

A Multiplexed Fluorescent Calcium and NFAT Reporter Gene Assay to Identify GPCR Agonists

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Abstract: Intracellular calcium response and resulting calcium signaling to an agonist-GPCR interaction are important for the measurement of compound activity in the GPCR drug development. The increase in cytosol calcium concentration can be measured by the fluorescent calcium indicator dye such as Fluo-4 in a quick assay (in 3-5 minutes) using the fluorescence imaging plate reader. The calcium signaling through the transcription factors such as NFAT that induces gene expression can be measured by the reporter gene assay that links to the expression of reporter enzyme such as the beta-lactamase that requires 5-hour incubation. We have evaluated a multiplexed assay that sequentially measures the calcium response to a GPCR agonist in a rapid fluorescent calcium dye assay, followed by a NFAT beta-lactamase assay, and compared them in the single assay format. We found that the agonist activity determined in the multiplexed assay were comparable with these determined in the single assay format and the Z' factors were all >0.5. Five active compounds were identified that were active in both calcium dye assay and beta-lactamase assay. Therefore, our results demonstrated the utility of this multiplexed calcium assay for screening of GPCR compounds that can cross validate the primary hits and help to eliminate the false positive compounds.

Keywords: FLIPR assay, fluorescent calcium assay, beta-lactamase reporter gene assay, GPCR, high throughput screening, multiplex assay.

INTRODUCTION

G-protein-coupled receptors (GPCRs) are a large family of trans-membrane receptors that bind to extracellular agonists to activate intracellular signal transduction pathways including calcium, cAMP, ERK-phosphorylation and beta-arrestin [1, 2]. The location of these receptors on the cell membrane and the extracellular interaction site of ligands with receptors allow easily accessing of therapeutic compounds that modify the biological functions of these receptors. GPCRs are traditional targets of drug therapies and there are more than 30% of the currently available medications that are targeting GPCRs [3, 4].

Intracellular calcium release stimulated by a GPCR agonist initiates an important signaling pathway that triggers a variety of cellular functions including cell secretion, muscle contraction and neurotransmitter release. Calcium mobilization also activates downstream calcium-dependent tran-

scription factors, such as nuclear factor of activated T-cells (NFAT) that regulates gene expression [1]. Because the calcium mobilization occurs early in the signaling pathway, it is a measurement point for assessing the quick response of compounds. The measurement of NFAT response on a GPCR agonist usually involves gene transcription that reports the late response of GPCR activation. Thus, a combination of an early response with a late response for a compound screening assay may help to identify additional information and property of the compounds acting on the GPCR.

Cell based assays with the high throughput screening (HTS) technology have become increasingly popular in drug discovery and development [5-7]. The new challenge in cell based assays includes generation of an appropriate cell model, adaptation to robotic screening platform, and elimination of false positive compounds. Currently, the main HTS assay for measurement of cytosol calcium concentration is based on the fluorescent calcium binding dyes such as Fluo-4 (excitation wavelength of 488 nm, emission wavelength of 520 nm). Other calcium indicator dyes are also available including Fura-2, Indo-1, Calcium Green, and Calcium Orange though they are not appropriate for HTS. The NFAT-reporter gene assay has also been used for the measurement of intracellular calcium response to a GPCR agonist such as the beta-lactamase assay with the GeneBLazer[®] technology

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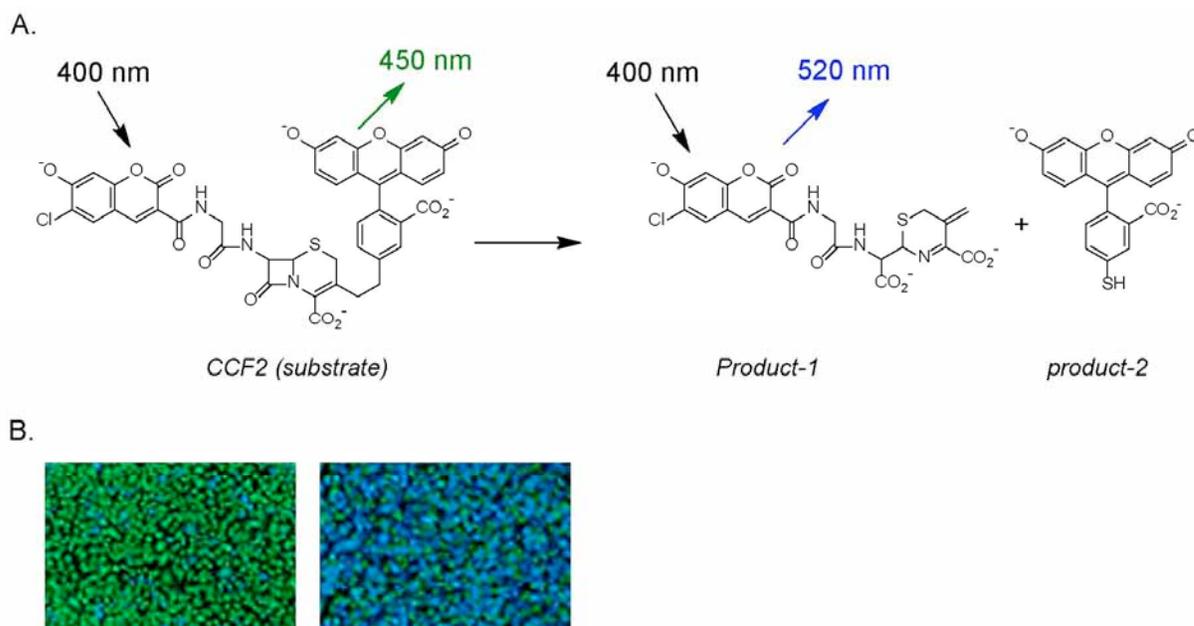


Fig. (1). Schematic diagram of coumarin cephalosporin fluorescein (CCF) cleaved by β -lactamase. After the substrate CCF-acetoxymethyl ester (AM) enters the cell, it is converted into CCF by endogenous esterases, thereby trapping it inside the cell. Excitation of CCF at 409 nm leads to FRET (A) from the coumarin moiety to the fluorescein derivative that produces green fluorescence detectable at 520 nm (green cells, B). In the presence of β -lactamase, the CCF substrate is cleaved to form two fluorophores that disrupts FRET. Excitation at 409 nm now produces a fluorescence detectable at 447 nm (blue cells, B) [8, 9].

[8, 9]. In this assay, increase in intracellular calcium by a GPCR agonist stimulates the production of beta-lactamase that turns the substrate from green color to blue color (Fig. 1). Here we report a multiplexing assay approach for GPCR compound screens that sequentially measures an intracellular calcium response detected by the calcium indicator Fluo-4 and the activity of beta-lactamase reporter in response to GPCR agonists or compounds. This multiplex assay enables measurements of GPCR-Ca signaling pathway at two different levels, which has not only saved time and cost for HTS but will also help to eliminate false positive hits.

MATERIALS AND METHODS

Materials

The CHO G α 15NFATbla-MT₁ cell line stably expressing the MT₁ (melatonin receptor 1) and β -lactamase under control of the NFAT response element was obtained from Donald Vanleeuwen (Pfizer Global R&D). The Fluo-4 NW (NW: no wash) Calcium Assay Kit was obtained from Molecular Probes/Life Technologies. The GeneBLAzer[®] assay kit was obtained from Invitrogen (Carlsbad, CA). The compounds were ordered from the Pfizer compound library. All cell culture components were obtained from Invitrogen (Carlsbad, CA), with the exception of Puromycin and Probenecid in NaOH solution, which were obtained from Sigma-Aldrich (St. Louis, MO). The assay buffer components used for the multiplexing experiment were Dulbecco's Modified Eagle Medium (DMEM, Invitrogen, #31053-028), dialyzed Fetal Bovine Serum (dFBS, Invitrogen #26400-036), Non-Essential Amino Acids (NEAA, Invitrogen, #11140-050), HEPES (Invitrogen, #15630-080), L-glutamine (Invitrogen, #35050-061), Na-Pyruvate (Invitrogen, #11360-070), and Probenecid in NaOH (Sigma Aldrich, P8761). Costar[®] (Cambridge, MA) 384-well plates with

clear bottoms (#3712) were used for seeding and assaying the CHO-MT₁ cells.

Cell Culture

MT₁ NFAT-bla Frozen CHO cells were frozen at 3×10^7 cells/ml in DMEM supplemented with 10% dFBS, 0.1 mM nonessential amino acids, 25 mM HEPES, Zeocin, Blastidicin, and Puromycin. The cells were grown in T-150 flasks (BD Biosciences, #15705-074) at 37 °C/5% CO₂, and frozen into vials. One vial was sufficient for seeding 5 plates at 12,000 cells per well.

Fluo-4 Calcium Dye Assay

The Fluo-4 NW dye was prepared according to the manufacturer's directions, with the exception of the buffer medium (included in the pack) being replaced by the assay medium containing phenol-red free DMEM, 0.1% dFBS, 0.1mM NEAA, 25mM HEPES, 3.97mM L-Glutamine, 1mM NaPyruvate, and 2.5mM Probenecid in NaOH. The cells were plated overnight in a 384-well plate in phenol-red DMEM, 5% dFBS, 0.1mM NEAA, and 25mM HEPES, at 12,000 cells/well in 40 μ l of solution. Cells were incubated overnight at 37 °C/5% CO₂ to allow the cells to adhere to the plate bottom. The following day, the media was removed from the wells, and replaced with 30 μ l of Fluo-4 NW dye solution. Cells were incubated at 37 °C/5% CO₂ for 1 hour, and assayed immediately. The assay medium added to the Fluo-4 NW dye in the stand-alone protocol contains 90% Hank's Balanced Salt Solution, 25mM HEPES, and 2.5mM Probenecid in NaOH solution.

Compound Plate Preparation

A set of 32 96-well master stock plates were prepared at a high concentration of 4mM, with a 10 point curve at a 1:4 dilution ratio. Two columns are high control of 2-

iodomelatonin at 1mM, and two columns are low control of 100% DMSO. Each master plate was stamped into four compound assaying plates at 0.5µl per well, and diluted with 50µl of the same assay medium used to dilute the Fluo-4 NW dye. The final high concentration of testing compound and final high control were 4µM and 1µM, respectively. Final DMSO concentration was 0.25%.

Multiplexed Fluo-4 Calcium and NFAT-Beta-Lactamas Reporter Gene Assay

The FLIPR liquid handling device from Molecular Devices (Sunnyvale, CA) had a 384 well-head, and was used for compound addition and testing with Fluo-4 NW dye for the FLIPR multiplexed and stand-alone screens. During compound testing, cells were stimulated with 10uL of the compound, and control wells were stimulated with 2-iodomelatonin. Each 384-well cell plate contained a control column of wells without cells creating a background subtraction column. Data was obtained for ~180 seconds after compound addition, and plotted as the difference between the maximum and minimum stimulation values ($\Delta\text{Max-Min}$). Tips were changed between each cell plate. For the β -lactamase stand-alone screen, compound was added to the cell plate using Molecular Devices FX robotic liquid handler. Following compound addition, the plates were incubated at 37 °C/5% CO₂ for 5 hours.

Cells were then loaded with GeneBLAzer[®] CCF4-AM dye. For the β -lactamase multiplexed and stand-alone screens, a loading solution was prepared consisting of 12µl of solution A, 60µl of Solution B, 925µl of Solution C, and 75µl of Probenecid in Phosphate buffer. Then, 10µl of the CCF4-AM dye solution was added to each well of the screening plates. The plates were incubated at 37 °C/5% CO₂ for 2 hours at room temperature (in the dark to prevent degradation by light) prior to reading fluorescence on the Analyst AD from Applied Biosystems (Foster City, CA) at an excitation filter of 425 nm and emission filters of 460/40 nm and 530/30 nm. Data are presented in blue/green ratio of each well divided by the average of the blue/green ratio of the un-stimulated control wells.

Data Analysis

Data for the Fluo-4 NW screen were analyzed in Spot-Fire[®] (TIBCO Software Inc., Palo Alto, CA), a valuable tool for removing outliers and presenting EC₅₀ values from the linear regression model of each agonist. Microsoft Excel (Redmond, WA) was used to compile all data from the multiplexed and stand-alone screens.

$$\%Activity = \frac{ExperimentalValue - LowControl}{HighControl - LowControl} \quad (1)$$

The % activity was calculated for the Fluo-4 NW and β -lactamase data sets. Two sets are considered: % activity above 30%, and above 50%. The multiplexed and stand-alone results were compared to analyze assay 'hit' consistency.

RESULTS

Multiplexing the FLIPR Calcium Assay with NFAT Beta-Lactamase Reporter Gene Assay

The FLIPR calcium assay with a Fluo-4 no wash dye kit in the single assay format used the standard assay buffer

consisting of 90% HBSS with 25mM HEPES, and 2.5mM Probenecid. However, this assay buffer was not adequate for the multiplexed assay because the cells did not survive the 5hr incubation for the subsequent NFAT beta-lactamase assay followed the FLIPR calcium assay. The growth media and overnight assay media for the MT1 CHO cells contained DMEM and dFBS. We found that the optimized assay buffer for the multiplexed assay consisted of DMEM without phenol-red, high glucose, Na-Pyruvate and L-glutamine and 0.1% dFBS with 2.5 mM Probenecid. The cells in this assay buffer produced optimal signal in the FLIPR calcium assay with the Fluo-4 no wash dye (Fig. 2). The slight reduction in the beta-lactamase signal in this assay buffer may be a result from the incubation with Probenecid for 5 hours. However, the EC₅₀ values of MT1 agonist remained same in the β -lactamase assay determined in the multiplexed assay format compared with that in the single assay format.

The use of frozen cells enables preparing large amount of cells in advance and pools all the cells into the same batch, which: simplifies the HTS process, minimizes the variation due to cell passage numbers, and differences in day-to-day cell culture [10]. The direct use of resuspended frozen MT₁ cells were also compared with the fresh cells for both FLIPR calcium assay and NFAT beta-lactamase assay in the single assay and multiplexed assay formatted. We found that the assay windows (signal-to-basal ratio) and EC₅₀ values of the MT₁ agonist were similar in the assays with frozen cells compared with the fresh cells. The results demonstrated the effectiveness of using frozen cells for compound screening (Fig. 3).

Assay Validation

Both FLIPR calcium assays with the Fluo-4 dye and beta-lactamase assay using the MT₁ CHO cells were carried out in the single assay format and multiplexed assay format to compare the EC₅₀ values of agonist and assay performance. As illustrated in the step-by-step assay procedure for the multiplexed assay (Fig. 4), cells in the assay plates, after one day culture, were incubated with Fluo-4 dye for one hour followed by the measurement of intracellular calcium response to the MT₁ agonist in the standard FLIPR assay. After an additional 5 hr incubation with the MT₁ agonist for the induction of the beta-lactamase reporter gene, the beta-lactamase substrate was added to the assay plates and followed by an additional 2 hr incubation. The beta-lactamase activity was then measured by detecting emissions at 465 and 520 nm from excitation at 409 nm. Thus, both assays were measured sequentially from the same assay plate.

Primary Screening of a Compound Library

The compound screens were performed in the multiplexed assay format in comparison with these in the single assay format. We found that all Z' factor values were greater than 0.5 in the screening test determined in the both assay in the single or multiplexed assay formats (Fig. 5). The assay performance was robust in all the assay formats. The EC₅₀ values and % activities for each screening format were comparable. The Fluo-4 multiplex and stand-alone formats had corresponding maximum fluorescence outputs; however, the blue/green ratio of the β -lactamase multiplexed format was slightly lower than the stand-alone protocol. This may be due to the low dFBS concentration in the assay buffer, or the addition of Probenecid (needed for the FLIPR calcium assay)

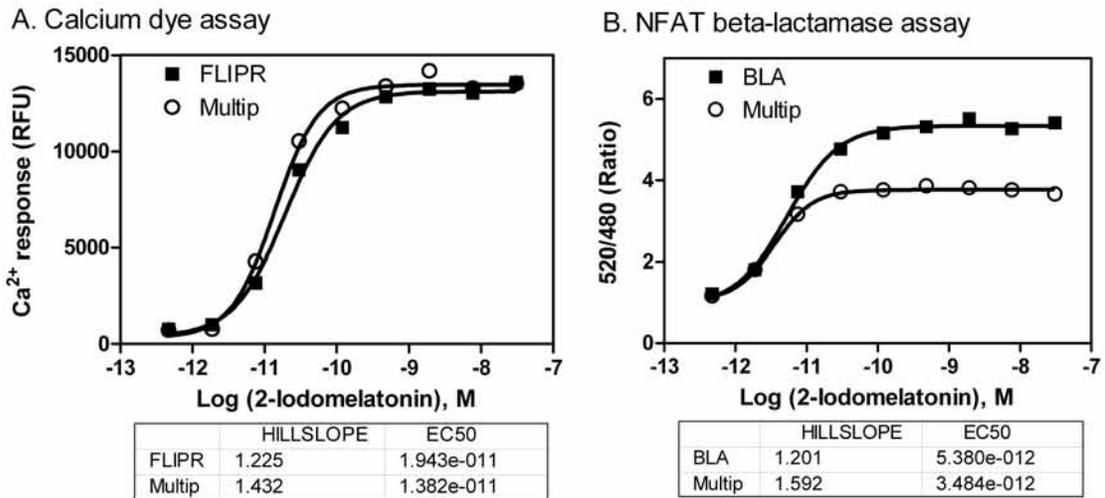


Fig. (2). Comparison of concentration responses of 2-iodomelatonin (a MT1 melatonin receptor agonist) determined in the single assay to these in the multiplexed assay. (A) EC₅₀ values of 2-iodomelatonin were 19.4 nM and 13.8 nM measured in the FLIPR calcium assay in the single assay format (FLIPR) and multiplexed assay format (Multip), respectively. (B) EC₅₀ values of 2-iodomelatonin were 5.38 pM and 3.48 pM determined in the NFAT beta-lactamase reporter gene assay in the single assay format (BLA) and multiplexed assay format (Multip), respectively.

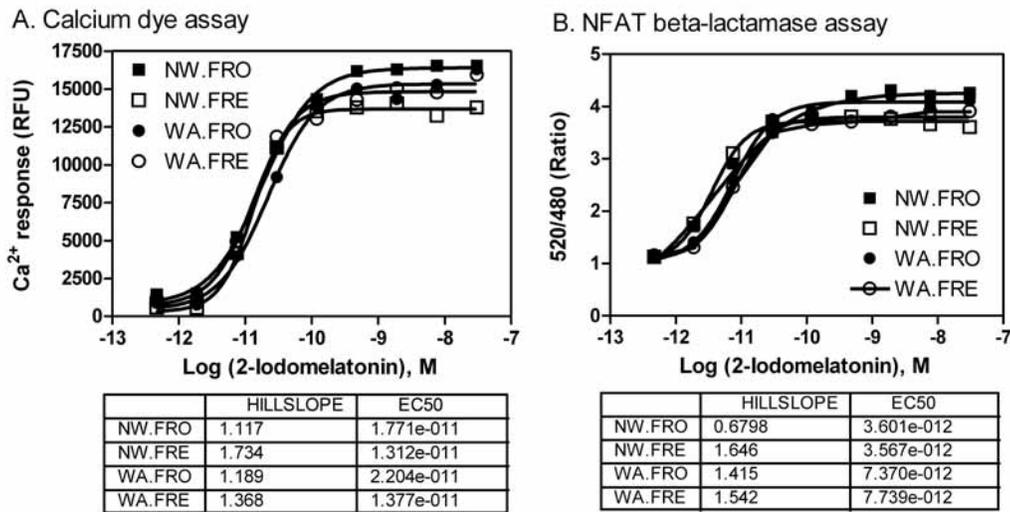


Fig. (3). Comparison of activities of 2-iodomelatonin determined in the multiplexed assay for the fresh-cells vs. frozen cells and wash vs. no-wash conditions. (A) The EC₅₀ values in the FLIPR calcium assay were 17.7, 13.1, 22.0, and 13.8 pM for the frozen cells (NW-FRO) and fresh cells (NW-FRE) in the no-wash assay and the frozen cells (WA-FRO) and fresh cells (WA-FRE) in the cell wash assay, respectively. (B) The EC₅₀ values in the NFAT-beta-lactamase reporter gene assay were 3.60, 3.56, 7.37 and 7.74 pM for the frozen cells (NW-FRO) and fresh cells (NW-FRE) in the no-wash assay and the frozen cells (WA-FRO) and fresh cells (WA-FRE) in the cell wash assay, respectively.

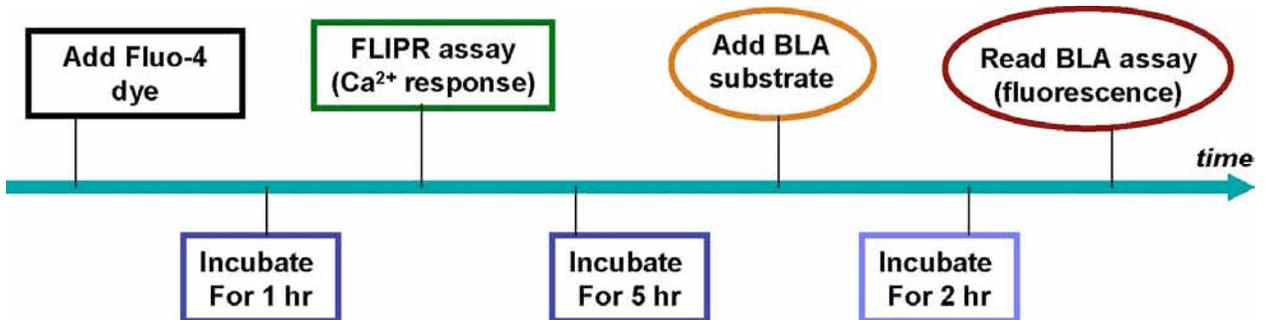


Fig. (4). A step by step illustration of assay procedure for the multiplexed FLIPR calcium and NFAT beta-lactamase assay.

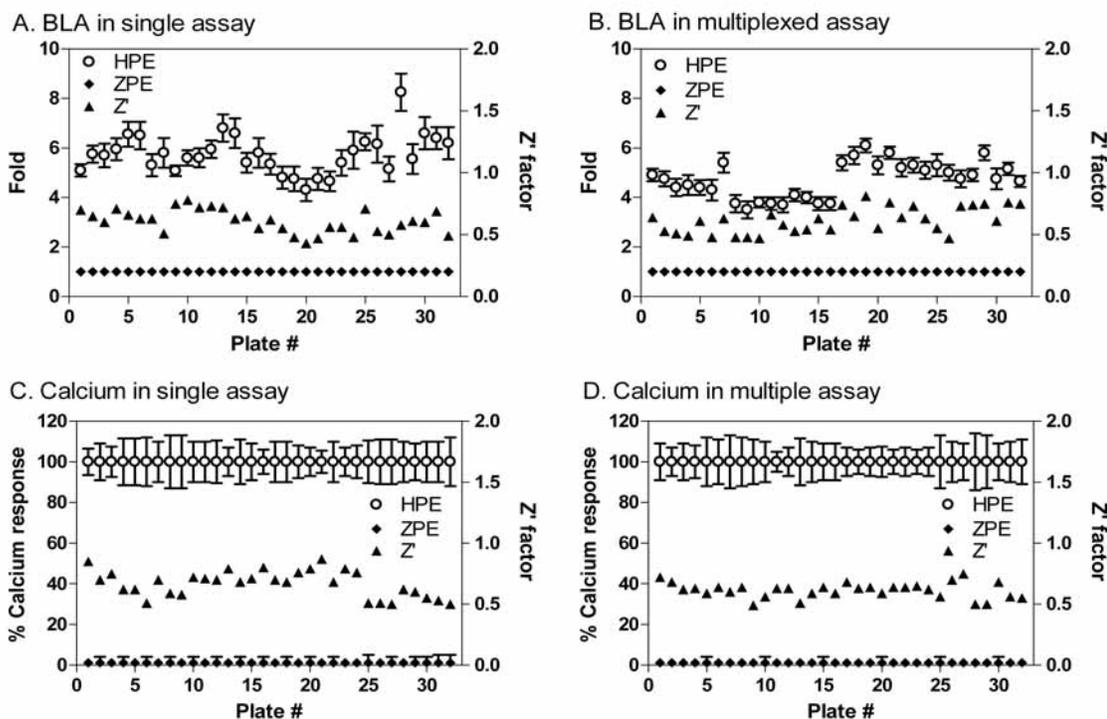


Fig. (5). HTS performance of the multiplexed assay. HPE (\diamond), the positive control wells in each plate were averaged and normalized to 100% effect in the four assay formats with error bars indicating the standard deviation of high control for each plate. ZPE (\blacksquare), the negative control wells were normalized to 0% effect. Z' Factor values (\blacktriangle) were calculated for the thirty-two compound plates tested. All Z' Factor values were greater than 0.5, indicating the robust results from this screen. (A) Results obtained from the NFAT beta-lactamase assay in the single assay format. (B) Results determined from the NFAT beta-lactamase assay in the multiplexed assay format. (C) Results obtained from the FLIPR calcium assay in the single assay format. (D) Results determined from the FLIPR calcium assay in the multiplexed assay.

for 5 hr incubation in the beta-lactamase assay. However, the EC_{50} values of the compounds were similar in the multiplexed assay compared with these in the single assay format of NFAT beta-lactamase assay (Fig. 6), indicating the results were comparable in both assays.

A total of 253 compounds were selected as the primary hits and the hit rate of $\sim 3\%$ at a cut-off of $>30\%$ activity or $\sim 1.7\%$ at $>50\%$ activity. Of these hits, 16 compounds were active in all four assay formats ($>30\%$ activity) including two in the multiplexed assay and two in the single assay format. For the beta-lactamase assay, there was a 70% correlation of these hits between the multiplexed assay format and single assay format, while an 85% correlation of the hits was observed for the FLIPR calcium assay in both assay formats. On the other hand, the multiplexed assay format showed a 73% hit rate correlation between the beta-lactamase assay and the FLIPR calcium assay (Tables 1, 2, and 3).

The EC_{50} values for each of the 16 hits that were active across all four assay formats are shown in Table 4. Five compounds of interest to the MT_1 project were also potent hits in all four assay formats (Fig. 6) that were promising as potential agonists of the MT_1 receptor.

DISCUSSION

The fluorescent calcium dye assay measures the quick calcium response to the binding of an agonist to a GPCR, while the NFAT beta-lactamase reporter gene assay meas-

ures the calcium signaling from GPCR activation then trans-activation of the NFAT reporter gene, beta-lactamase. Thus, the calcium dye assay only shows the early events of GPCR activation including agonist-GPCR binding, activation of Gq protein and PLC that release second messengers IP3 and DAG. The calcium is then released by the activation of IP3 channels in the intracellular storage site ER. On the other hand, a more complete intracellular calcium signaling pathway is needed for the activation of beta-lactamase transcription by the NFAT reporter gene. Once the cytosol calcium is increased by an agonist-GPCR interaction, calcium ions bind to calmodulin, a calcium sensor protein, which activates calcineurin, a serine/threonine phosphatase and rapidly dephosphorylates NFAT proteins. This allows NFAT enters nucleus where it binds to specific promoters and initiates gene transcription. The disadvantage of NFAT beta-lactamase assay is that it may produce a large amount of false positive hits in compound screens since so many other proteins and factors are involved in its signaling. However, a desired true GPCR activator should activate both of these mechanisms. Therefore, multiplexing the calcium dye assay with the NFAT beta-lactamase reporter gene assay enables a measurement of compound activities in two assays from the same cells. It also greatly saves the screening cost and time.

Since the calcium dye assay only takes a short time, including a 1 hour dye incubation and 3 to 5 minutes for agonist response recording, a simple physiological salt buffer is usually used for the experiment. The NFAT beta-lactamase

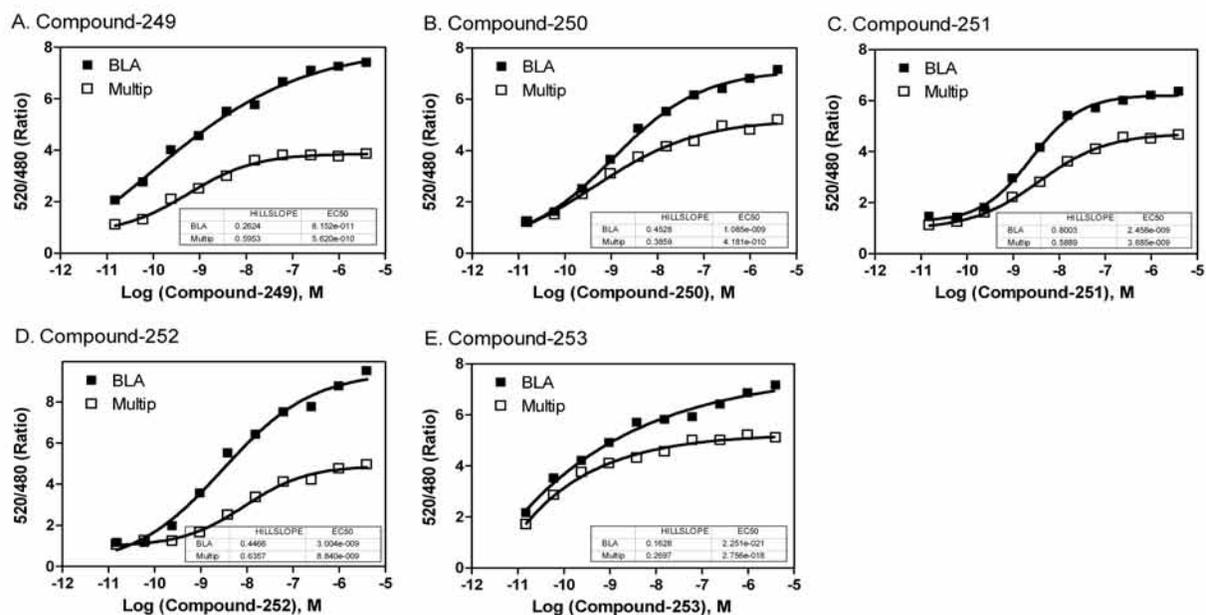


Fig. (6). Concentration responses of five representative active compounds identified from the compound library screen with this multiplexed assay. **(A)** The EC₅₀ values of Compound #249 in the NFAT beta-lactamase assay were 0.08 and 0.56 nM in the single assay format and multiplexed assay format, respectively. **(B)** The EC₅₀ values of Compound #250 in the NFAT beta-lactamase assay were 1.08 and 0.42 nM the single assay format and multiplexed assay format, respectively. **(C)** The EC₅₀ values of Compound #251 in the NFAT beta-lactamase assay were 2.46 and 3.69 nM in the single assay format and multiplexed assay format, respectively. **(D)** The EC₅₀ values of Compound #252 in the NFAT beta-lactamase assay were 3.00 and 8.84 nM in the single assay format and multiplexed assay format, respectively. **(E)** The EC₅₀ values of Compound #253 in the NFAT beta-lactamase assay were 2.25 and 2.76 nM in the single assay format and multiplexed assay format, respectively.

Table 1. Hit Correlation for all Four Type of Assays

Hit ratio	253 Compounds (All)		Hit ratio	52 Compounds (hits only)	
	Hit (30%)	Hit (50%)		Hit (30%)	Hit (50%)
4/4	6.37	4.78	4/4	30.7	23.1
3/4	2.39	3.19	3/4	11.5	15.3
2/4	3.98	2.39	2/4	19.2	11.5
1/4	7.97	5.98	1/4	38.4	28.8
0/4	79.3	83.7	0/4	0	21.1
Summary	100	100	Summary	100	100

Table 2. Assay Pairwise Correlation for Hits Above 30% Effect

		1	2	3	4
		BLA	M_MLA	FLIPR	M-FLIPR
1	BLA	26	18	21	17
2	M_BLA	18	26	21	19
3	FLIPR	21	21	37	28
4	M_FLIPR	17	19	28	33

Table 3. Assay Three and Four-Way Correlation for Hits Above 30% Effect

Combination A			Combination B			Combination C			Combination D			Combination E		
1	BLA	26	2	M_BLA	26	1	BLA	26	1	BLA	26	1	BLA	26
2	M_BLA	26	3	FLIPR	26	2	M_BLA	26	3	FLIPR	37	2	M_BLA	26

Table 3. contd....

Combination A			Combination B			Combination C			Combination D			Combination E		
3	FLIPR	37	4	M_FLIPR	33	4	M_FLIPR	33	4	M_FLIPR	33	3	FLIPR	37
												4	M_FLIPR	33
	Match	18		Match	19		Match	16		Match	17		Match	16

Table 4. EC₅₀ Values for the 16 Hits Across all Four Assay Formats

Compound #	BLA		M_BLA		FLIPR		M_FLIPR	
	EC ₅₀	%Activity						
40	18.9	113.9	31.9	87.7	4.54	75.6	5.20	83.3
144	248.9	84.2	807	36.8	8.19	103.5	15.6	85.2
148	0.097	100.4	0.371	103.8	0.046	113.9	0.106	89.6
202	38.5	96.6	50.7	65.1	1.36	91.1	0.575	82.7
205	69.8	51.7	109	37.9	31.4	73.8	6.87	94.5
234	772.6	59.4	1900	46.0	204.9	66.8	205	66.5
237	378.0	74.0	256	54.1	18.7	105.3	18.8	104.4
240	2.95	139.3	7.83	102.4	0.333	97.7	0.329	96.9
241	850.2	59.2	1100	30.4	355.7	52.1	356	51.9
243	871.4	65.9	679	39.1	364.6	74.5	365	73.8
248	1.10	116.3	8.48	74.9	0.082	105.2	0.082	104.0
249	0.599	119.9	0.671	77.1	0.065	76.8	0.065	78.0
250	0.810	111.0	0.764	102.6	0.009	96.2	0.009	97.7
251	0.955	97.5	0.350	98.0	0.037	105.9	0.037	107.5
252	0.950	147.2	0.433	104.5	0.056	99.1	0.056	95.6
253	0.020	91.2	0.093	115.7	0.040	115.2	0.040	116.9

assay requires long agonist incubation time (usually 5 hours) for the expression of reporter gene and thus cell culture medium over buffer is preferred. Thus, we have optimized the assay buffer for this multiplexed assay that consists of 0.1% dFBS, high glucose DMEM without phenol-red and with added Na-Pyruvate and L-glutamine, and 2.5 mM Probenecid. The addition of Probenecid is needed to prevent dye leaking out of cells through membrane transporters. The main advantages of using a single plate for two calcium assays for compound screens include savings in screening cost and time and more comparable results from two assays because the compound activities were determined from the same cells for the calcium dye and NFAT beta-lactamase reporter gene assays.

Cell culture to produce a large amount of cells for compound screens in a HTS format is a challenge because the day-to-day variation of yields and different cell passage numbers can cause increased variation in the screening results. Application of frozen cells has been used to minimize such a variation among the compound screening runs in different days. The frozen calls can be pooled for use in HTS that effectively minimizes the passage variations. The readily availability of frozen cells that have been stored in a freezer in aliquots allows compound screens in HTS formats without

being limited by the cell culture that is usually a bottleneck in cell based screens.

In conclusion, we have developed and validated a multiplexed GPCR-calcium response assay that sequentially measures agonist activities in a rapid calcium dye assay followed by a NFAT beta-lactamase reporter gene assay. The compound activities are determined in the same cells in a single assay plate for both calcium assays, resulting in more comparable results for the same compound. It also significantly saves the screening time and cost of reagents. Therefore, this multiplex calcium assay is a useful tool for GPCR screens of large size compound libraries in the drug discovery process.

CONFLICT OF INTEREST

The authors confirm that this article content has no conflicts of interest.

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ABBREVIATIONS

BLA	=	beta-lactamase
FLIPR	=	Fluorometric Imaging Plate Reader
GPCR	=	G-protein coupled receptor
HTS	=	high throughput screening
MT1 receptor	=	melatonin receptor 1
NFAT	=	Nuclear factor of activated T-cells
DAG	=	diacylglycerol
IP3	=	inositol trisphosphate

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