

# Complete Genome Sequence of *Serratia liquefaciens* Strain ATCC 27592

Wayne L. Nicholson,<sup>a</sup> Michael T. Leonard,<sup>a</sup> Patricia Fajardo-Cavazos,<sup>a</sup> Nedka Panayotova,<sup>b</sup> William G. Farmerie,<sup>b</sup> Eric W. Triplett,<sup>a</sup> Andrew C. Schuerger<sup>c</sup>

Department of Microbiology and Cell Science, University of Florida, Merritt Island, Florida, USA<sup>a</sup>; Interdisciplinary Center for Biotechnology Research, University of Florida, Gainesville, Florida, USA<sup>b</sup>; Department of Plant Pathology, University of Florida, Merritt Island, Florida, USA<sup>c</sup>

**We report the complete genome sequence of *Serratia liquefaciens* strain ATCC 27592, which was previously identified as capable of growth under low-pressure conditions. To the best of our knowledge, this is the first announcement of the complete genome sequence of an *S. liquefaciens* strain.**

Received 20 June 2013 Accepted 22 July 2013 Published 15 August 2013

Citation Nicholson WL, Leonard MT, Fajardo-Cavazos P, Panayotova N, Farmerie WG, Triplett EW, Schuerger AC. 2013. Complete genome sequence of *Serratia liquefaciens* strain ATCC 27592. *Genome Announc.* 1(4):e00548-13. doi:10.1128/genomeA.00548-13.

Copyright © 2013 Nicholson et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Wayne L. Nicholson, WLN@ufl.edu.

In a previous communication, it was reported that *Serratia liquefaciens* strain ATCC 27592 was able to grow under a combination of low-temperature (0°C), low-pressure (0.7 kPa), and CO<sub>2</sub>-enriched anoxic conditions that were intended to simulate the atmosphere of Mars (1). As part of an effort to further investigate the molecular basis of this response, we report here the complete genome sequence of the strain.

*S. liquefaciens* strain ATCC 27592 was obtained from the American Type Culture Collection (Manassas, VA), and its genome was sequenced at the University of Florida Interdisciplinary Center for Biotechnology Research (UF-ICBR) using the PacBio SMRT system (Pacific Biosciences, Menlo Park, CA). A total of 131,208 reads were obtained, with a mean read length of 4,015 bp. The initial PacBio reads were error corrected using the PacBio RS\_PreAssembler.1 module with a minimum subread length of 500 bp, a minimum read quality of 0.80, and a minimum seed read length of 5,000 bp. The error correction process yielded 20,974 reads, with an average length of 5,120 bp. A single scaffold was assembled directly from the error-corrected reads using Celera assembler (CA) version 7.0 software. The initial genome assembly was further refined using the PacBio RS\_Resequencing.1 module with Quiver consensus calling. This process removes sequencing errors that remain in the initial CA assembly and produces the final consensus genome sequence. The chromosome has 5,238,612 bp and an overall G+C content of 55.4%. We detected the presence of one 44,107-bp plasmid in the genome. Open reading frame (ORF) prediction and annotation were performed through the Rapid Annotations using Subsystems Technology (RAST) pipeline (2) using Glimmer (3). Of the 4,779 protein-coding ORFs that are present in the circular chromosome, 4,132 (86%) could be assigned by similarity to known annotated protein functions, while 647 (14%) were assigned to unknown protein functions. In addition, 2,762 ORFs (58%) were assigned to Clus-

ters of Orthologous Groups (COG) categories (4) through the Batch CD-Search tool (5). The rRNAs and tRNAs were identified using the “search\_for\_rNAs” script developed by Niels Larsen (2) and tRNAscan-SE (6), respectively. By these analyses, 79 tRNAs and 6 rRNA operons, comprising 5S, 16S, and 23S rRNA genes, were detected in the genome.

**Nucleotide sequence accession numbers.** The results of this whole-genome shotgun project have been deposited with GenBank under accession no. CP006252 (chromosome) and CP006253 (plasmid).

## ACKNOWLEDGMENTS

This work was supported by a grant from the NASA Planetary Protection Research Program (no. NNX12AJ84G) to A.C.S., P.F.-C., and W.L.N.

## REFERENCES

- Schuerger AC, Ulrich R, Berry BJ, Nicholson WL. 2013. Growth of *Serratia liquefaciens* under 7 mbar, 0°C, and CO<sub>2</sub>-enriched anoxic atmospheres. *Astrobiology* 13:115–131.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. *BMC Genomics* 9:75. doi:10.1186/1471-2164-9-75.
- Salzberg SL, Delcher AL, Kasif S, White O. 1998. Microbial gene identification using interpolated Markov models. *Nucleic Acids Res.* 26:544–548.
- Tatusov RL, Galperin MY, Natale DA, Koonin EV. 2000. The COG database: a tool for genome-scale analysis of protein functions and evolution. *Nucleic Acids Res.* 28:33–36.
- Marchler-Bauer A, Bryant SH. 2004. CD-Search: protein domain annotations on the fly. *Nucleic Acids Res.* 32:W327–W331. doi:10.1093/nar/gkh454.
- Schattner P, Brooks AN, Lowe TM. 2005. The tRNAscan-SE, snoscan and snoGPS web servers for the detection of tRNAs and snoRNAs. *Nucleic Acids Res.* 33:W686–W689. doi:10.1093/nar/gki366.