

Book Chapter

Cannabinoids and Terpenes as Chemotaxonomic markers in Cannabis

Sytze Elzinga¹, Justin Fishedick², Richard Podkolinski¹ and Jeffrey C Raber^{1*}

¹The Werc Shop, LLC., USA

²Excelsior Analytical Lab, Inc., USA

***Corresponding Author:** Jeffrey C Raber, The Werc Shop, LLC., Pasadena, CA 91107, USA

Published **January 21, 2020**

This Book Chapter is a republication of an article published by Jeffrey C Raber, et al. at Natural Products Chemistry & Research in July 2015. (Elzinga S, Fishedick J, Podkolinski R, Raber JC (2015) Cannabinoids and Terpenes as Chemotaxonomic Markers in Cannabis. Nat Prod Chem Res 3: 181.doi:10.4172/2329-6836.1000181)

How to cite this book chapter: Sytze Elzinga, Justin Fishedick, Richard Podkolinski, Jeffrey C Raber. Cannabinoids and Terpenes as Chemotaxonomic markers in Cannabis. In: Prime Archives in Chemistry. Hyderabad, India: Vide Leaf. 2020.

© The Author(s) 2020. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License(<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

In this paper, we present principal component analysis (PCA) results from a dataset containing 494 cannabis flower samples and 170 concentrate samples analyzed for 31 compounds. A continuum of chemical composition amongst cannabis strains was found instead of distinct chemotypes. Our data shows that some strains are much more reproducible in chemical composition than others. Strains labeled as indica were compared with those labeled as sativa and no evidence was found that these two cultivars are distinctly different chemotypes. PCA of “OG” and “Kush” type strains found that “OG” strains have relatively higher levels of α -terpineol, fenchol, limonene, camphene, terpinolene and linalool where “Kush” samples are characterized mainly by the compounds *trans*-ocimene, guaiol, β -eudesmol, myrcene and α -pinene. The composition of concentrates and flowers were compared as well. Although the absolute concentration of compounds in concentrates is much higher, the relative composition of compounds between flowers and concentrates is similar.

Keywords

Cannabis; Tetrahydrocannabinol; Cannabidiol; Marijuana; Terpenoids; Terpenes; Strains; Taxonomy

Introduction

Cannabis has been used for medicinal and recreational purposes for millennia. From a taxonomic perspective scientists have been debating the presence of multiple species for quite some time. Some scientist delineate 3 species, *C. sativa* (marijuana type), *C. indica* (hemp type) and *C. ruderalis* (wild type), where others see this as just a variety of 1 species and associated subspecies thereunder [1-4]. This distinction is exceptionally important as most US States new laws consider *Cannabis sativa* L. and subspecies as the only legal cultivars allowed.

It is commonly accepted in cannabis culture to make the distinction between sativa and indica strains [5]. **Indica plants** are said to be short, densely branched and have wider leaves and are claimed to be sedative and good for pain relief. **Sativa plants** are tall, loosely branched and have long, narrow leaves. Sativa is claimed to be uplifting, stimulating and recommended for daytime use. This is the typical information a patient or recreational user will hear when they visit a medicinal cannabis dispensary or recreational store. However, we only found one published study that compared indica and sativa strains in patients [6]. This study showed that cannabis was uniformly effective in relieving symptoms across a wide range of diagnostic categories. Indica strains appeared superior to sativa strains in improving energy and appetite. No statistical difference between sativa and indica was found for pain, mood, nausea, muscle spasms, seizures, ocular, insomnia, awareness or neuropathy. Strains were assigned based upon morphology. This study was not blinded and the observed differences could be a result of expectations by the patient.

If sativa and indica truly have different physiological effects upon consumption, some compound or interaction of compounds need to be responsible for this. It has been shown that cannabidiol (CBD) can influence the psychoactive effects of Δ^9 -tetrahydrocannabinol (THC) [7] and it has been postulated that the combination of phytocannabinoids and terpenes could result in complementary or synergistic results often referred to as the “entourage effect” [8].

In this paper we use PCA to investigate the analytical results for cannabinoids and terpenes in 494 cannabis flower samples and 170 cannabis concentrates. This analysis is performed in an attempt to investigate the potential existence of distinct cannabis chemotypes that could explain the different effects people experience from specific cannabis strains. Cannabinoids and terpenes were chosen as chemotype markers as they are considered to be the main physiologically active constituents in cannabis.

Researchers have looked at cannabis from a chemotaxonomic perspective as well. Small and Beckstead split *C. sativa* L. into three chemotypes based upon CBD/THC ratio [4]. Type 1 has a CBD/THC ratio of <0.5, type II has an intermediate CBD/THC ratio of 0.5-3.0 and type 3 has a ratio of >3.0. DeMeijer et al showed that this could be explained genetically by a model involving one locus, with two alleles. One allele codes for CBDA synthase where the other codes for THCA synthase. The alleles were shown to be co-dominant [9]. Pacifico et al later showed that classification using just the CBD/THC values will mask the existence of chemotypes with relatively high amounts of other cannabinoids [10].

Various authors have tried using the secondary metabolites in combination with PCA for forensic investigation of the geographical origin of the plant material [11-13]. The most elaborate study was performed by El Sohly et al who analyzed 157 samples from six geographical regions and classified them using statistical analysis of 175 GC/MS peaks. Although they managed to differentiate samples from different countries the success of this approach was limited as not only geographical location but many other cultivation variables influenced the chemical composition of the flowers. Much of the cannabis available in the western world is grown indoors often with strict control of variables. The use of controlled lighting cycles, specialized soil, fine-tuned nutrients and pest control eliminate many of the environmental variables and will make geographical assigning of the plant difficult if not impossible.

Fischedick et al analyzed 11 cultivars of cannabis for 36 compounds and managed to discriminate the various cultivars with PCA [14]. Higher levels of cannabinoids correlated positively to higher levels in terpenoids ($R^2=0.7688$). The authors of this paper showed that it is possible to grow cannabis with reproducible terpene and cannabinoid levels over different batches as long as environmental conditions and genetics are standardized. Alterations in grow cycle time, plant stress and different genotype can cause considerable differences in the chemical profile.

A study by Casano et al investigated the variability of terpene profiles in 16 plants from different strains of *C. sativa* L. [15]. They separated the samples into ‘mostly indica’ and ‘mostly sativa’ based upon the morphological appearance declared by cultivators of the strain. The study showed a large variation of relative content of terpenes between strains and suggest that terpene variation can be used as a tool for characterization of cannabis bio types. In this study ‘mostly indica’ strains were characterized by dominance of β -myrcene with limonene or α -pinene as the second most abundant terpenes. The ‘mostly sativa’ strains were characterized by more complex terpene profiles, with some strains having α -terpinolene or α -pinene as dominant, and some strains having β -myrcene as dominant with α -terpinolene or trans- β -ocimene as second most abundant.

To our knowledge this is the first paper reporting chemotypical differences using samples that are available to patients in the California medicinal cannabis dispensaries. Most of the previous papers use samples collected worldwide and based upon their reported cannabinoids levels, are not representative of the cannabis currently available in the United States to patients and recreational users. It is important to note that doctors often specifically recommend an indica or sativa strain to their patient, but that the scientific literature is lacking evidence to support these recommendations. In this paper we will take a PCA approach to investigate the variation between strains in the California medicinal marijuana market and also specifically look at the differences in composition between indica and sativa strains.

Methods

Chemicals

Methanol and water of analytical grade as well as terpene reference standards were purchased from Sigma Aldrich, St. Louis, MO, USA. Analytical standards for the cannabinoids were acquired from Restek, Bellefonte, PA, USA.

Origin of Samples

The samples used for this study have been submitted for analysis to our laboratory by California medicinal marijuana patients in the period from the beginning of 2012 to the end of 2013. The strain names for the samples were the names reported by the submitter at the time of submission.

Statistical Analyses

Strains were classified as indica, sativa, hybrid or unknown based upon the assignment by the cannabis strain database website Leafly.com on June 11, 2015. For concentrates, the whole data set was used and divided into three categories (high, medium, low CBD).

All data was modulated to express the various compounds as the contribution to the sum of compounds.

PCA analysis was performed in excel using a macro written by Tsugawa et al. and is available for free [16] (http://prime.psc.riken.jp/Metabolomics_Software/StatisticalAnalysisOnMicrosoftExcel/).

When scaling was performed the option “auto scale” was selected.

Analytical Measurements

Quantification of THC, CBD, cannabigerol (CBG), Δ^9 -tetrahydrocannabinolic acid (THCA), cannabidiolic acid (CBDA) and cannabigerolic acid (CBGA) was performed using a Shimadzu prominence UFLC system (Shimadzu Scientific Instruments, Columbia, MD, USA). The acidic analytical method as published by Hazekamp et al. was used [17]. Calculation of THCmax, CBDmax and CBGmax were performed as described in our previous paper [18]. Terpene content was determined using GC-FID according to the same approach as Fishedick et al using retention time comparison

with authentic reference, mass spectra, and literature data [14]. A 5% diphenyl and 95% dimethyl polysiloxane column (SHRX5, 15 meter, 0.25 mm ID, 0.25 μ m film thickness, Shimadzu Scientific Instruments, Columbia, MD, USA) and helium carrier gas (Airgas, Radnor, PA, USA) was used for separation. Table 1 list the terpenes that were analyzed in each sample.

Table 1: The terpenes analyzed.

Analyzed Terpenes			
α -Bisabolol	α -Cedrene	Limonene	α -Phelladerene
Borneol	β -Eudesmol	Linalool	α -Pinene
Camphene	(*) Fenchol	Menthol	β -Pinene
Camphor	Geraniol	Myrcene	Sabinene
Δ^1 -Carene	Guaiol	Nerol	α -Terpinene
β -Caryophyllene	α -Humulene	cis-Ocimene	α -terpineol
Caryophyllene oxide	Isoborneol	trans-Ocimene	Terpinolene

Results

Description of Flower Data Set

The dataset contained 494 samples. At least 8 different samples were present for each uniquely identifiable strain. A total of 35 different strains were present in the dataset. Table 2 shows the amount of replicates for each strain and the average, minimum and maximum THCmax concentration found in the samples. It can be noticed that the THCmax levels can vary widely even within one strain. In 14 out of 35 strains the difference between the minimum and maximum level found differed by more than a factor of 2 and in the highest case (OG Kush) even more than a factor of 5. This indicates that it will be exceptionally hard to predict the potency of a flower product based solely upon the strain name.

Table 2: Summary of the cannabis flower data set.

Strain Name	Replicates	Indica/Sativa/Hybrid or Unknown	OG or Kush	THCmax(%)		
				Average	Min	Max
1st Generation Diablo	16	Indica	-	20.8	16.2	24.2
Afghan Kush	10	Indica	Kush	17.6	14.7	22.0
Alien OG	8	Hybrid	OG	19.7	14.6	23.8
Black Mamba	9	Indica	-	21.0	19.4	22.8
Blackberry Kush	11	Indica	Kush	15.9	12.5	18.0
Blue Dream	31	Hybrid	-	16.9	12.2	21.2
Blue Dream Haze	9	Hybrid	-	17.4	13.6	21.0
Bubba Kush	9	Indica	Kush	15.5	10.2	19.4
ChemDawg	14	Hybrid	-	16.9	11.2	23.1
Fire OG	23	Hybrid	OG	17.3	9.8	20.2
Girl Scout Cookies	19	Hybrid	-	15.7	5.8	20.9
Grand Daddy Purple	14	Indica	-	16.9	12.2	23.3
Green Crack	16	Sativa	-	15.4	11.0	19.1
Harlequin	15	Sativa	-	5.0	2.5	12.6
Headband	8	Hybrid	-	15.5	5.4	22.1
Jack Herer	24	Sativa	-	16.9	13.1	21.4
LA Confidential	17	Indica	-	15.1	8.9	21.7
Larry OG	8	Hybrid	OG	17.1	6.2	24.3
Neptune OG	10	Indica	OG	18.0	12.6	22.8
NY Sour Diesel	10	Unknown	-	15.9	10.8	20.2
OG Herojuana	11	Unknown	OG	18.6	15.3	20.7
OG Kush	28	Hybrid	-	16.4	4.9	25.0
Platinum OG	8	Indica	OG	18.3	15.6	21.4
Pre '98 Bubba	9	Indica	-	14.4	9.3	20.3
Purple Kush	8	Indica	Kush	13.9	3.8	18.8
SFV OG	12	Hybrid	OG	18.9	14.1	23.3
Skywalker	15	Indica	-	19.1	9.6	24.5
Skywalker OG	19	Hybrid	OG	18.5	13.8	21.7
Sour Diesel	32	Sativa	-	16.6	7.7	22.0
Sour OG	10	Hybrid	OG	18.0	14.1	24.1
Strawberry Cough	8	Sativa	-	15.3	8.7	18.1
Tahoe OG	14	Hybrid	OG	17.5	13.2	21.6
Train wreck	16	Hybrid	-	14.0	5.9	19.8
True OG	13	Indica	OG	18.5	13.4	22.2
Velvet Kush	10	Unknown	Kush	21.6	20.0	23.1

Table 2 also shows the assignment to sativa, indica, hybrid or unknown. Thirteen (13) strains were assigned as indica, 5 were assigned as sativa and 14 as hybrid. Three (3) strains did not occur in the Leafly database that was used for the assignment.

Figure 1 shows the distribution of the THCmax(%) in the total data set.

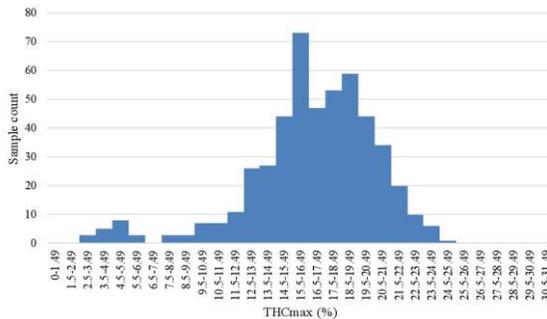


Figure 1: Distribution of THC max (%) in the Flower data set (N=494).

The average THCmax(%) in the data set was 16.8% and the median for THCmax was 17.1%. The distribution was not Gaussian but showed three peaks. The first peak is at 4.5-5.49%, which correlates with low THCmax content in high CBD strains. The second peak is at 15.5-16.49 and the third peak is at 18.5-19.49%. Information regarding cultivation condition was not present, but it is speculated that these two peaks represent the averages for outdoor and indoor cultivation methods.

Figure 2 shows the distribution of the CBDmax(%) in the total data set. The figure shows that 478 out of 494 samples had less than 1.49% CBD indicating that the medicinal marijuana market in California is dominated by high THC type strains. The average CBDmax(%) in the data set was 0.6% and the median was 0.3%.

The sum of terpenes was plotted against the sum of THCmax and CBDmax and a R^2 value of 0.4248 which is lower than the correlation of 0.77 found by Fishedick [14]. The graph can be found in supplementary information figure 1. This difference can be explained by the large variety of strains used in this study where the study by Fishedick was performed with a limited amount of strains grown and stored under standardized conditions.

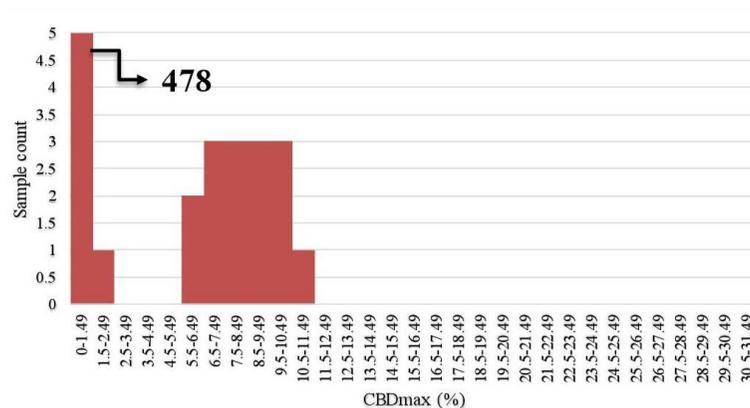


Figure 2: Distribution of CBD max (%) in the Flower data set (N=494).

For statistical analysis all data was expressed as a contribution to the sum of all compounds. This modulation of the data was performed as it is our experience the relative ratios of terpenes in a strain are more reproducible than the absolute concentration for a strain. Supplementary information figure 2 shows the effect of this conversion for the 8 main terpenes in 10 different Velvet Kush samples. Absolute data shows more variation than the standardized data. This effect is likely a result of trichome density [19]. Part of the plant exposed to more light will have a higher density of trichomes. Also during trimming of the dried female flowers more or less leafy material can be left behind influencing the absolute concentration as the trichomes are the cannabinoid and terpene producing parts of the plant.

It was noted that many flowers had a name containing “OG” or “Kush”. Kush is a reference to strains originating from the Hindu Kush region in Central Asia. The origin of the term “OG” is unknown. To investigate if this term has any relationship based on the chemical composition of the plant matter, PCA analysis was performed comparing OG with Kush type strains. Eleven (11) strains were assigned to the OG group and 5 strains were assigned to the Kush group.

PCA Analysis of Complete Flower Data Set

The full data set was analyzed without scaling and the scoring and loading plot can be found in figure 3 and 4. PC1 explained 86.3% of variance and PC2 explained 7.6%. Examination of the loading plot reveals cannabinoids are responsible for the differentiation of the samples. This is expected as the absolute concentration and variation of cannabinoids is much higher than that of the terpenes therefore without scaling the cannabinoids will dominate. A grouping of Harlequin (red) can be noticed. The loading plot indicates that these strains are differentiated due to a high CBDmax content. The original data showed that Harlequin is indeed a high CBD strain and fairly unique in this aspect. One OG Kush sample (purple) was mixed in with the Harlequin group and inspection revealed that this indeed was also a high CBD sample which was not characteristic of other OG Kush samples.

The same data set was analyzed with scaling. The scoring and loading plot can be found in figure 5 and 6. PC1 explained 22.1% of variance while PC2 explained 12.5%. Again a grouping of Harlequin (red) is observed and the loading plot reveals that relatively high quantities of CBD, α -pinene, guaiol and β -eudesmol are responsible for the separation in this plot. A second group is observed consisting of Trainwreck and Jack Herer. The loading plot reveals that this group is characterized by relatively high amounts of terpinolene, α -terpinene and α -phellandrene. It is worth noting that the monoterpene profile of these strains were similar to strains described as ‘sativa’ in other studies [14,15,20]. Plotting of additional principal components did not reveal any new groupings.

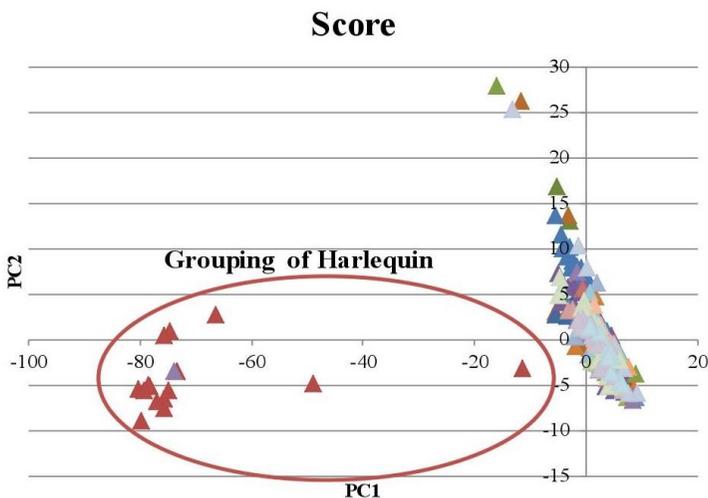


Figure 3: PCA scoring plot of full cannabis flower data set without scaling.

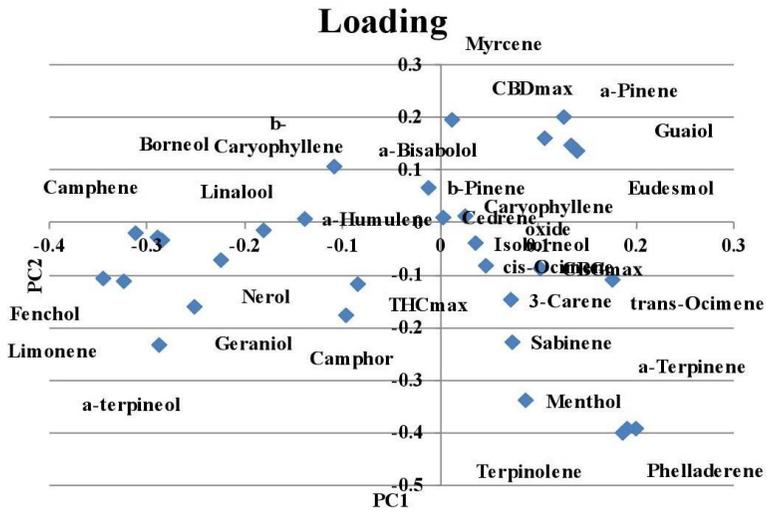


Figure 6: PCA loading plot of full cannabis flower data set with scaling.

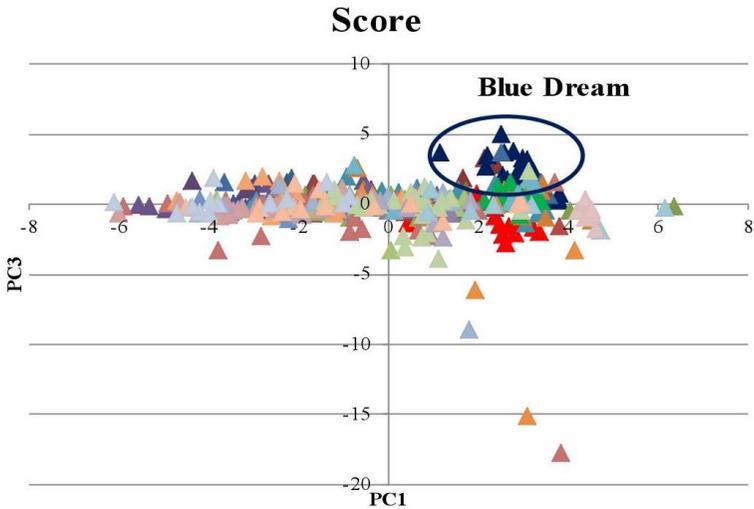


Figure 7: PCA (scaled) scoring plot after removing Jack Herer, Train wreck and Harlequin from the cannabis flower data set PC1 vs PC3.

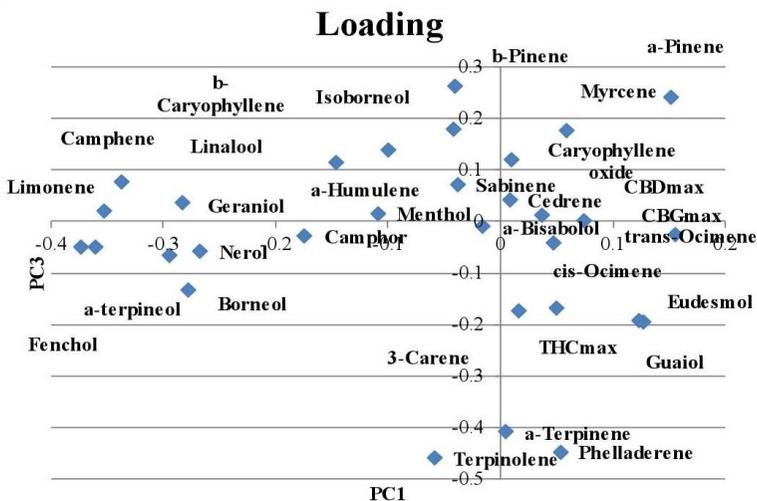


Figure 8: PCA (scaled) loading plot after removing Jack Herer, Train wreck and Harlequin from the cannabis flower data set PC1 vs PC3.

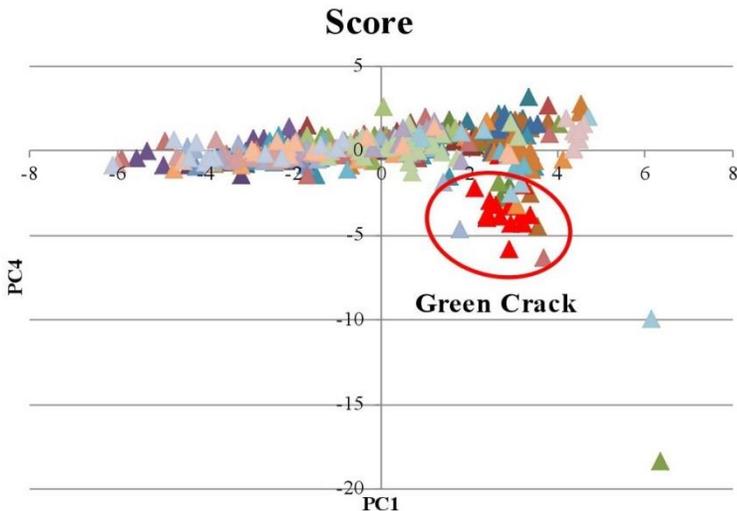


Figure 9: PCA (scaled) scoring plot after removing Jack Herer, Train wreck and Harlequin from the sample set PC1 vs PC4.

As Trainwreck, Jack Herer and Harlequin were very dominant in the separation, these strains were removed from the data set to see if additional groups could be detected. When scaled PCA

was performed, PC1 explained 20.5% of the variation, PC2 explained 10.9%, PC3 8.7% and PC4 7.9%. The PC1 vs PC2 plot did not show any specific grouping but a general scattered plot of all the samples. In the PC1 vs PC3 plot a small cluster of Blue Dream could be noticed (see figure 7). This group showed only slight separation and was caused by the relatively high concentration of α -pinene in these samples (figure 8). When PC1 vs PC4 is plotted a grouping of Green Crack can be noticed (figure 9). This group differentiates itself due to relatively high amounts of CBGmax, *cis*-ocimene and *trans*-ocimene (figure 10). No other groupings were noticed in any of the scoring plots.

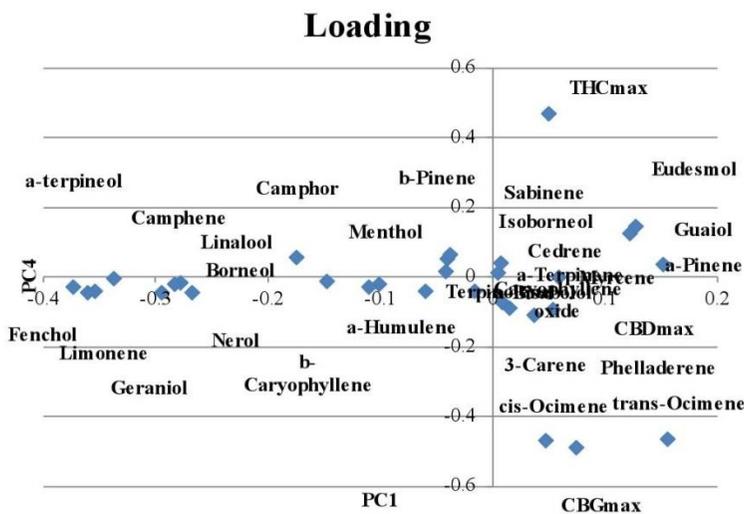


Figure 10: PCA (scaled) loading plot after removing Jack Herer, Train wreck and Harlequin from the sample set PC1 vs PC4.

Comparison of Indica vs Sativa

All unknown and hybrid samples were removed from the data set and the remaining samples were categorized as either indica (blue) or sativa (red). Scaled PCA was performed and the scoring and loading plot can be found in figure 11 and 12. PC1 explained 30.2% and PC2 explained 9.2% of the variation. In the scoring plot a mix of indica and sativa samples can be noticed, but approximately half of the indica samples separate from the

sativa samples. This group is dominant in the strains 1st Generation Diablo (1stGD), Black Mamba (BM), True OG (TrueOG), Neptune OG (NepOG) and Skywalker (SkyW). The loading plot reveals that this group has higher levels of limonene, fenchol, α -terpineol, camphene, linalool, THCmax, camphor, geraniol, β -pinene and β -caryophyllene.

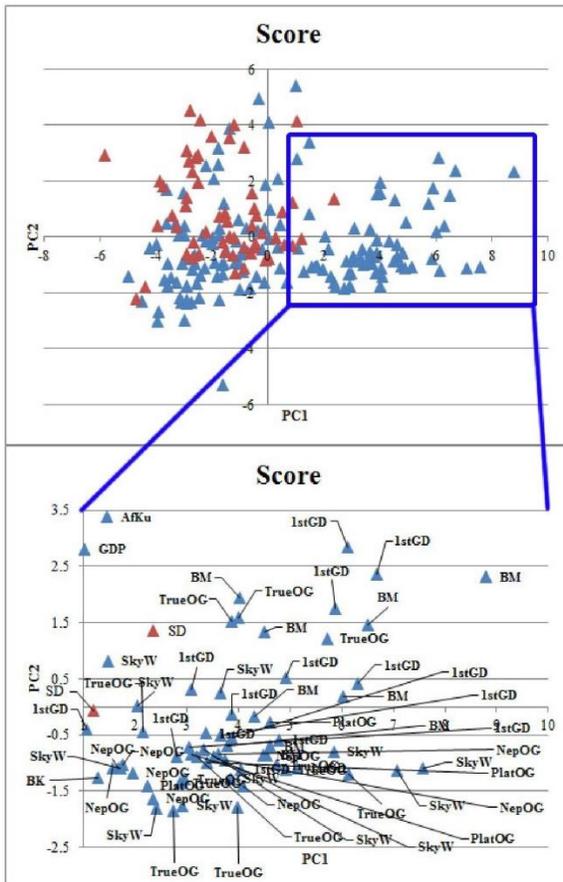


Figure 11: PCA (scaled) scoring plot for indica (blue) and sativa (red).

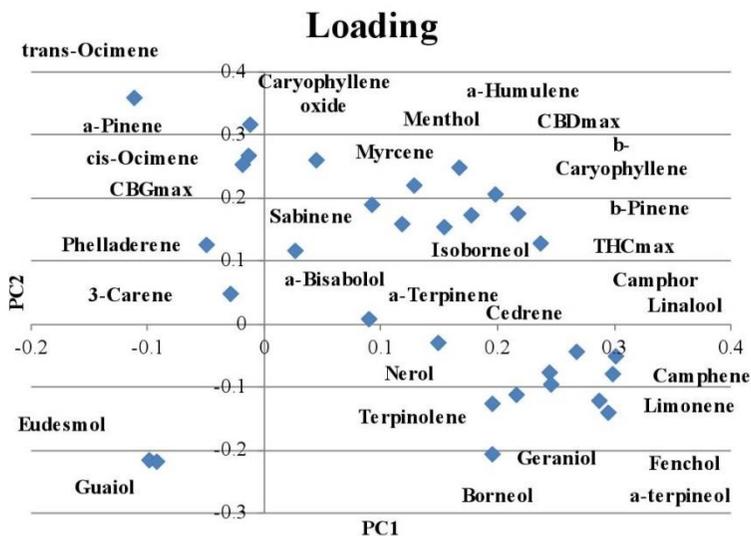


Figure 12: PCA (scaled) loading plot for indica and sativa.

OG vs Kush

A data set containing only OG type or Kush type samples was created from the original data (see table 1) and scaled PCA was performed. The scoring and loading plot can be found in figure 13 and 14. PC1 was 29.3%, PC2 was 11.5%. Separation of the two groups can be noticed although there is an overlapping section.

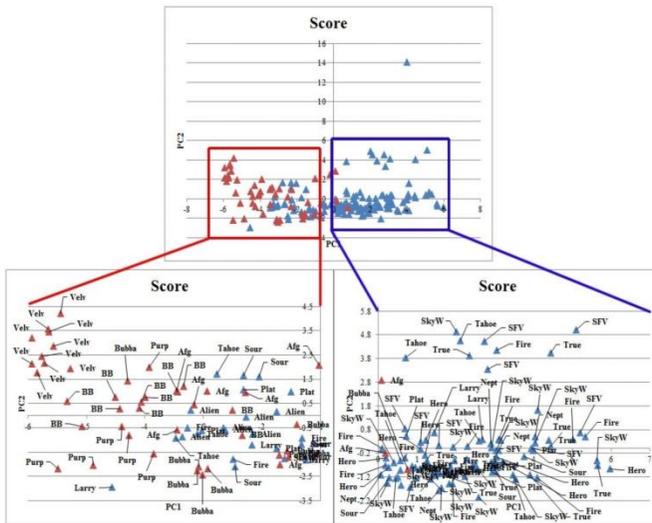


Figure 13: PCA (scaled) scoring plot for Kush (red) and OG (blue).

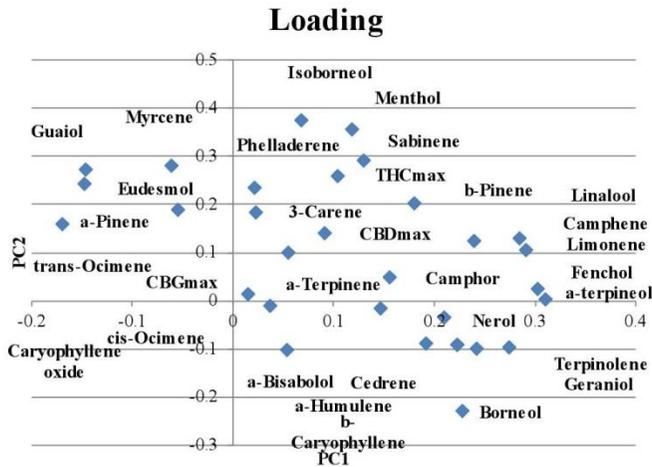


Figure 14: PCA (scaled) loading plot for Kush and OG.

The OG group is dominant in strains SFV OG (SFV), True OG (True), Tahoe OG (Tahoe), Fire OG (Fire), Neptune OG (Nept), Larry OG (Larry), Heroijuana OG (Hero), Platinum OG (Plat) and Skywalker OG (SkyW). This group is characterized mainly

by the compounds α -terpineol, fenchol, limonene, camphene, terpinolene and linalool.

The Kush group is dominant in strains Velvet Kush (Velv), Blackberry Kush (BB), Purple Kush (Purp). This group is characterized mainly by the compounds *trans*-Ocimene, Guaiol, β -Eudesmol, Myrcene and α -Pinene.

Description of Concentrate Data Set

The concentrate data set consisted of 170 samples. The samples were assigned to one of three groups. The high CBD group has a CBDmax/THCmax ratio of >5.0 , the medium CBD group had a ratio of $>0.95 < 5.0$ and the low CBD group had a ratio of <0.95 . No data was available regarding the production method of the various concentrates.

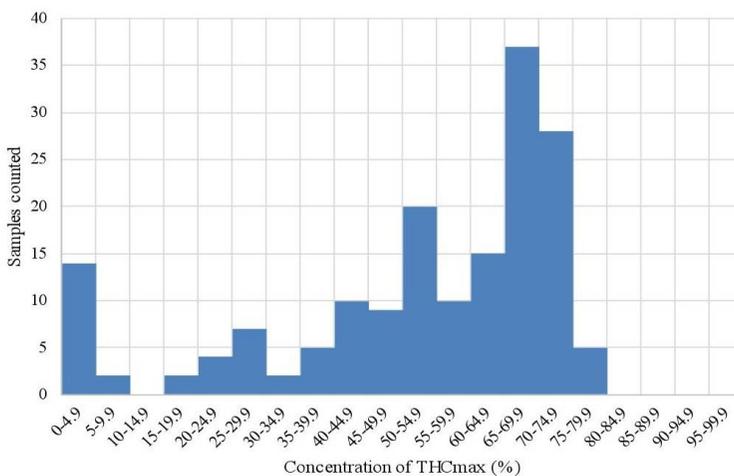


Figure 15: Distribution of THCmax(%) in the concentrate data set (N=170).

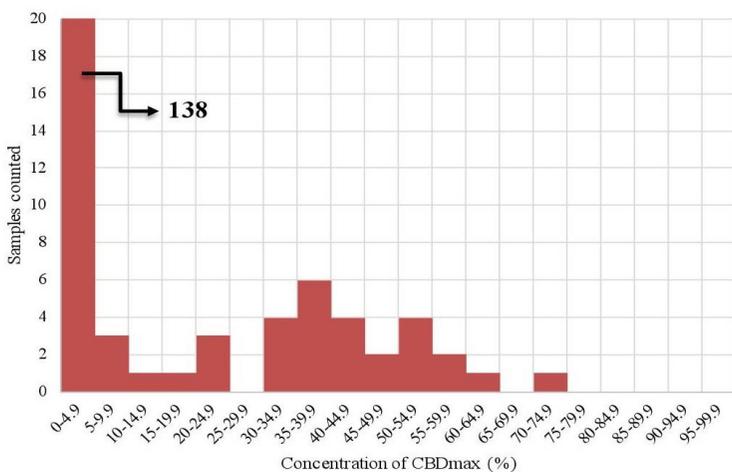


Figure 16: Distribution of CBDmax(%) in the concentrate data set (N=170).

The distribution of the THCmax and CBDmax in the concentrate samples can be found in figure 15 and 16. It was noticed that the frequency of certain THC levels occur more often. There appears to be a peak in frequency at 25-29.9%, 40-44.9, 50-54.9% and 65-69.9%. We speculate that this is caused by the difference in production methods of the concentrates. The concentrates peaking in frequency at 25-29.9% are expected to represent kief products, the peak at 40-44.9% is expected to represent hash products, the peak at 50-54.9% are expected to be mainly super critical CO₂ extracts and the peak at 65-69.9% are expected to be hydrocarbon extracts. As with the flowers, it can be noticed that the presence of high amounts of CBD is relatively rare. Out of 170 samples, 138 samples had less than 4.9% CBD.

PCA Analysis of the Concentrate Data Set

Figure 17 shows the PCA scoring plot for the data when scaling is not applied. PC1 explained 98.7% of the variation, PC2 explained 0.9%. Three distinct groups can be noticed. The loading plot (figure 18) shows that CBDmax and THCmax content are the main parameters that separate the groups. The presence of a low CBD, medium CBD and high CBD group

supports the model of 1 locus with 2 alleles by DeMeijer et al [9].

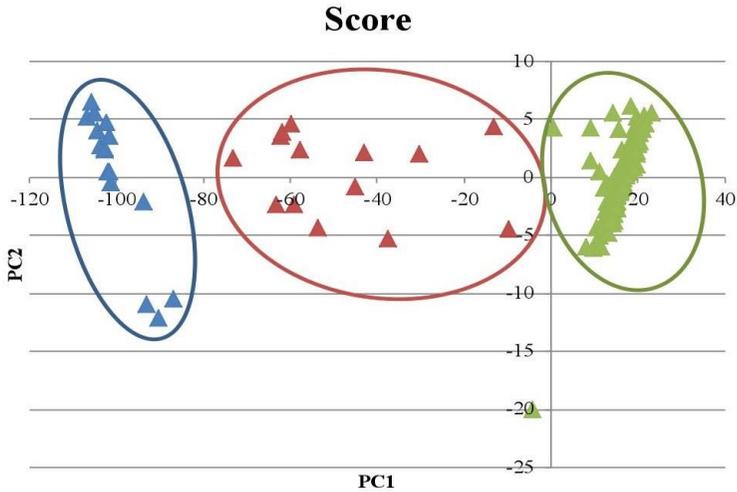


Figure 17: PCA (not scaled) scoring plot for concentrate data.

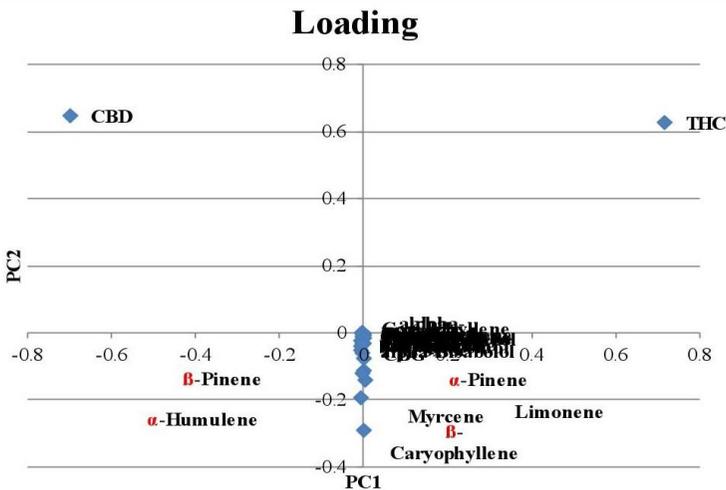


Figure 18: PCA (not scaled) loading plot for concentrate data.

Scaled PCA was performed as well and the scoring and loading plot can be found in supplementary information figure 3 and 4.

PC1 explains 21.4% of the variation and PC2 explains 16.2%. No distinct grouping could be observed in the scoring plots. This analysis was also repeated with just the terpene data, but this did not result in distinct groups. Three (3) samples separated from the majority of the samples in the right top field of the scoring plot. The loading plot reveals that samples in this region have higher levels of terpinolene, α -phellanderene, and α -terpinene. It is interesting to note that those are the same terpenoids that are typical for Jack Herer and Trainwreck strains. Most likely these concentrates originate from those strains.

PCA Analysis of Combined Flower and Concentrate Data

To see if the relative chemical composition changes during processing, PCA was plotted for all the concentrate and flower data. PC1 was 18.6%, PC2 was 13.2%. The scoring and loading plot can be found in figure 19 and 20. No clear grouping of flowers vs concentrates could be noticed but some concentrate samples separate from the rest of the samples. Inspection of the loading plot and the sample data revealed these samples have higher relative ratios of β -eudesmol, guaiol, α -bisabolol and isoborneol. These terpenes elute relatively late in GC analyses and it is speculated that these concentrates have been heated at some time during processing, evaporating part of the more volatile terpenoids and therefore changing the relative ratio in favor of the less volatile terpenes. However, the large majority of the flowers and concentrate samples are dispersed among each other in the scoring plot indicating a similar relative composition of compounds.

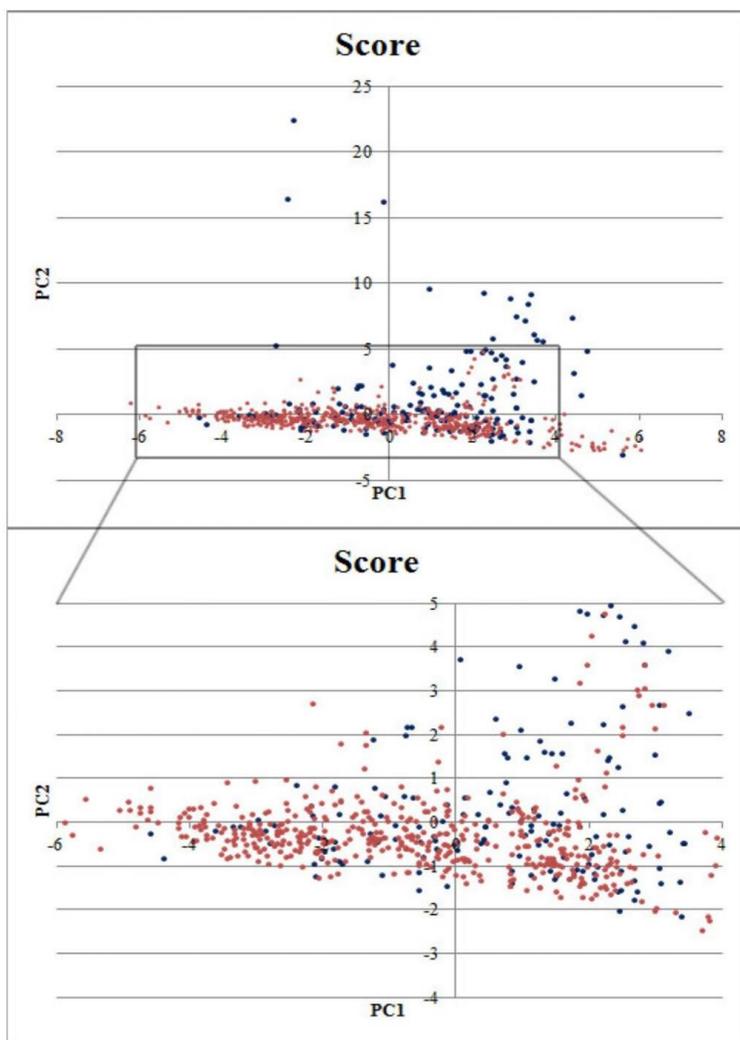


Figure 19: PCA (scaled) scoring plot for scaled flower (red) and concentrate (blue) data.

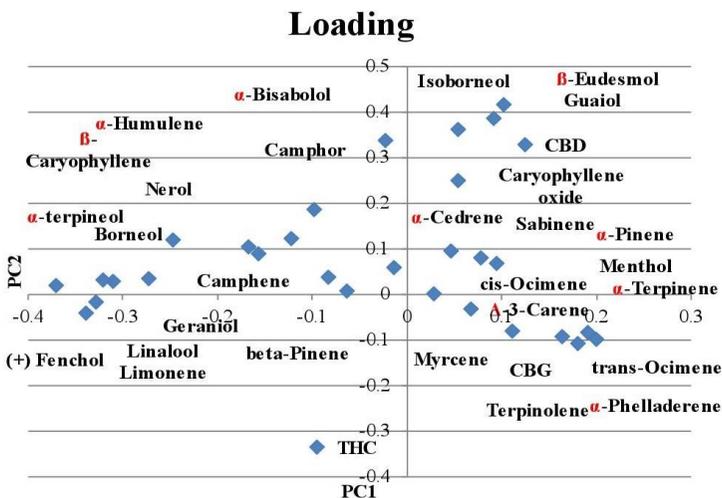


Figure 20: PCA (scaled) loading plot for scaled flower and concentrate data.

Discussion and Conclusions

Cannabis testing labs regularly receive the question “Which strain has the most THC?”, but as was shown in table 1, THCmax content can be highly variable for a strain. In some cases the level of THCmax could be 5 times higher in the highest sample compared to the lowest sample. The data presented shows that strain name cannot be used as an indication of potency. The observed variation in THC content is most likely a result of cultivation conditions. This data indicates the importance of testing for potency so the consumer knows what to expect. Strain name is also not a clear indicator of chemical composition. Variations in chemical composition of samples with the same strain name describe the importance of broad based chemical profiling. By providing patients with more information regarding the composition of the cannabis they can rely on chemical composition for reproducible physiological results instead of strain names who have been shown to not necessarily correlate with compounds present in the cannabis flowers.

PCA of the flower data did not reveal tight clustering of specific chemotypes but indicates a continuum of varied chemical composition. In most cases the replicates of a specific strain did not cluster showing a highly variable chemical composition even within a strain name. Some strains specifically Harlequin, Jack Herer, Trainwreck, Blue Dream and Green Crack showed much better clustering and seem to have a more distinct chemical profile than the majority of the strains. Perhaps these strains are more easily identified by their smell than other strains as they had relatively high concentrations of specific terpenes. As our testing perspective here is one from the patient point of view it would be interesting to investigate chemically distinct strains for their physiological properties to see to which extent chemotype might influence physiological effects.

In the comparison of samples characterized as sativa or indica most of the samples overlapped. No distinct sativa group clustered independently from indica strains, however approximately half of the indica samples separated from the overlapping group. It is likely that many of the strains labeled as sativa are not in fact pure sativa strains as represented by hemp cultivars (with relatively higher CBD levels compared to THC) but rather high potency narrow leaflet drug type cultivars with higher THC levels compared to CBD. This data also indicates another distinct chemotype referred to as an indica with high levels of limonene, fenchol, α -terpineol, camphene, linalool, THCmax, camphor geraniol, β -pinene and β -caryophyllene exists. This chemotype is represented in strains like 1st Generation Diablo, Black Mamba, Skywalker, Neptune OG and True OG. The observed data does not support the classification between indica and sativa as it is commonly presented currently in cannabis culture. A new classification system is needed to further the medical utility of cannabis products for patients to enable them to communicate better with physicians and health care providers.

Varieties with the terms OG and Kush in them are popular in the California medicinal cannabis market most likely due to their relatively high THC potency and strong pungent odor. These strains are also characterized by relatively high levels of

terpenoids with alcohol substitutions. In the comparison of strains with the term Kush vs OG a differentiation between the two categories could be noticed. OG strains had relatively higher levels of α -terpineol, fenchol, limonene, camphene, terpinolene and linalool whereas Kush samples were characterized mainly by the compounds *trans*-ocimene, guaiol, β -eudesmol, myrcene and α -pinene. Higher levels of sesquiterpenoid alcohols have been reported to be a potential distinguishing characteristic of the wide leaflet drug type strains originating from Hindu Kush region of Afghanistan and Pakistan [21].

In media outlets cannabis concentrates are referred to as potentially more dangerous than herbal cannabis due to higher potency. Although the average THC_{max} concentration of concentrates (52.5%) is much higher than the average concentration in flowers (16.8%) the relative composition is similar. It is therefore expected that physiological effects should be similar for concentrates and flower if the dose is corrected for the concentration difference.

References

1. Beutler JA, Der Marderosian AH. Chemotaxonomy of Cannabis I. Crossbreeding between Cannabis sativa and C. ruderalis, with analysis of cannabinoid content. Economic Botany. 1978; 32: 387-394.
2. Small E, Jui PY, Lefkovitch LP. A numerical taxonomic analysis of Cannabis with special reference to species delimitation. Systematic Botany. 1976; 1: 67-84.
3. Hillig KW, Mahlberg PG. A chemotaxonomic analysis of cannabinoid variation in Cannabis (Cannabaceae). American Journal of Botany. 2004; 91: 966-975.
4. Small E, Beckstead HD. Cannabinoid phenotypes in Cannabis sativa. Nature. 1973; 245: 147-148.
5. Erkelens JL, Hazekamp A. That which we call Indica, by any other name would smell as sweet. Cannabinoids. 2014; 9: 9-15.
6. Leveroni Corral V. Differential effects of medicinal marijuana based on strain and route of administration. Journal of Cannabis Therapeutics. 2001; 1: 43-59.

7. Zuardi AW, Shirakawa I, Finkelfarb E, Karniol IG. Action of Cannabidiol on the Anxiety and Other Effects Produced by Δ^9 -tetrahydrocannabinol in Normal Subjects. *Psychopharmacology*. 1982; 76: 245-250.
8. Russo EB. Taming THC: Potential Cannabis Synergy and Phytocannabinoid-terpenoid entourage effects. *Br J Pharmacol*. 2011; 163: 1344-1364.
9. De Meijer EPM, Bagatta M, Carboni A, Crucitti P, Moliterni VMC, et al. The Inheritance of Chemical Phenotype in *Cannabis sativa* L. *Genetics*. 2003; 163: 335-346.
10. Pacifico D, Miselli F, Micheler M, Carboni A, Ranalli P, et al. Genetics and marker-assisted selection of the chemotype in *Cannabis sativa* L. *Molecular Breeding*. 2006; 17: 257-268.
11. ElSohly MA, Stanford DF, Murphy TP. Chemical fingerprinting of cannabis as a means of source identification. In: ElSohly MA, editor. *Marijuana and the cannabinoids*, 1st edn. Totowa: Humana Press Inc. 2007; 51-66.
12. Ahmad UK, Muniandy Y, Hassan MS. Physical Analysis and Chemical Profiling of Illicit Herbal Cannabis using Multivariate Analysis. *Malaysian Journal of Forensic Sciences*. 2014; 5: 26-34.
13. Ilias Y, Rudaz S, Mathieu P, Christen P, Veuthey JL. Extraction and analysis of different Cannabis samples by headspace solid-phase microextraction combined with gas chromatography-mass spectrometry. *Journal of Separation Sciences*. 2005; 28: 2293-2300.
14. Fishedick JT, Hazekamp A, Erkelens T, Choi YH, Verpoorte R. Metabolic fingerprinting of *Cannabis sativa* L., cannabinoids and terpenoids for chemotaxonomic and drug standardization purposes. *Phytochemistry*. 2010; 71: 2058-2073.
15. Casano S, Grassi G, Martini V, Michelozzi M. Variations in Terpene Profiles of Different Strains of *Cannabis sativa* L. *ISHS Acta Horticulturae* 925: XXVIII International Horticultural Congress on Science and Horticulture for People (IHC2010): A New Look at Medicinal and Aromatic Plants Seminar. 2011; 115-121.

16. Tsugawa H, Cajka T, Kind T, Ma Y, Higgins B, et al. (2015) MS-DIAL: data independent MS/MS deconvolution for comprehensive metabolome analysis. *Nature Methods*, 12, 523-526.
17. Hazekamp A, Peltenburg A, Verpoorte R, Giroud C. Chromatographic and Spectroscopic Data of Cannabinoids from *Cannabis sativa* L. *Journal of Liquid Chromatography & Related Technologies*. 2005; 28: 2361-2382.
18. Elzinga S, Ortiz O, Raber JC. The Conversion and Transfer of Cannabinoids from Cannabis to Smoke Stream in Cigarettes. *Natural Product Chemistry & Research*. 2015; 3: 163.
19. Nappyana N, Agnolet S, Muntendam R, Van Dam A, Schneider B, et al. Analysis of cannabinoids in laser-microdissected trichomes of medicinal *Cannabis sativa* using LCMS and cryogenic NMR. *Phytochemistry*. 2013; 87: 51-59.
20. Hazekamp A, Fisededick JT. Cannabis - from cultivar to chemovar. *Drug Test Anal*. 2012; 4: 660-667.
21. Hillig K. A chemotaxonomic analysis of terpenoids variation in *Cannabis*. *Biochemical Systematics and Ecology*. 2004; 32: 875-891.