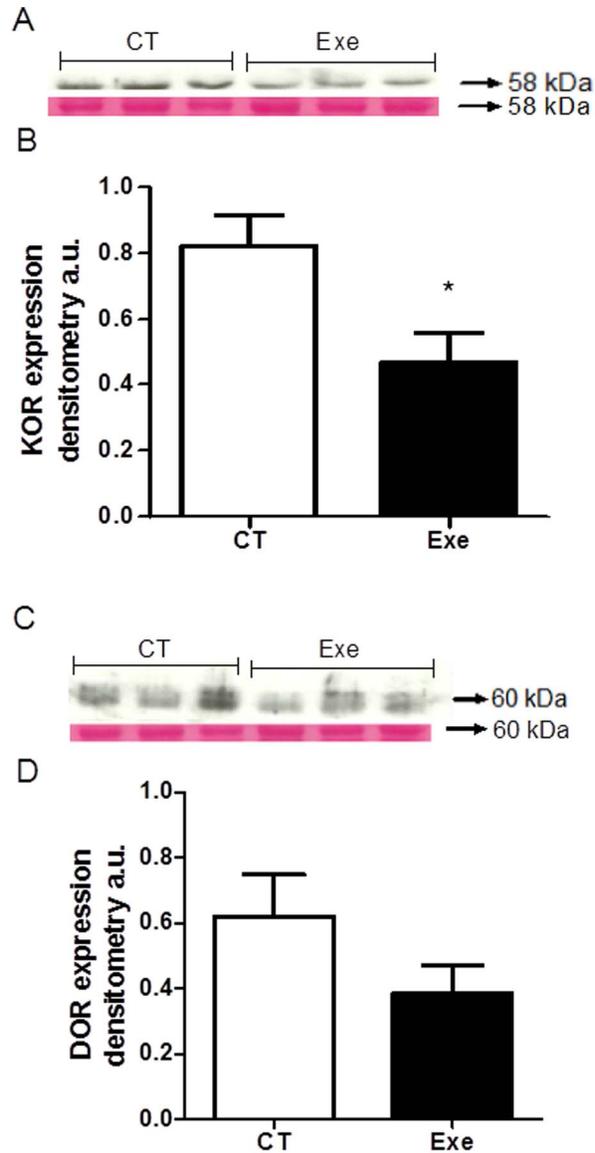
## Increased myocardial capillary density and HSP72 expression are not involved in exercise-induced cardioprotection

Our results indicated that the left ventricle capillary density did not increase following four independent exercise training sessions. This finding was expected because an exercise-induced remodeling of the vascular network in the heart would likely require several weeks of endurance training [7], [31]. Given that as few as 3–5 consecutive days of exercise have consistently been reported to produce significant cardioprotection, it is clear that the exercise-induced cardioprotection observed in the current experiments was mediated by mechanisms other than increased angiogenesis in the heart [13], [31], [32].

Over-expression of HSP72 in the heart has been suggested to promote cardioprotection against an IR insult [33]. Furthermore, several weeks of endurance exercise training can significantly increase the protein abundance of HSP72 in the heart of rodents [34]. Nonetheless, reports have clearly demonstrated that the prevention of exercise-induced HSP72 expression in the heart does not prevent exercise-induced cardioprotection against IR injury [32], [35]. Therefore, it appears unlikely that an exercise-induced increase in the myocardial levels of HSP72 was responsible for the exercise-induced cardioprotection observed in the current experiments.

## Increased antioxidant enzyme activity may participate in exercise-induced cardioprotection

Similar to Dickson et al. [13], our results indicate that a limited number of exercise sessions did not alter the cardiac gene expression of SOD isoforms and catalase. However, Calvert et al. [11] reported that exercise increased the cardiac expression of CuZnSOD. The apparent discrepancy between our results and those from Calvert et al. seems to be related to differences in the animal model used, the training methodology, and the approach used to evaluate antioxidant activity. In the Calvert et al. study, the protective effects of exercise against myocardial IR injury were investigated by employing voluntary exercise training in mice for 4 weeks, and antioxidant protein expression was analyzed using a Western blotting approach. In our study, the rats were trained at a previously established exercise intensity of  $\sim 70\%$  of  $VO_{2max}$  for 4 sessions (less than 1 week), and the cardiac gene expression of CuZnSOD was analyzed via quantitative RT-PCR. Our results revealed that aerobic exercise training increased the MnSOD and catalase activities in the heart. Given that increased myocardial ROS production plays an important role in promoting IR-induced myocardial cell death, it follows that exercise-induced improvements in cardiac antioxidant capacity could be a potential mediator of the cardioprotection observed in the current experiments [36]. Similarly, other independent studies have reported increased MnSOD and catalase activities in the heart following exercise training [11], [36–38]. Nonetheless, although exercise promotes an enhanced myocardial antioxidant capacity, it remains controversial whether increased MnSOD levels in the heart are required for exercise-induced cardioprotection. For example, using an antisense



**Figure 6. OR subtype expression after IR injury.** Immunoblots of the control (CT; n=4) and exercised (Exe; n=5) group bands for the kappa opioid receptor (KOR) (A) and delta opioid receptor (DOR) (C). The bars represent the KOR (B) and DOR (D) expression levels normalized by *Ponceau* densitometry. a.u., arbitrary units. \* P<0.05 vs. CT.

doi:10.1371/journal.pone.0113541.g006

oligonucleotide against MnSOD to prevent exercise-induced increases in myocardial MnSOD activity, Yamashita et al. [39] demonstrated that an increase in myocardial MnSOD activity is required to provide training-induced protection against IR-induced myocardial infarction. In contrast, Lennon et al. [40], using the same MnSOD gene silencing approach, reported that prevention of the exercise-induced increase in myocardial MnSOD did not result in a loss of training-induced protection against IR-mediated arrhythmias. Another study

concluded that exercise-induced increases in MnSOD contributed to exercise-induced cardioprotection, but other factors must also contribute to this protection [41]. Therefore, based on our results and the work of others [11], [39], [40], improved myocardial antioxidants may contribute to exercise-induced cardioprotection.

### Opioid receptors are involved in exercise-mediated cardioprotection

The current experiments provide evidence that endogenous opioids contribute to the cardioprotective effects of exercise. These findings agree with previous studies that also reported an opioid system-mediated cardioprotective effect of exercise [13], [14], [21]. In this regard, Galvao et al. [21] showed that the effect of chronic exercise training on decreasing IS seems to occur, at least in part, through OR stimulation and not by increasing myocardial perfusion. Concerning the acute effect of exercise-induced cardioprotection, Dickson et al. [13] demonstrated that the administration of a non-selective OR antagonist before exercise abrogated exercise-induced cardioprotection. Our results showed that when trained animals were treated with OR antagonists prior to ischemia, exercise-induced cardioprotection was abolished. It is feasible that the activation of ORs over the course of multiple exercise sessions could initiate a signaling cascade that contributes to the cardioprotective effects of exercise. Taken together, these results underscore the importance of the opioid system, not only as an initial trigger of acute myocardial adaptation to exercise but also as an intermediate pathway contributing to cardioprotection.

### The delta OR is the specific subtype involved in exercise-mediated cardioprotection

To determine which opioid receptor subtype was involved in exercise-induced cardioprotection, we treated exercise-trained animals with specific antagonists prior to exposure to an IR insult. The current study is the first investigation to demonstrate the importance of DOR in exercise-induced cardioprotection. Based on the IPC model of cardioprotection results, we hypothesized that the protective effects of DOR may be linked to the activation of protein kinase C (PKC), which, in turn, opens sarcolemma/mitochondrial  $K^+$  ATP channels [20], leading to shortening of the cardiac action potential duration by accelerating phase III repolarization [42]. This would inhibit  $Ca^{2+}$  entry into the cell via L-type channels and prevent  $Ca^{2+}$  overload and the mitochondrial permeability transition pore (MPTP) from opening [7]. The MPTP typically opens during reperfusion, resulting in mitochondrial swelling and protein release, ultimately resulting in necrotic cell death [7]. Nevertheless, further investigations to confirm this sequence of events and to unravel the precise intracellular pathway of DOR-mediated cardioprotection following exercise are required.

We also examined whether exercise training resulted in increased expression of specific ORs in the heart. Our results revealed that exercise did not increase the protein abundance of DOR or KOR in the heart. In fact, exercise training resulted in a significant decrease in the myocardial levels of KOR (and apparently DOR as well). Although the current investigation is the first study to report the impact of exercise training on the protein abundance of specific ORs in the heart, Dickson et al. [13] reported an increase in both KOR and DOR mRNA expression in the heart immediately following exercise. It should be noted however that this increase in cardiac mRNA expression returned to baseline within 8 h following exercise and apparently did not result in increased protein synthesis of these ORs.

The fact that exercise resulted in a decrease in the protein levels of both KOR and DOR in the heart is noteworthy. Specifically, exercise reduced the cardiac protein expression of KOR by ~44%, whereas DOR expression was diminished by ~37%. A previous study by Aitchison et al. [43] demonstrated that activation of KOR signaling promotes an anti-cardioprotective state, resulting in an increased IR-induced IS in the rat heart compared with controls. Therefore, the exercise-induced reduction in cardiac levels of KOR may serve as a cardioprotective adaptation. Indeed, the opposing outcomes of DOR and KOR stimulation could establish a balance in the opioid signaling system. Therefore, exercise-induced cardioprotection could be produced not only through DOR stimulation but also through KOR downregulation. Interestingly, KOR and DOR have been consistently implicated in IPC- and remote IPC-induced cardioprotection [15], [20], [44–52], reinforcing a potential common cardioprotective link between exercise, ORs and IPC. Exercise may induce cardioprotection via DOR and KOR in a signal transduction pathway similar to IPC-induced cardioprotection. However, further research describing the signaling pathways of DOR and KOR and their interactions is required to provide additional insight regarding the OR mechanism(s) of exercise-induced cardioprotection.

Finally, MOR expression in the heart was also investigated in the present study. Nonetheless, using a Western blot approach, there were no detectable levels of MOR protein in the heart. This finding agrees with previous studies that also did not detect MOR expression in the rat heart [43], [53], [54]. Together, these findings suggest that MOR do not play a role in the modulation of IS.

### Study limitations

The results of the present study should be interpreted while considering certain limitations, particularly aspects related to methodology and protocol. First, the best method for measuring myocardial IS, as well as the use of TTC as the single approach to quantitatively estimate myocardial injury, is still extensively debated in the literature. TTC is an inexpensive, rapid, and reliable method for myocardial IS measurement and has been used as the sole quantitative method in several investigations involving IS assessment [13], [39], [55], [56]. Moreover, TTC has been extensively utilized as the gold standard technique to validate new imaging methods of infarct area quantification, such as cardiac magnetic resonance

imaging [57–61], contrast echocardiography [62–66], and two-dimensional speckle tracking imaging [67]. Nevertheless, the use of biochemical techniques, such as a dosage of troponin-I, may also be reliable for estimating myocardial IS and could confirm and emphasize our results concerning the effects of OR antagonists in exercise-induced cardioprotection. Second, the use of a single fixed dose of antagonists is always a problem in pharmacological testing. In our study, based on the large dose of OR antagonists used, the low density of ORs in cardiac tissue, previous results from Schultz et al. [17], [44] and preliminary data from our lab, we assumed that 3 mg/kg of OR antagonists would be the appropriate dose to demonstrate the involvement of the opioid system in exercise-induced cardioprotection. Although the ideal protocol would involve establishing a dose-response curve for all antagonists, ethical concerns related to the large number of animals used in the study and the increase in operational costs limited the feasibility of more experiments. Third, the effect of OR antagonists on antioxidant enzyme activity is a very interesting issue that was not addressed in the present study. Indeed, increased myocardial antioxidant capacity is considered an important mechanism involved in exercise-induced cardioprotection, and OR antagonists may theoretically interfere with enzyme expression. However, no evidence exists that any of the OR antagonists used in the current experiments are capable of inhibiting antioxidant enzyme activity. Further, it appears unlikely that any of these OR blockers are capable of crossing the mitochondrial inner membrane to inhibit MnSOD, the only antioxidant enzyme that has consistently been shown to be important in exercise-induced cardioprotection [68–70]. Nevertheless, further studies are warranted to better determine the effects of OR antagonists on antioxidant enzyme activities.

## Summary and Conclusions

Although it is well established that regular bouts of endurance exercise result in cardioprotection against IR injury, the mechanisms responsible for this cardioprotection remain under debate. Although an increase in myocardial antioxidants (e.g., MnSOD) may contribute to exercise-induced cardioprotection, the current experiments demonstrate that endogenous opioids also contribute to exercise-induced cardioprotection. Importantly, our results reveal for the first time that the DOR subtype plays an essential role in exercise-induced cardioprotection. Indeed, pharmacological blockade of DOR resulted in a significant loss of exercise-induced cardioprotection against IR-induced injury. However, additional experiments are required to determine if a significant interaction exists between exercise-induced changes in DOR signaling and increased myocardial antioxidants in the heart.

## Supporting Information

**Figure S1.**

[doi:10.1371/journal.pone.0113541.S001](https://doi.org/10.1371/journal.pone.0113541.S001) (TIF)

**Figure S2.**

[doi:10.1371/journal.pone.0113541.S002](https://doi.org/10.1371/journal.pone.0113541.S002) (TIF)

**Author Contributions**

Conceived and designed the experiments: JPB KSV AD MAL. Performed the experiments: JPB KSV AD VHO. Analyzed the data: JPB KSV AD SKP VHO RSF ET MAL. Contributed reagents/materials/analysis tools: JPB KSV AD SKP VHO RSF ET MAL. Wrote the paper: JPB KSV AD SKP VHO RSF ET MAL.

**References**

1. Powers SK, Lennon SL, Quindry J, Mehta JL (2002) Exercise and cardioprotection. *Curr Opin Cardiol* 17: 495–502.
2. Powers SK, Quindry JC, Kavazis AN (2008) Exercise-induced cardioprotection against myocardial ischemia-reperfusion injury. *Free Radic Biol Med* 44: 193–201.
3. Berlin JA, Colditz GA (1990) A meta-analysis of physical activity in the prevention of coronary heart disease. *Am J Epidemiol* 132: 612–628.
4. Paffenbarger RS Jr, Hyde RT, Wing AL, Hsieh CC (1986) Physical activity, all-cause mortality, and longevity of college alumni. *N Engl J Med* 314: 605–613.
5. Sattelmair J, Pertman J, Ding EL, Kohl HW 3rd, Haskell W, et al. (2011) Dose response between physical activity and risk of coronary heart disease: a meta-analysis. *Circulation* 124: 789–795.
6. Calvert JW, Lefer DJ (2013) Role of beta-adrenergic receptors and nitric oxide signaling in exercise-mediated cardioprotection. *Physiology (Bethesda)* 28: 216–224.
7. Golbidi S, Laher I (2011) Molecular mechanisms in exercise-induced cardioprotection. *Cardiol Res Pract* 2011: 972807.
8. Quindry JC, Hamilton KL (2013) Exercise and cardiac preconditioning against ischemia reperfusion injury. *Curr Cardiol Rev* 9: 220–229.
9. Powers SK, Sollanek KJ, Wiggs MP, Demirel HA, Smuder AJ (2014) Exercise-induced improvements in myocardial antioxidant capacity: the antioxidant players and cardioprotection. *Free Radic Res* 48: 43–51.
10. Frasier CR, Sloan RC, Bostian PA, Gonzon MD, Kurowicki J, et al. (2011) Short-term exercise preserves myocardial glutathione and decreases arrhythmias after thiol oxidation and ischemia in isolated rat hearts. *J Appl Physiol* (1985) 111: 1751–1759.
11. Calvert JW, Condit ME, Aragon JP, Nicholson CK, Moody BF, et al. (2011) Exercise protects against myocardial ischemia-reperfusion injury via stimulation of beta(3)-adrenergic receptors and increased nitric oxide signaling: role of nitrite and nitrosothiols. *Circ Res* 108: 1448–1458.
12. Lee Y, Min K, Talbert EE, Kavazis AN, Smuder AJ, et al. (2012) Exercise protects cardiac mitochondria against ischemia-reperfusion injury. *Med Sci Sports Exerc* 44: 397–405.
13. Dickson EW, Hogrefe CP, Ludwig PS, Ackermann LW, Stoll LL, et al. (2008) Exercise enhances myocardial ischemic tolerance via an opioid receptor-dependent mechanism. *Am J Physiol Heart Circ Physiol* 294: H402–408.
14. Michelsen MM, Stottrup NB, Schmidt MR, Lofgren B, Jensen RV, et al. (2012) Exercise-induced cardioprotection is mediated by a bloodborne, transferable factor. *Basic Res Cardiol* 107: 260.
15. Wang GY, Wu S, Pei JM, Yu XC, Wong TM (2001) Kappa- but not delta-opioid receptors mediate effects of ischemic preconditioning on both infarct and arrhythmia in rats. *Am J Physiol Heart Circ Physiol* 280: H384–391.
16. Schultz JE, Rose E, Yao Z, Gross GJ (1995) Evidence for involvement of opioid receptors in ischemic preconditioning in rat hearts. *Am J Physiol* 268: H2157–2161.

17. **Schultz JJ, Hsu AK, Gross GJ** (1997) Ischemic preconditioning is mediated by a peripheral opioid receptor mechanism in the intact rat heart. *J Mol Cell Cardiol* 29: 1355–1362.
18. **Tsang A, Hausenloy DJ, Yellon DM** (2005) Myocardial postconditioning: reperfusion injury revisited. *Am J Physiol Heart Circ Physiol* 289: H2–7.
19. **Kato R, Foex P** (2002) Myocardial protection by anesthetic agents against ischemia-reperfusion injury: an update for anesthesiologists. *Can J Anaesth* 49: 777–791.
20. **Schultz JE, Gross GJ** (2001) Opioids and cardioprotection. *Pharmacol Ther* 89: 123–137.
21. **Galvao TF, Matos KC, Brum PC, Negrao CE, Luz PL, et al.** (2011) Cardioprotection conferred by exercise training is blunted by blockade of the opioid system. *Clinics (Sao Paulo)* 66: 151–157.
22. **Mattfeldt T, Mall G, Gharehbaghi H, Moller P** (1990) Estimation of surface area and length with the orientator. *J Microsc* 159: 301–317.
23. **Estato V, Obadia N, Carvalho-Tavares J, Freitas FS, Reis P, et al.** (2013) Blockade of the renin-angiotensin system improves cerebral microcirculatory perfusion in diabetic hypertensive rats. *Microvasc Res* 87: 41–49.
24. **Chaves EA, Fortunato RS, Carvalho DP, Nascimento JH, Oliveira MF** (2013) Exercise-induced cardioprotection is impaired by anabolic steroid treatment through a redox-dependent mechanism. *J Steroid Biochem Mol Biol* 138: 267–272.
25. **Chaves EA, Pereira-Junior PP, Fortunato RS, Masuda MO, de Carvalho AC, et al.** (2006) Nandrolone decanoate impairs exercise-induced cardioprotection: role of antioxidant enzymes. *J Steroid Biochem Mol Biol* 99: 223–230.
26. **Bradford MM** (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248–254.
27. **Crapo JD, McCord JM, Fridovich I** (1978) Preparation and assay of superoxide dismutases. *Methods Enzymol* 53: 382–393.
28. **Aebi H** (1984) Catalase in vitro. *Methods Enzymol* 105: 121–126.
29. **Dostanic I, Schultz Jel J, Lorenz JN, Lingrel JB** (2004) The alpha 1 isoform of Na,K-ATPase regulates cardiac contractility and functionally interacts and co-localizes with the Na/Ca exchanger in heart. *J Biol Chem* 279: 54053–54061.
30. **Lowry OH, Rosebrough NJ, Farr AL, Randall RJ** (1951) Protein measurement with the Folin phenol reagent. *J Biol Chem* 193: 265–275.
31. **White FC, Bloor CM, McKirnan MD, Carroll SM** (1998) Exercise training in swine promotes growth of arteriolar bed and capillary angiogenesis in heart. *J Appl Physiol* (1985) 85: 1160–1168.
32. **Hamilton KL, Powers SK, Sugiura T, Kim S, Lennon S, et al.** (2001) Short-term exercise training can improve myocardial tolerance to I/R without elevation in heat shock proteins. *Am J Physiol Heart Circ Physiol* 281: H1346–1352.
33. **Hutter MM, Sievers RE, Barbosa V, Wolfe CL** (1994) Heat-shock protein induction in rat hearts. A direct correlation between the amount of heat-shock protein induced and the degree of myocardial protection. *Circulation* 89: 355–360.
34. **Quindry JC, Hamilton KL, French JP, Lee Y, Murlasits Z, et al.** (2007) Exercise-induced HSP-72 elevation and cardioprotection against infarct and apoptosis. *J Appl Physiol* (1985) 103: 1056–1062.
35. **Taylor RP, Harris MB, Starnes JW** (1999) Acute exercise can improve cardioprotection without increasing heat shock protein content. *Am J Physiol* 276: H1098–1102.
36. **Powers SK, Demirel HA, Vincent HK, Coombes JS, Naito H, et al.** (1998) Exercise training improves myocardial tolerance to in vivo ischemia-reperfusion in the rat. *Am J Physiol* 275: R1468–1477.
37. **Husain K** (2003) Interaction of physical training and chronic nitroglycerin treatment on blood pressure, nitric oxide, and oxidants/antioxidants in the rat heart. *Pharmacol Res* 48: 253–261.
38. **Ji LL, Gomez-Cabrera MC, Vina J** (2006) Exercise and hormesis: activation of cellular antioxidant signaling pathway. *Ann N Y Acad Sci* 1067: 425–435.
39. **Yamashita N, Hoshida S, Otsu K, Asahi M, Kuzuya T, et al.** (1999) Exercise provides direct biphasic cardioprotection via manganese superoxide dismutase activation. *J Exp Med* 189: 1699–1706.

40. **Lennon SL, Quindry JC, Hamilton KL, French JP, Hughes J, et al.** (2004) Elevated MnSOD is not required for exercise-induced cardioprotection against myocardial stunning. *Am J Physiol Heart Circ Physiol* 287: H975–980.
41. **French JP, Hamilton KL, Quindry JC, Lee Y, Upchurch PA, et al.** (2008) Exercise-induced protection against myocardial apoptosis and necrosis: MnSOD, calcium-handling proteins, and calpain. *Faseb J* 22: 2862–2871.
42. **Noma A** (1983) ATP-regulated K<sup>+</sup> channels in cardiac muscle. *Nature* 305: 147–148.
43. **Aitchison KA, Baxter GF, Awan MM, Smith RM, Yellon DM, et al.** (2000) Opposing effects on infarction of delta and kappa opioid receptor activation in the isolated rat heart: implications for ischemic preconditioning. *Basic Res Cardiol* 95: 1–10; discussion 11.
44. **Schultz JJ, Hsu AK, Gross GJ** (1997) Ischemic preconditioning and morphine-induced cardioprotection involve the delta (delta)-opioid receptor in the intact rat heart. *J Mol Cell Cardiol* 29: 2187–2195.
45. **Tsuchida A, Miura T, Tanno M, Nozawa Y, Kita H, et al.** (1998) Time window for the contribution of the delta-opioid receptor to cardioprotection by ischemic preconditioning in the rat heart. *Cardiovasc Drugs Ther* 12: 365–373.
46. **Wu S, Li HY, Wong TM** (1999) Cardioprotection of preconditioning by metabolic inhibition in the rat ventricular myocyte. Involvement of kappa-opioid receptor. *Circ Res* 84: 1388–1395.
47. **Valtchanova-Matchouganska A, Ojewole JA** (2003) Mechanisms of opioid delta (delta) and kappa (kappa) receptors' cardioprotection in ischaemic preconditioning in a rat model of myocardial infarction. *Cardiovasc J S Afr* 14: 73–80.
48. **Fryer RM, Auchampach JA, Gross GJ** (2002) Therapeutic receptor targets of ischemic preconditioning. *Cardiovasc Res* 55: 520–525.
49. **Surendra H, Diaz RJ, Harvey K, Tropak M, Callahan J, et al.** (2013) Interaction of delta and kappa opioid receptors with adenosine A1 receptors mediates cardioprotection by remote ischemic preconditioning. *J Mol Cell Cardiol* 60: 142–150.
50. **Zhang SZ, Wang NF, Xu J, Gao Q, Lin GH, et al.** (2006) Kappa-opioid receptors mediate cardioprotection by remote preconditioning. *Anesthesiology* 105: 550–556.
51. **Peart JN, Gross ER, Gross GJ** (2005) Opioid-induced preconditioning: recent advances and future perspectives. *Vascul Pharmacol* 42: 211–218.
52. **Zhang Y, Irwin MG, Wong TM, Chen M, Cao CM** (2005) Remifentanyl preconditioning confers cardioprotection via cardiac kappa- and delta-opioid receptors. *Anesthesiology* 102: 371–378.
53. **Pugsley MK** (2002) The diverse molecular mechanisms responsible for the actions of opioids on the cardiovascular system. *Pharmacol Ther* 93: 51–75.
54. **Wittert G, Hope P, Pyle D** (1996) Tissue distribution of opioid receptor gene expression in the rat. *Biochem Biophys Res Commun* 218: 877–881.
55. **Hamilton KL, Staib JL, Phillips T, Hess A, Lennon SL, et al.** (2003) Exercise, antioxidants, and HSP72: protection against myocardial ischemia/reperfusion. *Free Radic Biol Med* 34: 800–809.
56. **Lessa MA, Tibirica E** (2006) Pharmacologic evidence for the involvement of central and peripheral opioid receptors in the cardioprotective effects of fentanyl. *Anesth Analg* 103: 815–821.
57. **Verani MS, Jeroudi MO, Mahmarian JJ, Boyce TM, Borges-Neto S, et al.** (1988) Quantification of myocardial infarction during coronary occlusion and myocardial salvage after reperfusion using cardiac imaging with technetium-99m hexakis 2-methoxyisobutyl isonitrile. *J Am Coll Cardiol* 12: 1573–1581.
58. **Wu KC, Kim RJ, Bluemke DA, Rochitte CE, Zerhouni EA, et al.** (1998) Quantification and time course of microvascular obstruction by contrast-enhanced echocardiography and magnetic resonance imaging following acute myocardial infarction and reperfusion. *J Am Coll Cardiol* 32: 1756–1764.
59. **Grieve SM, Mazhar J, Callaghan F, Kok CY, Tandy S, et al.** (2014) Automated quantification of myocardial salvage in a rat model of ischemia-reperfusion injury using 3D high-resolution magnetic resonance imaging (MRI). *J Am Heart Assoc* 3.
60. **Jin J, Teng G, Feng Y, Wu Y, Jin Q, et al.** (2007) Magnetic resonance imaging of acute reperfused myocardial infarction: intraindividual comparison of ECIII-60 and Gd-DTPA in a swine model. *Cardiovasc Intervent Radiol* 30: 248–256.

61. **Yang Y, Graham JJ, Connelly K, Foltz WD, Dick AJ, et al.** (2012) MRI manifestations of persistent microvascular obstruction and acute left ventricular remodeling in an experimental reperfused myocardial infarction. *Quant Imaging Med Surg* 2: 12–20.
62. **Chen X, Cui K, Xiu J, Lin H, Lao Y, et al.** (2009) Evaluation and simplified measurement of infarct size by myocardial contrast echocardiography in a rat model of myocardial infarction. *Int J Cardiovasc Imaging* 25: 713–716.
63. **Filusch A, Buss S, Hardt S, Katus HA, Kuecherer HF, et al.** (2008) Evaluation cardioprotective effects of atorvastatin in rats by real time myocardial contrast echocardiography. *Echocardiography* 25: 974–981.
64. **Han B, Wei M** (2008) Proximal coronary hemodynamic changes evaluated by intracardiac echocardiography during myocardial ischemia and reperfusion in a canine model. *Echocardiography* 25: 312–320.
65. **dos Santos L, Mello AF, Antonio EL, Tucci PJ** (2008) Determination of myocardial infarction size in rats by echocardiography and tetrazolium staining: correlation, agreements, and simplifications. *Braz J Med Biol Res* 41: 199–201.
66. **Park SW, Lee SY, Park SJ, Lee SC, Gwon HC, et al.** (2004) Quantitative assessment of infarct size in vivo by myocardial contrast echocardiography in a murine acute myocardial infarction model. *Int J Cardiol* 97: 393–398.
67. **Fu Q, Xie M, Wang J, Wang X, Lv Q, et al.** (2009) Assessment of regional left ventricular myocardial function in rats after acute occlusion of left anterior descending artery by two-dimensional speckle tracking imaging. *J Huazhong Univ Sci Technolog Med Sci* 29: 786–790.
68. **Quindry JC, Schreiber L, Hosick P, Wrieden J, Irwin JM, et al.** (2010) Mitochondrial KATP channel inhibition blunts arrhythmia protection in ischemic exercised hearts. *Am J Physiol Heart Circ Physiol* 299: H175–183.
69. **Hamilton KL, Quindry JC, French JP, Staib J, Hughes J, et al.** (2004) MnSOD antisense treatment and exercise-induced protection against arrhythmias. *Free Radic Biol Med* 37: 1360–1368.
70. **Chen Z, Siu B, Ho YS, Vincent R, Chua CC, et al.** (1998) Overexpression of MnSOD protects against myocardial ischemia/reperfusion injury in transgenic mice. *J Mol Cell Cardiol* 30: 2281–2289.