

Mechanisms of Thermal Adaptation Revealed From the Genomes of the Antarctic *Archaea* *Methanogenium frigidum* and *Methanococoides burtonii*

Neil F.W. Saunders,¹ Torsten Thomas,^{1,10} Paul M.G. Curmi,² John S. Mattick,³ Elizabeth Kuczek,³ Rob Slade,³ John Davis,³ Peter D. Franzmann,⁴ David Boone,⁵ Karl Rusterholtz,⁵ Robert Feldman,⁶ Chris Gates,⁶ Shellie Bench,⁶ Kevin Sowers,⁷ Kristen Kadner,⁸ Andrea Aerts,⁸ Paramvir Dehal,⁸ Chris Detter,⁸ Tijana Glavina,⁸ Susan Lucas,⁸ Paul Richardson,⁸ Frank Larimer,⁹ Loren Hauser,⁹ Miriam Land,⁹ and Ricardo Cavicchioli^{1,11}

¹School of Biotechnology and Biomolecular Sciences, The University of New South Wales, Sydney, NSW 2052, Australia;

²School of Physics, The University of New South Wales, Sydney, NSW 2052, Australia and Centre for Immunology, St. Vincent's Hospital, Sydney, NSW 2010, Australia; ³Australian Genome Research Facility, Institute for Molecular Bioscience, University of Queensland, Brisbane, Qld 4072, Australia; ⁴CSIRO Land and Water, Floreat, Western Australia, 6014, Australia; ⁵Department of Biology, Portland State University, Portland, Oregon 97201 USA; ⁶Genomics Applications, Amersham Biosciences, Sunnyvale, California 94086-4520, USA; ⁷Center of Marine Biotechnology, University of Maryland Biotechnology Institute, Baltimore, Maryland 21202, USA; ⁸DOE Joint Genome Institute, Walnut Creek, California 94598, USA; ⁹IUT Genome Science and Technology, Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, USA

We generated draft genome sequences for two cold-adapted *Archaea*, *Methanogenium frigidum* and *Methanococoides burtonii*, to identify genotypic characteristics that distinguish them from *Archaea* with a higher optimal growth temperature (OGT). Comparative genomics revealed trends in amino acid and tRNA composition, and structural features of proteins. Proteins from the cold-adapted *Archaea* are characterized by a higher content of noncharged polar amino acids, particularly Gln and Thr and a lower content of hydrophobic amino acids, particularly Leu. Sequence data from nine methanogen genomes (OGT 15°–98°C) were used to generate ILLI modeled protein structures. Analysis of the models from the cold-adapted *Archaea* showed a strong tendency in the solvent-accessible area for more Gln, Thr, and hydrophobic residues and fewer charged residues. A cold shock domain (CSD) protein (CspA homolog) was identified in *M. frigidum*, two hypothetical proteins with CSD-folds in *M. burtonii*, and a unique winged helix DNA-binding domain protein in *M. burtonii*. This suggests that these types of nucleic acid binding proteins have a critical role in cold-adapted *Archaea*. Structural analysis of tRNA sequences from the *Archaea* indicated that GC content is the major factor influencing tRNA stability in hyperthermophiles, but not in the psychrophiles, mesophiles or moderate thermophiles. Below an OGT of 60°C, the GC content in tRNA was largely unchanged, indicating that any requirement for flexibility of tRNA in psychrophiles is mediated by other means. This is the first time that comparisons have been performed with genome data from *Archaea* spanning the growth temperature extremes from psychrophiles to hyperthermophiles.

The analysis of thermal adaptation across the full spectrum of known growth temperatures has been hampered by the lack of genome sequences for cold-adapted organisms. Of the 16 complete archaeal genomes (December 2002), 13 are for thermophiles or hyperthermophiles, and the remainder are for mesophiles. Even though *Archaea* contribute significantly to biomass in the predominantly cold biosphere (e.g., ~10²⁸ cells in the world's oceans; Karner et al. 2001), the only free living

isolates that have been formally characterized are a handful of methanogens (Franzmann et al. 1992, 1997; Simankova et al. 2001; Chong et al. 2002; von Klein et al. 2002). Consequently, our understanding of cold adaptation in this domain of life is limited (Cavicchioli et al. 2000).

The methanogen with the lowest known optimum growth temperature (OGT) is *Methanogenium frigidum* (15°C), which is unable to grow above 18°C (Franzmann et al. 1997). It was isolated from Ace Lake in the Vestfold Hills region of Antarctica, where the bottom waters are saturated in methane and permanently 1°–2°C. Ace Lake was also the source of *Methanococoides burtonii* (Franzmann et al. 1992). Its ability to grow on methyl-substrates and tolerate a broader range of growth temperatures (< 4° to 29°C) has led to its use for studies on protein adaptation (Thomas and Cavicchioli 1998,

¹⁰Present address: Nucleics Pty Ltd, PO Box 620, Randwick, 2031, NSW, Australia.

¹¹Corresponding author.

E-MAIL r.cavicchioli@unsw.edu.au; FAX 61-2-93852742.

Article and publication are at <http://www.genome.org/cgi/doi/10.1101/gr.1180903>. Article published online before print in June 2003.

2000, 2002; Thomas et al. 2001; Siddiqui et al. 2002) and gene regulation (Lim et al. 2000).

The methanogens stand out as the only group of organisms that have species capable of growth at 0°C (*M. frigidum* and *M. burtonii*) and 110°C (*Methanopyrus kandleri*). Moreover, there are complete genome sequences for *M. kandleri* (OGT 98°C; Slesarev et al. 2002), *Methanocaldococcus jannaschii* (85°C; Bult et al. 1996), *Methanothermobacter thermautotrophicus* (65°C; Smith et al. 1997), *Methanosarcina acetivorans* (37°C; Galagan et al. 2002), *Methanosarcina mazei* (37°C; Deppenmeier et al. 2002), and high-quality draft sequences for *Methanococcus maripaludis* (37°C) and *Methanosarcina barkeri* (35°C).

In this study we report the draft sequencing of *M. frigidum* and *M. burtonii*. These data provide the missing link for enabling a rigorous assessment of thermal adaptation at the genomic level. Using comparative genomics with the methanogen and/or all available archaeal genomes, we identified genome-wide characteristics of cold adaptation. In particular, we identified trends in amino acid and tRNA composition, and structural and compositional features of protein homology models, that distinguish the genomes of the cold-adapted *Archaea* from other *Archaea*. In addition, we highlight the value of careful processing of draft genomes for identifying targets for functional studies, by identifying unique or hypothetical genes that may be novel signatures of cold adaptation.

RESULTS

Draft Genome Statistics of *M. frigidum* and *M. burtonii*

Descriptive statistics from the current genome assemblies are presented in Table 1. Genome size has not been measured experimentally for these two organisms. However, based on the coverage and on the increase in assembly size as reads were added to the assembly, we estimate genome sizes of approximately 2–2.5 Mbp and 2.8–3 Mbp for *M. frigidum* and *M. burtonii*, respectively. These estimates fall within the known range of genome sizes for methanogens (1.6–5.7 Mbp). The number of candidate protein-coding genes in each case (*M. frigidum* 1815, *M. burtonii* 2676) agrees well with the estimated genome sizes and with the coding density of other prokaryotic genomes. Due to the draft state of the genomes, we have defined three sets of gene data: predicted coding regions, putative open reading frames (ORFs), and stringent ORFs (see Methods).

In *M. burtonii*, 81% of the coding regions had a BLAST hit to the nonredundant protein database (nrdb). This reflects the high number of sequences with similarity to the genomes of *M. acetivorans* and *M. mazei*. The proportion of sequences

from *M. burtonii* with at least one BLAST hit to a sequence from these genomes is 73% and 72%, respectively. In contrast, the *M. frigidum* sequence contains a far higher proportion of coding regions with no significant BLAST hit to the nrdb (~34%). The proportion of *M. frigidum* sequences with BLAST hits to other methanogen genomes is also lower than for *M. burtonii* (53% for both *M. acetivorans* and *M. mazei*). This indicates a substantial level of novel coding potential in the *M. frigidum* genome.

Predicted Genes Unique to *M. frigidum* and *M. burtonii*

Five predicted genes were identified that were present in both *M. frigidum* and *M. burtonii* and unique to these two organisms (no BLAST hit to the nrdb at $E \leq e^{-10}$). Threading against the SCOP database of protein folds was used to assign putative functions. One of the pairs of genes (*M. frigidum* Contig474.292.11, *M. burtonii* Scaffold8.gene3587) was identified as a 3-helical bundle DNA/RNA-binding protein. The best threading template, with z-scores of 28.92 (*M. frigidum*) and 40.04 (*M. burtonii*), was the ArsR-like transcriptional regulator SmtB from *Synechococcus* and the top nine templates were all members of the 'winged helix' DNA-binding domain superfamily.

Bacterial Cold Shock Protein (Csp) Homologs

Proteins homologous to *E. coli* CspA and *Bacillus* CspB are associated with cold adaptation in a broad range of bacteria, but are not widely dispersed in *Archaea* (Cavicchioli et al. 2000, 2002). Using bacterial Csp sequences as BLAST queries, we identified a Csp in *M. frigidum* but not in *M. burtonii*. We also searched the genome and nucleotide databases for other archaeal csp genes. Apart from those recently annotated in the *Halobacterium* genome (Kennedy et al. 2001) and an uncultured marine crenarchaeote (Beja et al. 2002), only one other was identified in *Haloferax volcanii* (GenBank accession AF442116). Residues known to be involved in RNA-binding in the bacterial Csp proteins are conserved in the archaeal sequences (Fig. 1).

Csp proteins contain the conserved cold shock domain (CSD; Somerville 1999). The absence of a Csp homolog in *M. burtonii* prompted us to search for the CSD-fold in sequences which did not have primary sequence similarity to Csp proteins. The S1 RNA-binding domain of polynucleotide phosphorylase was identified as the best threading template for two sequences from *M. burtonii* (Scaffold3.gene1629 and Scaffold5.gene2863, z-scores 29.53 and 32.66, respectively). The top three threading templates in both cases belonged to the CSD-fold family.

Amino Acid Composition

The amino acid composition of predicted archaeal proteomes was analyzed using principal components analysis (PCA). PCA can be applied to a multivariate data set in order to reduce the complexity of the data and to determine whether there are underlying trends that explain the observed variation (for discussion of the method, see Kreil and Ouzounis 2001).

The first two principal components (PCs) accounted for 71% of the variance in amino acid composition. PCs 1 and 2 correlated strongly with genome GC content and OGT (correlation coefficients for PC scores vs. these variables were -0.95 and -0.95, respectively). The two PCs were plotted for

Table 1. Genome Assembly Statistics for *M. frigidum* and *M. burtonii*

Organism	<i>M. frigidum</i>	<i>M. burtonii</i>
Total sequence reads	9870	41,957
Contigs	1413	287 (114 scaffolds)
Total assembled bases	1,597,795	2,668,325
Coverage	~2×	~9.4×
Estimated genome size	~2–2.5 Mbp	~2.8 Mbp
Protein coding regions	1815	2676
Hits to nr database	1200	2168
Putative full-length ORFs	1096	2557

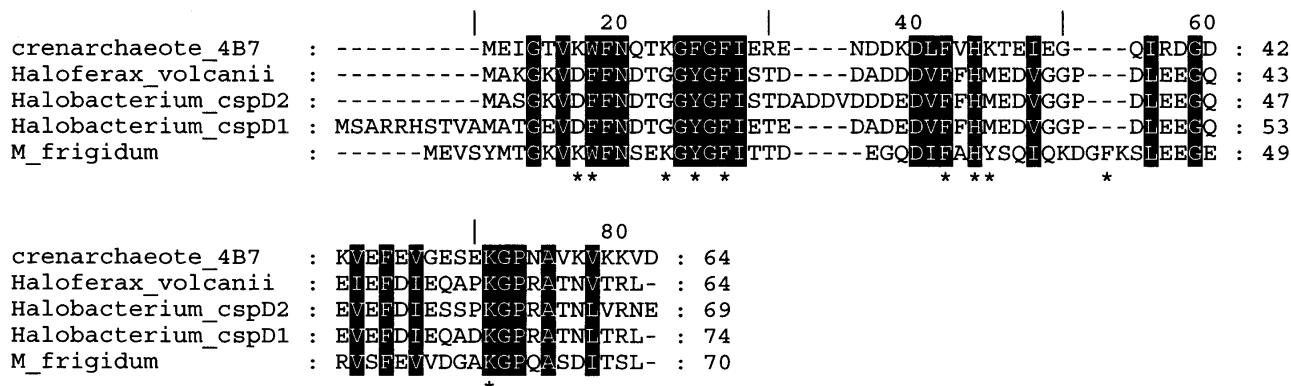


Figure 1 Multiple alignment (CLUSTALW) of archaeal Csp sequences. (*) Conserved residues in *M. frigidum* Csp involved with RNA binding.

each organism (Fig. 2A). Organisms with the highest scores were exclusively hyperthermophiles, with OGTs of 83°–103°C, and those with the lowest scores were the cold-adapted methanogens and *Halobacterium*. The presence of *Halobacterium* as an outlier can be attributed to the overrepresentation of acidic amino acids as an adaptation of this halophilic mesophile to growth in highly saline environments (Kennedy et al. 2001).

Loadings for PCs 1 and 2 were plotted for each amino acid (Fig. 2B). The influence of GC content on PC 1 was apparent, with clusters at the extremes of the axis that contain amino acids encoded by high (Gly, Ala, Arg, Val, Pro) and low (Tyr, Lys, Phe, Ile, Asn) GC codons. On the PC 2 axis, the strongest PC loadings were observed for Gln, Thr, and to a lesser extent Asp and His, whereas at the other extreme, Leu exhibited the strongest loading with weaker contributions from Trp and Glu. These amino acids were analyzed further using regression analysis. High regression coefficients for amino acid composition versus OGT were observed for Leu (0.77), Gln (−0.80), and Thr (−0.79), all with *P*-values significant at 0.01%. Regression coefficients for Trp, His, and Asp were 0.69, −0.63, and −0.58, respectively, all significant at 1%. Regression of Glu content versus OGT was less significant (coefficient, 0.36; 0.12% confidence level).

Comparative Homology Modeling of Proteins From Methanogens

Sequences of predicted proteins from the methanogen genomes were used to generate 1111 three-dimensional models. The proportion of sequences that could be modeled varied from 2.3% (*M. frigidum*) to 7.5% (*M. maripaludis*; Table 2). The complete set of models from each organism was analyzed for structural features previously associated with cold adaptation (Russell 2000; Zecchinon et al. 2001; Cavicchioli et al. 2002): (1) H-bonds/100 residues, (2) salt-bridges/100 residues, (3) fraction of atoms/residues in loops, (4) fraction of Pro in loops, (5) percentage of completely inaccessible residues, (6) fraction of accessible or inaccessible area composed of charged residues, and (7) fraction of accessible or inaccessible area composed of hydrophobic residues. The contributions of Gln, Thr, and Leu residues to the accessible or inaccessible area of the modeled proteins were also analyzed.

Significant trends were observed across the entire OGT range for the accessibility of charged and hydrophobic residues. The mean fraction of accessible area composed of

charged residues increased with OGT (Fig. 3A). A similar trend was observed for the contribution of charged residues to the total inaccessible area (Fig. 3A), but the mean inaccessible fraction was ~50% that of the accessible fraction. The contribution of hydrophobic residues to the accessible area decreased with increasing OGT (Fig. 3B), although there was no significant trend in the inaccessible area (data not shown). The summed amino acid composition of charged or hydrophobic residues did not correlate significantly with OGT, highlighting the importance of their structural context. The number of predicted salt-bridges decreased significantly at lower OGT for several models; however, this was not a consistent finding in all models (data not shown). No significant trends were identified in any of the models for the other variables tested.

The compositional decrease in Gln and Thr content with OGT seen in the PCA analysis (Fig. 2B) was also reflected by a decrease in their contribution to both accessible and inaccessible areas (Fig. 3C,D). Gln had a preference for accessible regions, whereas Thr distributed approximately equally between accessible and inaccessible areas. The accessibility of Leu residues showed no significant trend with OGT (data not shown), despite that observed for hydrophobic residues (Fig. 3B). Moreover, the proportion of hydrophobic residues in the solvent-accessible fraction increased at low OGT (Fig. 3B), whereas Leu (hydrophobic residue) strongly associated with hyperthermophiles in PCA (Fig. 2B). All of the trends described for the models were observed for both all-atoms and only-side-chain atoms.

Composition and Structure of tRNA in Archaeal Genomes

The genome sequence data from *M. frigidum* and *M. burtonii* contained 28 and 46 predicted tRNA sequences, respectively. The mean GC content was calculated for these and for predicted tRNA sequences from all available archaeal genome data, and mean GC content was plotted versus OGT for both stems and total tRNA (Fig. 4). There was a clear nonlinear relationship between OGT and tRNA GC content, with a significant elevation in GC content in the hyperthermophiles. However, GC content in tRNA from the cold-adapted *Archaea* was comparable to that of mesophiles and moderate thermophiles. These trends are mainly due to tRNA in the stem regions.

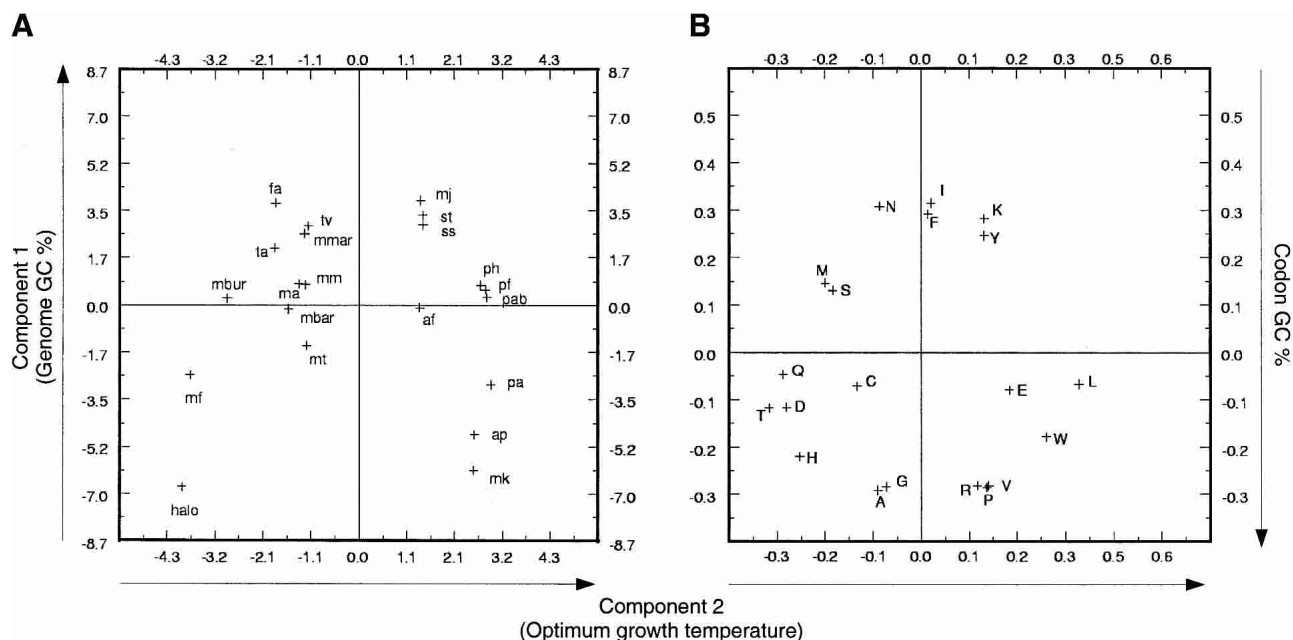


Figure 2 PCA of amino acid composition in archaeal proteomes. (A) Component scores for organisms; (B) component loadings for amino acids. af, *Archaeoglobus fulgidus*; ap, *Aeropyrum pernix*; fa, *Ferroplasma acidarmanus*; halo, *Halobacterium* sp. NRC-1; ma, *Methanosarcina acetivorans*; mbar, *Methanosarcina barkeri*; mbur, *Methanococcoides burtonii*; mf, *Methanogenium frigidum*; mj, *Methanocaldococcus jannaschii*; mk, *Methanopyrus kandleri*; mm, *Methanosarcina mazei*; mmar, *Methanococcus maripaludis*; mt, *Methanobacter thermautotrophicus*; pa, *Pyrobaculum aerophilum*; pab, *Pyrococcus abyssii*; pf, *Pyrococcus furiosus*; ph, *Pyrococcus horikoshii*; ss, *Sulfolobus solfataricus*; st, *Sulfolobus tokodaii*; ta, *Thermoplasma acidophilum*; tv, *Thermoplasma volcanium*.

DISCUSSION

Amino Acid Composition and Structural Context

Our aim in this study was to make maximum use of novel genome sequence data from the cold-adapted *Archaea* and all currently available sequence data from archaeal genomes to address the question of thermal adaptation, particularly in the low temperature range. The analysis of amino acid composition was achieved through PCA, a method which has previously been applied using different genome data sets (Kreil and Ouzounis 2001; Tekaiia et al. 2002). Importantly, the inclusion of our genome data shows that the amino acid composition in proteins from cold-adapted *Archaea* is sufficiently different to distinguish them from both mesophilic and hyper/thermophilic organisms. Strong evidence for decreased Gln and increased Glu content (Kreil and Ouzounis 2001; Tekaiia et al. 2002), and moderate evidence for a decrease in

Thr, His, and Ser content at high OGTs was reported (Kreil and Ouzounis 2001). Our results show that at least in the *Archaea*, there is an almost linear trend in the content of Gln, Thr, and Leu over the complete range of OGTs from psychrophile to hyperthermophile. The trend for increased Leu content with OGT has not been previously described. This may be due to the inclusion of data sets for the cold-adapted *Archaea* and the availability of more archaeal genomes.

We performed large-scale comparative modeling using protein sequences from the methanogen genomes and generated the largest number of models (141) from cold-adapted organisms described to date. The use of proteins from a common phylogenetic and metabolic group (Slesarev et al. 2002) minimized problems associated with genetic diversity and physiology that have been faced by other studies. The main factor, which may affect amino acid composition, was the variation in genome GC content (31%–61%), but this would

have more influence on asymmetrical substitution patterns and was accounted for in the PCA analysis. The models were analyzed for structural features that have been proposed as important for protein function at low temperature (Russell 2000; Zecchinon et al. 2001; Cavicchioli et al. 2002). Rather than focus on the details of specific subsets of proteins, we calculated mean values for the set of models from each organism in order to determine whether general trends were apparent.

The most significant findings

Table 2. Summary of Comparative Homology Modeling for Protein Sequences from Nine Methanogens

Organism	Putative ORFs	Top BLAST hits to PDB	Prospect templates	Prospect model files	Models generated
<i>M. frigidum</i>	1096	30	29	28	25
<i>M. burtonii</i>	2676	146	141	139	116
<i>M. acetivorans</i>	4540	226	223	220	182
<i>M. barkeri</i>	3145	180	175	173	142
<i>M. maripaludis</i>	1729	168	163	160	130
<i>M. mazei</i>	3371	210	206	203	168
<i>M. thermautotrophicus</i>	1873	173	169	164	124
<i>M. jannaschii</i>	1729	165	159	157	123
<i>M. kandleri</i>	1687	129	124	121	101

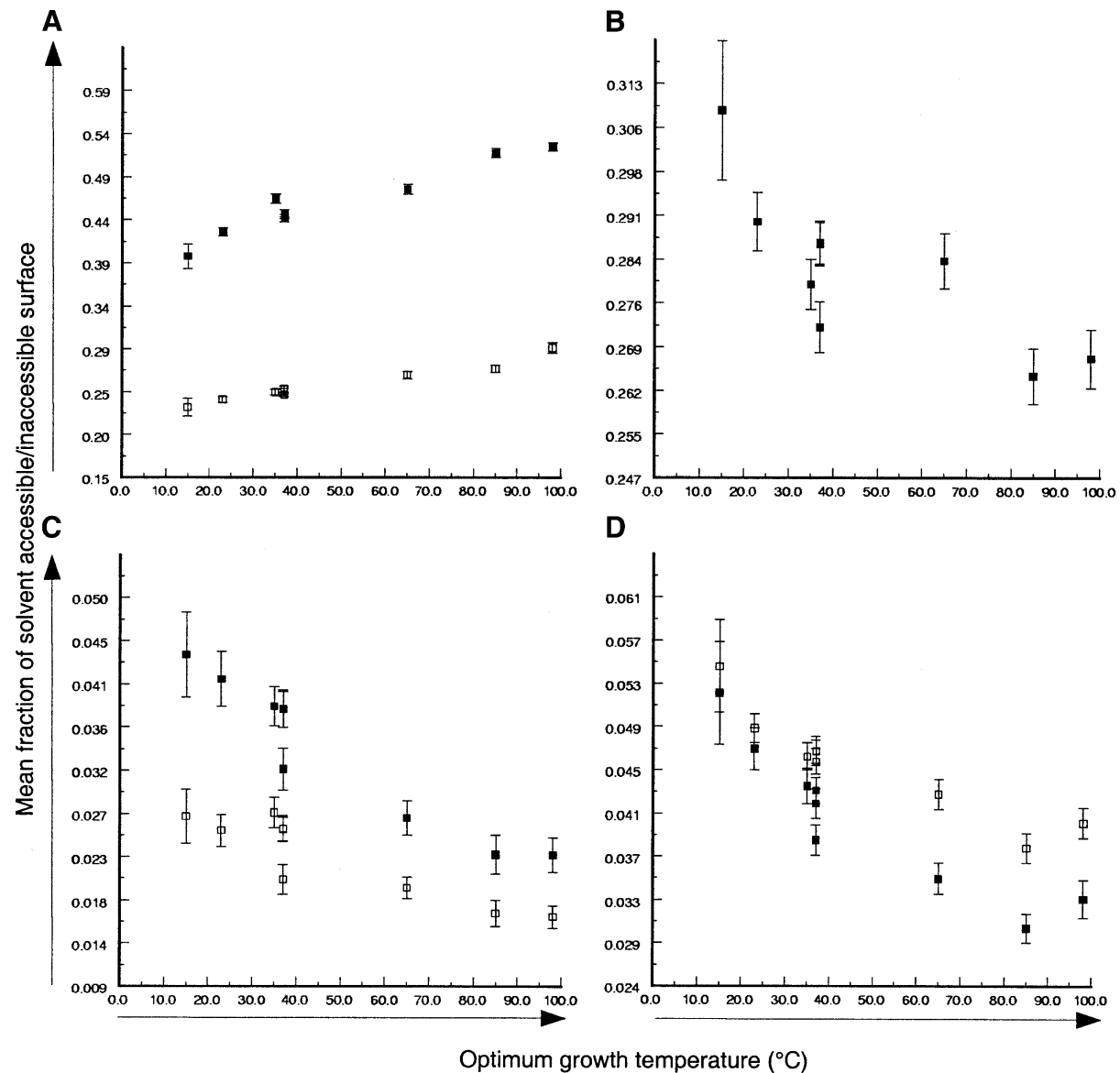


Figure 3 Contribution of amino acids to the surfaces of modeled proteins from methanogens. (A) Mean fraction of solvent-accessible (■) and buried (□) surface that is charged residues. (B) Mean fraction of solvent-accessible surface that is hydrophobic residues. (C) Mean fraction of solvent-accessible (■) and buried (□) surface that is Gln. (D) Mean fraction of solvent-accessible (■) and buried (□) surface that is Thr. Error bars are standard error of the mean (s.e.m.).

relate to the solvent accessibility of charged and hydrophobic residues. The contribution of charged amino acids to both accessible and inaccessible area is lowest in proteins from the cold-adapted *Archaea* and increases with increasing OGT. This reflects a general increase in the proportion of Lys+Arg+Glu in hyperthermophiles. However, these amino acids contribute approximately twofold greater area to the solvent-accessible area than to the inaccessible area, and the increase in contribution to accessibility with OGT is also more pronounced for the accessible area. In contrast, the exposure of hydrophobic residues in the accessible area was greatest for proteins from the cold-adapted *Archaea* and decreased with increasing OGT, indicating that exposure of hydrophobic residues to the solvent is linked to thermal adaptation. The increased exposure

of hydrophobic residues and decreased charge is likely to destabilize the surface of proteins from cold-adapted *Archaea*. Surface energy and consequent thermal expansion have previously been correlated to overall protein stability (Palma and Curmi 1999; Rees 2001; Rees and Robertson 2001). This is consistent with the findings from studies of individual cold-active proteins (Russell 2000; Zecchinon et al. 2001; Cavicchioli et al. 2002) and supports the concept that increased flexibility may reduce the activation energy of the protein-substrate transition state and increase catalytic efficiency at low temperature (Zecchinon et al. 2001; Siddiqui et al. 2002). Moreover, the reduced fraction of charged surface may also reduce the effects of cold denaturation which will also act on the accessible surface of the protein (Graziano et al. 1997).

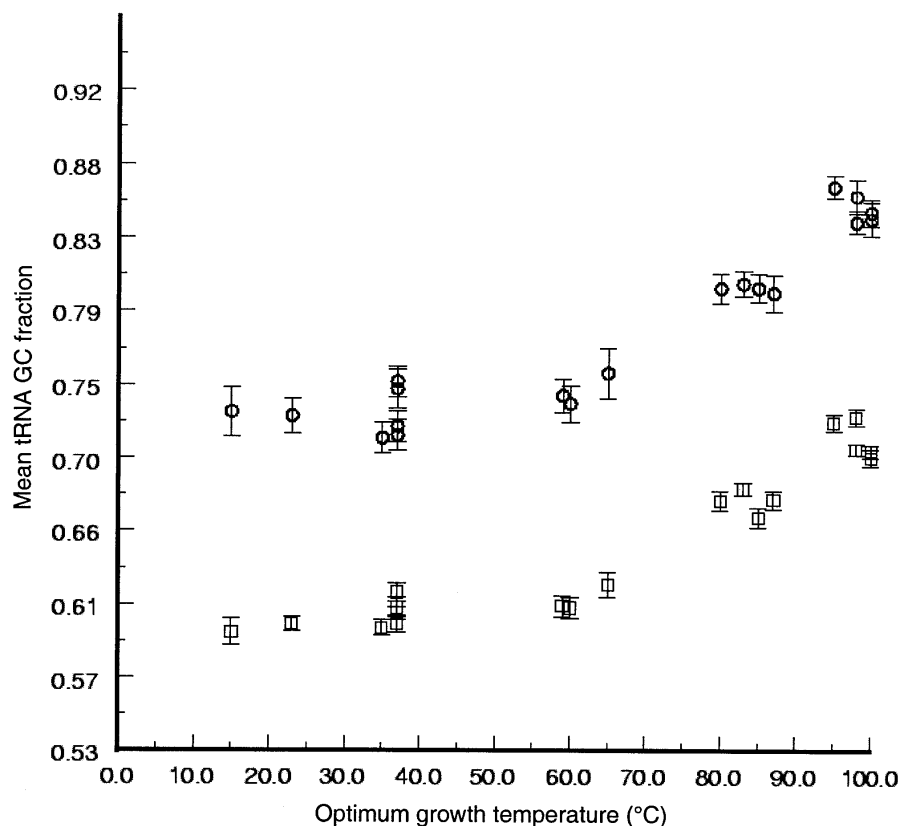


Figure 4 Mean GC fraction of predicted tRNA sequences from archaeal genomes. Total GC (□), stem GC (○). Error bars: s.e.m.

The lack of a trend for hydrophobic residues in the inaccessible area suggests a more complex relationship between internal hydrophobic interactions and thermal characteristics.

We analyzed the positioning of Gln, Thr, and Leu in the models to determine whether their marked compositional bias had a structural basis. The increased proportion of Gln and Thr in the cold-adapted *Archaea* is reflected in an increase in their contribution to both solvent-accessible and -inaccessible area. However, Gln makes up a significantly higher proportion of the solvent-accessible area as OGT decreases (Fig. 3C), whereas Thr contributes more equally to both areas (Fig. 3D). No trends in solvent accessibility were observed for Leu despite its decreasing abundance with decreasing OGT. It was reported that fewer polar noncharged residues and more hydrophobic residues occur in proteins from thermophilic *Methanococcus* species (Haney et al. 1999). It was also reported that the proportion of solvent-accessible charge increases and that of polar noncharged residues decreases in proteins from hyperthermophiles (Cambillau and Claverie 2000). Clearly, these trends are not limited to a particular thermal class but occur from psychrophiles to hyperthermophiles and so reflect adaptations in amino acid composition at all growth temperatures. Moreover, we found that the polar noncharged amino acids which contributed the most to the protein accessible area were Gln and Thr. Gln residues are susceptible to deamidation, and Thr may catalyze peptide bond cleavage (Wright 1991). The increase in Gln and Thr may compensate for the general reduction in surface charged residues. This would minimize problems with aggrega-

tion that would otherwise occur if they were only replaced by hydrophobic residues. The increased abundance of Gln and Thr may also reflect a tolerance in cold-active proteins, in contrast to their thermal lability in proteins from hyper/thermophiles.

The largest structural study of psychrophilic enzymes prior to this study examined 21 proteins using both crystal structures and homology models (Gianese et al. 2001). A tendency was observed for the replacement of Arg, Glu, and Lys at exposed sites in the direction thermophile → psychrophile. In a later comparative study, it was concluded that although specific structural features were important for function in individual protein families, characteristics generally conserved in the crystal structures of cold-adapted proteins included a decrease in the number of ion pairs, side chain contribution to exposed surface, and apolar fraction of the buried surface (Gianese et al. 2002). Our modeling study suggests that although specific structural features can be important adaptations in individual cases, the most important general trends in proteins across the entire range of OGTs relate to the interaction of the accessible surface area with the intracellular solvent.

Predicting Novel Genes for Cold-Adaptation

Five predicted genes were found to be present in both *M. frigidum* and *M. burtonii* that had no significant BLAST hit in the nrdb. One of these gene pairs very likely belongs to the superfamily of 'winged helix' DNA-binding proteins (Gajiwala and Burley 2000). Proteins of this superfamily in all three domains of life are transcription factors which regulate a wide variety of cellular processes. The unique presence of this gene pair encoding winged-helix structures in the two cold-adapted *Archaea* is therefore the strongest indicator of its possible function.

The *csp* gene in *M. frigidum* was the first such gene identified in a cold-adapted archaeon. Aside from its presence in the mesophilic halophiles, the other *Archaeon* in which it was found also inhabits cold environments (Beja et al. 2002). Moreover, the gene was absent from the mesophilic methanogens and hyper/thermophilic *Archaea*. The fact that the protein also contained all ten residues (conserved or conserved substitutions) that are involved in RNA binding (Sommerville 1999) implies it is likely to bind nucleic acid and have a cellular function in cold adaptation.

Interestingly, no *csp* gene was found in *M. burtonii*, but two hypothetical proteins were predicted to have a CSD-fold. Despite the lack of obvious primary sequence similarity between members of the CSD-fold family, they appear to have evolved common structural properties which may have been retained from an ancestral protein (Graumann and Marahiel

1998). Some members of the CSD-fold family also perform similar cellular functions; for example, the S1 domain can functionally complement an *E. coli* quadruple *csp* deletion mutant (Xia et al. 2001). In this context it is tempting to speculate that the novel CSD-fold proteins may have a role in cold adaptation, similar to that of the Csp protein in *M. frigidum*. All four of these putative nucleic acid binding proteins are clearly attractive targets for experimental study.

tRNA Flexibility

Previous studies (Galtier and Lobry 1997) have shown, and our analyses confirm, that although there is no relationship between genome GC content and OGT, the GC content of structural RNAs is elevated in hyper/thermophiles. However, without the inclusion of extensive data from psychrophiles, it had not been possible to predict whether the GC content would continue to decrease with low OGT. Our analysis shows that lowered GC content is not an adaptation in the cold-adapted *Archaea*. The tRNAs from these organisms are comparable in GC content to those from mesophiles and moderate thermophiles, suggesting a threshold temperature of around 60°C, above which GC-mediated structural integrity increases in importance. Based on genome sequences, our data are assembled from complete (or nearly complete) sets of tRNA, which would avoid biases introduced from sampling specific tRNA species. Moreover, although our data are exclusively for *Archaea*, the inclusion of tRNA from the complete genomes of psychrophilic, mesophilic, and hyperthermophilic bacteria did not change the trends we observed (data not shown).

It is likely that a minimum GC content is required for the structural integrity of tRNA, even at low OGT. However, studies of psychrophilic bacteria indicate that the maintenance of flexibility at low temperature is important for tRNA function, and that this is largely achieved through posttranscriptional incorporation of dihydrouridine (Dalluge et al. 1997), in contrast to the incorporation of different modified nucleosides in archaeal hyperthermophiles (McCloskey et al. 2001). The recent identification of the gene encoding dihydrouridine synthase (Bishop et al. 2002) enabled us to search our genome sequences, and candidate genes were identified in both organisms (data not shown). To examine this further, nucleoside modifications in tRNA from *M. burtonii* were examined by liquid chromatography mass spectrometry (LC-MS), and a significant amount of dihydrouridine was detected, a component which had not previously been identified in other methanogens (McCloskey et al. 2001; K. Noon, R. Guymon, P. Crain, J. McCloskey, M. Thomm, J. Lim, R. Cavicchioli, in prep.). This further highlights the specific adaptation strategies which have evolved in cold-adapted *Archaea*.

METHODS

Genome Sequencing and Assembly

M. burtonii was grown at 23°C (Franzmann et al. 1992) and *M. frigidum* at 15°C (Franzmann et al. 1997). Small-insert (2–4 and 4–7 kb) libraries of *M. frigidum* DNA, prepared by *Sau3AI* partial digestion of genomic DNA cloned in pUC18, were sequenced at the Australian Genome Research Facility and Amersham Biosciences. *M. burtonii* small- and large-insert libraries were prepared and sequenced at the Joint Genome Institute (JGI; http://www.jgi.doe.gov/Internal/protos_index.html). Sequencing was carried out by using dye terminator reactions and ABI 377, ABI 3700, and MegaBACE 4000 sequencers. Base

calling and shotgun assembly were performed using Phred (Ewing et al. 1998) and Phrap (P. Green, unpubl.) and the JGI JAZZ assembler for *M. burtonii*. The Staden package (Dear and Staden 1991) was used where inspection and editing of the assembly were required.

Prediction and Analysis of ORFs

Critica (Badger and Olsen 1999), Glimmer (Delcher et al. 1999), and Generation were used to identify putative coding regions, and contigs were searched using tBLASTN to identify genes that may have been missed. Because our genome sequences are at a draft stage, we defined three data sets. Coding regions were sequences derived from raw output of gene finding. These sequences can contain nonstandard amino acid letters (due to, e.g., frameshifts), but were used for initial BLAST searches. Putative ORFs were coding regions that contained in-frame sequence between start and stop codons. These may be truncated due to errors in sequencing or ORF prediction, but can be used for, for example, threading. Stringent ORFs were defined as ORFs that had a BLAST hit ($< 1e^{-10}$) in the COG database (Tatusov et al. 2001) where the sequences differ in length by $< 10\%$. These were considered genuine ORFs and used where full-length, error-free sequences were required. The naming scheme for *M. burtonii* ORFs was ScaffoldXX.geneYY. *M. frigidum* ORFs were named ContigXX.YY.ZZ, where XX is the contig number and YY.ZZ represents the start and end positions of the ORF (bp) within the contig. Scripts for data analysis were written using the BioPerl modules (Stajich et al. 2002). The *M. burtonii* and *M. frigidum* data are available at the JGI (http://www.jgi.doe.gov/JGI_microbial/html/index.html) or at our Web site (<http://psychro.bioinformatics.unsw.edu.au/>), and sequences are to be submitted to the public databases.

Analysis of Amino Acid Composition

Predicted protein sequences in Fasta format from complete archaeal genomes were downloaded from the NCBI. Sequence data were obtained from the JGI (*M. barkeri* and *Ferroplasma acidarmanus*) and from the University of Washington Genome Center (*M. maripaludis*, <http://www.genome.washington.edu/UWGC/methanococcus/>). In the latter case, Critica was used to predict coding regions, as only contig sequence data were available. Values for OGT and genome GC content were found at the TIGR CMR (<http://www.tigr.org/tigr-scripts/CMR2/CMRHomePage.spl>) and from a previous study (Galtier and Lobry 1997). For draft genomes, stringent ORF data sets were used and translated to protein sequence. Amino acid composition for each sequence set was calculated using a Perl script. Very similar compositions were obtained regardless of whether the stringent ORF or larger putative ORF data sets were used. Principal components analysis (PCA) was performed using the packages *prcomp* and *princomp*, part of the R statistical analysis package (Ihaka and Gentleman 1996). The input data were a matrix of 20 columns (percentage composition of each amino acid in the sequence set) with 21 rows (organisms).

Threading

PROSPECT version 2.0 beta (Xu and Xu 2000) was used for threading. Folds were assigned to novel proteins by threading sequences against templates from the SCOP database (Murzin et al. 1995) using global-local alignment. To identify potential cold shock domain (CSD-like) folds in proteins, all putative ORFs from *M. frigidum* and *M. burtonii* were threaded against a nonredundant set of CSDs from SCOP using global alignment. Sequences annotated as hypothetical or conserved hypothetical and with a z-score of >6 were then threaded against the complete set of SCOP domains using global-local align-

ment. Threading output was ranked by z-score and examined for significant scores against CSDs.

Comparative Homology Modeling and Analysis

Putative ORFs from the five complete and four incomplete methanogen genomes were used to search the current release of the PDB using BLAST (cutoff e^{-10}). A parser was written in Perl to extract query sequences with >40% identity and length $\pm 30\%$ or 30 residues of the BLAST hit, plus the PDB code of the top hit. A nonredundant list of PDB codes was compiled, and the corresponding structures downloaded from the PDB. The PROSPECT program make_template was used to generate threading templates not already present in the supplied PROSPECT database. PROSPECT was used to align query sequences to their best PDB template, taking into account sequence similarity (PSI-BLAST matrix) and predicted secondary structure (PHD prediction). Output from the threading alignment was processed using modellerProspect, to generate input files for MODELLER version 6v2 (Marti-Renom et al. 2000). Secondary structure in the protein models was analyzed using DSSP (Kabsch and Sander 1983). Salt bridges were calculated using the WWW interface to WHATIF (Rodriguez et al. 1998). The program naccess (Hubbard and Thornton 1993) was used for area accessibility calculations using the defaults for probe size (1.40 Å) and van der Waals radii. The total solvent-accessible and -inaccessible areas were calculated in each case. The contribution of a residue type to either the solvent-accessible or -inaccessible area was defined as the summed accessible or inaccessible area of that residue type divided by the total accessible or inaccessible area, respectively.

Prediction and Analysis of tRNA

tRNA sequences in archaeal genomes were predicted using tRNAScan-SE (Lowe and Eddy 1997). RNAfold, part of the Vienna RNA package (Hofacker et al. 1994) was used to predict tRNA secondary structure.

ACKNOWLEDGMENTS

Thanks to Jim McCloskey for data on *M. burtonii* tRNA, Greg Tyrelle, Steve Harrop, Mark Tanaka, and Tassia Kolesnikow for helpful discussions. This work was supported by the Australian Research Council and the United States Department of Energy.

The publication costs of this article were defrayed in part by payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 USC section 1734 solely to indicate this fact.

REFERENCES

- Badger, J.H. and Olsen, G.J. 1999. CRITICA: Coding region identification tool invoking comparative analysis. *Mol. Biol. Evol.* **16**: 512–524.
- Beja, O., Koonin, E.V., Aravind, L., Taylor, L.T., Seitz, H., Stein, J.L., Bensen, D.C., Feldman, R.A., Swanson, R.V., and DeLong, E.F. 2002. Comparative genomic analysis of archaeal genotypic variants in a single population and in two different oceanic provinces. *Appl. Environ. Microbiol.* **68**: 335–345.
- Bishop, A.C., Xu, J., Johnson, R.C., Schimmel, P., and de Crecy-Lagard, V. 2002. Identification of the tRNA-dihydrouridine synthase family. *J. Biol. Chem.* **277**: 25090–25095.
- Bult, C.J., White, O., Olsen, G.J., Zhou, L., Fleischmann, R.D., Sutton, G.G., Blake, J.A., FitzGerald, L.M., Clayton, R.A., Gocayne, J.D., et al. 1996. Complete genome sequence of the methanogenic archaeon, *Methanococcus jannaschii*. *Science* **273**: 1058–1073.
- Cambillau, C. and Claverie, J.-M. 2000. Structural and genomic correlates of hyperthermostability. *J. Biol. Chem.* **275**: 32383–32386.
- Cavicchioli, R., Thomas, T., and Curmi, P.M.G. 2000. Cold stress response in Archaea. *Extremophiles* **4**: 321–331.
- Cavicchioli, R., Saunders, N., and Thomas, T. 2002. In *Encyclopedia of life support systems (EOLSS)*, Eolss Publishers, Oxford, UK (<http://www.eolss.net>).
- Chong, S.C., Liu, Y., Cummins, M., Valentine, D.L., and Boone, D.R. 2002. *Methanogenium marinum* sp. nov., a H₂-using methanogen from Skan Bay, Alaska, and kinetics of H₂ utilization. *Antonie van Leeuwenhoek* **81**: 263–270.
- Dalluge, J.J., Hamamoto, T., Horikoshi, K., Morita, R.Y., Stetter, K.O., and McCloskey, J.A. 1997. Posttranscriptional modification of tRNA in psychrophilic bacteria. *J. Bacteriol.* **179**: 1918–1923.
- Dear, S. and Staden, R. 1991. A sequence assembly and editing program for efficient management of large projects. *Nucleic Acids Res.* **19**: 3907–3911.
- Delcher, A.L., Harmon, D., Kasif, S., White, O., and Salzberg, S.L. 1999. Improved microbial gene identification with GLIMMER. *Nucleic Acids Res.* **27**: 4636–4641.
- Deppenmeier, U., Johann, A., Hartsch, T., Merkl, R., Schmitz, R.A., Martinez-Arias, R., Henne, A., Wiewer, A., Baumer, S., Jacobi, C., et al. 2002. The genome of *Methanosarcina mazei*: Evidence for lateral gene transfer between bacteria and archaea. *J. Mol. Microbiol. Biotechnol.* **4**: 453–461.
- Ewing, B., Hillier, L., Wendl, M.C., and Green, P. 1998. Base-calling of automated sequencer traces using phred. I. Accuracy assessment. *Genome Res.* **8**: 175–185.
- Franzmann, P.D., Springer, N., Ludwig, W., Conway de Macario, E., and Rohde, M. 1992. A methanogenic archaeon from Ace Lake, Antarctica: *Methanococcoides burtonii* sp. nov. *Syst. Appl. Microbiol.* **15**: 573–581.
- Franzmann, P.D., Liu, Y., Balkwill, D.L., Aldrich, H.C., Conway de Macario, E., and Boone D.R. 1997. *Methanogenium frigidum* sp. nov., a psychrophilic, H₂-using methanogen from Ace Lake, Antarctica. *Int. J. Syst. Bacteriol.* **47**: 1068–1072.
- Gajiwala, K.S. and Burley, S.K. 2000. Winged helix proteins. *Curr. Opin. Struct. Biol.* **10**: 110–116.
- Galagan, J.E., Nusbaum, C., Roy, A., Endrizzi, M.G., Macdonald, P., FitzHugh, W., Calvo, S., Engels, R., Smirnov, S., Atnoor, D., et al. 2002. The genome of *M. acetivorans* reveals extensive metabolic and physiological diversity. *Genome Res.* **12**: 532–542.
- Galtier, N. and Lobry, J.R. 1997. Relationships between genomic G+C content, RNA secondary structures, and optimal growth temperature in prokaryotes. *J. Mol. Evol.* **44**: 632–636.
- Gianese, G., Argos, P., and Pascarella, S. 2001. Structural adaptation of enzymes to low temperatures. *Protein Eng.* **14**: 141–148.
- Gianese, G., Bossa, F., and Pascarella, S. 2002. Comparative structural analysis of psychrophilic and meso- and thermophilic enzymes. *Proteins* **47**: 236–249.
- Graumann, P.L. and Marahiel, M.A. 1998. A superfamily of proteins that contain the cold-shock domain. *Trends Biochem. Sci.* **23**: 286–290.
- Graziano, G., Catanzano, F., Riccio, A., and Barone G. 1997. A reassessment of the molecular origin of cold denaturation. *J. Biochem.* **122**: 395–401.
- Haney, P.J., Badger, J.H., Buldak, G.L., Reich, C.I., Woese, C.R., and Olsen, G.J. 1999. Thermal adaptation analyzed by comparison of protein sequences from mesophilic and extremely thermophilic *Methanococcus* species. *Proc. Natl. Acad. Sci.* **96**: 3578–3583.
- Hofacker, I.L., Fontana, W., Stadler, P.F., Bonhoeffer, L.S., Tacker, M., and Schuster, P. 1994. Fast folding and comparison of RNA secondary structures. *Monatsh. Chem.* **125**: 167–188.
- Hubbard, S.J. and Thornton, J.M. 1993. naccess computer program, Department of Biochemistry and Molecular Biology, University College London, UK.
- Ihaka, R. and Gentleman, R. 1996. R: A language for data analysis and graphics. *J. Comp. Graph. Stat.* **5**: 299–314.
- Kabsch, W. and Sander, C. 1983. Dictionary of protein secondary structure: Pattern recognition of hydrogen-bonded and geometrical features. *Biopolymers* **22**: 2577–2637.
- Karner, M.B., DeLong, E.F., and Karl, D.M. 2001. Archaeal dominance in the mesopelagic zone of the Pacific Ocean. *Nature* **409**: 507–510.
- Kennedy, S.P., Ng, W.V., Salzberg, S.L., Hood, L., and DasSarma, S. 2001. Understanding the adaptation of *Halobacterium* species NRC-1 to its extreme environment through computational analysis of its genome sequence. *Genome Res.* **11**: 1641–1650.
- Kreil, D.P. and Ouzounis, C.A. 2001. Identification of thermophilic species by the amino acid compositions deduced from their genomes. *Nucleic Acids Res.* **29**: 1608–1615.
- Lim, J., Thomas, T., and Cavicchioli, R. 2000. Low temperature regulated DEAD-box RNA helicase from the Antarctic archaeon, *Methanococcoides burtonii*. *J. Mol. Biol.* **297**: 553–567.

- Lowe, T.M. and Eddy, S.R. 1997. tRNAscan-SE: A program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res.* **25**: 955–964.
- Marti-Renom, M.A., Stuart, A., Fiser, A., Sánchez, R., Melo, F., and Sali, A. 2000. Comparative protein structure modeling of genes and genomes. *Annu. Rev. Biophys. Biomol. Struct.* **29**: 291–325.
- McCloskey, J.A., Graham, D.E., Zhou, S., Crain, P.F., Ibba, M., Konisky, J., Soll, D., and Olsen, G.J. 2001. Posttranscriptional modification in archaeal tRNAs: Identities and phylogenetic relations of nucleotides from mesophilic and hyperthermophilic *Methanococcales*. *Nucleic Acids Res.* **29**: 4699–4706.
- Murzin, A.G., Brenner, S.E., Hubbard, T., and Chothia, C. 1995. SCOP: A structural classification of proteins database for the investigation of sequences and structures. *J. Mol. Biol.* **247**: 536–540.
- Palma, R. and Curmi, P.M.G. 1999. Computational studies on mutant protein stability: The correlation between surface thermal expansion and protein stability. *Protein Sci.* **8**: 913–920.
- Rees, D.C. 2001. Crystallographic analyses of hyperthermophilic proteins. *Methods Enzymol.* **334**: 423–437.
- Rees, D.C. and Robertson, A.D. 2001. Some thermodynamic implications for the thermostability of proteins. *Protein Sci.* **10**: 1187–1194.
- Rodriguez, R., Chinae, G., Lopez, N., Pons, T., and Vriend, G. 1998. Homology modeling, model and software evaluation: Three related resources. *Comput. Appl. Biosci.* **14**: 523–528.
- Russell, N.J. 2000. Toward a molecular understanding of cold activity of enzymes from psychrophiles. *Extremophiles* **4**: 83–90.
- Siddiqui, K.S., Cavicchioli, R., and Thomas, T. 2002. Thermodynamic activation properties of elongation factor 2 (EF-2) proteins from psychrotolerant and thermophilic Archaea. *Extremophiles* **6**: 143–150.
- Simankova, M.V., Parshina, S.N., Tourova, T.P., Kolganova, T.V., Zehnder, A.J.B., and Nozhevnikova, A.N. 2001. *Methanosarcina lacustris* sp. nov., a new psychrotolerant methanogenic archaeon from anoxic lake sediments. *Syst. Appl. Microbiol.* **24**: 362–367.
- Slesarev, A.I., Mezhevaya, K.V., Makarova, K.S., Polushin, N.N., Shcherbinina, O.V., Shakhova, V.V., Belova, G.I., Aravind, L., Natale, D.A., Rogozin, I.B., et al. 2002. The complete genome of hyperthermophile *Methanopyrus kandleri* AV19 and monophyly of archaeal methanogens. *Proc. Natl. Acad. Sci.* **99**: 4644–4649.
- Smith, D.R., Doucette-Stamm, L.A., Deloughery, C., Lee, H., Dubois, J., Aldredge, T., Bashirzadeh, R., Blakely, D., Cook, R., Gilbert, K., et al. 1997. Complete genome sequence of *Methanobacterium thermoautotrophicum* 8H: Functional analysis and comparative genomics. *J. Bacteriol.* **179**: 7135–7155.
- Sommerville, J. 1999. Activities of cold-shock domain proteins in translation control. *Bioessays* **21**: 319–325.
- Stajich, J.E., Block, D., Boule, K., Brenner, S.E., Chervitz, S.A., Dagdigian, C., Fuellen, G., Gilbert, J.G., Korf, I., Lapp, H., et al. 2002. The Bioperl toolkit: Perl modules for the life sciences. *Genome Res.* **12**: 1611–1618.
- Tatusov, R.L., Natale, D.A., Garkavtsev, I.V., Tatusova, T.A., Shankavaram, U.T., Rao, B.S., Kiryutin, B., Galperin, M.Y., Fedorova, N.D., and Koonin, E.V. 2001. The COG database: New developments in phylogenetic classification of proteins from complete genomes. *Nucleic Acids Res.* **29**: 22–28.
- Tekaia, F., Yeramian, E., and Dujon, B. 2002. Amino acid composition of genomes, lifestyles of organisms, and evolutionary trends: A global picture with correspondence analysis. *Gene* **297**: 51–60.
- Thomas, T. and Cavicchioli, R. 1998. Archaeal cold-adapted proteins: Structural and evolutionary analysis of the elongation factor 2 proteins from psychrophilic, mesophilic and thermophilic methanogens. *FEBS Lett.* **439**: 281–286.
- Thomas, T. and Cavicchioli, R. 2000. Effect of temperature on stability and activity of elongation factor 2 proteins from Antarctic and thermophilic methanogens. *J. Bacteriol.* **182**: 1328–1332.
- Thomas, T. and Cavicchioli, R. 2002. Cold adaptation of archaeal elongation factor 2 (EF-2) proteins. *Curr. Prot. Pep. Sci.* **3**: 223–230.
- Thomas, T., Kumar, N., and Cavicchioli, R. 2001. Effects of ribosomes and intracellular solutes on activities and stabilities of elongation factor 2 proteins from psychrotolerant and thermophilic methanogens. *J. Bacteriol.* **183**: 1974–1982.
- von Klein, D., Arab, H., Volker, H., and Thomm, M. 2002. *Methanosarcina baltica*, sp. nov., a novel methanogen isolated from the Gotland Deep of the Baltic Sea. *Extremophiles* **6**: 103–110.
- Wright, H.T. 1991. Nonenzymatic deamidation of asparaginyl and glutaminyl residues in proteins. *Crit. Rev. Biochem. Mol. Biol.* **26**: 1–52.
- Xia, B., Ke, H. and Inouye, M. 2001. Acquisition of cold sensitivity by quadruple deletion of the *cspA* family and its suppression by PNase S1 domain in *Escherichia coli*. *Mol. Microbiol.* **40**: 179–188.
- Xu, Y. and Xu, D. 2000. Protein threading using PROSPECT: Design and evaluation. *Prot.* **40**: 343–354.
- Zecchinon, L., Claverie, P., Collins, T., D'Amico, S., Delille, D., Feller, G., Georgette, D., Gratia, E., Hoyoux, A., Meuwis, M.-A., et al. 2001. Did psychrophilic enzymes really win the challenge? *Extremophiles* **5**: 313–321.

WEB SITE REFERENCES

- http://www.jgi.doe.gov/Internal/protos_index.html; JGI sequencing protocols.
- http://www.jgi.doe.gov/JGI_microbial/html/index.html; JGI microbial genomes.
- <http://psychro.bioinformatics.unsw.edu.au/>; Cavicchioli lab bioinformatics site.
- <http://www.genome.washington.edu/UWGC/methanococcus/>; UWGC *Methanococcus* site.
- <http://www.tigr.org/tigr-scripts/CMR2/CMRHomePage.spl>; TIGR CMR database.

Received January 15, 2003; accepted in revised form April 22, 2003.



Mechanisms of Thermal Adaptation Revealed From the Genomes of the Antarctic *Archaea Methanogenium frigidum* and *Methanococcoides burtonii*

Neil F.W. Saunders, Torsten Thomas, Paul M.G. Curmi, et al.

Genome Res. 2003 13: 1580-1588

Access the most recent version at doi:[10.1101/gr.1180903](https://doi.org/10.1101/gr.1180903)

References

This article cites 54 articles, 24 of which can be accessed free at:
<http://genome.cshlp.org/content/13/7/1580.full.html#ref-list-1>

Creative Commons License

This article is distributed exclusively by Cold Spring Harbor Laboratory Press for the first six months after the full-issue publication date (see <http://genome.cshlp.org/site/misc/terms.xhtml>). After six months, it is available under a Creative Commons License (Attribution-NonCommercial 3.0 Unported License), as described at <http://creativecommons.org/licenses/by-nc/3.0/>.

Email Alerting Service

Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article or [click here](#).

A green banner advertisement for Gene Link. On the left is the Gene Link logo, which consists of four green diamond shapes arranged in a square. To the right of the logo, the text reads "All Modifications and Oligo Types Synthesized" in white, bold font. Below this, in a smaller white font, are the words "Long Oligos • Fluorescent • Chimeric • DNA • RNA • Antisense". On the right side of the banner, there is a stylized image of a DNA double helix. Above the DNA, the text "Oligo Modifications?" is written in a cursive font, and below it, "Your wish is our command." is written in a smaller, sans-serif font.

To subscribe to *Genome Research* go to:
<http://genome.cshlp.org/subscriptions>
