

Mini Review

SIRT1 Activators and Their Effects on Atherosclerosis Progression

Tingting Feng^{1,2}, Peng Liu¹, Yanni Xu^{1*}, Shuyi Si^{1*}

¹NHC Key Laboratory of Biotechnology of Antibiotics, Chinese Academy of Medical Sciences & Peking Union Medical College (CAMS & PUMC), Beijing, China

²Department of Clinical Pharmacy, Shanghai Jiao Tong University School of Medicine, Shanghai, China

***Corresponding authors:** Yanni Xu, NHC Key Laboratory of Biotechnology of Antibiotics, National Center for New Microbial Drug Screening, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences & Peking Union Medical College (CAMS & PUMC), Beijing, 100050, China. Tel: +861063180623; Email: xuyanniwendeng@hotmail.com

Shuyi Si, NHC Key Laboratory of Biotechnology of Antibiotics, National Center for New Microbial Drug Screening, Institute of Medicinal Biotechnology, Chinese Academy of Medical Sciences & Peking Union Medical College (CAMS & PUMC), Beijing, 100050, China. Tel: +861063180604; Email: sisymb@hotmail.com

Citation: Feng T, Liu P, Xu Y, Si S (2018) SIRT1 Activators and Their Effects on Atherosclerosis Progression. *Cardiol Res Cardiovasc Med*: CRCM-138. DOI: 10.29011/CRCM-138.000038

Received Date: 21 August, 2018; **Accepted Date:** 29 August, 2018; **Published Date:** 07 September, 2018

Abstract

Atherosclerosis is the main cause of Cardiovascular Disease (CVD), which is the leading cause of death worldwide. Hyperlipidemia and associated lipid metabolic disorders are the major risk factors involved in the progression of atherosclerosis. Sirtuins (SIRT1-SIRT7) are a family of NAD⁺-dependent deacetylases that catalyze the deacetylation of both histone and non-histone protein substrates. Among sirtuins, Sirtuin 1 (SIRT1) has been intensively investigated due to its broad regulatory effects on lifespan and cellular metabolism. Accumulating recent evidence has shown that activating SIRT1 genetically and pharmacologically has broad atheroprotective effects. Discovery of SIRT1-activating compounds (STACs, such as SIRT1720, resveratrol, SRT3025, E1231, etc) are practicable and emerge as new therapeutic strategies to treat atherosclerosis and its related cardiovascular disease which are the main lethal geriatric diseases.

Keywords: Atherosclerosis; Cardiovascular Disease (CVD); Resveratrol (RSV); SIRT1 Activators

NF-κB : Nuclear Factor-Kappa B
PCSK9 : Proprotein Convertase Subtilisin/Kexin Type 9
RSV : Resveratrol

Abbreviations

Ac-LXRα : Acetylated LXRα
CVD : Cardiovascular Disease
eNOS : Endothelial Nitric Oxide Synthase
H₂S : Hydrogen Sulfide
LDL-C : Low-Density Lipoprotein Cholesterol
LDLR : Low Density Lipoprotein Receptor
Lox-1 : Lectin-Like Ox-LDL Receptor 1
LXR : Liver X Receptor
NAD⁺ : Nicotinamide Adenine Dinucleotide
NBS-1 : Nijmegen Breakage Syndrome-1

Introduction

The yeast silencing information regulator complex (Sir complex) regulates transcriptional repression or silence, thus playing an important role in lifespan extension [1]. In mammals, there are seven members in the sirtuin family (SIRT1-SIRT7) which are ubiquitously expressed and share a highly conserved deacetylase domain [2]. Among them, SIRT1 is the most extensively studied. SIRT1 regulates a wide range of cellular processes which are crucial to cell survival, apoptosis, cell growth, cell senescence, and metabolism through deacetylating histones and many non-histone proteins [3]. Accumulating studies have demonstrated that activating SIRT1 expression/activity have extensive cardiovascular protecting effects [2,4]. For example, endothelial SIRT1 inhibits

endothelial apoptosis, and improves vascular endothelial function through increasing Endothelial Nitric Oxide Synthase (eNOS) expression, thereby exerting anti-atherosclerotic effects [5]. At the vascular smooth muscle cell level, SIRT1 protects against DNA damage and inhibits atherosclerosis, in part, through activation of the repair protein Nijmegen Breakage Syndrome-1 (NBS-1) [6]. SIRT1 may also have a prominent role in vascular biology, and regulate aspects of age-dependent atherosclerosis. Li et al. demonstrated that SIRT1 is a positive regulator of Liver X Receptor (LXR) activation and cholesterol efflux, suggesting that the interaction between diet and genetic factors could affect the progression of age-associated atherosclerosis through direct deacetylation of LXR by SIRT1 [7]. SIRT1 may also prevent atherothrombosis by downregulating the endothelial expression of tissue factor [8,9]. Additionally, SIRT1 inhibits inflammation by negatively regulates nuclear factor-kappa B (NF-κB) signaling pathway thus exerting anti-atherosclerosis effects [10].

Due to the cardioprotective effects, several SIRT1 activators have been studied both *in vivo* and *in vitro*. Most of these molecules are phenolic compounds, such as resveratrol (RSV), curcumin, etc [3]. Though RSV was questioned with some researchers proposing that it was dependent on the fluorophore utilized to label the evaluated peptide, but later, it was proved to be an allosteric activator of SIRT1 [11]. The anti-atherosclerotic function of RSV was evaluated in several animal models. RSV decreases oxidative stress, inhibits cardiac hypertrophy, promotes cell survival, and inhibits cell apoptosis [3]. Some synthetic SIRT1-Activating Compounds (STACs) have been developed and successfully tested [12], such as SRT1720, SRT1460, etc. SRT1720 was proved to attenuate angiotensin II-induced atherosclerosis in Apolipoprotein E knockout (*ApoE*^{-/-}) mice through inhibiting vascular inflammatory response [13]. Also, SRT1720 could reverse vascular endothelial dysfunction, excessive superoxide production, and inflammation with aging [14]. SRT3025 provided atheroprotection in *ApoE*^{-/-} mice by reducing hepatic Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9) secretion and enhancing Low Density Lipoprotein Receptor (LDLR) expression [15]. Hydrogen Sulfide (H₂S) has a protective role in the pathogenesis of atherosclerosis. Recent study showed that H₂S was a novel SIRT1 activator, and endogenous H₂S directly sulphydrated SIRT1, increased its deacetylation activity and stability, thus reducing atherosclerotic plaque formation [16]. Our recent study showed a new small molecule compound E1231 as a SIRT1 activator [17]. E1231, 1- {4-[2-(5-methylfuran-2-yl) quinoline-4-carbonyl] piperazin-1-yl} ethanone, may regulate cholesterol and lipid metabolism and thus play anti-atherosclerotic role in both foam macrophage and *ApoE*^{-/-} mice. This study also revealed the anti-atherosclerotic mechanism of E1231 was through SIRT1-LXRα-ABCA1 (ATP-binding cassette transporter A1) pathway. According to recent studies, the atherosclerosis protective effects of SIRT1 activators are summarized in Figure 1.

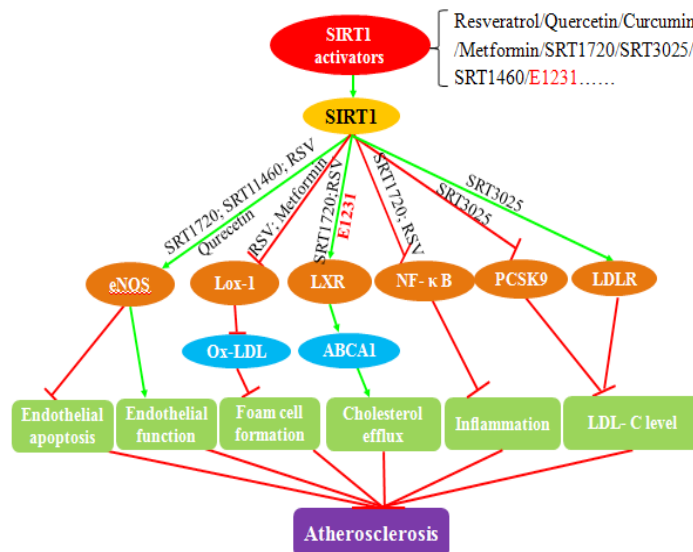


Figure 1: SIRT1 activators and their effects on atherosclerosis progression. RSV: Resveratrol; eNOS: Endothelial Nitric Oxide Synthase; Lox-1: Lectin-Like Ox-LDL Receptor 1; LXR: Liver X Receptor; NF-κB: Nuclear Factor-Kappa B; PCSK9: Proprotein Convertase Subtilisin/Kexin Type 9; LDLR: Low Density Lipoprotein Receptor; LDL-C: Low-Density Lipoprotein Cholesterol.

Our study was based on a high-throughput screening technology using HTRF (homogeneous time-resolved fluorescence) to identify SIRT1 small molecule activators. Activated SIRT1 deacetylates the substrate, causing the loss of FRET and a subsequent reduction in assay signal. This assay involved two steps: the enzymatic step and the following detection step. During the enzymatic step, the substrate-d2 containing a single acetylated lysine was incubated with the SIRT1 enzyme with or without candidate compound. Next, the deacetylation activity was quantified using an anti-acetyl MAb labeled with Eu³⁺ cryptate. With this method, a novel activator named E1231 was identified, which was distinguished from previously reported SIRT1 activators structurally, such as SRT1720, SRT3025, SRT1460, RSV, etc. In order to exclude the influence of active SIRT1 with peptide substrate containing a covalently attached fluorophore, the compound was further tested for functional activity in a SIRT1 cell-based (HepG2 cells) deacetylation assay. The protein expression levels of ac-p53 (a well-known SIRT1 substrate) [2] and p53 were tested. A selective SIRT1 inhibitor EX527, 6-chloro-2,3,4,9-tetra-hydro-1-H-carbazole-1-carboxamide [18,19], abrogated the deacetylation activity, indicating that the effect was dependent on SIRT1. Furthermore, the energetics of E1231 binding to purified SIRT1 enzyme was studied by SPR (surface plasmon resonance). The dissociation constant (K_d) was 9.61 μM, indicating a high affinity between them. Binding to the protein may induce a conformational change that leads to the exposure of an allosteric binding site.

The binding sites were simulated by Discovery studio software and are now under investigating by mutated SIRT1 proteins. We hypothesized that E1231 promoted the deacetylation activity of LXR and enhanced the activity of downstream ABCA1 in view of LXR as the direct deacetylation target of SIRT1 [7]. Thus, the first aim was to ensure where E1231 could elevate the activity of LXRs. Co-immunoprecipitation assay indicated that it could deacetylate LXR α . Moreover, RNA interference (RNAi) and luciferase reporter gene assays indicated E1231-induced LXR α targeted ABCA1 expression was SIRT1-dependent. This stimulates the ABCA1 promoter *in vitro*, thus promoting cholesterol efflux and inhibiting lipid accumulation in macrophages. More importantly, *ApoE*^{-/-} mice fed an atherogenic diet treated with E1231 decreased total cholesterol and triglyceride levels in both serum and liver, and reduced atherosclerotic plaque development compared with untreated mice, suggesting that E1231 can serve as a promising SIRT1 activator which modulates lipid and cholesterol metabolic disorder-related diseases. The principal results are summarized in Figure 2.

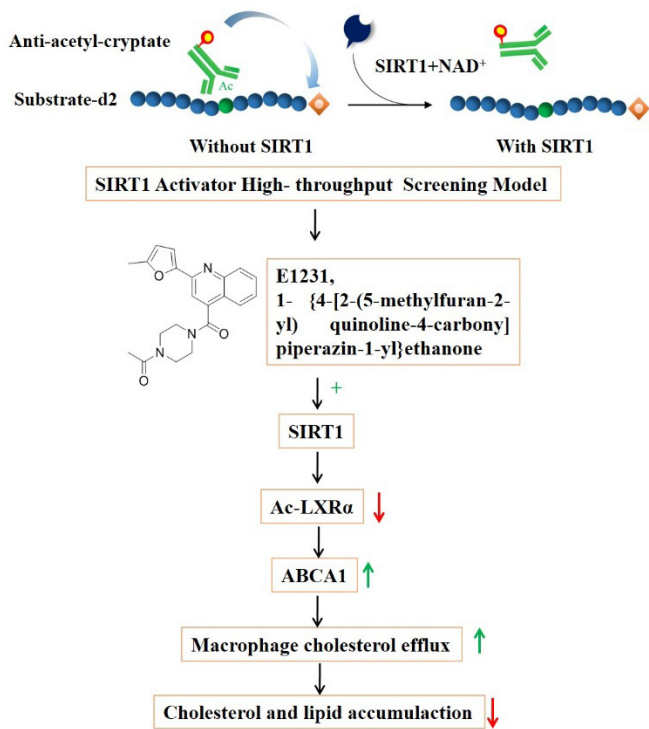


Figure 2: Working model of depicting identification of E1231 and its mechanism on regulating cholesterol and lipid metabolism. Ac-LXR α : Acetylated LXR α ; Nicotinamide Adenine Dinucleotide: NAD⁺; + (green color) indicates activate; green upward arrow indicates the increase; red upward arrow indicates the decrease.

In summary, our recent study and many others studies have demonstrated the necessity to harness high throughput compound screening to identify putative SIRT1 activators with atheroprotective

functions. Further studies are warranted to translate the pre-clinical findings to clinical settings for therapeutic purposes.

Acknowledgments

This work was supported by grants from the National Natural Science Foundation of China (grant numbers 81573482, 81621064, 81703503, 81273515 and 81503065), the Health and Medical Creation Program of CAMS (grant numbers 2016-I2M-1-011), the Key New Drug Creation and Manufacturing Program (grant No. 2018ZX09711001-003-006 and 2018ZX09735001-002-001) and the Interdisciplinary Program of Shanghai Jiao Tong University (grant No. YG2017QN26).

References

- Pan H, Finkel T (2017) Key proteins and pathways that regulate lifespan. *J Bio Chem* 292: 6452-6460.
- Ma L, Li Y (2015) SIRT1: role in cardiovascular biology. *Clinica chimica acta. International Journal of Clinical Chemistry* 440: 8-15.
- Trevino-Saldana N, Garcia-Rivas G (2017) Regulation of Sirtuin-Mediated Protein Deacetylation by Cardioprotective Phytochemicals. *Oxidative medicine and cellular longevity* 2017: 1750306.
- Xu S, Bai P, Jin ZG (2016) Sirtuins in Cardiovascular Health and Diseases. *Trends Endocrinol Metab* 27: 677-678.
- Zhang W, Huang Q, Zeng Z, Wu J, Zhang Y, et al. (2017) Sirt1 Inhibits Oxidative Stress in Vascular Endothelial Cells. *Oxidative medicine and cellular longevity* 2017: 7543973.
- D'Onofrio N, Servillo L, Balestrier ML (2018) SIRT1 and SIRT6 Signaling Pathways in Cardiovascular Disease Protection. *Antioxid Redox Signal* 28: 711-732.
- Li X, Zhang S, Blander G, Tse JG, Krieger M, et al. (2007) SIRT1 deacetylates and positively regulates the nuclear receptor LXR. *Mol Cell* 28: 91-106.
- Corbi G, Bianco A, Turchiarelli V, Cellurale M, Fatica F, et al. (2013) Potential mechanisms linking atherosclerosis and increased cardiovascular risk in COPD: focus on Sirtuins. *Int J Mol Sci* 14: 12696-12713.
- Breitenstein A, Stein S, Holy EW, Camici GG, Lohmann C, et al. (2011) Sirt1 inhibition promotes *in vivo* arterial thrombosis and tissue factor expression in stimulated cells. *Cardiovas Res* 89: 464-472.
- Kitada M, Ogura Y, Koya D (2016) The protective role of Sirt1 in vascular tissue: its relationship to vascular aging and atherosclerosis. *Aging* 8: 2290-2307.
- Hubbard BP, Gomes AP, Dai H, Li J, Case AW, et al. (2013) Evidence for a common mechanism of SIRT1 regulation by allosteric activators. *Science* 339: 1216-1219.
- Smith JJ, Kenney RD, Gagne DJ, Frushour BP, Ladd W, et al. (2009) Small molecule activators of SIRT1 replicate signaling pathways triggered by calorie restriction *in vivo*. *BMC Syst Biol* 3: 31.
- Chen YX, Zhang M, Cai Y, Zhao Q, Dai W, et al. (2015) The Sirt1 activator SRT1720 attenuates angiotensin II-induced atherosclerosis in apoE(-)/(-) mice through inhibiting vascular inflammatory response. *Biochem Biophys Res Commun* 465: 732-738.
- Gano LB, Donato AJ, Pasha HM, Hearon CM Jr, Sindler AL, et al.

- (2014) The SIRT1 activator SRT1720 reverses vascular endothelial dysfunction, excessive superoxide production, and inflammation with aging in mice, *American journal of physiology*. *Am J Physiol Heart Circ Physiol* 307: H1754-H1763.
15. Miranda MX, Van Tits LJ, Lohmann C, Arsiwala T, Winnik S, et al. (2015) The Sirt1 activator SRT3025 provides atheroprotection in ApoE^{-/-} mice by reducing hepatic Pcsk9 secretion and enhancing Ldlr expression. *Eur Heart J* 36: 51-59.
16. Du C, Lin X, Xu W, Zheng F, Cai J, et al. (2018) Sulfhydrated Sirtuin-1 Increasing Its Deacetylation Activity Is an Essential Epigenetics Mechanism of Anti-Atherogenesis by Hydrogen Sulfide. *Antioxid Redox Signal*.
17. Feng T, Liu P, Wang X, Luo J, Zuo X, et al. (2018) SIRT1 activator E1231 protects from experimental atherosclerosis and lowers plasma cholesterol and triglycerides by enhancing ABCA1 expression. *Atherosclerosis* 274: 172-181.
18. Dai H, Sinclair DA, Ellis JL, Steegborn C (2018) Sirtuin activators and inhibitors: Promises, achievements, and challenges. *Pharmacol Ther* 188: 140-154.
19. Villalba JM, Alcain FJ (2012) Sirtuin activators and inhibitors. *BioFactors* 38: 349-359.