



Review

# Gene-Diet Interactions in Type 2 Diabetes: The Chicken and Egg Debate

Ángeles Ortega <sup>1,2</sup>, Genoveva Berná <sup>1,2,3</sup>, Anabel Rojas <sup>1,2,3</sup>, Franz Martín <sup>1,2,3,\*</sup>  
and Bernat Soria <sup>1,3,\*</sup>

<sup>1</sup> Centro Andaluz de Biología Molecular y Medicina Regenerativa–CABIMER, 41092 Seville, Spain; maortega@upo.es (A.O.); gberamo@upo.es (G.B.); anabel.rojas@cabimer.es (A.R.)

<sup>2</sup> Universidad Pablo Olavide, 41013 Seville, Spain

<sup>3</sup> Centro de Investigación Biomédica en Red de Diabetes y Enfermedades Metabólicas Asociadas (CIBERDEM), 28029 Madrid, Spain

\* Correspondence: fmarber@upo.es (F.M.); bernat.soria@cabimer.es (B.S.);  
Tel.: +34-954977944 (F.M.); +34-954468004 (B.S.)

Academic Editors: Lynnette Ferguson and Virginia R. Parslow

Received: 21 March 2017; Accepted: 26 May 2017; Published: 2 June 2017

**Abstract:** Consistent evidence from both experimental and human studies indicates that Type 2 diabetes mellitus (T2DM) is a complex disease resulting from the interaction of genetic, epigenetic, environmental, and lifestyle factors. Nutrients and dietary patterns are important environmental factors to consider in the prevention, development and treatment of this disease. Nutritional genomics focuses on the interaction between bioactive food components and the genome and includes studies of nutrigenetics, nutrigenomics and epigenetic modifications caused by nutrients. There is evidence supporting the existence of nutrient-gene and T2DM interactions coming from animal studies and family-based intervention studies. Moreover, many case-control, cohort, cross-sectional cohort studies and clinical trials have identified relationships between individual genetic load, diet and T2DM. Some of these studies were on a large scale. In addition, studies with animal models and human observational studies, in different countries over periods of time, support a causative relationship between adverse nutritional conditions during in utero development, persistent epigenetic changes and T2DM. This review provides comprehensive information on the current state of nutrient-gene interactions and their role in T2DM pathogenesis, the relationship between individual genetic load and diet, and the importance of epigenetic factors in influencing gene expression and defining the individual risk of T2DM.

**Keywords:** Type 2 diabetes; nutrients; nutrigenetic; nutrigenomic; epigenetic; pancreatic  $\beta$ -cell

## 1. Introduction

Diabetes is considered to be one of the biggest global health burdens of the current century. The International Diabetes Federation (IDF) Atlas 2015 [1] estimates that 415 million adults suffer from this disease. In addition, diabetes prevalence is constantly increasing. There are three main types of diabetes (Type 1, Type 2 and gestational diabetes). Type 2 diabetes (T2DM) is the most common type of diabetes. According to IDF, in high-income countries, up to 91% of adults with diabetes have the Type 2 form. The disease usually appears in adults, mostly due to an inappropriate lifestyle, however, T2DM is increasing in children and adolescents.

T2DM is a very complex disease, for which the hallmarks are  $\beta$ -cell failure and insulin resistance (IR). A combination of genetic, epigenetic, environmental, and lifestyle factors, such as diet, are responsible for the onset and development of T2DM [2]. Genome-wide association studies have identified many genetic variant loci involved in T2DM, however, these variants can only explain

between approximately 10–15% of the heritability of the disease [3]. Strong evidence in the field proposes epigenetic factors as regulators of gene expression in T2DM. Epigenetic marks might be influenced by environmental factors that ultimately have an impact on the risk of T2DM [4].

The nutrient-gene interactions, as well as the relationship between individual genetic load and diet play an important role in the pathophysiology of T2DM [5]. Thus, a better knowledge of nutrigenomics and nutrigenetics of T2DM, together with a better understanding of the epigenetic marks are instrumental in the development of precision medicine approaches to prevent, detect and treat this disease.

In this review, we will examine the nutrient-gene interactions and their role in T2DM pathogenesis. We will discuss how an individual's genetic component can also impact on the risk of T2DM, depending on the type of diet they follow. Finally, since epigenetic variations may interact through diet to influence gene expression and define the individual risk of T2DM, we will illustrate the importance of epigenetic factors in explaining the missing heritability cause of T2DM.

## 2. Effects of Nutrients on Gene Expression in the Pathogenesis of T2DM

Dietary nutrition plays an important role in the pathogenesis of T2DM, making the identification and analysis of nutrient-gene interactions a necessary step in the understanding of this chronic disease. Nutrigenomics, the area of nutrition that uses molecular tools to elucidate the influence of nutrients over the genome, proteome and metabolome, provides a genetic understanding of how common dietary components lead to up and/or down-regulation of gene(s) and changes in protein expression levels [6]. The nutrient gene interaction may modulate the gene expression via different mechanisms: (i) Directly; (ii) Through their metabolites; (iii) By activating various signalling molecules of complex metabolic pathways [7].

### 2.1. Polyphenol-Gene Interactions in T2DM Pathogenesis

Polyphenols are a large and heterogeneous group of plant products found in fruits, vegetables, legumes, cereals and chocolate, which may be further classified into flavonoids, lignans, phenolic acids, and stilbenes [8]. Numerous studies have reported beneficial effects of polyphenol-rich foods in lowering the risk of T2DM and improving inflammation and glycaemia markers in Type 2 diabetic subjects [9,10]. Dietary polyphenolic compounds may exert hypoglycemic effects in multiple ways, such as diminished carbohydrate digestion and glucose absorption, inhibition of glucose release, stimulation of insulin secretion and protection of pancreatic  $\beta$ -cells against glucotoxicity, increased glucose uptake in peripheral tissues by modulating intracellular signalling, antioxidant activity and inhibition of advanced glycation end product formation [9,11,12].

#### 2.1.1. Flavonoid-Gene Interactions in DM Pathogenesis

Dietary flavonoids include flavones, flavonols, flavanols, flavanones, isoflavones, and anthocyanins. Numerous studies strongly support the protective effects of polyphenols on glucose homeostasis, and on some specific molecules such as flavanols, quercetin, luteolin and others that have been shown to impact on different steps of intracellular signalling pathways (insulin secretion, insulin signalling and glucose uptake, enhancing mitochondrial status, suppression of inflammatory cytokine production and reactive oxygen species (ROS)/reactive nitrogen). In addition, flavonoids, in a similar way to phenolic acids and tannins, have the property of inhibiting the  $\alpha$ -glucosidase and  $\alpha$ -amylase enzymes responsible for carbohydrate digestion [9,11–15].

Consistently, human urinary excretion studies on T2DM case-controlled pairs selected from the Nurses' Health Study (NHS) and NHSII have indicated that specific flavonoids, including flavanones (naringenin and hesperetin) and flavonols (quercetin and isorhamnetin), as well as the phenolic acid, caffeic acid, but not flavan-3-ols and ferulic acid, were associated with a lower risk of developing T2DM in a time-dependent manner [16]. In an average 8.8 year follow-up study in women after ingestion of total or individual flavonoids and flavonoid-rich foods it was shown that whereas total flavonols

and flavones, quercetin, kaempferol, myricetin, apigenin, and luteolin did not significantly increase diabetes risk, apple or tea consumption was inversely associated with T2DM risk [17]. This is in accordance with the Health Professionals Follow-Up Study which also found that higher consumption of anthocyanins, particularly from pears, apples and blueberries, but not total flavonoid intake or other flavonoid subclasses, were inversely associated with T2DM [18]. By contrast, the Iowa Women's Health prospective cohort study in postmenopausal women suggested that only food constituent flavonoid consumption was not associated with diabetes risk. However, in the same study, regular consumption of red wine, white wine, beer, and liquors was inversely associated with diabetes, suggesting that it is the alcohol per se and not the polyphenols which may protect against diabetes [19]. The conflicting results of those studies might be a consequence of the difficulties in evaluating the contribution of other dietary constituents, or improper timing and quality of dietary assessment.

Flavan-3-ols are present in fruit, tea, cocoa and chocolate (Table 1). Catechins are flavanols compounds present mainly in green tea. In streptozotocin (STZ)-induced diabetic rats, catechin hydrate treatment reduced the elevated level of serum glucose and prevented vascular endothelial dysfunction through the activation of endothelial phosphatidylinositol 3-kinase (PI3K) signal and subsequent activation of eNOS signalling systems [20].

One of the most common flavanols is epigallocatechin gallate (EGCG). In rats, EGCG has been demonstrated to affect numerous processes, including more efficient insulin secretory function, increased viability of  $\beta$ -cells under conditions of glucotoxicity, glucose uptake and tolerance, IR, oxidative stress, inflammation and mitochondrial function. These effects of EGCG involved augmented insulin receptor substrate-2 (IRS2) signalling, although other mechanisms might be relevant too. The reductive glucolipotoxic effect of EGCG occurs through the incrementation of AMP-activated protein kinase (AMPK) signalling to inhibit the activities of lipogenic enzymes and ameliorate mitochondrial function [21]. Similarly, the positive effects of EGCG on IR in dexamethasone-induced rat L6 cells was essentially dependent on the AMPK and PI3K/Akt activation pathway [22], while in differentiated rat L6 myotubes, EGCG and epicatechin enhanced insulin-mediated glucose uptake in vitro and translocation of glucose transporter (GLUT) 4 [23]. EGCG has also been shown to protect insulin-producing  $\beta$ -cells from pro-inflammatory cytokine-induced cytotoxicity via the modulation of B cell CLL/lymphoma 2 (BCL-2) expression [24].

In animal studies, EGCG supplementation (1% in diet) in obese db/db mice improved glucose tolerance, increased glucose-stimulated insulin secretion from pancreatic  $\beta$ -cells and preserved islet morphology by reducing the expression of the endoplasmic reticulum stress-associated islet marker genes *DNA-damage inducible transcript 3 (Ddit3)* and its downstream target protein *phosphatase 1, regulatory subunit 15A (Ppp1r15a)*, as well as *cyclin-dependent kinase inhibitor 1A (Cdkn1a)* [25]. In rodent models of T2DM, EGCG enhanced oral glucose tolerance in severely diabetic mice and in moderately diabetic rats, and increased glucose-stimulated insulin secretion. EGCG upregulated glucokinase, acyl CoA oxidase-1 (ACO-1) and carnitine palmitoyl transferase-1 (CPT-1) mRNA expression in liver and adipose tissues of obese db/db mice in a dose-dependent manner. Conversely, exposure of H4IIE hepatoma cells to EGCG decreased the expression of genes involved in gluconeogenesis and the synthesis of fatty acids, triacylglycerol and cholesterol, whereas genes involved in glucose transport (*Glut 1*) and glycolysis (phosphofructokinase) were induced in [26].

In humans, results from epidemiological studies on EGCG are contradictory. Whereas some studies have demonstrated that an acute, high dose of EGCG concentrated green tea supplement could control postprandial hyperglycemia [27], long-term studies in Type 2 diabetic adults did not reveal a hypoglycemic effect [28]. A recent meta-analysis of clinical human studies with catechins in controlled trials on glycemic control showed that green tea catechins (EGCG) reduced fasting glucose over a median of 12 weeks, whereas fasting insulin, glycated haemoglobin (HbA1c), and homeostatic model assessment of IR (HOMA-IR) were not affected [29].

Naringin and hesperidin, the two major flavanones, are present in citrus fruits and also have hypoglycemic and hypolipidemic action in T2DM by partly regulating the fatty acid and

cholesterol metabolism and affecting the gene expression of glucose-regulating enzymes (Table 1). In C57BL/KsJ-db/db mice, both hesperidin and naringin significantly increased hepatic glucokinase mRNA level via peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ) and upregulated adipocyte *Glut 4*. Naringin also lowered hepatic mRNA expression of phosphoenolpyruvate carboxykinase PEPCK and glucose-6-phosphatase G6Pase [30].

T2DM rats fed a high-fat diet (HFD) and administered low-dose STZ injection naringin, were shown to improve  $\beta$ -cell function, ameliorating hyperglycemia, hyperinsulinemia and IR by increasing PPAR- $\gamma$  and heat shock proteins (HSP) expression in the livers of diabetic rats [31]. Despite this promising data, more clinical studies in humans to support the anti-diabetic potential of these flavanones are necessary.

The most abundant flavonoids are flavonols, which are widely distributed throughout plant-based foods. Quercetin is one of the most important dietary flavonols and is found in red wine and numerous fruits, vegetables and nuts (Table 1). The Protective effects of quercetin have been related to decreasing oxidative stress with preservation of pancreatic  $\beta$ -cell integrity in STZ-induced diabetic rats, possibly by decreasing lipid peroxidation, nitric oxide (NO) production and by increasing antioxidant enzyme activity [32]. In addition, quercetin seems to be beneficial for the treatment of diabetic neuropathy as it has been shown to protect cultured dorsal root ganglion neurons of rats against high glucose-induced apoptosis via Nrf-2/HO-1 activation and nuclear factor kappa B (NF- $\kappa$ B) inhibition [33]. The inhibitory effect on inflammatory pathways (activation of NF- $\kappa$ B signalling and reduction in serum level of both TNF- $\alpha$  and C-reactive protein (CRP)) without affecting glucose levels, would explain the potential of quercetin to prevent diabetic vascular complications in both animal models with insulin deficiency and resistance by its anti-inflammatory effect rather than its metabolic effects [34]. In a different study, dietary supplementation with 0.5% quercetin in the diet for 2 weeks in STZ-induced T2DM BALB/c mice decreased blood glucose and enhanced serum insulin concentrations by inhibiting expression of *cyclin-dependent kinase inhibitor p21 (WAF1/Cip1) (Cdkn1a)* associated with cell proliferation in the liver and pancreas [35].

Quercetin also showed anti-inflammatory effects in male C57Bl/6j mice and obese Zucker rats. In male C57Bl/6j mice, supplementation with 0.8% quercetin in a HFD for 8 weeks decreased the circulating markers of inflammation interferon-gamma (IFN- $\gamma$ ), interleukin-1 $\alpha$  (IL-1 $\alpha$ ), and interleukin-4 (IL-4) [36]. In obese Zucker rats, administration of both 2 and 10 mg/kg of body weight of quercetin for 10 weeks improved dyslipidemia, hypertension, and hyperinsulinemia, but only the highest dose of quercetin produced anti-inflammatory effects as it increased the plasma concentration of adiponectin, reduced NO $_x$  levels in plasma and lowered TNF- $\alpha$  production in visceral adipose tissue [37].

The hypoglycemic effects of the major dietary flavones apigenin and luteolin, found in celery, parsley and many herbs, may be related to the protection of pancreatic  $\beta$ -cells against glucotoxicity (Table 1). In addition, together with quercetin, apigenin, and luteolin protected RINmF5 cells from cytokine-induced pancreatic-cell damage by inhibiting inducible nitric oxide synthase (iNos) gene expression through suppression of NF- $\kappa$ B activation, and also prevented IL-1 $\beta$ - and IFN- $\gamma$ -mediated inhibition of insulin secretion [38]. However, it is not clear whether this effect also occurs in the islets in vivo.

The major dietary isoflavones are daidzein and genistein which are primarily present in soybeans, soy foods and legumes, and have been shown to reduce hyperinsulinemia (Table 1). Studies in rats with diet-induced obesity showed that soy isoflavones in a HFD significantly stimulated insulin secretion, decreased PPAR- $\gamma$ , GLUT 2, and SREBP-1 expression, and ameliorating hyperinsulinemia observed during obesity [39]. In STZ-induced diabetic mice, dietary intake of genistein significantly improved hyperglycemia, glucose tolerance, and circulating insulin levels, preserving islet  $\beta$ -cell proliferation, survival, and mass. The insulin-secreting activity and proliferative effect of genistein in pancreatic  $\beta$ -cells is mediated, at least partially, via activation of the cAMP/PKA-dependent ERK1/2 signalling pathway [40].

In humans, the antidiabetic effect of the isoflavones is still poorly known. However, data from a human study showed that genistein administration at 54 mg/day in postmenopausal women decreased fasting glucose and increased glucose tolerance and insulin sensitivity [41].

Previous studies have shown that anthocyanins and anthocyanidins stimulate insulin secretion and have protective effects on  $\beta$ -cells in vitro (Table 1) by stimulating important factors for insulin gene transcription and reducing reactive oxygen species (ROS)-mediated apoptosis and necrosis [42]. Intake of anthocyanins in T2DM mouse models was found to inhibit elevation of blood glucose levels and improve insulin sensitivity. The antidiabetic effects of anthocyanins were likely mediated by upregulation of *solute carrier family 2 members 4 (Slc2a4)* (gene encoding for GLUT 4 transporter), down-regulation of retinol-binding protein and the related inflammatory adipocytokines [43]. Similar results were obtained when T2DM mice received a dietary bilberry extract rich in anthocyanins. These mice displayed AMPK activation and *Slc2a4* in white adipose tissue while in liver glucose production and lipid content were suppressed. Inactivation of acetyl-CoA carboxylase and upregulation of PPAR- $\alpha$ , acyl-CoA oxidase, and carnitine palmitoyltransferase-1A were also observed in the liver of T2DM mice fed with anthocyanins [44]. In humans, consumption of anthocyanin-rich foods, particularly blueberries and apples or pears, was associated with a lower risk of T2DM, without a significant association between total flavonoid intake or other types of flavonoids [18].

**Table 1.** Flavonoids-gene interactions in the development of type 2 diabetes (T2DM).

Flavonoids	Genes and Gene Products Affected	Function	Experimental Model	Reference
<b>Flavan-3-ols</b>				
catechins	$\uparrow$ <i>PI3K</i> , $\uparrow$ eNOS signalling system	$\downarrow$ Hyperglycemia	STZ-diabetic rats	[20]
epigallocatechingallate (EGCG)	$\uparrow$ <i>Irs2</i> , $\uparrow$ <i>Akt</i> , $\uparrow$ <i>Foxo1</i> , $\uparrow$ <i>Pdx1</i>	$\uparrow$ Viability of $\beta$ -cell, $\uparrow$ Insulin secretion	RIN-m5F cells	[21]
	$\uparrow$ <i>AMPK</i> $\uparrow$ <i>PI3K/Akt</i> pathway	$\downarrow$ IR	dexamethasone-induced rat L6 cells	[22]
	$\uparrow$ <i>PI3K</i> phosphorylation	$\uparrow$ Glucose uptake	differentiated rat L6 myotubes	[23]
	$\uparrow$ <i>Bcl-2</i>	$\downarrow$ Apoptosis, $\uparrow$ Glucose uptake	RINm5F cells	[24]
	$\downarrow$ <i>L-cpt-1</i> , $\downarrow$ <i>Ddit3</i> , $\downarrow$ <i>Ppp1r15a</i> , $\downarrow$ <i>Cdkn1a</i>	$\uparrow$ Insulin secretion, preserve islet structure	db/db mice	[25]
	$\uparrow$ Glucokinase, $\uparrow$ Acyl CoA oxidase-1 (ACO-1) $\uparrow$ Carnitine palmitoyl transferase-1 (CPT-1) $\downarrow$ Phosphofruktokinase $\downarrow$ <i>Glut 1</i>	$\uparrow$ Insulin secretion $\downarrow$ Gluconeogenesis $\uparrow$ Glycolysis	db/db mice and ZDF rats	[26]
<b>Flavanones</b>				
naringin or hesperidin	$\uparrow$ <i>Gk</i> (liver), $\uparrow$ <i>Glut 4</i> , $\uparrow$ PPAR- $\gamma$	$\downarrow$ Hyperglycemia	C57BL/KsJ-db/db mice	[30]
naringin	$\uparrow$ PPAR- $\gamma$ , $\uparrow$ <i>Hsp</i>	$\downarrow$ Hyperglycemia, $\downarrow$ Hyperinsulinemia, $\downarrow$ IR, $\uparrow$ $\beta$ -cell function	HFD-STZ-induced T2D rats	[31]
	$\downarrow$ PEPCK $\downarrow$ G6Pase	$\downarrow$ Hyperglycemia, $\downarrow$ Hyperinsulinemia, $\downarrow$ IR, $\uparrow$ $\beta$ -cell function	HFD-STZ-induced T2D rats	[30]

Table 1. Cont.

Flavonoids	Genes and Gene Products Affected	Function	Experimental Model	Reference
<b>Flavonols</b>				
quercetin	↑ Antioxidant enzyme activity ↓ Nitric oxide (NO) production	↓ Lipid peroxidation, ↑ $\beta$ -cell preservation	STZ-induced DM mice	[32]
	↑ <i>Nrf-2/HO-1</i> activation ↓ NF- $\kappa$ B	↓ High glucose induced apoptosis	Dorsal root ganglion neurons of rats	[33]
	↓ <i>Cdkn1a</i>	↓ Hyperglycemia, ↑ Insulin plasma levels ↑ Pancreatic cell proliferation	STZ-induced DM mice	[35]
	↓ interferon- $\gamma$ , ↓ IL-1 $\alpha$ , ↓ IL-4	↓ Circulating markers of inflammation	C57Bl/6j mice	[36]
	↑ adiponectin, ↓ NOx levels in plasma, ↓ TNF- $\alpha$	↓ Dyslipidemia ↓ Hypertension ↓ Hyperinsulinemia	obese Zucker rats	[37]
<b>Flavones</b>				
luteolin, apigenin	↓ iNos ↓ NF- $\kappa$ B	↓ Apoptosis ↓ IL-1 $\beta$ - and IFN- $\gamma$ -mediated inhibition of insulin secretion	RINm5F cells	[38]
<b>Isoflavones</b>				
daidzein and genistein	↓ PPAR- $\gamma$ ↓ <i>Glut-2</i> ↓ SREBP-1	↑ Insulin secretion ↓ Hyperinsulinemia	rats and pancreatic islets of rats in a HFD	[39]
genistein	↑ cAMP/PKA-dependent ERK1/2 signalling pathway	↓ Hyperglycemia ↑ Glucose tolerance, ↑ Insulin plasma levels ↑ $\beta$ -cell preservation	STZ-induced DM mice	[40]
<b>Anthocyanins</b>				
	↑ heme oxygenase-1 (HO-1) gene ↑ ERK1/2 and PI3K/Akt signalling	↑ Insulin secretion ↓ Apoptosis	pancreatic $\beta$ INS-1 cells and primary islets	[42]
	↑ <i>Glut 4</i> ↓ retinol binding protein 4 (RBP4)	↓ Hyperglycemia ↑ Insulin sensitivity	T2D KK-A(y) mice	[43]
	↑ <i>Glut 4</i> ↑ AMPK, ↑ PPAR- $\alpha$ , ↓ Acetyl-CoA carboxylase ↑ Acyl-CoA oxidase (ACO), ↑ Carnitine palmitoyltransferase-1A-(CPT-1)	↓ Hyperglycemia ↑ Insulin sensitivity	T2D KK-A(y) mice	[44]

### 2.1.2. Phenolic Acid-Gene Interactions in T2DM Pathogenesis

The most common phenolic acids are caffeic acid, chlorogenic acid (the major phenolic compound present in coffee) and ferulic acid (which is esterified to hemicelluloses in cereals) [14].

Most of the epidemiological studies showed an inverse association of caffeinated, decaffeinated coffee and caffeine intake with T2DM in a dose-response manner (1–6 cups/day), compared with no or rare coffee consumption, supporting the hypothesis that habitual coffee consumption is associated with a substantially lower risk of T2DM [45,46]. Regarding tea intake, several meta-analyses also reported potential beneficial effects, although it is not clear whether they are in a dose-response manner [47–50]. These beneficial effects on T2DM include an association with decreased IR [50,51], but not with increased insulin secretion and postprandial effects rather than in fasting glucose metabolism [52,53].

The protective effects of coffee consumption on T2DM involve multiple mechanisms. Recent studies have suggested that chlorogenic acid produced from caffeic acid and quinic acid reduces the ability to inhibit  $\alpha$ -amylase and  $\alpha$ -glucosidase activities, which are key enzymes linked to T2DM [54]. In mouse preadipocyte 3T3-L1 cells, coffee reduced the accumulation of lipids during

differentiation and inhibited the expression of differentiation marker genes such as *PPAR- $\gamma$* , *adiponectin*, *CCAAT-enhancer-binding protein alpha (C/EBP $\alpha$ )*, *Glut 4*, and *lipoprotein lipase (LPL)* [55]. In db/db mice, chlorogenic acid has been suggested to lower the levels of fasting plasma glucose and HbA1c during late diabetes through an adiponectin receptor signalling pathway, elevating the adiponectin level in visceral fat and the adiponectin receptors in the liver and muscles. In addition, chlorogenic acid would inhibit gluconeogenesis by down-regulating hepatic glucose-6-phosphatase (G6Pase) activity, increasing glucose transport in skeletal muscles via up-regulation of AMPK and improving disordered lipid metabolism via up-regulation of hepatic PPAR- $\alpha$  [56].

Regarding ferulic acid, previous studies have reported an important antioxidant activity and hypoglycemic effect in different T2DM diabetic mice models. In high fat and fructose-induced Type 2 diabetic rats, ferulic acid restored normal glucose homeostasis, improving insulin sensitivity and hepatic glycogenesis and inhibiting gluconeogenesis and the expression of insulin signalling inhibitors, such as the gluconeogenic enzyme genes PEPCCK and G6Pase [57]. Besides restoring blood glucose, serum insulin, glucose tolerance and insulin tolerance to a normal range, it also reduces the over-expression of hepatic *solute carrier family 2 (facilitated glucose transporter), member 2 Slc2a2* (which encodes for glucose transporter GLUT 2) by impairing the binding of SREBP1c, HNF1 $\alpha$  and HNF3 $\beta$  transcription factors with the *Slc2a2* promoter [58].

### 2.1.3. Other Bioactive Compound-Gene Interactions in T2DM Pathogenesis (Polyphenols)

Numerous studies support the effect of resveratrol (trans-3,5,4'-trihydroxystilbene), found in grapes, peanuts, blueberries and red wine, in reducing diabetic complications in many organs and tissues including liver and pancreatic  $\beta$ -cells and in different animal models of diabetes [59]. Other effects include the improvement of glucose homeostasis, a decrease in IR, the protection of pancreatic  $\beta$ -cells, an improvement in insulin secretion and an amelioration of metabolic disorders [60]. Resveratrol may act as an anti-inflammatory molecule in diabetes and other chronic diseases associated with chronic activation of NF- $\kappa$ B, blocking the NF- $\kappa$ B-dependent expression of the inflammatory cytokines IL-6, IL-8, and MCP-1 [61]. In addition, resveratrol may prevent diabetes by augmenting brain-derived neurotrophic factor (BDNF), when levels are low in T2DM, and may enhance production of the anti-inflammatory lipid, lipoxin A4 [62]. Another potential mechanism may be by inducing resveratrol-mediated changes in the gut microbiome, as recent studies in obese mice suggested, showing that faecal transplantation from healthy resveratrol-fed donor improved glucose homeostasis [63].

Controversy arises when human clinical studies are considered. Although some studies have not found clear effects [64,65], many others have described the beneficial actions of resveratrol. In obese insulin-resistant adults after four weeks of resveratrol treatment, it has been observed that postprandial glucose decreased and glucose tolerance was not dose-dependent [66]. In Type 2 diabetic subjects, resveratrol supplementation significantly improved fasting blood glucose, HbA1c, total cholesterol, triglyceride, and low density lipoprotein concentrations. The intake of resveratrol also decreased IR and oxidative stress and improved insulin signalling via the AKT pathway in Type 2 diabetic subjects [67,68].

Curcumin in addition to resveratrol has recently been suggested as a potential bioactive product in the field of diabetic nutrition. Treatment with either curcumin (1–100 pM) or resveratrol (0.1–10  $\mu$ M) was reported to enhance pancreatic  $\beta$ -cell function, promoting insulin secretion in mouse  $\beta$ -cell lines as well as in human islets. In both humans and mice, curcumin might be acting as an inhibitor of phosphodiesterases, which degrades cAMP and cGMP, and therefore improves islet insulin secretion [69]. A 9-month study of curcumin treatment in a pre-diabetic population appeared to improve overall function of  $\beta$ -cells, indicated by an increased HOMA- $\beta$  and reduced C-peptide, and significantly prevented T2DM development in the pre-diabetic population. In addition, curcumin intervention significantly increased adiponectin levels, which is inversely correlated to the T2DM risk [70]. Previous studies have also shown an anti-inflammation activity of curcumin by downregulating other inflammatory cytokines, such as tumor necrosis factor-alpha (TNF- $\alpha$ ), leptin and resistin, because the inhibition of the transcription factors NF- $\kappa$ B and Wnt/ $\beta$ -catenin and the

activation of peroxisome proliferator-activated receptor- $\gamma$  and Nrf2 cell-signalling pathways [71]. This regulation would lead to its antidiabetic effect on IR, hyperglycemia, and hyperlipidemia.

Despite the many beneficial actions of curcumin and resveratrol, controversy rises when human clinical studies are considered [64,65]. Poor solubility and absorption of resveratrol when given orally and poor curcumin stability under certain cell culture conditions emphasize the need to optimise the experimental set-up.

## 2.2. Vitamin-Gene Interactions in T2DM Pathogenesis

Vitamin D deficiency affects insulin secretion, IR, and  $\beta$ -cell dysfunction since it modifies the expression of numerous genes involved in immune responses, chemotaxis, cell death and pancreatic  $\beta$ -cell function/phenotype [72,73]. Administration of vitamin D improves impaired glucose tolerance and insulin secretion in vitamin D deficient rats by modulating the generation and effects of cytokines [74]. In addition, vitamin D enhances insulin sensitivity by stimulating the expression of insulin receptors [75] and by promoting the expression of PPAR- $\delta$  [76] (Table 2). Furthermore, vitamin D decreases the effects of systemic inflammation in T2DM and protects against  $\beta$ -cell cytokine-induced apoptosis by directly modulating the expression and activity of cytokines affecting IR, such as IL-1, IL-6, and TNF- $\alpha$  in monocytes from T2DM patients [77]. Some studies support the indirect effects of vitamin D on  $\beta$ -cell function via its well-recognised role in regulating extracellular calcium and calcium influx [78].

In humans, lower vitamin D levels represent a risk factor for incident T2DM [79] and hypovitamin D levels with increased parathyroid hormone (PTH) levels seems to be an independent predictor of  $\beta$ -cell dysfunction, IR and glycemia [80]. Patients with T2DM with established hypovitaminosis improve glycaemia and insulin secretion by Vitamin D replenishment, not only through a direct action on pancreatic  $\beta$ -cell function but also via regulation of plasma calcium levels, which regulate insulin synthesis and secretion [81,82]. Currently, some clinical trials have suggested the combined effects of vitamin D and calcium supplementation for an improvement in the glycemic status of vitamin D insufficient diabetic patients [83]. However, a study selection trial that compared vitamin D3 supplementation with placebo or a non-vitamin D supplement in adults with normal glucose tolerance, prediabetes, or T2DM showed no effect of vitamin D3 supplementation on glucose homeostasis or diabetes prevention. The moderate degree of heterogeneity among subjects and the short-term follow-up duration of this trial could make the potential effects of vitamin D3 supplementation in diabetic patients inconclusive [84,85].

Vitamin A is essential for pancreas development, islet formation and function [86]. However, there is controversy about the effectiveness of vitamin A, retinol and its carrier proteins, retinol binding protein (RBP) and transthyretin for treating diabetes since the effects on insulin secretion seem to be dependent on its metabolites. Retinoic acid is a potent inducer of pancreatic differentiation [87]. Vitamin A (all-trans-retinol) is required for both maintenance of pancreatic  $\beta$ -cell and  $\alpha$ -cell mass and for glucose-stimulated insulin secretion in adult mice [88], although studies on the use of supplements in T2DM humans have shown inconclusive results [89]. RBP might affect glucose homeostasis through the activity of GLUT. This is supported by the observation that, enhanced serum RBP4 levels appear to be a trigger for the development of systemic IR both in animal models and in humans [90,91]. Moreover, the antioxidant properties of carotenoids (pro-vitamin A compounds) may also protect against T2DM [92], although previous studies found little evidence for an association between dietary intake of lycopene, a powerful antioxidant carotenoid compound, and the risk of T2DM in women [93].

Regarding the antidiabetic effects of Vitamin E and tocopherols, although results from animal studies suggested an association between Vitamin E consumption and a lower risk of T2DM [94], several randomised interventional clinical trials not only failed to confirm that there was a beneficial effect in preventing or treating T2DM but suggested an aggravation of some diabetes affections [95–97].

Biotin is also required for the normal expression of critical carbohydrate metabolism genes and for glucose homeostasis, such as *forkhead box A2 (Foxa2)*, the *pancreatic and duodenal homeobox factor transcription factor (Pdx1)*, *hepatocyte nuclear factor 4alpha (Hnf4 $\alpha$ )*, *insulin (Ins)*, *glucokinase (Gck)*, *Calcium*

Voltage-Gated Channel Subunit Alpha1 D (*Cacna1d*) and acetyl-CoA carboxylase [98] (Table 2). In the Type 2 diabetic rat model induced by HFD and low-dose of STZ, biotin supplementation exerted antioxidant, anti-hyperlipidaemic, anti-inflammatory and anti-hyperglycaemic effects, increasing the level of insulin, probably through modulation of PPAR- $\gamma$ , IRS-1 and NF- $\kappa$ B proteins [99].

The role of ascorbic acid in diabetes treatment remains unclear since it can also be a pro-oxidant under certain in vitro conditions and it can glycate proteins despite its beneficial preventive antioxidant properties. In vitro studies suggested that appropriate concentrations of vitamin C in pancreatic  $\beta$ -cells may have beneficial effects since the number of  $\alpha$ - and  $\beta$ -cells increased after vitamin C supplementation [100]. Different studies have found lower levels of ascorbic acid in the plasma of diabetic patients and experimental diabetic animal models [101,102] and a strong inverse association between plasma vitamin C levels and T2DM risk [103]. Administration of a high dose of vitamin C inhibited insulin secretion in rat islets in a dose-dependent manner [104].

Clinical trials of vitamin C supplementation in diabetic populations have shown controversial results. In some Type 2 diabetic patient studies, administration of vitamin C increased plasma vitamin C levels and whole body glucose disposal thus improving both fasting blood glucose and HbA1c levels [105,106]. However, in others, administration of oral vitamin C was unable to completely replenish the plasma level of vitamin C and did not improve endothelial dysfunction or IR [107] or even suggested that supplements of vitamin C may actually increase the risk of cardiovascular disease mortality [108].

Despite their antioxidant properties and effects in the reduction of diabetic complications in diabetic mice models [109], supplementation of B vitamins in general has not been proposed for the primary prevention of T2DM. In fact, recent studies have suggested that long-term exposure to high levels of the B vitamins niacin, thiamin and riboflavin may correlate with prevalence of obesity and diabetes. In fact, niacin may induce glucose intolerance, IR and liver injury [110]. Furthermore, it has been observed that riboflavin in non-toxic doses prevents cytokines-induced p38 phosphorylation and IL-6 upregulation on insulinoma NIT-1 cells and isolated rodent islets [111]. Nicotinamide, known to promote differentiation of mouse embryonic stem cells into insulin producing cells [112–114], also induced insulin gene expression by increasing *v-maf avian musculoaponeurotic fibrosarcoma oncogene homolog A (MafA)* expression in INS1-1  $\beta$ -cells [115] (Table 2).

**Table 2.** Vitamin-gene interactions in the development of T2DM.

Vitamins	Genes and Gene Products Affected	Function	Experimental Model	References
<b>vitamin D</b>				
	↑ Human insulin receptor gene	↑ Insulin sensitivity	U-937 human promonocytic cells	[75]
	↓ TNF- $\alpha$ , IL-6, IL-1, and IL-8	↓ $\beta$ -cell cytokine-induced apoptosis	CD14 <sup>+</sup> monocytes isolated from peripheral blood insulin-treated T2DM patients	[77]
<b>Biotin</b>				
	↑ <i>Foxa2</i> , ↑ <i>Pdx-1</i> ↑ <i>Hnf-4<math>\alpha</math></i> ↑ Ins, ↑ Gk, ↑ <i>Cacna1d</i> , ↑ <i>Acac</i>	↑ Insulin secretion ↑ Islet function	BALB/cAnN Hsd mice	[98]
	↑ PPAR- $\gamma$ ↑ IRS-1 ↓ NF- $\kappa$ B	↑ Antioxidant ↓ Hyperlipidaemia ↓ Inflammation ↓ Hyperglycaemia	T2DM rat model induced by high-fat diet (HFD) and low-dose STZ	[99]
<b>Riboflavin</b>				
	↓ IL-6 upregulation	↓ Cytokines-induced p38 phosphorylation	Insulinoma NIT-1 cells and isolated rodent islets	[111]
<b>Nicotinamide</b>				
	↑ <i>MafA</i>	↑ Insulin synthesis	INS-1 cells, pancreatic islets	[115]
	↑ <i>Pdx-1</i>	Pancreatic $\beta$ -cell differentiation	Mouse embryonic stem cells	[112–114]

### 2.3. Amino Acid-Gene Interactions in T2DM Pathogenesis

Dietary protein and amino acids also modulate insulin secretion and/or contribute to the maintenance of  $\beta$ -cell function [116,117]. The mechanisms function by influencing gene and protein expression in pancreatic islets and activating the PI3K/PKB/mTOR pathway, which results in improved insulin release [118,119]. In this respect, it has been suggested that dietary components of germinated brown rice improves glycemic control by downregulating the gluconeogenic genes *Fbp1* and *Pck1* in Type 2 diabetic rats [120]. In general, the mechanisms proposed for the amino acid effects on glucose homeostasis and blood glucose control included a direct insulinotropic effect on insulin secretion from the  $\beta$ -cell, the role of precursors for liver glucose synthesis, and a potential effect on insulin signalling [121]. Recent studies have suggested that supplementation with casein hydrolysate would attenuate the NLRP3-ASC inflammasome activity, a molecular platform mediating the processing of IL-1 $\beta$  in response to infection and stress conditions, improving insulin sensitivity and glucose tolerance in vitro and in mice fed with HFD [122].

Overall, the role of dietary protein on glucose control through endogenous production of glucose varies depending on the experimental conditions, the protein and carbohydrate content of the meal and the window of metabolic investigation after the meal. In T2DM patients, protein ingestion augmented postprandial insulin release and led to the attenuation of postprandial elevation in circulating glucose concentration [123,124]. Regarding the effects of animal protein intake on T2DM risk, several prospective cohort studies have found associations between animal protein intake and T2DM risk highly dependent on the source and type of animal protein and presence of others food components. Epidemiological studies indicate that higher intake of low-fat and fat-free fermented dairy foods are associated with lower T2DM risk [125–127] but more epidemiological evidence is necessary to elucidate the real association among intake of protein supplements and T2DM risk.

Amino acids are potentiators of glucose-induced insulin release [117] promoting slow cytosolic calcium oscillations in mouse pancreatic islets [116]. Four amino acids, namely arginine, leucine, isoleucine and valine are largely responsible for the observed effects. Branched-chain amino acids (BCAAs) isoleucine, leucine and valine are important as nutrient signals and metabolic regulators [128]. Epidemiological studies have shown that BCAAs plasma concentrations and metabolism are altered in T2DM. In obese and T2DM animal models, low circulating levels of adiponectin impair the BCAAs catabolism, which leads to accumulation of these amino acids through a down-regulation of mitochondrial phosphatase 2C (PP2Cm) expression mediated by the AMPK signalling pathway [129].

Increased plasma BCAAs have been associated with IR and levels of HbA1c. Accumulation of BCAAs has been suggested to promote mitochondrial dysfunction, linked to stress kinase stimulation and  $\beta$ -cell apoptosis which are frequently related to IR and T2DM. Furthermore, elevated circulating BCAAs levels have been found to be a reliable prediction of T2DM especially in Caucasians and Hispanics and not only in presymptomatic individuals, but also in those classified as prediabetic or diagnosed as Type 2 diabetic [130–132]. Despite the numerous studies, it is not clear yet whether increased BCAAs levels are simply markers of IR or whether they are direct contributors to IR and loss of action [133–136].

Among BCAAs, L-leucine is unique in inhibiting NO synthesis from L-arginine in endothelial cells, activating glutamine: fructose-6-phosphate aminotransferase (GFAT) [137] and may modulate cardiovascular homeostasis in IR activating the mechanistic target of rapamycin (serine/threonine kinase) (mTOR) complex, a key regulator in cell growth and proliferation [138–140]. Leucine also appeared to enhance pancreatic  $\beta$ -cell insulin secretion by its ability to allosterically activate glutamate dehydrogenase, which has also been reported for other amino acids, such as phenylalanine [141].

Despite numerous studies having suggested a causal role of high levels of BCAAs in impairing insulin signalling or IR [142,143], other observations indicate that BCAAs activation of the mammalian target of rapamycin complex 1 (mTORC1), which generates an inhibitory feedback loop on insulin receptor substrate proteins, is not necessary or sufficient to trigger IR [144]. Recent work using untargeted metabolomics in a T2DM rat model consistently found that BCAA levels were not elevated

until six months post-onset of diabetes, supporting the idea that the increase in BCAA level is not enough to elicit IR and T2DM in that model [145].

Elevated circulating levels of the aromatic amino acids phenylalanine and tyrosine have also been linked to the development of T2DM, acting as indirect markers of insulin sensitivity, not only in T2DM patients but also in presymptomatic individuals [130,131,146–148]. On the other hand, in those clinical trials, glycine was found to be reduced, possibly due to an increase in the gluconeogenesis or glutathione consumption by increased oxidative stress [130,146,147,149,150].

The insulinotropic effect of arginine in the presence of glucose could be explained by directly depolarising the plasma membrane of  $\beta$ -cells, activating the  $\text{Ca}^{2+}$  channels resulting in the influx of  $\text{Ca}^{2+}$  which triggers insulin exocytosis [116,151].

Arginyl-fructose, which is widely distributed in red ginseng, has antidiabetic effects attributed to it by suppressing carbohydrate absorption in the gastrointestinal tract [152]. Arginyl-fructose has potential as a pharmacological agent for glycemic control since it significantly reduces postprandial blood glucose levels in rodents and humans. In contrast, glucose-related biomarkers including HbA1c, insulin, and C-peptide levels were not significantly improved by dietary supplementation with arginyl-fructose [153,154].

#### 2.4. Dietary Fat-Gene Interactions and Their Role in T2DM

Although numerous qualitative analyses on the intake of different types of fat have found inconsistent associations between intake and T2DM risk [155,156], consumption of a high fat diet (HFD), mainly in saturated fatty acids (SFAs), is believed to be associated with an increased risk of T2DM. In addition, a correlation between a higher intake of SFAs and decreased insulin sensitivity has been demonstrated in several cross-sectional studies [125,157–159]. In animal models, an increase in plasma fatty acid concentration is associated with an increase in intracellular fatty acyl-CoA and diacylglycerol concentrations. As a consequence, insulin activation of IRS-1 and IRS-1-associated PI3-kinase activity is inhibited, resulting in a decrease in insulin-stimulated glucose transport activity [160]. In addition, T2DM C57BL/6J mice fed with HFD upregulated proteins involved in pancreatic  $\beta$ -cell proliferation and downregulation of glutathione peroxidase gene (*Gpx1*) implicated in the antioxidant defenses of  $\beta$ -cells and in the regulation of *MafA* expression, which is important in the regulation of insulin expression [161]. Reports supporting the potential unhealthy effects of palm oil due to the high palmitic acid content in the development of T2DM have suggested that chronic exposure of pancreatic islets to palm oil inhibited glucose-induced expression of prepro-insulin, as well as *Pdx1* and *MafA* expression, causing  $\beta$ -cell failure [162,163]. The mechanism of downregulation of insulin gene expression by palmitate involves the inhibition of PDX1 nuclear translocation, and therefore blocking *MafA* gene expression [164]. Besides preproinsulin mRNA expression, palmitate also reduces *Slc2a2* and *Gck* probably through the inhibition of *Pdx1* mRNA [165]. Elevated levels of fatty acids lipotoxicity have been described to be mediated by de novo synthesis of ceramide which can be formed from palmitate but not from oleate, explaining the fact that both palmitate and oleate inhibit insulin secretion but only palmitate impairs insulin gene expression [166,167]. In addition, saturated fatty acid palmitate but not unsaturated oleate, induces the activation of the NLRP3-ASC inflammasome causing caspase-1, IL-1 $\beta$  and IL-18 production leading to reduced glucose tolerance and insulin sensitivity [168]. On the other hand, monounsaturated fatty acid-enriched HFD improves insulin sensitivity in mice, attenuates adipose IL-1 $\beta$  secretion and maintains adipose AMPK activation when compared to SFAs-HFD-fed mice [169]. Moreover, we recently reported that an extra virgin olive oil intervention ameliorate non-alcoholic steatohepatosis was induced by a high fat “Western-style” diet in mice [170]. Overall, mice and in vitro experiments suggest that high levels of palmitate cause an alteration to genes involved in lipid metabolism, inflammation and oxidative stress [171]. The possible mechanism of SFAs to induce inflammation and IR in adipocytes, which would imply involvement of oxidative stress through ROS in a toll-like receptor-4 and 2 (Tlr4, Tlr2), mediated inflammatory signalling.

Furthermore, activation of c-jun N-terminal kinase (JNK) also appeared to be essential to Tlr2-, as well as Tlr4-induced IR and oxidative stress [172–174].

Intervention studies in humans where SFAs intake was reduced via replacement with unsaturated fatty acids showed contradictory data that was observed in animal models: whilst some favored both liver and visceral fat accumulation with a detrimental role in T2DM development and progression [175], others showed improved insulin sensitivity but no effect on insulin secretion [176] whilst others showed no significant effects [177]. Recently, the EPIC-InterAct case-cohort study in people with incident T2DM reported that different SFAs have different effects on T2DM incidence [178]. Specifically, myristic, palmitic, and stearic acids were all positively associated with incident T2DM while pentadecanoic acid, heptadecanoic acid and arachidic acid were inversely associated with incident T2DM. Lauric, myristic, and palmitic acids can raise total and LDL cholesterol, increase coagulation, induce IR, and promote inflammation.

Regarding monounsaturated fatty acids (MUFAs), oleic acid has been suggested to improve glucose control and insulin sensitivity [179]. The effects observed when replacing saturated fatty acids with oleic acid may be due to partial removal of the SFAs biological effect, any constituent of the diet, or, to the combination of foods/nutrients rather than to oleic acid. However, some studies indicate oleic acid has some effects on transcription factors involved in lipid homeostasis, such as SREBPs [180]. Similarly, palmitoleic acid also increases the transcription activity of SREBP1c, apart from enhancing the binding of SREBP1c to FAS promoter and decreasing the phosphorylation of NF- $\kappa$ B, p65 and the expression of proinflammatory cytokines [181]. In vitro approaches indicated that *cis*-palmitoleic acid can influence pancreatic  $\beta$ -cell survival, insulin secretion, skeletal muscle insulin response and adipocyte metabolism. On the other hand, prospective studies have shown a positive association between higher circulating palmitoleic acid and improved insulin sensitivity or lower incident type T2DM [182,183].

T2DM is associated with increased blood concentrations of inflammatory biomarkers, including C-reactive protein (CRP), which leads to a low-grade inflammation that is the mechanism underlying IR. Epidemiological, human intervention, animal and cell culture studies have supported a beneficial role for dietary *n*-3 polyunsaturated fatty acids (PUFAs) in low-grade chronic inflammation situations, among others, by reducing CRP [184,185]. In overweight/obese patients with impaired fasting glucose or impaired glucose tolerance, *n*-3 PUFAs reduced glycemia and fasting plasma insulin [186], while in T2DM patients with omega-3 supplementation, a high ratio of eicosapentaenoic acid/docosahexaenoic acid (EPA/DHA) contributed to a greater decreasing tendency in plasma insulin and HbA<sub>1c</sub>, although with no statistically significant results [187]. The anti-inflammatory and preventive inflammation-driven disease effect of *n*-3 PUFAs is related to NLRP3 inflammasome activation and subsequent inhibition of caspase-1 activation and IL-1 $\beta$  secretion [188]. In addition, EPA has been suggested to repress Srebp-1c, which may lead to a lower lipid accumulation in pancreatic cells and an enhancement of insulin secretory mechanism [189,190].

Linoleic acid is the most prevalent omega-6 PUFAs in the human diet, found in some studies to increase insulin sensitivity [191]. Although it has been shown to activate NF- $\kappa$ B [192], dietary intakes were not associated with CRP, IL-6, soluble TNF receptor (sTNFR) 1, or sTNFR2 concentrations in humans [193], therefore having only a limited effect on inflammation [194].

### 3. Diet-Gene Interaction and Risk of T2DM

For a decade, genome-wide association studies (GWAS) on T2DM have been conducted in a variety of populations of different ancestries and more than 200 T2DM-related genetic variants have been identified by GWAS so far [5,195]. Not all, but many of the gene variants found are located in genes related to the insulin secretion pathway, insulin signalling, pancreatic  $\beta$ -cell dysfunction, and IR pathway [196]. However, gene variants have been described in genes that apparently would not be candidates for greater susceptibility to T2DM. In addition to the genetic architecture of T2DM susceptibility, environmental factors are suggested to play a key role in the etiology of T2DM. Dietary factors in particular may interact with genetic variants to modulate the risk of T2DM [197].

All this makes it very complex to establish T2DM risk values according to allelic variants, due to: (i) the existence of genetic variants that increase T2DM risk independently of the type of diet (ii) genetic variants related to diet that modify some glucose metabolism systems, such as fasting glucose levels and IR, but do not modify the risk of T2DM (iii) genetic variants that present a greater risk of T2DM but this risk is modified depending on the type of diet (iv) genetic polymorphisms that modify risk according to other parameters such as ethnic class and obesity. All this makes it somewhat difficult to clearly establish cause-effect and therefore, the association between a gene variant and the risk of developing T2DM.

### 3.1. Gene Variants Associated with Insulin Stimulus-Secretion Coupling and T2DM

Among the gene variants strongly associated with the risk of developing T2DM in various human populations are several genes involved in the WNT-mediated signalling pathway, which induces the expression of genes involved in pancreas development and in glucose homeostasis, such as incretin-expressing genes that potentiate insulin secretion. One of the most important is the *TCF7L2* gene encoding the transcription factor TCF7-like 2 located on chromosome 10 [198]. In several studies in white European [199], Indian [200], Japanese [201], Mexican American [202], Chinese [203] and African populations [204], *TCF7L2* showed a strong association with the odds of developing T2DM being increased by 30–50%. Several genetic polymorphisms have been described in the *TCF7L2* gene associated with an increased risk of T2DM (rs12255372 G<T, rs7903146 C>T), which interact with the diet modifying susceptibility to T2DM. In this sense, it is important to differentiate between the improvement in parameters related to glucose metabolism and the risk of T2DM. The Malmö Diet and Cancer Study (MDCS) cohort conducted by Hindy et al. 2016 showed that high fibre intake improved fasting plasma glucose and insulin levels, an improvement in IR values (HOMA-IR) and lower levels of glycosylated haemoglobin. However, the T2DM risk in individuals carrying the risk allele *TCF7L2* rs12255372 or *TCF7L2* rs7903146 was higher when fibre consumption was high. In contrast, individuals who did not carry the risk allele showed a decrease in T2DM risk with fibre intake [205].

No interaction was observed between polymorphisms and consumption of carbohydrates, fats or proteins and the incidence of T2DM [206] in MDCS cohort.

On the other hand, an EPIC sub cohort study (EPIC-InterAct study) was carried out and an interaction was noted between *TCF7L2* variant rs12255372 and coffee intake, with an inverse association between coffee consumption and T2DM among carriers of the diabetes risk allele (T) [207].

In MDCS cohort other allelic variants of genes encoding transcription factors involved in the WNT signalling pathway with greater susceptibility to T2DM that is modified by the diet are *ZBED3* rs4457053 G>A and *NOTCH2* rs10923931 G>T in which fibre consumption protects against the susceptibility to T2DM with higher fibre consumption in the risk allele carriers (T) for the case of *NOTCH2* and in individuals homozygous for the risk factor (GG) in the case of *ZBED3* [205].

The protein encoded by the gene solute carrier family 30 (zinc transporter) member 8 (*SLC30A8*) is a zinc efflux transporter protein necessary for the accumulation of zinc in intracellular vesicles. The expression of this gene is very high in pancreas, especially in pancreatic islets. Zinc is required for insulin biosynthesis and the maturation of insulin secretory granules [208]. Genetic polymorphisms have been described in this gene that increase T2DM risk. Thus, the polymorphism rs13266634 C>T was associated with lowered  $\beta$ -cell function and a 14% increase in diabetes abundance per risk (C) allele [209]. A significant interaction between rs13266634 and serum levels of trans- $\beta$ -carotene and  $\gamma$ -tocopherol was identified. Higher levels of trans- $\beta$ -carotene nutrient factor appeared to have a protective effect in individuals with the risk allele. In contrast, high levels of  $\gamma$ -tocopherol produced adverse effects in individuals with the risk allele [210]. Additionally, in a 14-cohort meta-analysis study, an interaction was observed between the allelic variant *SLC30A8* rs11558471 A>G, conferring higher levels of fasting plasma glucose to A allele carriers, and zinc consumption. In a 14-cohort meta-analysis study, total zinc intake had a stronger inverse association with fasting glucose levels in

individuals carrying the glucose-raising A allele of rs11558471 *SLC30A8*, compared with individuals carrying the G allele [211].

There exists a common genetic variant in the glucose-dependent insulinotropic polypeptide receptor gene (*GIPR* rs10423928 T>A) which is related with a reduced insulin release and an increase in T2DM risk. In the Swedish population-based Malmö Diet and Cancer cohort, it has been studied the relationship between the mentioned genetic variant, macronutrients and fiber intake, body mass index (BMI) and T2DM risk. It has been observed that when AA-genotype people follow high-fat, low-carbohydrate diets, the T2DM risk decreases. On the other hand, two thirds of the people homozygous for the T-allele resulted benefitted when they follow high-carbohydrate, low-fat diets [212].

A meta-analysis of 14 cohort studies for the interactions between the glucokinase regulatory protein (*GCKR*) variant rs780094 G>A and whole grain intake demonstrated a possible relationship between the gene variant and whole grain intake. In fact, people with the insulin-raising allele rs780094 that followed a larger whole grain intake had a smaller reduction in fasting insulin compared to subjects without the variant [213].

The *S100A9* gene is located in region 1q21, a high susceptibility region to T2DM, and codes for the calcium binding protein S100A9 (calgranulin B). The variant *S100A9* (rs3014866 C>T) was associated with protection against T2DM development in several populations, such as individuals carrying the less frequent allele (T) who had a lower T2DM risk than those who did not carry the allele (CC homozygous individuals). In the intervention trial study in 3 diverse populations (CORDIOPREV: Coronary diet Intervention with Olive Oil and Cardiovascular prevention; GOLDN: Genetics of Lipids Lowering Drugs and Diet network and BPRHS: Boston Puerto Rican health Study) related to the interaction of polymorphism with diet, individuals with the non-protective variant (CC) had a higher HOMA-IR value than individuals carrying the allele (T) when the association between intake of saturated fatty acids (SFAs) versus carbohydrates was high, and that this difference disappeared when intake of SFAs/carbohydrates was low [214]. This may be one of the reasons why individuals with the CC polymorphism are at increased risk of T2DM and that diet may reduce this risk.

The transient receptor potential vanilloid 1 (*TRPV1*) gene is involved in energy and glucose metabolism. TRPV1 activation increases insulin sensitivity and potentiates glucose-stimulated insulin secretion. A Korean Genome Epidemiology Study demonstrated that individuals with the minor alleles of the *TRPV1* single nucleotide polymorphisms (SNPs) rs161364 T>C and rs8065080 T>C were negatively associated with the prevalence of T2DM. They also determined that carriers of the minor allele of both SNPs have a lower risk of diabetes with a high-fat diet but individuals with the major alleles are at a higher risk of T2DM when consuming high-fat diets [215].

In a small case-control study examining interactions between magnesium intake and loci in *TRPM6* (transient receptor potential cation channel, subfamily M, member 6) associated with T2DM, in older women carried out over 10 years, it reported that women who were carriers of 2 rare alleles from nonsynonymous SNPs in *TRPM6* (rs3750425 and rs2274924) had nearly 5 times the odds of T2DM when their magnesium intake was less than 250 mg/d [216].

The SNP rs3786897 G>A in the peptidase D (*PEPD*) gene has been associated with the risk of T2DM in Asian individuals [217]. PEPD protein plays an important role in the recycling of collagen metabolisms, while it is shown to have a profound impact on  $\beta$ -cell function [218]. In a case-control study examining the genetic effect of the *PEPD* rs3786897 A allele on the risk of T2DM may be abolished when n-3 PUFA intake is high [219] (Table 3).

**Table 3.** List of gene-nutrient interactions in the development of T2DM.

Genes Related with Stimulus-Secretion Coupling					
Gene	SNP	T2D Risk	Types of Studies/Dietary Factors	Effect of Interaction between SNP and Dietary Factors	References
TCF7L2	rs12255372 G>T	Allele-T risk increase	MDCS cohort/fibre intake	Higher risk in individuals carrying the allele-T. Smaller risk in individuals homozygous GG	[205]
			EPIC InterAct study/ Coffee Intake	Smaller risk in individuals carrying the allele-T	[207]
	rs7903146 C>T	Allele-T risk increase	MDCS cohort/fibre intake	Higher risk in individuals carrying the allele-T. Smaller risk Individuals homozygous CC	[205]
ZBED3	rs4457053 G>A	Allele-G risk increase	MDCS cohort/fibre intake	Smaller risk in individuals homozygous for the risk factor (GG)	[205]
NOTCH2	rs10923931 G>T	Allele-T risk increase	MDCS cohort/fibre intake	Smaller risk in individuals carrying the allele-T	[205]
SLC30A8	rs13266634 C>T	Allele-C risk increase	NHANES cohort study/ <i>trans</i> -β-carotene intake	Smaller risk in individuals carrying the allele-C	[209,210]
			NHANES cohort study/ γ-tocopherol intake	Higher risk in individuals carrying the allele-C	[209,210]
	rs11558471 A>G	Allele-A increases levels of fasting plasma glucose	14-Cohort meta-analysis study/ zinc intake	Smaller levels of fasting plasma glucose in individuals carrying the allele-A	[211]
GIPR	rs10423928 T>A	Allele-A risk increase	MDCS cohort/fats and carbohydrates intake	Smaller risk in individuals homozygous for de risk factor AA consuming high-fat, low-carbohydrate diets. High-carbohydrate, low-fat diets benefit of the population homozygous for the T-allele	[212]
GCKR	rs780094 C>T	Allele-T risk increase	14-cohort meta-analysis study/whole-grain intake	Smaller risk in individuals carrying the allele-T	[213]
S100A9	rs3014866 C>T	Allele-T risk decrease	Intervention trial in CORDIOPERV, GOLDN and BPRHS/SFAs:carbohydrates ration	Ration high: Individuals with the non-protective variant (CC) had a higher HOMA-IR. Ration low: Individuals with the non-protective variant (CC) had a normal HOMA-IR.	[214]
TRPV1	rs161364 T>C	Allele-C risk decrease	Cohort study/high fat diet intake	Smaller risk in individuals carrying the allele-C Higher risk in individuals carrying the allele-T	[215]
	rs8065080 T>C				
TRPM6	rs2274924 C>T	Allele-T risk increase	Cases-control study/magnesium intake <250 mg/day	Higher risk in women carrying the allele-T	[216]
	rs3750425 C>T				
PEPD	rs3786897 G>A	Allele-A risk increase	Cases-control study/ High n-3 PUFAs	The risk disappears in individuals carrying the allele-A	[219]
Genes Related with Insulin Signaling					
Gene	SNP	T2D	Dietary Factors	Effect of Interaction between SNP and Dietary Factors	References
PPAR-γ	rs180282 C>G	Allele-G risk increase	DESIR cohort study/high fat intake increases	Higher risk in individuals carrying the allele-T	[220]
ADIPOQ	rs1501299 G>T	Allele-T risk increase	Cohort study/carbohydrate intake	Higher fasting blood glucose and HbA1C concentrations in individuals carrying allele-T	[221]
	rs2241766 T>G	Allele-G risk increase	MARINA trial study/ high n-3 PUFAs	Smaller risk in individuals carrying the allele-G	[222]
IRS1	rs7578326 A>G	Allele-G lower risk of IR and lower fasting insulin	GOLDN and BPRHS cohort studies/low total fat and SFA:carbohydrate ratio intake	Decrease resistance to insulin and plasma insulin	[223]
	rs2943641 C>T	Allele-T lower risk of IR and lower fasting insulin	GOLDN and BPRHS cohort studies/low fat and SFA:carbohydrate ratio intake Cohort study/vitamin D intake	Decrease resistance to insulin and plasma insulin Smaller risk in individuals carrying the allele-T	[223] [224]
CAV2	rs2270188 G>T	Allele-T risk increase	Cases-cohort study/high fat or SFA intake	Higher risk in individuals carrying the allele-T.	[225]
Other Genes					
Gene	SNP	T2D	Dietary Factors	Effect of Interaction between SNP and Dietary Factors	References
FTO	rs9939609 T>A	Allele-A risk increase	Cases-control study/adherence to Mediterranean diet	The risk disappears in individuals carrying the allele-A	[226]
	rs8050136 C>A	Allele-A risk increase	Cases-control study/adherence to Mediterranean diet	The risk disappears in individuals carrying the allele-A	[226]
CLOCK	rs4580704 C>G	Allele-G risk decrease	PREDIMED trial study/high MUFA intake	Lower risk in individuals carrying the allele-G	[227]
CRY1	rs2287161 G>C	Allele-C risk increase	Cohorts Mediterranean and North American study/high carbohydrate intake	Increase in HOMA-IR index, fasting insulin and a decrease in QUICKI	[228]
MC4R	rs17782313 T>C	Allele-C risk increase	Systematic review study/adherence to Mediterranean diet	The risk disappears in individuals carrying the allele-C	[226,229]
PLIN	rs11482 G>A	Allele-A risk increase	cross-sectional study/SFAs:carbohydrates ration	Ration high: women homozygous for AA increased HOMA-IR	[230]
FABP2	rs1799883 G>A	Allele-A risk increase	11-cases-control meta-analysis study/MUFA intake	Decrease in HOMA-IR index	[231]

### 3.2. Gene Variants Related to IR and T2DM

Peroxisome proliferator-activated receptor- $\gamma$  (*PPAR- $\gamma$* ) is a transcription factor that directly regulates target genes, mediating lipid metabolism and adipocyte differentiation. The genetic variant Pro12Ala rs180282 in *PPAR- $\gamma$*  has been associated with the T2DM risk and BMI [232]. The less frequent *PPAR- $\gamma$*  Ala12 variant reduces the risk of T2DM and is positively associated with insulin sensitivity. DESIR (Data from an Epidemiological study on the Insulin Resistance Syndrome) cohort study indicates high fat intake increases T2DM risk in Pro homozygous individuals [220].

Adiponectin (encoded by *ADIPOQ*) is an important adipocytokine that is secreted by adipocytes and plays a key role in the inflammatory response that is associated with insulin-resistant states and T2DM. In recent years, the association between SNPs of the *ADIPOQ* gene and T2DM has been reported in the Asian population in particular [233]. Interactions of variants of the adiponectin gene with carbohydrate intake [29] and fat intake [30] have been explored. Significant dose-response interactions were identified between the *ADIPOQ* rs1501299 G>T polymorphism and the dietary intake of carbohydrates in a prospective study of 673 patients with T2DM. This previous study demonstrated a connection between the gene-nutrient interactions of rs1501299 G>T polymorphism and the level of carbohydrate intake modulating fasting plasma blood glucose and glycosylated hemoglobin [221]. Other gene-nutrient interactions include rs2241766 T>G and a diet high in *n-3* PUFAs. The MARINA trial study demonstrated that individuals who had a high intake of *n-3* PUFAs showed a decreased risk of T2DM [222].

The *IRS1* gene encodes insulin receptor substrate 1 (*IRS1*), a protein central to insulin signalling pathways. Two genetic variants (rs7578326 and rs2943641) near *IRS1* were identified by GWAS to be associated with T2DM [223,234]. *IRS1* variants rs7578326 G allele and rs2943641 T allele were associated with a lower risk of IR and lower fasting insulin in two independent populations of different ancestries (GOLDN cohort study and the BPRHS cohort study). These associations appeared to be modulated by dietary factors, especially the dietary SFAs-to-carbohydrate ratio, MUFAs, and carbohydrate quantity and quality. Thus, low consumption of MUFAs, low total fat consumption and a low SFAs to carbohydrate ratio decrease resistance to insulin and plasma insulin, but not when consumption is high [223]. Another longitudinal cohort study also explored the relationship between *IRS1* rs2943641 and circulating levels of 25-hydroxyvitamin D 25(OH)D. Puerto Rican adult females homozygous for the minor allele rs2943641T with higher circulating 25(OH)D showed a lower risk of IR and T2DM compared to carriers of the major allele (C) [224].

The *CAV2* gene encode the major protein components of caveolae. These are invaginations of the plasma membrane. These proteins are important for several cellular functions as signal transduction, lipid metabolism, cellular proliferation, apoptosis, differentiation and trafficking. The case-control study of the European prospective Investigation into Cancer and Nutrition-Post dam cohort demonstrated a significant relationship between the *CAV2* rs2270188 G>T polymorphism and fat and SFAs intake with respect to T2DM. Homozygous individuals of the rare T-allele showed a 100% greater risk of T2DM when daily fat intake was increased from 30% to 40% in energy value, and an increase in dietary SFAs from 10% to 20% energy value predicted approximately 200% greater risk of T2DM. However, homozygotes for the G allele and heterozygotes do not have increased risk of T2DM with higher fat or SFAs consumption [225] (Table 3).

### 3.3. Other Genetic Variants and T2DM

The fat mass and obesity-associated gene (*FTO*) is located on chromosome 16 and contains nine exons. Actually, it has been described several SNPs of this gene that have been regularly associated with obesity risk. Nevertheless, there isn't a consistent association between obesity risk alleles and T2DM. A meta-analysis of 62 case-control studies from different regions (including Asia, Europe and North America) demonstrated that the rs9939609 T>A and rs8050136 C>A SNPs contributed to an increased risk of T2DM [235]. A case-control study, with 3430 T2DM patients and 3622 healthy people without difference in their BMI, showed steady gene-diet interactions with adherence to the Mediterranean diet for individuals carrying the *FTO*-rs9939609 variant. Moreover, in the presence of a

low Mediterranean diet adherence, people of the variant alleles showed a higher T2DM risk. On the contrary, with a high Mediterranean diet adherence, these associations were not observed [226].

Dysregulation and genetic variations in the Circadian Locomotor Output Cycles Kaput genes (*CLOCK*), which are responsible for the circadian system, have been associated with T2DM [236]. A longitudinal study, undertaken with 7098 participants in the PREDIMED trial analysed the association between the SNP *CLOCK* rs4580704 C>G and the incidence of T2DM. G-allele carriers presented lower T2DM risk than CC homozygous subjects. The protective association of the G-allele against T2DM incidence was stronger and of higher statistical significance in the Mediterranean diet (High MUFAs) intervention group than in the control group (Low MUFAs) [227]. The interaction between the cryptochrome 1 (*CRY1*) rs2287161 G>C variant and several diabetes-related traits (fasting glucose and insulin, HOMA-IR and QUICKI index) has been studied in The Mediterranean and North American Cohort. The authors showed that a significant increase in HOMA-IR index and fasting insulin, as well as a decrease in QUICKI was associated with a higher carbohydrate intake in people homozygous for the minor C allele [228].

The melanocortin-4 receptor (*MC4R*) gene is a receptor for the  $\alpha$ -melanocyte stimulating hormone ( $\alpha$ -MSH) which regulates food intake and energy homeostasis. In a systematic review study a consistent interaction between adherence to a Mediterranean diet and the *MC4R* rs17782313 T>C was demonstrated. When adherence to the Mediterranean diet was low, carriers of the variant allele for *MC4R* rs17782313 (C) had a higher risk of T2DM than homozygous participants for the major allele (T). When adherence to the Mediterranean diet was high, however, there was no association between this polymorphism and T2DM [226,229].

The *Perilipin* gene (*PLIN*) regulates adipose metabolism and has been associated with several risk factors for diabetes, including obesity, weight gain and IR, in both American [237] and Chinese women [238]. Although an interaction between the allelic variants and an increased risk of T2DM has not been found, the analysis of the interaction between these variants and diet showed a significant gene-diet interaction between the *PLIN* 11482 G>A and the *PLIN* 14995 A>T polymorphisms with dietary fat and carbohydrate intake. A cross-sectional study of Asian women with these SNPs showed women homozygous for the minor allele A are at increased insulin resistance when SFA intake is high and carbohydrate intake is low. Notably, these gene-fat interactions were observed only for SFAs, but not for MUFAs or PUFAs [230].

Intestinal fatty acid-binding protein 2 (*FABP2*) Ala54Thr polymorphism (rs1799883) is associated with risk of T2DM. Carriers of the Thr54 allele are more susceptible to T2DM than those which are homozygous for Ala54 [231]. Fat intake and insulin sensitivity have also been studied [239] and it was shown that insulin sensitivity improved in subjects with the Thr54 allele of the *FABP2* polymorphism when SFAs were replaced by MUFAs and carbohydrates in 11 case-controlled meta-analysis studies (Table 3).

#### 4. The Role of Epigenetics in the Onset of T2DM

As previously mentioned, T2DM occurs as a consequence of the interaction between genetic and environmental factors. One of the most important environmental factors to consider is diet. GWAS have identified about 100 loci that influence the risk of developing T2DM. Most of them are responsible for alterations in the development of pancreatic islets and/or their function and only a few act through insulin signalling [240]. However, these variants have a reduced effect on the risk of T2DM and are only able to explain at most 10–15% of T2DM inheritance [4]. Thus, if T2DM is a disease in which the hereditary component is important and the genetic variants can explain only a minimal part of that inheritance, this means epigenetic modifications could explain the relationship between environmental factors and the risk of developing T2DM. The proposed explanation is that epigenetic changes occurring throughout life caused by environmental factors (such as diet) can modify gene expression and affect susceptibility to T2DM. In this regard, one of the major changes that has occurred in the Western world and that undoubtedly contributes to the increase in the prevalence of T2DM is an increased caloric intake and reduced energy expenditure, the latter caused by a sedentary lifestyle.

Epigenetics is a form of genomic control that produces changes in the structure of chromatin, without generating changes in the DNA sequence. These changes in chromatin originated by external stimuli are capable of activating or silencing gene expression. There are three types of epigenetic mechanisms: (i) DNA methylation; (ii) post-transcriptional modification of histones; (iii) Mechanisms based on non-coding RNA (microRNA & long non-coding RNA). The first modifies the accessibility to the genes by inducing transcriptional repression. The second includes acetylation, methylation, phosphorylation, ubiquitination and SUMOylation of certain histone amino acids. These processes induce chromatin condensation and consequently, the phenomena of gene activation and/or silencing. Finally, non-coding RNAs can regulate gene expression by acting on the protein synthesis machinery at the translational and post-transcriptional levels. Of special note is the role of microRNAs (miRNAs) and long non-coding RNAs (LncRNAs). Numerous studies have shown that epigenetic modifications are able to induce IR and damage  $\beta$ -cells, thereby causing insufficient insulin release and T2DM [241].

It has been established in experimental studies that patients with T2DM have reduced  $\beta$ -cell mass [242]. In addition, this is known to occur due to external factors, sometimes of a nutritional nature and may make it difficult to maintain the state of differentiation of  $\beta$ -cells. The control of  $\beta$ -cell mass during development and in adulthood is regulated by master genes that act as a balance to regulate  $\beta$ - and  $\alpha$ -cell mass. In this regard, paired box gene 4 (*Pax 4*) is necessary for the existence of mature  $\beta$ -cells and Arista-less-related gene homeobox (*Arx*) for the formation of  $\alpha$ -cells [243]. Thus, in an experimental study with pancreatic islets from patients with T2DM, the *PAX4* gene is hypermethylated and therefore silenced [244]. In contrast, in rodents, the *Arx* gene is methylated and repressed in  $\beta$ -cells [245]. Variations in DNA methylation also affect insulin release processes. For example, the incretin hormone glucagon-like peptide-1 (GLP-1) increases insulin release acting through its receptor (GLP1R). It has been confirmed that in isolated pancreatic islets of patients with T2DM, hypermethylation of the *GLP1R* gene occurs, thus reducing its expression [246].

Post-transcriptional modifications of histones are also able to regulate  $\beta$ -cell mass. It has been shown that in animals lacking histone deacetylase 5 and 9 expression,  $\beta$ -cell mass is increased. Conversely, in mice, with increased deacetylase 4 and 5 expression, there is decreased  $\beta$ -cell mass [247]. Moreover, it is known that although  $\beta$ -cell mass is established before adulthood, it can be modified according to metabolic demands, such as obesity and pregnancy. Indeed, failures in this compensatory capacity can lead to the onset of diabetes [248]. Interestingly, epigenetic modifications can regulate this adaptive proliferation response. This would be done, as it has been demonstrated in mice, by modifying the expression of the cyclin dependent kinase inhibitor 2A gene (*Cdkn2a*) by post-transcriptional modifications of histones [249]. Another fundamental master gene for the maintenance of the  $\beta$ -cell phenotype, and for its correct functioning, is *Pdx1*. It has been shown in mice that the association between this gene and the histone deacetylases varies according to the levels of circulating glucose [250]. Histones also play a role in insulin secretion in response to glucose. For example, histone methyltransferase Set7/9 promotes the expression of important genes such as v-maf musculoaponeurotic fibrosarcoma oncogene family, *MafA* and *Slc2a2* for insulin secretion. In fact, a depletion of Set7/9 in mouse pancreatic islets induces a decrease in insulin release in response to glucose [251].

miRNAs regulate processes related to insulin signalling as well as inflammatory processes. Both phenomena are directly involved in the onset of IR. The role of miRNAs in the development of IR and the onset of T2DM has been studied in different metabolic tissues (liver, adipose, muscle and pancreas) both in humans and mice. In addition, it has been suggested that the microvesicular transport of miRNAs could be a mechanism by which miRNAs act as local and systemic mediators of IR [252]. Studies indicate that the miRNA 143–145 cluster intervenes in IR produced by obesity. Mice that do not express this cluster are protected against IR induced by obesity caused by a high caloric intake. In contrast, conditioned overexpression of miRNA worsens obesity-induced IR [253]. The let-7 family of miRNAs also participates in the regulation of IR. Mice fed a HFD and overexpression of let-7 had impaired glucose tolerance and IR, despite having normal insulin production and secretion levels [254]. Moreover, the administration of anti-miRNA let-7 partially diminished the

effects of HFD on IR in experimental studies on mice and humans [255]. The miRNA 200 family also participates in the regulation of insulin sensitivity. Specifically, inhibition of miRNA 200a in the hypothalamus of ob/ob mice increases leptin receptor expression, decreasing body weight and increasing insulin sensitivity [256]. miRNA 223 knockout mice exhibit higher IR when fed an HFD. In addition, they present a higher degree of inflammatory stress [257]. It has been known for several years that the miRNA29 family negatively regulates the insulin signalling pathway in adipocytes [258]. In addition, increased miRNA29 expression in the livers of diet-induced obese mice and Zucker diabetic rats has been demonstrated. Most interestingly, miRNA29 levels were normalised after treatment with pioglitazone, in both models [259]. miRNA 375 is particularly important for blood glucose regulation. It is expressed at high levels in  $\beta$ -cells, contributes to the maintenance of the  $\beta$ -cell phenotype and enhances insulin synthesis in IR mouse models. In this respect, ob/ob mice show increased miRNA 375 expression. Conversely, mice lacking miRNA 375 have low insulin synthesis and hyperglycemia [260,261]. In addition, experimental studies have shown that miRNA 375 is abundantly expressed during human pancreatic islet differentiation [262].

Aside from dietary modifications, certain nutrients are capable of affecting miRNA expression. It is known that chronic hyperglycemia and hyperlipidemia are capable of modifying gene expression and inducing  $\beta$ -cell damage or dysfunction leading, ultimately, to apoptosis. This process is known as glucotoxicity and lipotoxicity. Excess glucose and fatty acids lead to stress on the rough endoplasmic reticulum of the  $\beta$ -cells, which in turn causes an increase in the production of reactive oxygen species (ROS) [263]. In other tissues it has been shown that ROS can induce modifications in DNA methylation [264], which could also occur in  $\beta$ -cells. Elevated levels of glycosylated hemoglobin indicating a chronic blood glucose elevation, have been found to correlate positively with increases in methylation of the promoter regions of the *INS* and *PDX1* genes in human pancreatic islets [265,266]. Regarding miRNAs, it has been shown in rat pancreatic islets that prolonged exposure of  $\beta$ -cells to SFAs, such as palmitate, increases the expression levels of miRNA 34a and miRNA 146a [267]. Also, glucose is able to modify the expression of miRNA 9, 30d, 124a, 130a, 132, 133, 212 and 335 [268].

#### 4.1. Epigenetic Modifications as Biomarkers of T2DM

Epigenetic modifications can be used as epigenetic biomarkers to detect recent  $\beta$ -cell death. In experimental studies, in mice and humans, it has been shown that the insulin promoter has low methylation levels in endocrine cells, but its methylation level is higher in the rest of the cells in the body [269]. Dying cells release genomic DNA into the bloodstream. When a high proportion of lowly methylated versus highly methylated insulin promoter is found, this is an indicator of recent  $\beta$ -cell death [3]. In the GOLDN Cohort, it was observed that the methylation levels of the ATP binding cassette gene subfamily G member 1 (*ABCG1*, important for cholesterol transport) were significantly associated with fasting insulin levels and IR values in patients with T2DM [270]. In an observational study in the Twins-UK Cohort, it was noted that methylation changes occurred in mucosa-associated lymphoid tissue lymphoma translocation gene 1 (*MALT1*) and G-protein receptor 6 (*GPR6*) gene which indicated T2DM progression [271]. Different degrees of DNA methylation in the adipose tissue of patients with T2DM compared to healthy individuals has been found in peroxisome proliferator activated receptor gamma (*PPAR- $\gamma$* ), insulin receptor substrate 1 (*IRS1*) and transcription factor 7-like 2 (*TCF7L2*) genes [272]. Finally, decreased methylation of the septin 9 (*SEPT9*) gene has been identified in an experimental study using pancreatic islets of patients with T2DM [273].

The presence of miRNAs have also been proposed as potential biomarkers for different diseases including T2DM. This idea is supported by several facts: (i) They are present in human fluids (e.g., plasma, serum, urine, saliva and tears); and (ii) They are very stable at room temperature and show good resistance to freeze-thaw changes. In T2DM there is the additional advantage that compared to other traditional biomarkers (blood, glucose and insulin levels), certain miRNAs have a greater postprandial stability [252]. Nonetheless, the use of miRNAs as biomarkers of T2DM poses three problems. The first is that it is very difficult to determine the precise origin of most circulating

miRNAs. Because pancreatic islets make up a tiny fraction of our body's cells, the amount of plasma miRNAs that come from the pancreatic islets is probably negligible. Secondly, the expression of a specific miRNA is affected by numerous physiological and pathological conditions and, accordingly, it varies. Finally, it is important to have information that relates the plasma miRNAs to SNPs associated with T2DM. Studies are beginning to appear that lead us to believe that miRNAs could be used as biomarkers of T2DM. It has been shown that members of the let-7 family of miRNAs are able to modulate insulin sensitivity and glucose metabolism. A nutritional intervention study found that in healthy young women, reducing the dietary glycemic load increased plasma levels of let-7b by up to eight times [274]. In addition, plasma values of let-7a and 7f are significantly lower in patients with T2DM. In a case-control study, it was found that in treated patients, the levels were normalised when compared to those of the control subjects [275].

#### 4.2. Fetal Developmental Memory

It is known that a large portion of epigenetic footprints are established during the fetal development period. Thus, an adverse intrauterine environment can be “memorized” by generating specific methylation patterns. These patterns may increase the risk of developing T2DM in adulthood, something akin to “metabolic reprogramming”. Within the impact of maternal nutrition, it is important to consider the lipid profile, circulating blood glucose, overweight and obesity, and malnutrition. The proposed explanation is that transient nutritional changes occurring throughout the prenatal period, and possibly affecting the future risk of developing T2DM, are produced as a consequence of epigenetic changes. This hypothesis is based on three assumptions: (i) Epigenetic modifications can occur in response to environmental factors, including food; (ii) Epigenetic changes can be maintained stable throughout life; (iii) These changes can be transmitted to the next generation through gametes.

In animal models, the effects of different nutritional models on the development of T2DM in the F1 and F2 generations have been studied in young female rodents in gestation and during lactation. The nutritional models usually employed are low protein diets, caloric malnutrition and HFD, although other animal models include intrauterine growth retardation (IUGR). Thompson et al. (2010) found that the pancreatic islets of rats subjected to IUGR have an alteration in gene expression, together with different methylation patterns at approximately 1400 CpG sites [276]. In this same IUGR model, a permanent reduction in *Pdx1* expression is produced in  $\beta$ -cells due to epigenetic changes occurring during development. This reduction in *Pdx1* expression has been correlated with the onset of T2DM [277]. On the other hand, Sandovici et al. [278] found that when low protein diets were fed to rats during pregnancy and lactation, there was an increase in methylation of *Hnf4 $\alpha$*  gene promoter, as well as a reduction in the active marks of histone H3 (H3K3me) acetylation in pancreatic islets. Furthermore, the glucose intolerance produced in mice subjected to a caloric malnutrition model is transmitted to the F1 generations and not to the F2 generations [279]. This is due to the appearance of different regions that are methylated and are resistant to epigenetic erasure during the early stages of embryogenesis [280]. In HFD models maintained during pregnancy, an increased risk of obesity and T2DM has also been observed. When female mice were fed an HFD from the fourth week prior to pregnancy until the end of lactation, male mice born in the first generation developed obesity and IR when they reached adulthood [281]. Another widely used model to study fetal environment effects on the development of T2DM includes the induction of gestational diabetes in pregnant females in order to generate a hyperglycemic environment in the uterus. In this case, the offspring of these mothers had a defect in insulin secretion leading to glucose intolerance. In addition, the male offspring of the females born in an environment of intrauterine hyperglycemia also developed the same phenotype [282]. In both cases, the defective insulin secretion was due to hypermethylation of the DNA methylation region, together with a concomitant reduction in the expression of insulin growth factor-2 and H19.

Although there is less information available, there is one study that identifies the existence of a possible “metabolic reprogramming” of paternal origin. Ng et al. crossed male rats subjected to a HFD with control female rats and found that their offspring had altered  $\beta$ -cell function as well as a

differential expression of 642 genes in their pancreatic islets [283]. Moreover, the study by Dunn et al. found that the second generation of male offspring also developed the same alterations [284]. The existence of transmission of a metabolic phenotype mediated by dietary modifications up to the second generation and through the paternal line suggests that epigenetic mechanisms are contributing to this process. To verify whether this generational transmission of phenotypes is mediated by epigenetic factors linked to the germ line, Dunn et al. analysed the F3 generation phenotypes. It was found that the F3 generation females, who were born to the F2 generation males, also showed the same phenotype [281]. Another study also indicates that the induction of HFD-mediated obesity in male mice causes glucose intolerance in their offspring [285]. Finally, in models where prediabetes was induced (with HFD and low-dose of STZ) in male mice, their offspring developed IR and glucose intolerance. When studying gene expression in the islets, a differential expression of 402 genes in the islets of the offspring was found. In addition, when genome-wide DNA methylation analysis was performed there were more than 800 regions showing different methylation patterns in the islets of the offspring. Most interestingly, this study demonstrated that prediabetes in the fathers altered methylation patterns in their sperm. Finally, a significant portion of the hypermethylated and hypomethylated intragenic regions overlapped in the sperm and pancreatic islet cells.

In humans, the most solid data comes from the analysis of what occurred during the famine in Holland between 1944 and 1945. There was a period of approximately five months during which there was a significant food restriction that affected pregnant women in different periods of their gestation. In addition, these women had adequate food before and after this short period. The study and follow-up of the children, after reaching adulthood, who were affected by this famine during their intrauterine life provides much information on how feeding in the prenatal period may subsequently affect the risk of developing T2DM.

The cohort study of Ravelli et al. showed that adults who were born near this period of famine had lower glucose tolerance. The authors of this study concluded that poor intrauterine nutrition could cause permanent changes in the glucose homeostasis process and that these changes were much more evident when those individuals developed obesity [286]. Subsequently, De Rooij et al. found, in the Dutch Famine Birth Cohort, that this glucose intolerance was mediated by a defect in insulin secretion [287]. Moreover, it has been proven, in the cohort study of Hillier et al. that the impact of such alterations can even be transmitted to the next generation [288]. In a recent study in a cohort of people exposed to famine in Bangladesh during their intrauterine developmental stage, their increased predisposition to T2DM was shown to be due to epigenetic changes (mainly changes in DNA methylation). These epigenetic changes vary according to the intrauterine development time frame during which famine was experienced [289]. A similar conclusion was reached by Heijmans et al., in the Dutch Hunger Winter Families Study [290]. In addition, these authors found that the effects of famine on epigenetic modifications were more critical if famine occurred during the embryonic developmental period.

Furthermore, studies in the Dutch Famine Birth Cohort found that when the fetuses were subjected to insufficient feeding during any period of their gestation, there was an increase in the LDL:HDL ratio upon reaching adulthood [291]. This increases the risk of developing T2DM, as it is well established that an increase in LDL levels predisposes to T2DM.

In contrast, a population-based cohort study with women who develop obesity during pregnancy, have IR. The excess nutrients consumed by the mother reach the fetus which in turn releases its own insulin, thus increasing the fetal uptake of glucose and fats causing these infants to be born with a higher weight [292]. When these children reach adulthood, they are exposed to an increased risk of developing obesity and T2DM. Studies by Wellen et al. indicate that in cell culture lines, the ATP-citrate lyase enzyme is essential for histone acetylation [293]. This enzyme is critical in the conversion of citrate through glycolysis into acetyl-CoA. Curiously, the acetylation of histones is more evident in periods when there are sufficient glucose inputs. There is another historical cohort from Northern Sweden (Överkalix cohort) that demonstrates that overfeeding in children may increase the risk of T2D in their grandchildren [294]. Finally, as occurs with male rats, a historical cohort study based on

the Dutch Famine Population showed that the children of men who experienced prenatal malnutrition are more obese and therefore at greater risk of developing T2DM [295].

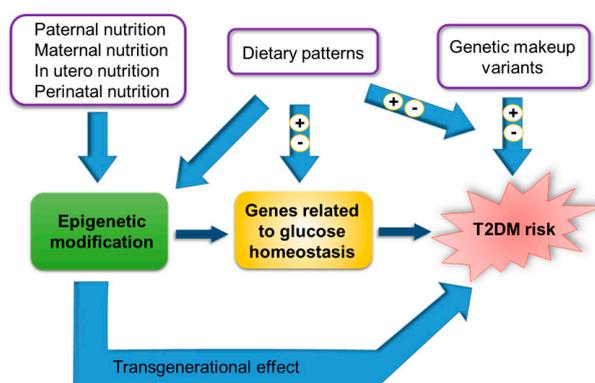
## 5. Conclusions

T2DM is a multifactorial disease which arises from complicated interactions between the genetic makeup and environmental factors. Among these factors, many studies (experimental, case-control, cohort and nutritional interventions) have shown the impact of diet on the development of T2DM. Several cohort studies have also shown that diet can generate epigenetic modifications throughout the lifetime that affect gene expression and consequently, susceptibility to T2DM. Moreover, these epigenetic modifications might play a relevant role in the inheritance of T2DM.

Nutrition is a very important factor that modulates expression of genes involved in metabolic pathways related to T2DM pathogenesis. As previously discussed, different macro and micronutrients, as well as food bioactive compounds positively affect the expression of genes involved in insulin synthesis, stimulus-secretion coupling, protection of  $\beta$ -cells against gluco-lipotoxicity, inflammation and oxidative stress, and IR. However, this jigsaw is not so easy to solve. Nutrigenetics give us the shape of the pieces, but we still need the drawing of the pieces of the puzzle and the resulting picture we want to reproduce.

The information about the pieces is provided by nutrigenomics. Linkage analysis, candidate gene approaches, genome-wide association studies and sequencing have provided us with plenty of information that allows a better understanding of the genetic landscape of T2DM. This disease has a strong genetic predisposition and the discovery of the existence of genetic variants associated with T2DM and their related traits has shed light on the different individual risks. There are cohort and nutritional intervention studies showing that people with higher genetic predisposition should avoid particular dietetic patterns that are more harmful in the variation of a specific T2DM-related phenotype.

The picture to be reproduced in our puzzle appears when the epigenetic factors come into the scene. Nutrigenetics and nutrigenomics are not able to completely explain the complex relationship between environmental factors and genetic susceptibility to T2DM. In fact, the overall risk explained by genetic variations is small. Epigenetic could be the missing link that gives shape to the puzzle. The rationale is that nutrients and dietetic patterns experienced during one's lifetime, including the prenatal and neonatal periods, induce epigenetic modifications that affect gene expression and change the individual disease risk present in our genetic makeup (Figure 1). In addition, these epigenetic modifications could be behind the possible transgenerational inheritance of T2DM.



**Figure 1.** Schematic diagram of the role of nutritional, genetic and epigenetic factors in the development of T2DM. Nutrients, dietary patterns and genetic makeup (SNPs) have a direct impact on T2DM risk. Parental nutrition, prenatal and perinatal nutrition induce epigenetic modifications that increase the susceptibility of T2DM development during adulthood. The epigenetic modifications can be inherited to following generations. +: Increase; -: decrease.

In this review, we have tried to provide clues in order to demonstrate that the relationship between diet and genes in T2DM is not easy to understand, nor is it easy to pinpoint which came first: genes or diet (or vice versa). At the moment, there are three actors: (i) Nutrient-gene interactions; (ii) Individual genetic variants; (iii) Epigenetic modifications. Considering these three actors, we could likely write the script and it should not take long, because considering the global pandemic of T2DM and the broad spectrum of T2DM patients, the movie is currently being shown without the screenwriters. This approach will result in patient centered, more accurate precision-medicine.

**Acknowledgments:** This work was supported by Grants from ISCIII cofounded by Fondos FEDER (FIS PI14/01015; RD/0019/0028 and RD16/0011/0034 to BS; PI14/0804 to AR), grants from MINECO (INNPRONTA Program to NewBiotechnic SA and AGL2014-54585-R to FM), “Ramón y Cajal” program from the Spanish Ministry of Economy and Competitiveness (RYC-2013-14533 to AR) and Fundación Progreso y Salud from the Junta de Andalucía which supported publishing in open access.

**Author Contributions:** All authors contributed to previous studies in which this review is based. Franz Martín, Bernat Soria, Genoveva Berná, Anabel Rojas and Ángeles Ortega conceived and designed this article. The first version was written by Ángeles Ortega and Genoveva Berná, subsequently Anabel Rojas, Franz Martín and Bernat Soria revised and rewritten the final version. Anabel Rojas, Franz Martín and Bernat Soria contributed to the costs. All the authors are fully responsible for the content.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

T2DM	Type 2 diabetes Mellitus
STZ	Streptozotocin
PI3K	Endothelial phosphatidylinositol 3-kinase
EGCG	Epigallocatechin gallate
Ins	Insulin
IR	Insulin resistance
IRS	Insulin receptor substrate
AMPK	AMP-activated protein kinase
GLUT	Glucose transporter
Ppp1r15a	Protein phosphatase 1, regulatory subunit 15A
Cdkn1a	Cyclin-dependent kinase inhibitor 1A
ACO-1	Acyl CoA oxidase-1
CPT-1	Carnitine palmitoyl transferase-1
HbA1c	Glycated hemoglobin
PPAR	Peroxisome proliferator-activated receptor
PEPCK	Phosphoenolpyruvate carboxykinase
HFD	High-fat diet
HSP	Heat shock proteins
NO	Nitric oxide
NF-κB	Nuclear factor kappa B
CRP	C-reactive protein
IFN-γ	Interferon-gamma
IL	Interleukin
G6Pase	Glucose-6-phosphatase
RBP	Retinol binding protein
Pdx1	Duodenal homeobox factor transcription factor
Hnf	Hepatocyte nuclear factor
Gk	Glucokinase
Cacna1d	Calcium Voltage-Gated Channel Subunit Alpha1 D
MafA	v-Maf avian musculoaponeurotic fibrosarcoma oncogene homolog A
BCAAs	Branched-chain amino acids
SFAs	Saturated fatty acids
ROS	Reactive oxygen species

MUFAs	Monounsaturated fatty acids
PUFAs	Polyunsaturated fatty acids
EPA	Eicosapentaenoic acid
DHA	Docosahexaenoic acid
GWAS	Genome-wide association studies
IRS	Insulin receptor substrate
FTO	Fat mass and obesity-associated
SNPs	Single nucleotide polymorphisms
CLOCK	Circadian Locomotor Output Cycles Kaput
MC4R	Melanocortin-4 receptor
PLIN	Perilipin
FABP	Fatty acid-binding protein
miRNAs	microRNAs
LncRNAs	Long non-coding RNAs
Pax	Paired box gene
Arx	Arista-less-related gene homeobox
GLP	Glucagon-like peptide
IUGR	Intrauterine growth retardation
LDL	Low-density lipoprotein
HDL	High-density lipoprotein

## References

- International Diabetes Federation. *IDF Diabetes Atlas*, 7th ed.; International Diabetes Federation: Brussels, Belgium, 2015.
- Schulze, M.B.; Hu, F.B. Primary prevention of diabetes: What can be done and how much can be prevented? *Annu. Rev. Public Health* **2005**, *26*, 445–467. [[CrossRef](#)] [[PubMed](#)]
- Bernstein, D.; Golson, M.L.; Kaestner, K.H. Epigenetic control of  $\beta$ -cell function and failure. *Diabetes Res. Clin. Pract.* **2017**, *123*, 24–36. [[CrossRef](#)] [[PubMed](#)]
- Stančáková, A.; Laakso, M. Genetics of Type 2 diabetes. *Endocr. Dev.* **2016**, *31*, 203–220. [[PubMed](#)]
- Berná, G.; Oliveras-López, M.J.; Jurado-Ruíz, E.; Tejedó, J.; Bedoya, F.; Soria, B.; Martín, F. Nutrigenetics and nutrigenomics insights into diabetes etiopathogenesis. *Nutrients* **2014**, *6*, 5338–5369. [[CrossRef](#)] [[PubMed](#)]
- Rana, S.; Kumar, S.; Rathore, N.; Padwad, Y.; Bhushana, S. Nutrigenomics and its impact on life style associated metabolic diseases. *Curr. Genom.* **2016**, *17*, 261–278. [[CrossRef](#)] [[PubMed](#)]
- Sales, N.M.R.; Pelegrini, P.B.; Goersch, M.C. Nutrigenomics: Definitions and advances of this new science. *J. Nutr. Metab.* **2014**, *2014*, 202759. [[CrossRef](#)] [[PubMed](#)]
- Manach, C.; Scalbert, A.; Morand, C.; Rémésy, C.; Jiménez, L. Polyphenols: Food sources and bioavailability. *Am. J. Clin. Nutr.* **2004**, *79*, 727–747. [[PubMed](#)]
- Xiao, J.B.; Högger, P. Dietary polyphenols and Type 2 diabetes: Current insights and future perspectives. *Curr. Med. Chem.* **2015**, *22*, 23–38. [[CrossRef](#)] [[PubMed](#)]
- Shidfar, F.; Heydari, I.; Hajimiresmaiel, S.J.; Hosseini, S.; Shidfar, S.; Amiri, F. The effects of cranberry juice on serum glucose, apoB, apoA-I, Lp(a), and paraoxonase-1 activity in type 2 diabetic male patients. *J. Res. Med. Sci.* **2012**, *17*, 355–360. [[PubMed](#)]
- Lin, D.; Xiao, M.; Zhao, J.; Li, Z.; Xing, B.; Li, X.; Kong, M.; Li, L.; Zhang, Q.; Liu, Y.; et al. An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. *Molecules* **2016**, *21*, 1374. [[CrossRef](#)] [[PubMed](#)]
- Hanhineva, K.; Törrönen, R.; Bondia-Pons, I.; Pekkinen, J.; Kolehmainen, M.; Mykkänen, H.; Poutanen, K. Impact of dietary polyphenols on carbohydrate metabolism. *Int. J. Mol. Sci.* **2010**, *11*, 1365–1402. [[CrossRef](#)] [[PubMed](#)]
- Kerimi, A.; Williamson, G. At the interface of antioxidant signalling and cellular function: Key polyphenol effects. *Mol. Nutr. Food Res.* **2016**, *60*, 1770–1788. [[CrossRef](#)] [[PubMed](#)]
- Kim, Y.; Keogh, J.; Clifton, P. Polyphenols and glycemic control. *Nutrients* **2016**, *8*, 17. [[CrossRef](#)] [[PubMed](#)]
- Babu, P.V.A.; Liu, D.; Gilbert, E.R. Recent advances in understanding the anti-diabetic actions of dietary flavonoids. *J. Nutr. Biochem.* **2013**, *24*, 1777–1789. [[CrossRef](#)] [[PubMed](#)]

16. Sun, Q.; Wedick, N.M.; Tworoger, S.S.; Pan, A.; Townsend, M.K.; Cassidy, A.; Franke, A.A.; Rimm, E.B.; Hu, F.B.; van Dam, R.M. Urinary excretion of select dietary polyphenol metabolites is associated with a lower risk of Type 2 diabetes in proximate but not remote follow-up in a prospective investigation in 2 cohorts of US women. *J. Nutr.* **2015**, *145*, 1280–1288. [[CrossRef](#)] [[PubMed](#)]
17. Song, Y.; Manson, J.E.; Buring, J.E.; Sesso, H.D.; Liu, S. Associations of dietary flavonoids with risk of Type 2 diabetes, and markers of insulin resistance and systemic inflammation in women: A prospective study and cross-sectional analysis. *J. Am. Coll. Nutr.* **2005**, *24*, 376–384. [[CrossRef](#)] [[PubMed](#)]
18. Wedick, N.M.; Pan, A.; Cassidy, A.; Rimm, E.B.; Sampson, L.; Rosner, B.; Willett, W.; Hu, F.B.; Sun, Q.; van Dam, R.M. Dietary flavonoid intakes and risk of Type 2 diabetes in US men and women. *Am. J. Clin. Nutr.* **2012**, *95*, 925–933. [[CrossRef](#)] [[PubMed](#)]
19. Nettleton, J.A.; Harnack, L.J.; Scrafford, C.G.; Mink, P.J.; Barraj, L.M.; Jacobs, D.R. Dietary flavonoids and flavonoid-rich foods are not associated with risk of Type 2 diabetes in postmenopausal women. *J. Nutr.* **2006**, *136*, 3039–3045. [[PubMed](#)]
20. Bhardwaj, P.; Khanna, D.; Balakumar, P. Catechin averts experimental diabetes mellitus-induced vascular endothelial structural and functional abnormalities. *Cardiovasc. Toxicol.* **2014**, *14*, 41–51. [[CrossRef](#)] [[PubMed](#)]
21. Cai, E.P.; Lin, J.-K. Epigallocatechin gallate (EGCG) and rutin suppress the glucotoxicity through activating IRS2 and AMPK signalling in rat pancreatic beta cells. *J. Agric. Food Chem.* **2009**, *57*, 9817–9827. [[CrossRef](#)] [[PubMed](#)]
22. Zhang, Z.F.; Li, Q.; Liang, J.; Dai, X.Q.; Ding, Y.; Wang, J.B.; Li, Y. Epigallocatechin-3-O-gallate (EGCG) protects the insulin sensitivity in rat L6 muscle cells exposed to dexamethasone condition. *Phytomedicine* **2010**, *17*, 14–18. [[CrossRef](#)] [[PubMed](#)]
23. Ueda-Wakagi, M.; Mukai, R.; Fuse, N.; Mizushima, Y.; Ashida, H. 3-O-Acyl-epicatechins increase glucose uptake activity and GLUT4 translocation through activation of PI3K signalling in skeletal muscle cells. *Int. J. Mol. Sci.* **2015**, *16*, 16288–16299. [[CrossRef](#)] [[PubMed](#)]
24. Zhang, Z.; Ding, Y.; Dai, X.; Wang, J.; Li, Y. Epigallocatechin-3-gallate protects pro-inflammatory cytokine induced injuries in insulin-producing cells through the mitochondrial pathway. *Eur. J. Pharmacol.* **2011**, *670*, 311–316. [[CrossRef](#)] [[PubMed](#)]
25. Ortsäter, H.; Grankvist, N.; Wolfram, S.; Kuehn, N.; Sjöholm, A. Diet supplementation with green tea extract epigallocatechin gallate prevents progression to glucose intolerance in db/db mice. *Nutr. Metab.* **2012**, *9*, 11. [[CrossRef](#)] [[PubMed](#)]
26. Wolfram, S.; Raederstorff, D.; Preller, M.; Wang, Y.; Teixeira, S.R.; Riegger, C.; Weber, P. Epigallocatechin gallate supplementation alleviates diabetes in rodents. *J. Nutr.* **2006**, *136*, 2512–2518. [[PubMed](#)]
27. Tsuneki, H.; Ishizuka, M.; Terasawa, M.; Wu, J.-B.; Sasaoka, T.; Kimura, I. Effect of green tea on blood glucose levels and serum proteomic patterns in diabetic (db/db) mice and on glucose metabolism in healthy humans. *BMC Pharmacol.* **2004**, *4*, 18. [[CrossRef](#)] [[PubMed](#)]
28. Mackenzie, T.; Leary, L.; Brooks, W.B. The effect of an extract of green and black tea on glucose control in adults with Type 2 diabetes mellitus: Double-blind randomized study. *Metabolism* **2007**, *56*, 1340–1344. [[CrossRef](#)] [[PubMed](#)]
29. Zheng, X.-X.; Xu, Y.-L.; Li, S.-H.; Hui, R.; Wu, Y.-J.; Huang, X.-H. Effects of green tea catechins with or without caffeine on glycemic control in adults: A meta-analysis of randomized controlled trials. *Am. J. Clin. Nutr.* **2013**, *97*, 750–762. [[CrossRef](#)] [[PubMed](#)]
30. Jung, U.J.; Lee, M.-K.; Park, Y.B.; Kang, M.A.; Choi, M.-S. Effect of citrus flavonoids on lipid metabolism and glucose-regulating enzyme mRNA levels in type-2 diabetic mice. *Int. J. Biochem. Cell Biol.* **2006**, *38*, 1134–1145. [[CrossRef](#)] [[PubMed](#)]
31. Sharma, A.K.; Bharti, S.; Ojha, S.; Bhatia, J.; Kumar, N.; Ray, R.; Kumari, S.; Arya, D.S. Up-regulation of PPAR $\gamma$ , heat shock protein-27 and -72 by naringin attenuates insulin resistance,  $\beta$ -cell dysfunction, hepatic steatosis and kidney damage in a rat model of Type 2 diabetes. *Br. J. Nutr.* **2011**, *106*, 1713–1723. [[CrossRef](#)] [[PubMed](#)]
32. Coskun, O.; Kanter, M.; Korkmaz, A.; Oter, S. Quercetin, a flavonoid antioxidant, prevents and protects streptozotocin-induced oxidative stress and beta-cell damage in rat pancreas. *Pharmacol. Res.* **2005**, *51*, 117–123. [[CrossRef](#)] [[PubMed](#)]

33. Shi, Y.; Liang, X.; Zhang, H.; Wu, Q.; Qu, L.; Sun, Q. Quercetin protects rat dorsal root ganglion neurons against high glucose-induced injury in vitro through Nrf-2/HO-1 activation and NF- $\kappa$ B inhibition. *Acta Pharmacol. Sin.* **2013**, *34*, 1140–1148. [[CrossRef](#)] [[PubMed](#)]
34. Mahmoud, M.F.; Hassan, N.A.; El Bassossy, H.M.; Fahmy, A. Quercetin protects against diabetes-induced exaggerated vasoconstriction in rats: Effect on low grade inflammation. *PLoS ONE* **2013**, *8*, e63784. [[CrossRef](#)] [[PubMed](#)]
35. Kobori, M.; Masumoto, S.; Akimoto, Y.; Takahashi, Y. Dietary quercetin alleviates diabetic symptoms and reduces streptozotocin-induced disturbance of hepatic gene expression in mice. *Mol. Nutr. Food Res.* **2009**, *53*, 859–868. [[CrossRef](#)] [[PubMed](#)]
36. Stewart, L.K.; Soileau, J.L.; Ribnicky, D.; Wang, Z.Q.; Raskin, I.; Poulev, A.; Majewski, M.; Cefalu, W.T.; Gettys, T.W. Quercetin transiently increases energy expenditure but persistently decreases circulating markers of inflammation in C57BL/6J mice fed a high-fat diet. *Metabolism* **2008**, *57*, S39–S46. [[CrossRef](#)] [[PubMed](#)]
37. Rivera, L.; Morón, R.; Sánchez, M.; Zarzuelo, A.; Galisteo, M. Quercetin ameliorates metabolic syndrome and improves the inflammatory status in obese Zucker rats. *Obesity* **2008**, *16*, 2081–2087. [[CrossRef](#)] [[PubMed](#)]
38. Kim, E.-K.; Kwon, K.-B.; Song, M.-Y.; Han, M.-J.; Lee, J.-H.; Lee, Y.-R.; Lee, J.-H.; Ryu, D.-G.; Park, B.-H.; Park, J.-W. Flavonoids protect against cytokine-induced pancreatic  $\beta$ -cell damage through suppression of nuclear factor  $\kappa$ B activation. *Pancreas* **2007**, *35*, e1–e9. [[CrossRef](#)] [[PubMed](#)]
39. Noriega-López, L.; Tovar, A.R.; Gonzalez-Granillo, M.; Hernández-Pando, R.; Escalante, B.; Santillán-Doherty, P.; Torres, N. Pancreatic insulin secretion in rats fed a soy protein high fat diet depends on the interaction between the amino acid pattern and isoflavones. *J. Biol. Chem.* **2007**, *282*, 20657–20666. [[CrossRef](#)] [[PubMed](#)]
40. Fu, Z.; Zhang, W.; Zhen, W.; Lum, H.; Nadler, J.; Bassaganya-Riera, J.; Jia, Z.; Wang, Y.; Misra, H.; Liu, D. Genistein induces pancreatic beta-cell proliferation through activation of multiple signalling pathways and prevents insulin-deficient diabetes in mice. *Endocrinology* **2010**, *151*, 3026–3037. [[CrossRef](#)] [[PubMed](#)]
41. Villa, P.; Costantini, B.; Suriano, R.; Perri, C.; Macrì, F.; Ricciardi, L.; Panunzi, S.; Lanzone, A. The differential effect of the phytoestrogen genistein on cardiovascular risk factors in postmenopausal women: Relationship with the metabolic status. *J. Clin. Endocrinol. Metab.* **2009**, *94*, 552–558. [[CrossRef](#)] [[PubMed](#)]
42. Zhang, B.; Kang, M.; Xie, Q.; Xu, B.; Sun, C.; Chen, K.; Wu, Y. Anthocyanins from Chinese bayberry extract protect  $\beta$  cells from oxidative stress-mediated injury via HO-1 upregulation. *J. Agric. Food Chem.* **2011**, *59*, 537–545. [[CrossRef](#)] [[PubMed](#)]
43. Sasaki, R.; Nishimura, N.; Hoshino, H.; Isa, Y.; Kadowaki, M.; Ichi, T.; Tanaka, A.; Nishiumi, S.; Fukuda, I.; Ashida, H.; et al. Cyanidin 3-glucoside ameliorates hyperglycemia and insulin sensitivity due to downregulation of retinol binding protein 4 expression in diabetic mice. *Biochem. Pharmacol.* **2007**, *74*, 1619–1627. [[CrossRef](#)] [[PubMed](#)]
44. Takikawa, M.; Inoue, S.; Horio, F.; Tsuda, T. Dietary anthocyanin-rich bilberry extract ameliorates hyperglycemia and insulin sensitivity via activation of AMP-activated protein kinase in diabetic mice. *J. Nutr.* **2010**, *140*, 527–533. [[CrossRef](#)] [[PubMed](#)]
45. Ding, M.; Bhupathiraju, S.N.; Chen, M.; van Dam, R.M.; Hu, F.B. Caffeinated and decaffeinated coffee consumption and risk of Type 2 diabetes: A systematic review and a dose-response meta-analysis. *Diabetes Care* **2014**, *37*, 569–586. [[CrossRef](#)] [[PubMed](#)]
46. Jiang, X.; Zhang, D.; Jiang, W. Coffee and caffeine intake and incidence of Type 2 diabetes mellitus: A meta-analysis of prospective studies. *Eur. J. Nutr.* **2014**, *53*, 25–38. [[CrossRef](#)] [[PubMed](#)]
47. Van Dieren, S.; Uiterwaal, C.S.P.M.; van der Schouw, Y.T.; van der A, D.L.; Boer, J.M.A.; Spijkerman, A.; Grobbee, D.E.; Beulens, J.W.J. Coffee and tea consumption and risk of Type 2 diabetes. *Diabetologia* **2009**, *52*, 2561–2569. [[CrossRef](#)] [[PubMed](#)]
48. Yang, J.; Mao, Q.-X.; Xu, H.-X.; Ma, X.; Zeng, C.-Y. Tea consumption and risk of Type 2 diabetes mellitus: A systematic review and meta-analysis update. *BMJ Open* **2014**, *4*, e005632. [[CrossRef](#)] [[PubMed](#)]
49. Yang, W.-S.; Wang, W.-Y.; Fan, W.-Y.; Deng, Q.; Wang, X. Tea consumption and risk of Type 2 diabetes: A dose-response meta-analysis of cohort studies. *Br. J. Nutr.* **2014**, *111*, 1329–1339. [[CrossRef](#)] [[PubMed](#)]
50. Pham, N.M.; Nanri, A.; Kochi, T.; Kuwahara, K.; Tsuruoka, H.; Kurotani, K.; Akter, S.; Kabe, I.; Sato, M.; Hayabuchi, H.; et al. Coffee and green tea consumption is associated with insulin resistance in Japanese adults. *Metabolism* **2014**, *63*, 400–408. [[CrossRef](#)] [[PubMed](#)]

51. Arnlöv, J.; Vessby, B.; Risérus, U. Coffee consumption and insulin sensitivity. *JAMA* **2004**, *291*, 1199–1201. [[CrossRef](#)] [[PubMed](#)]
52. Yarmolinsky, J.; Mueller, N.T.; Duncan, B.B.; Bisi Molina, M.D.C.; Goulart, A.C.; Schmidt, M.I. Coffee Consumption, Newly Diagnosed Diabetes, and Other Alterations in Glucose Homeostasis: A Cross-Sectional Analysis of the Longitudinal Study of Adult Health (ELSA-Brasil). *PLoS ONE* **2015**, *10*, e0126469. [[CrossRef](#)] [[PubMed](#)]
53. Van Dam, R.M.; Dekker, J.M.; Nijpels, G.; Stehouwer, C.D.A.; Bouter, L.M.; Heine, R.J. Hoorn study Coffee consumption and incidence of impaired fasting glucose, impaired glucose tolerance, and Type 2 diabetes: The Hoorn Study. *Diabetologia* **2004**, *47*, 2152–2159. [[CrossRef](#)] [[PubMed](#)]
54. Oboh, G.; Agunloye, O.M.; Adefegha, S.A.; Akinyemi, A.J.; Ademiluyi, A.O. Caffeic and chlorogenic acids inhibit key enzymes linked to Type 2 diabetes (in vitro): A comparative study. *J. Basic Clin. Physiol. Pharmacol.* **2015**, *26*, 165–170. [[CrossRef](#)] [[PubMed](#)]
55. Aoyagi, R.; Funakoshi-Tago, M.; Fujiwara, Y.; Tamura, H. Coffee inhibits adipocyte differentiation via inactivation of PPAR $\gamma$ . *Biol. Pharm. Bull.* **2014**, *37*, 1820–1825. [[CrossRef](#)] [[PubMed](#)]
56. Jin, S.; Chang, C.; Zhang, L.; Liu, Y.; Huang, X.; Chen, Z. Chlorogenic acid improves late diabetes through adiponectin receptor signalling pathways in db/db mice. *PLoS ONE* **2015**, *10*, e0120842.
57. Narasimhan, A.; Chinnaiyan, M.; Karundevi, B. Ferulic acid exerts its antidiabetic effect by modulating insulin-signalling molecules in the liver of high-fat diet and fructose-induced type-2 diabetic adult male rat. *Appl. Physiol. Nutr. Metab.* **2015**, *40*, 769–781. [[CrossRef](#)] [[PubMed](#)]
58. Narasimhan, A.; Chinnaiyan, M.; Karundevi, B. Ferulic acid regulates hepatic GLUT2 gene expression in high fat and fructose-induced type-2 diabetic adult male rat. *Eur. J. Pharmacol.* **2015**, *761*, 391–397. [[CrossRef](#)] [[PubMed](#)]
59. Bagul, P.K.; Banerjee, S.K. Application of resveratrol in diabetes: Rationale, strategies and challenges. *Curr. Mol. Med.* **2015**, *15*, 312–330. [[CrossRef](#)] [[PubMed](#)]
60. Szkudelski, T.; Szkudelska, K. Resveratrol and diabetes: From animal to human studies. *Biochim. Biophys. Acta Mol. Basis Dis.* **2015**, *1852*, 1145–1154. [[CrossRef](#)] [[PubMed](#)]
61. Zagotta, I.; Dimova, E.Y.; Debatin, K.-M.; Wabitsch, M.; Kietzmann, T.; Fischer-Posovszky, P. Obesity and inflammation: Reduced cytokine expression due to resveratrol in a human in vitro model of inflamed adipose tissue. *Front. Pharmacol.* **2015**, *6*, 79. [[CrossRef](#)] [[PubMed](#)]
62. Diaz-Gerevini, G.T.; Repossi, G.; Dain, A.; Tarres, M.C.; Das, U.N.; Eynard, A.R. Beneficial action of resveratrol: How and why? *Nutrition* **2016**, *32*, 174–178. [[CrossRef](#)] [[PubMed](#)]
63. Sung, M.M.; Kim, T.T.; Denou, E.; Soltys, C.-L.M.; Hamza, S.M.; Byrne, N.J.; Masson, G.; Park, H.; Wishart, D.S.; Madsen, K.L.; et al. Improved glucose homeostasis in obese mice treated with resveratrol is associated with alterations in the gut microbiome. *Diabetes* **2017**, *66*, 418–425. [[CrossRef](#)] [[PubMed](#)]
64. Timmers, S.; de Ligt, M.; Phielix, E.; van de Weijer, T.; Hansen, J.; Moonen-Kornips, E.; Schaart, G.; Kunz, I.; Hesselink, M.K.C.; Schrauwen-Hinderling, V.B.; et al. Resveratrol as add-on therapy in subjects with well-controlled Type 2 diabetes: A randomized controlled trial. *Diabetes Care* **2016**, *39*, 2211–2217. [[CrossRef](#)] [[PubMed](#)]
65. Bo, S.; Ponzio, V.; Ciccone, G.; Evangelista, A.; Saba, F.; Goitre, I.; Procopio, M.; Pagano, G.F.; Cassader, M.; Gambino, R. Six months of resveratrol supplementation has no measurable effect in type 2 diabetic patients. A randomized, double blind, placebo-controlled trial. *Pharmacol. Res.* **2016**, *111*, 896–905. [[CrossRef](#)] [[PubMed](#)]
66. Crandall, J.P.; Oram, V.; Trandafirescu, G.; Reid, M.; Kishore, P.; Hawkins, M.; Cohen, H.W.; Barzilai, N. Pilot study of resveratrol in older adults with impaired glucose tolerance. *J. Gerontol. A Biol. Sci. Med. Sci.* **2012**, *67*, 1307–1312. [[CrossRef](#)] [[PubMed](#)]
67. Bhatt, J.K.; Thomas, S.; Nanjan, M.J. Resveratrol supplementation improves glycemic control in Type 2 diabetes mellitus. *Nutr. Res.* **2012**, *32*, 537–541. [[CrossRef](#)] [[PubMed](#)]
68. Brasnyó, P.; Molnár, G.A.; Mohás, M.; Markó, L.; Laczy, B.; Cseh, J.; Mikolás, E.; Szijártó, I.A.; Mérei, A.; Halmai, R.; et al. Resveratrol improves insulin sensitivity, reduces oxidative stress and activates the Akt pathway in type 2 diabetic patients. *Br. J. Nutr.* **2011**, *106*, 383–389. [[CrossRef](#)] [[PubMed](#)]
69. Rouse, M.; Younès, A.; Egan, J.M. Resveratrol and curcumin enhance pancreatic  $\beta$ -cell function by inhibiting phosphodiesterase activity. *J. Endocrinol.* **2014**, *223*, 107–117. [[CrossRef](#)] [[PubMed](#)]

70. Chuengsamarn, S.; Rattanamongkolgul, S.; Luechapudiporn, R.; Phisalaphong, C.; Jirawatnotai, S. Curcumin extract for prevention of Type 2 diabetes. *Diabetes Care* **2012**, *35*, 2121–2127. [[CrossRef](#)] [[PubMed](#)]
71. Aggarwal, B.B. Targeting inflammation-induced obesity and metabolic diseases by curcumin and other nutraceuticals. *Annu. Rev. Nutr.* **2010**, *30*, 173–199. [[CrossRef](#)] [[PubMed](#)]
72. Chiu, K.C.; Chu, A.; Go, V.L.W.; Saad, M.F. Hypovitaminosis D is associated with insulin resistance and beta cell dysfunction. *Am. J. Clin. Nutr.* **2004**, *79*, 820–825. [[PubMed](#)]
73. Oh, Y.S.; Jun, H.-S.; Jun, H.-S. Role of bioactive food components in diabetes prevention: Effects on Beta-cell function and preservation. *Nutr. Metab. Insights* **2014**, *7*, 51–59. [[PubMed](#)]
74. Cade, C.; Norman, A.W. Rapid normalization/stimulation by 1,25-dihydroxyvitamin D<sub>3</sub> of insulin secretion and glucose tolerance in the vitamin D-deficient rat. *Endocrinology* **1987**, *120*, 1490–1497. [[CrossRef](#)] [[PubMed](#)]
75. Maestro, B.; Molero, S.; Bajo, S.; Dávila, N.; Calle, C. Transcriptional activation of the human insulin receptor gene by 1,25-dihydroxyvitamin D<sub>3</sub>. *Cell Biochem. Funct.* **2002**, *20*, 227–232. [[CrossRef](#)] [[PubMed](#)]
76. Dunlop, T.W.; Väisänen, S.; Frank, C.; Molnár, F.; Sinkkonen, L.; Carlberg, C. The human peroxisome proliferator-activated receptor delta gene is a primary target of 1 $\alpha$ ,25-dihydroxyvitamin D<sub>3</sub> and its nuclear receptor. *J. Mol. Biol.* **2005**, *349*, 248–260. [[CrossRef](#)] [[PubMed](#)]
77. Giulietti, A.; van Etten, E.; Overbergh, L.; Stoffels, K.; Bouillon, R.; Mathieu, C. Monocytes from type 2 diabetic patients have a pro-inflammatory profile. *Diabetes Res. Clin. Pract.* **2007**, *77*, 47–57. [[CrossRef](#)] [[PubMed](#)]
78. Wolden-Kirk, H.; Overbergh, L.; Christesen, H.T.; Brusgaard, K.; Mathieu, C. Vitamin D and diabetes: Its importance for beta cell and immune function. *Mol. Cell. Endocrinol.* **2011**, *347*, 106–120. [[CrossRef](#)] [[PubMed](#)]
79. Afzal, S.; Bojesen, S.E.; Nordestgaard, B.G. Low 25-hydroxyvitamin D and risk of Type 2 diabetes: A prospective cohort study and metaanalysis. *Clin. Chem.* **2013**, *59*, 381–391. [[CrossRef](#)] [[PubMed](#)]
80. Kramer, C.K.; Swaminathan, B.; Hanley, A.J.; Connelly, P.W.; Sermer, M.; Zinman, B.; Retnakaran, R. Prospective associations of vitamin D status with  $\beta$ -Cell function, insulin sensitivity, and glycemia: The impact of parathyroid hormone status. *Diabetes* **2014**, *63*, 3868–3879. [[CrossRef](#)] [[PubMed](#)]
81. Palomer, X.; González-Clemente, J.M.; Blanco-Vaca, F.; Mauricio, D. Role of vitamin D in the pathogenesis of Type 2 diabetes mellitus. *Diabetes Obes. Metab.* **2008**, *10*, 185–197. [[CrossRef](#)] [[PubMed](#)]
82. Leung, P. The potential protective action of vitamin d in hepatic insulin resistance and pancreatic islet dysfunction in Type 2 diabetes Mellitus. *Nutrients* **2016**, *8*, 147. [[CrossRef](#)] [[PubMed](#)]
83. Tabesh, M.; Azadbakht, L.; Faghihmani, E.; Tabesh, M.; Esmailzadeh, A. Effects of calcium-vitamin D co-supplementation on metabolic profiles in vitamin D insufficient people with Type 2 diabetes: A randomised controlled clinical trial. *Diabetologia* **2014**, *57*, 2038–2047. [[CrossRef](#)] [[PubMed](#)]
84. Seida, J.C.; Mitri, J.; Colmers, I.N.; Majumdar, S.R.; Davidson, M.B.; Edwards, A.L.; Hanley, D.A.; Pittas, A.G.; Tjosvold, L.; Johnson, J.A. Clinical review: Effect of vitamin D<sub>3</sub> supplementation on improving glucose homeostasis and preventing diabetes: A systematic review and meta-analysis. *J. Clin. Endocrinol. Metab.* **2014**, *99*, 3551–3560. [[CrossRef](#)] [[PubMed](#)]
85. Nakashima, A.; Yokoyama, K.; Yokoo, T.; Urashima, M. Role of vitamin D in diabetes mellitus and chronic kidney disease. *World J. Diabetes* **2016**, *7*, 89. [[CrossRef](#)] [[PubMed](#)]
86. Matthews, K.A.; Rhoten, W.B.; Driscoll, H.K.; Chertow, B.S. Vitamin A deficiency impairs fetal islet development and causes subsequent glucose intolerance in adult rats. *J. Nutr.* **2004**, *134*, 1958–1963. [[PubMed](#)]
87. Delaspre, F.; Massumi, M.; Salido, M.; Soria, B.; Ravassard, P.; Savatier, P.; Skoudy, A. Directed pancreatic acinar differentiation of mouse embryonic stem cells via embryonic signalling molecules and exocrine transcription factors. *PLoS ONE* **2013**, *8*, e54243. [[CrossRef](#)] [[PubMed](#)]
88. Trasino, S.E.; Benoit, Y.D.; Gudas, L.J. Vitamin A deficiency causes hyperglycemia and loss of pancreatic  $\beta$ -cell mass. *J. Biol. Chem.* **2015**, *290*, 1456–1473. [[CrossRef](#)] [[PubMed](#)]
89. Valdés-Ramos, R.; Guadarrama-López, A.L.; Martínez-Carrillo, B.E.; Benítez-Arciniega, A.D. Vitamins and Type 2 diabetes mellitus. *Endocr. Metab. Immune Disord. Drug Targets* **2015**, *15*, 54–63. [[CrossRef](#)] [[PubMed](#)]
90. Yang, Q.; Graham, T.E.; Mody, N.; Preitner, F.; Peroni, O.D.; Zabolotny, J.M.; Kotani, K.; Quadro, L.; Kahn, B.B. Serum retinol binding protein 4 contributes to insulin resistance in obesity and Type 2 diabetes. *Nature* **2005**, *436*, 356–362. [[CrossRef](#)] [[PubMed](#)]

91. Graham, T.E.; Yang, Q.; Blüher, M.; Hammarstedt, A.; Ciaraldi, T.P.; Henry, R.R.; Wason, C.J.; Oberbach, A.; Jansson, P.-A.; Smith, U.; et al. Retinol-binding protein 4 and insulin resistance in lean, obese, and diabetic subjects. *N. Engl. J. Med.* **2006**, *354*, 2552–2563. [[CrossRef](#)] [[PubMed](#)]
92. Sugiura, M.; Nakamura, M.; Ogawa, K.; Ikoma, Y.; Yano, M. High-serum carotenoids associated with lower risk for developing Type 2 diabetes among Japanese subjects: Mikkabi cohort study. *BMJ Open Diabetes Res. Care* **2015**, *3*, e000147. [[CrossRef](#)] [[PubMed](#)]
93. Wang, L.; Liu, S.; Manson, J.E.; Gaziano, J.M.; Buring, J.E.; Sesso, H.D. The consumption of lycopene and tomato-based food products is not associated with the risk of Type 2 diabetes in women. *J. Nutr.* **2006**, *136*, 620–625. [[PubMed](#)]
94. Sena, C.M.; Nunes, E.; Gomes, A.; Santos, M.S.; Proença, T.; Martins, M.I.; Seica, R.M. Supplementation of coenzyme Q10 and  $\alpha$ -tocopherol lowers glycosylated hemoglobin level and lipid peroxidation in pancreas of diabetic rats. *Nutr. Res.* **2008**, *28*, 113–121. [[CrossRef](#)] [[PubMed](#)]
95. Ward, N.C.; Wu, J.H.Y.; Clarke, M.W.; Puddey, I.B.; Burke, V.; Croft, K.D.; Hodgson, J.M. The effect of vitamin E on blood pressure in individuals with Type 2 diabetes: A randomized, double-blind, placebo-controlled trial. *J. Hypertens.* **2007**, *25*, 227–234. [[CrossRef](#)] [[PubMed](#)]
96. Miller, E.R.; Pastor-Barriuso, R.; Dalal, D.; Riemersma, R.A.; Appel, L.J.; Guallar, E. Meta-analysis: High-dosage vitamin E supplementation may increase all-cause mortality. *Ann. Intern. Med.* **2005**, *142*, 37–46. [[CrossRef](#)] [[PubMed](#)]
97. Xu, R.; Zhang, S.; Tao, A.; Chen, G.; Zhang, M. Influence of vitamin E supplementation on glycaemic control: A meta-analysis of randomised controlled trials. *PLoS ONE* **2014**, *9*, e95008. [[CrossRef](#)] [[PubMed](#)]
98. Lazo de la Vega-Monroy, M.L.; Larrieta, E.; German, M.S.; Baez-Saldana, A.; Fernandez-Mejia, C. Effects of biotin supplementation in the diet on insulin secretion, islet gene expression, glucose homeostasis and beta-cell proportion. *J. Nutr. Biochem.* **2013**, *24*, 169–177. [[CrossRef](#)] [[PubMed](#)]
99. Sahin, K.; Tuzcu, M.; Orhan, C.; Sahin, N.; Kucuk, O.; Ozercan, I.H.; Juturu, V.; Komorowski, J.R. Anti-diabetic activity of chromium picolinate and biotin in rats with Type 2 diabetes induced by high-fat diet and streptozotocin. *Br. J. Nutr.* **2013**, *110*, 197–205. [[CrossRef](#)] [[PubMed](#)]
100. Kaplan, B.; Gönül, B.; Erdogan, D.; Elbeg, S. Effects of limited food intake and vitamin C supplementation on pancreatic glucagon and insulin in guinea pigs. *Eur. J. Histochem.* **2007**, *51*, 137–144. [[PubMed](#)]
101. Sinclair, A.J.; Taylor, P.B.; Lunec, J.; Girling, A.J.; Barnett, A.H. Low plasma ascorbate levels in patients with Type 2 diabetes mellitus consuming adequate dietary vitamin C. *Diabet. Med.* **1994**, *11*, 893–898. [[CrossRef](#)] [[PubMed](#)]
102. Wells, W.W.; Dou, C.Z.; Dybas, L.N.; Jung, C.H.; Kalbach, H.L.; Xu, D.P. Ascorbic acid is essential for the release of insulin from scorbutic guinea pig pancreatic islets. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 11869–11873. [[CrossRef](#)] [[PubMed](#)]
103. Harding, A.-H.; Wareham, N.J.; Bingham, S.A.; Khaw, K.; Luben, R.; Welch, A.; Forouhi, N.G. Plasma vitamin C level, fruit and vegetable consumption, and the risk of new-onset Type 2 diabetes mellitus: the European prospective investigation of cancer–norfolk prospective study. *Arch. Intern. Med.* **2008**, *168*, 1493. [[CrossRef](#)] [[PubMed](#)]
104. Bergsten, P.; Moura, A.S.; Atwater, I.; Levine, M. Ascorbic acid and insulin secretion in pancreatic islets. *J. Biol. Chem.* **1994**, *269*, 1041–1045. [[PubMed](#)]
105. Eriksson, J.; Kohvakka, A. Magnesium and ascorbic acid supplementation in diabetes mellitus. *Ann. Nutr. Metab.* **1995**, *39*, 217–223. [[CrossRef](#)] [[PubMed](#)]
106. Paolisso, G.; Balbi, V.; Volpe, C.; Varricchio, G.; Gambardella, A.; Saccomanno, F.; Ammendola, S.; Varricchio, M.; D’Onofrio, F. Metabolic benefits deriving from chronic vitamin C supplementation in aged non-insulin dependent diabetics. *J. Am. Coll. Nutr.* **1995**, *14*, 387–392. [[CrossRef](#)] [[PubMed](#)]
107. Chen, H.; Karne, R.J.; Hall, G.; Campia, U.; Panza, J.A.; Cannon, R.O.; Wang, Y.; Katz, A.; Levine, M.; Quon, M.J. High-dose oral vitamin C partially replenishes vitamin C levels in patients with Type 2 diabetes and low vitamin C levels but does not improve endothelial dysfunction or insulin resistance. *Am. J. Physiol. Heart Circ. Physiol.* **2006**, *290*, H137–H145. [[CrossRef](#)] [[PubMed](#)]
108. Lee, D.-H.; Folsom, A.R.; Harnack, L.; Halliwell, B.; Jacobs, D.R. Does supplemental vitamin C increase cardiovascular disease risk in women with diabetes? *Am. J. Clin. Nutr.* **2004**, *80*, 1194–1200. [[PubMed](#)]

109. Alam, M.M.; Iqbal, S.; Naseem, I. Ameliorative effect of riboflavin on hyperglycemia, oxidative stress and DNA damage in type-2 diabetic mice: Mechanistic and therapeutic strategies. *Arch. Biochem. Biophys.* **2015**, *584*, 10–19. [[CrossRef](#)] [[PubMed](#)]
110. Zhou, S.-S.; Li, D.; Zhou, Y.-M.; Sun, W.-P.; Liu, Q.-G. B-vitamin consumption and the prevalence of diabetes and obesity among the US adults: Population based ecological study. *BMC Public Health* **2010**, *10*, 746. [[CrossRef](#)] [[PubMed](#)]
111. Cobianchi, L.; Fornoni, A.; Pileggi, A.; Molano, R.D.; Sanabria, N.Y.; Gonzalez-Quintana, J.; Bocca, N.; Marzorati, S.; Zahr, E.; Hogan, A.R.; et al. Riboflavin inhibits IL-6 expression and p38 activation in islet cells. *Cell Transplant.* **2008**, *17*, 559–566. [[CrossRef](#)] [[PubMed](#)]
112. Soria, B.; Roche, E.; Berná, G.; León-Quinto, T.; Reig, J.A.; Martín, F. Insulin-secreting cells derived from embryonic stem cells normalize glycemia in streptozotocin-induced diabetic mice. *Diabetes* **2000**, *49*, 157–162. [[CrossRef](#)] [[PubMed](#)]
113. León-Quinto, T.; Jones, J.; Skoudy, A.; Burcin, M.; Soria, B. In vitro directed differentiation of mouse embryonic stem cells into insulin-producing cells. *Diabetologia* **2004**, *47*, 1442–1451. [[CrossRef](#)] [[PubMed](#)]
114. Vaca, P.; Berná, G.; Araujo, R.; Carneiro, E.M.; Bedoya, F.J.; Soria, B.; Martín, F. Nicotinamide induces differentiation of embryonic stem cells into insulin-secreting cells. *Exp. Cell Res.* **2008**, *314*, 969–974. [[CrossRef](#)] [[PubMed](#)]
115. Ye, D.Z.; Tai, M.-H.; Linning, K.D.; Szabo, C.; Olson, L.K. MafA expression and insulin promoter activity are induced by nicotinamide and related compounds in INS-1 pancreatic beta-cells. *Diabetes* **2006**, *55*, 742–750. [[CrossRef](#)] [[PubMed](#)]
116. Martín, F.; Soria, B. Amino acid-induced [Ca<sup>2+</sup>]<sub>i</sub> oscillations in single mouse pancreatic islets of Langerhans. *J. Physiol.* **1995**, 361–371. [[CrossRef](#)]
117. Bolea, S.; Pertusa, J.A.; Martín, F.; Sanchez-Andrés, J.V.; Soria, B. Regulation of pancreatic beta-cell electrical activity and insulin release by physiological amino acid concentrations. *Pflugers Arch.* **1997**, *433*, 699–704. [[CrossRef](#)] [[PubMed](#)]
118. De Oliveira, C.A.M.; Latorraca, M.Q.; de Mello, M.A.R.; Carneiro, E.M. Mechanisms of insulin secretion in malnutrition: Modulation by amino acids in rodent models. *Amino Acids* **2011**, *40*, 1027–1034. [[CrossRef](#)] [[PubMed](#)]
119. Menni, C.; Fauman, E.; Erte, I.; Perry, J.R.B.; Kastenmüller, G.; Shin, S.-Y.; Petersen, A.-K.; Hyde, C.; Psatha, M.; Ward, K.J.; et al. Biomarkers for Type 2 diabetes and impaired fasting glucose using a nontargeted metabolomics approach. *Diabetes* **2013**, *62*, 4270–4276. [[CrossRef](#)] [[PubMed](#)]
120. Imam, M.U.; Ismail, M. Nutrigenomic effects of germinated brown rice and its bioactives on hepatic gluconeogenic genes in type 2 diabetic rats and HEPG2 cells. *Mol. Nutr. Food Res.* **2013**, *57*, 401–411. [[CrossRef](#)] [[PubMed](#)]
121. Azzout-Marniche, D.; Gaudichon, C.; Tomé, D. Dietary protein and blood glucose control. *Curr. Opin. Clin. Nutr. Metab. Care* **2014**, *17*, 349–354. [[CrossRef](#)] [[PubMed](#)]
122. Healy, N.P.; Kirwan, A.M.; McArdle, M.A.; Holohan, K.; Nongonierma, A.B.; Keane, D.; Kelly, S.; Celkova, L.; Lyons, C.L.; McGillicuddy, F.C.; et al. A casein hydrolysate protects mice against high fat diet induced hyperglycemia by attenuating NLRP3 inflammasome-mediated inflammation and improving insulin signalling. *Mol. Nutr. Food Res.* **2016**, *60*, 2421–2432. [[CrossRef](#)] [[PubMed](#)]
123. Gannon, M.C.; Nuttall, J.A.; Damberg, G.; Gupta, V.; Nuttall, F.Q. Effect of Protein Ingestion on the Glucose Appearance Rate in People with Type 2 diabetes. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 1040–1047. [[CrossRef](#)] [[PubMed](#)]
124. Manders, R.J.F.; Hansen, D.; Zorenc, A.H.G.; Dendale, P.; Kloek, J.; Saris, W.H.M.; van Loon, L.J.C. Protein co-ingestion strongly increases postprandial insulin secretion in Type 2 diabetes patients. *J. Med. Food* **2014**, *17*, 758–763. [[CrossRef](#)] [[PubMed](#)]
125. Gijsbers, L.; Ding, E.L.; Malik, V.S.; de Goede, J.; Geleijnse, J.M.; Soedamah-Muthu, S.S. Consumption of dairy foods and diabetes incidence: A dose-response meta-analysis of observational studies. *Am. J. Clin. Nutr.* **2016**, *103*, 1111–1124. [[CrossRef](#)] [[PubMed](#)]
126. Gao, D.; Ning, N.; Wang, C.; Wang, Y.; Li, Q.; Meng, Z.; Liu, Y.; Li, Q. Dairy products consumption and risk of Type 2 diabetes: Systematic review and dose-response meta-analysis. *PLoS ONE* **2013**, *8*, e73965. [[CrossRef](#)] [[PubMed](#)]

127. Comerford, K.B.; Pasin, G. Emerging evidence for the importance of dietary protein source on glucoregulatory markers and Type 2 diabetes: Different effects of dairy, meat, fish, egg, and plant protein foods. *Nutrients* **2016**, *8*, 446. [[CrossRef](#)] [[PubMed](#)]
128. Bifari, F.; Nisoli, E. Branched-chain amino acids differently modulate catabolic and anabolic states in mammals: A pharmacological point of view. *Br. J. Pharmacol.* **2017**, *174*, 1366–1377. [[CrossRef](#)] [[PubMed](#)]
129. Lian, K.; Du, C.; Liu, Y.; Zhu, D.; Yan, W.; Zhang, H.; Hong, Z.; Liu, P.; Zhang, L.; Pei, H.; et al. Impaired adiponectin signalling contributes to disturbed catabolism of branched-chain amino acids in diabetic mice. *Diabetes* **2015**, *64*, 49–59. [[CrossRef](#)] [[PubMed](#)]
130. Floegel, A.; Stefan, N.; Yu, Z.; Muehlenbruch, K.; Drogan, D.; Joost, H.-G.; Fritsche, A.; Haring, H.-U.; Hrabce de Angelis, M.; Peters, A.; et al. Identification of serum metabolites associated with risk of Type 2 diabetes using a targeted metabolomic approach. *Diabetes* **2013**, *62*, 639–648. [[CrossRef](#)] [[PubMed](#)]
131. Newgard, C.B.; An, J.; Bain, J.R.; Muehlbauer, M.J.; Stevens, R.D.; Lien, L.F.; Haqq, A.M.; Shah, S.H.; Arlotto, M.; Slentz, C.A.; et al. A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metab.* **2009**, *9*, 311–326. [[CrossRef](#)] [[PubMed](#)]
132. Lee, C.C.; Watkins, S.M.; Lorenzo, C.; Wagenknecht, L.E.; Il'yasova, D.; Chen, Y.-D.I.; Haffner, S.M.; Hanley, A.J. Branched-chain amino acids and insulin metabolism: The insulin resistance atherosclerosis study (IRAS). *Diabetes Care* **2016**, *39*, 582–588. [[CrossRef](#)] [[PubMed](#)]
133. Giesbertz, P.; Daniel, H. Branched-chain amino acids as biomarkers in diabetes. *Curr. Opin. Clin. Nutr. Metab. Care* **2016**, *19*, 48–54. [[CrossRef](#)] [[PubMed](#)]
134. Lynch, C.J.; Adams, S.H. Branched-chain amino acids in metabolic signalling and insulin resistance. *Nat. Rev. Endocrinol.* **2014**, *10*, 723–736. [[CrossRef](#)] [[PubMed](#)]
135. Batch, B.C.; Hyland, K.; Svetkey, L.P. Branch chain amino acids: Biomarkers of health and disease. *Curr. Opin. Clin. Nutr. Metab. Care* **2014**, *17*, 86–89. [[CrossRef](#)] [[PubMed](#)]
136. Langenberg, C.; Savage, D.B. An amino acid profile to predict diabetes? *Nat. Med.* **2011**, *17*, 418–420. [[CrossRef](#)] [[PubMed](#)]
137. Yang, Y.; Wu, Z.; Meininger, C.J.; Wu, G. l-Leucine and NO-mediated cardiovascular function. *Amino Acids* **2015**, *47*, 435–447. [[CrossRef](#)] [[PubMed](#)]
138. Kwon, G.; Marshall, C.A.; Pappan, K.L.; Remedi, M.S.; McDaniel, M.L. Signalling elements involved in the metabolic regulation of mTOR by nutrients, incretins, and growth factors in islets. *Diabetes* **2004**, *53* (Suppl. S3), S225–S232. [[CrossRef](#)] [[PubMed](#)]
139. Xu, G.; Kwon, G.; Cruz, W.S.; Marshall, C.A.; McDaniel, M.L. Metabolic regulation by leucine of translation initiation through the mTOR-signalling pathway by pancreatic beta-cells. *Diabetes* **2001**, *50*, 353–360. [[CrossRef](#)] [[PubMed](#)]
140. Dickson, L.M.; Rhodes, C.J. Pancreatic beta-cell growth and survival in the onset of Type 2 diabetes: A role for protein kinase B in the Akt? *AJP Endocrinol. Metab.* **2004**, *287*, E192–E198. [[CrossRef](#)] [[PubMed](#)]
141. Liu, Z.; Jeppesen, P.B.; Gregersen, S.; Chen, X.; Hermansen, K. Dose- and glucose-dependent effects of amino acids on insulin secretion from isolated mouse islets and clonal INS-1E beta-cells. *Rev. Diabet. Stud.* **2008**, *5*, 232–244. [[CrossRef](#)] [[PubMed](#)]
142. Yoon, M.-S. The Emerging Role of Branched-chain amino acids in insulin resistance and metabolism. *Nutrients* **2016**, *8*, 405. [[CrossRef](#)] [[PubMed](#)]
143. Jang, C.; Oh, S.F.; Wada, S.; Rowe, G.C.; Liu, L.; Chan, M.C.; Rhee, J.; Hoshino, A.; Kim, B.; Ibrahim, A.; et al. A branched-chain amino acid metabolite drives vascular fatty acid transport and causes insulin resistance. *Nat. Med.* **2016**, *22*, 421–426. [[CrossRef](#)] [[PubMed](#)]
144. Leibowitz, G.; Cerasi, E.; Ketzinel-Gilad, M. The role of mTOR in the adaptation and failure of beta-cells in Type 2 diabetes. *Diabetes. Obes. Metab.* **2008**, *10* (Suppl. S4), 157–169. [[CrossRef](#)] [[PubMed](#)]
145. Piccolo, B.D.; Graham, J.L.; Stanhope, K.L.; Fiehn, O.; Havel, P.J.; Adams, S.H. Plasma amino acid and metabolite signatures tracking diabetes progression in the UCD-T2DM rat model. *Am. J. Physiol. Endocrinol. Metab.* **2016**, *310*, E958–E969. [[CrossRef](#)] [[PubMed](#)]
146. Wang-Sattler, R.; Yu, Z.; Herder, C.; Messias, A.C.; Floegel, A.; He, Y.; Heim, K.; Campillos, M.; Holzapfel, C.; Thorand, B.; et al. Novel biomarkers for pre-diabetes identified by metabolomics. *Mol. Syst. Biol.* **2012**, *8*, 615. [[CrossRef](#)] [[PubMed](#)]

147. Würtz, P.; Mäkinen, V.-P.; Soininen, P.; Kangas, A.J.; Tukiainen, T.; Kettunen, J.; Savolainen, M.J.; Tammelin, T.; Viikari, J.S.; Rönnemaa, T.; et al. Metabolic signatures of insulin resistance in 7098 young adults. *Diabetes* **2012**, *61*, 1372–1380. [[PubMed](#)]
148. Klein, M.S.; Shearer, J. Metabolomics and Type 2 diabetes: Translating basic research into clinical application. *J. Diabetes Res.* **2016**, *2016*, 1–10. [[CrossRef](#)] [[PubMed](#)]
149. Sekhar, R.V.; McKay, S.V.; Patel, S.G.; Guthikonda, A.P.; Reddy, V.T.; Balasubramanyam, A.; Jahoor, F. Glutathione synthesis is diminished in patients with uncontrolled diabetes and restored by dietary supplementation with cysteine and glycine. *Diabetes Care* **2011**, *34*, 162–167. [[CrossRef](#)] [[PubMed](#)]
150. Lustgarten, M.S.; Price, L.L.; Phillips, E.M.; Fielding, R.A. Serum glycine is associated with regional body fat and insulin resistance in functionally-limited older adults. *PLoS ONE* **2013**, *8*, e84034. [[CrossRef](#)] [[PubMed](#)]
151. Rorsman, P.; Braun, M. Regulation of Insulin Secretion in Human Pancreatic Islets. *Annu. Rev. Physiol.* **2013**, *75*, 155–179. [[CrossRef](#)] [[PubMed](#)]
152. Ha, K.-S.; Jo, S.-H.; Kang, B.-H.; Apostolidis, E.; Lee, M.S.; Jang, H.-D.; Kwon, Y.-I. In vitro and in vivo antihyperglycemic effect of 2 amadori rearrangement compounds, arginyl-fructose and arginyl-fructosyl-glucose. *J. Food Sci.* **2011**, *76*, H188–H193. [[CrossRef](#)] [[PubMed](#)]
153. Lee, K.-H.; Ha, K.-S.; Jo, S.-H.; Lee, C.M.; Kim, Y.-C.; Chung, K.-H.; Kwon, Y.-I. Effect of long-term dietary arginyl-fructose (AF) on hyperglycemia and HbA1c in diabetic db/db mice. *Int. J. Mol. Sci.* **2014**, *15*, 8352–8359. [[CrossRef](#)] [[PubMed](#)]
154. Park, S.E.; Kim, O.-H.; Kwak, J.H.; Lee, K.-H.; Kwon, Y.-I.; Chung, K.H.; Lee, J.H. Antihyperglycemic effect of short-term arginyl-fructose supplementation in subjects with prediabetes and newly diagnosed Type 2 diabetes: Randomized, double-blinded, placebo-controlled trial. *Trials* **2015**, *16*, 521. [[CrossRef](#)] [[PubMed](#)]
155. De Souza, R.J.; Mente, A.; Maroleanu, A.; Cozma, A.I.; Ha, V.; Kishibe, T.; Uleryk, E.; Budyłowski, P.; Schönemann, H.; Beyene, J.; et al. Intake of saturated and trans unsaturated fatty acids and risk of all cause mortality, cardiovascular disease, and Type 2 diabetes: Systematic review and meta-analysis of observational studies. *BMJ* **2015**, *351*, h3978. [[CrossRef](#)] [[PubMed](#)]
156. Morio, B.; Fardet, A.; Legrand, P.; Lecerf, J.-M. Involvement of dietary saturated fats, from all sources or of dairy origin only, in insulin resistance and Type 2 diabetes. *Nutr. Rev.* **2016**, *74*, 33–47. [[CrossRef](#)] [[PubMed](#)]
157. Zhou, Y.; Tian, C.; Jia, C. Association of fish and n-3 fatty acid intake with the risk of Type 2 diabetes: A meta-analysis of prospective studies. *Br. J. Nutr.* **2012**, *108*, 408–417. [[CrossRef](#)] [[PubMed](#)]
158. Lindström, J.; Ilanne-Parikka, P.; Peltonen, M.; Aunola, S.; Eriksson, J.G.; Hemio, K.; Hämäläinen, H.; Härkönen, P.; Keinänen-Kiukkaanniemi, S.; Laakso, M.; et al. Sustained reduction in the incidence of Type 2 diabetes by lifestyle intervention: Follow-up of the Finnish Diabetes Prevention Study. *Lancet* **2006**, *368*, 1673–1679. [[CrossRef](#)]
159. Wallin, A.; Di Giuseppe, D.; Orsini, N.; Patel, P.S.; Forouhi, N.G.; Wolk, A. Fish consumption, dietary long-chain n-3 fatty acids, and risk of Type 2 diabetes: Systematic review and meta-analysis of prospective studies. *Diabetes Care* **2012**, *35*, 918–929. [[CrossRef](#)] [[PubMed](#)]
160. Yu, C.; Chen, Y.; Cline, G.W.; Zhang, D.; Zong, H.; Wang, Y.; Bergeron, R.; Kim, J.K.; Cushman, S.W.; Cooney, G.J.; et al. Mechanism by which fatty acids inhibit insulin activation of insulin receptor substrate-1 (IRS-1)-associated phosphatidylinositol 3-kinase activity in muscle. *J. Biol. Chem.* **2002**, *277*, 50230–50236. [[CrossRef](#)] [[PubMed](#)]
161. Qiu, L.; List, E.O.; Kopchick, J.J. Differentially expressed proteins in the pancreas of diet-induced diabetic mice. *Mol. Cell. Proteom.* **2005**, *4*, 1311–1318. [[CrossRef](#)] [[PubMed](#)]
162. Ritz-Laser, B.; Meda, P.; Constant, I.; Klages, N.; Charollais, A.; Morales, A.; Magnan, C.; Ktorza, A.; Philippe, J. Glucose-induced preproinsulin gene expression is inhibited by the free fatty acid palmitate. *Endocrinology* **1999**, *140*, 4005–4014. [[CrossRef](#)] [[PubMed](#)]
163. Poutout, V.; Hagman, D.; Stein, R.; Artner, I.; Robertson, R.P.; Harmon, J.S. Regulation of the insulin gene by glucose and fatty acids. *J. Nutr.* **2006**, *136*, 873–876. [[PubMed](#)]
164. Hagman, D.K.; Hays, L.B.; Parazzoli, S.D.; Poutout, V. Palmitate inhibits insulin gene expression by altering PDX-1 nuclear localization and reducing MafA expression in isolated rat islets of Langerhans. *J. Biol. Chem.* **2005**, *280*, 32413–32418. [[CrossRef](#)] [[PubMed](#)]
165. Yoshikawa, H.; Tajiri, Y.; Sako, Y.; Hashimoto, T.; Umeda, F.; Nawata, H. Effects of free fatty acids on  $\beta$ -cell functions: A possible involvement of peroxisome proliferator-activated receptors  $\alpha$  or pancreatic/duodenal homeobox. *Metabolism* **2001**, *50*, 613–618. [[CrossRef](#)] [[PubMed](#)]

166. Moore, P.C.; Ugas, M.A.; Hagman, D.K.; Parazzoli, S.D.; Poitout, V. Evidence against the involvement of oxidative stress in fatty acid inhibition of insulin secretion. *Diabetes* **2004**, *53*, 2610–2616. [[CrossRef](#)] [[PubMed](#)]
167. Kelpe, C.L.; Moore, P.C.; Parazzoli, S.D.; Wicksteed, B.; Rhodes, C.J.; Poitout, V. Palmitate inhibition of insulin gene expression is mediated at the transcriptional level via ceramide synthesis. *J. Biol. Chem.* **2003**, *278*, 30015–30021. [[CrossRef](#)] [[PubMed](#)]
168. Wen, H.; Gris, D.; Lei, Y.; Jha, S.; Zhang, L.; Huang, M.T.-H.; Brickey, W.J.; Ting, J.P.-Y. Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signalling. *Nat. Immunol.* **2011**, *12*, 408–415. [[CrossRef](#)] [[PubMed](#)]
169. Finucane, O.M.; Lyons, C.L.; Murphy, A.M.; Reynolds, C.M.; Klinger, R.; Healy, N.P.; Cooke, A.A.; Coll, R.C.; McAllan, L.; Nilaweera, K.N.; et al. Monounsaturated fatty acid-enriched high-fat diets impede adipose NLRP3 inflammasome-mediated IL-1 $\beta$  secretion and insulin resistance despite obesity. *Diabetes* **2015**, *64*, 2116–2128. [[CrossRef](#)] [[PubMed](#)]
170. Jurado-Ruiz, E.; Varela, L.M.; Luque, A.; Berná, G.; Cahuana, G.; Martinez-Force, E.; Gallego-Durán, R.; Soria, B.; de Roos, B.; Romero Gómez, M.; et al. An extra virgin olive oil rich diet intervention ameliorates the nonalcoholic steatohepatitis induced by a high-fat "Western-type" diet in mice. *Mol. Nutr. Food Res.* **2017**, *61*, 1600549. [[CrossRef](#)] [[PubMed](#)]
171. Mancini, A.; Imperlini, E.; Nigro, E.; Montagnese, C.; Daniele, A.; Orrù, S.; Buono, P. Biological and Nutritional Properties of Palm Oil and Palmitic Acid: Effects on Health. *Molecules* **2015**, *20*, 17339–17361. [[CrossRef](#)] [[PubMed](#)]
172. Davis, J.; Gabler, N.; Walker-Daniels, J.; Spurlock, M. The c-Jun N-terminal kinase mediates the induction of oxidative stress and insulin resistance by palmitate and Toll-like receptor 2 and 4 ligands in 3T3-L1 adipocytes. *Horm. Metab. Res.* **2009**, *41*, 523–530. [[CrossRef](#)] [[PubMed](#)]
173. Legrand-Poels, S.; Esser, N.; L'homme, L.; Scheen, A.; Paquot, N.; Piette, J. Free fatty acids as modulators of the NLRP3 inflammasome in obesity/Type 2 diabetes. *Biochem. Pharmacol.* **2014**, *92*, 131–141. [[CrossRef](#)] [[PubMed](#)]
174. Marchetti, P.; Bugliani, M.; Boggi, U.; Masini, M.; Marselli, L. The pancreatic beta cells in human Type 2 diabetes. *Adv. Exp. Med. Biol.* **2012**, *771*, 288–309. [[PubMed](#)]
175. Rosqvist, F.; Iggman, D.; Kullberg, J.; Cedernaes, J.; Johansson, H.-E.; Larsson, A.; Johansson, L.; Ahlström, H.; Arner, P.; Dahlman, I.; et al. Overfeeding polyunsaturated and saturated fat causes distinct effects on liver and visceral fat accumulation in humans. *Diabetes* **2014**, *63*, 2356–2368. [[CrossRef](#)] [[PubMed](#)]
176. Vessby, B.; Uusitupa, M.; Hermansen, K.; Riccardi, G.; Rivellese, A.A.; Tapsell, L.C.; Näslén, C.; Berglund, L.; Louheranta, A.; Rasmussen, B.M.; et al. Substituting dietary saturated for monounsaturated fat impairs insulin sensitivity in healthy men and women: The KANWU Study. *Diabetologia* **2001**, *44*, 312–319. [[CrossRef](#)] [[PubMed](#)]
177. Jebb, S.A.; Lovegrove, J.A.; Griffin, B.A.; Frost, G.S.; Moore, C.S.; Chatfield, M.D.; Bluck, L.J.; Williams, C.M.; Sanders, T.A.; RISCK Study Group. Effect of changing the amount and type of fat and carbohydrate on insulin sensitivity and cardiovascular risk: The RISCK (Reading, Imperial, Surrey, Cambridge, and Kings) trial. *Am. J. Clin. Nutr.* **2010**, *92*, 748–758. [[CrossRef](#)] [[PubMed](#)]
178. Forouhi, N.G.; Koulman, A.; Sharp, S.J.; Imamura, F.; Kröger, J.; Schulze, M.B.; Crowe, F.L.; Huerta, J.M.; Guevara, M.; Beulens, J.W.; et al. Differences in the prospective association between individual plasma phospholipid saturated fatty acids and incident Type 2 diabetes: The EPIC-InterAct case-cohort study. *Lancet Diabetes Endocrinol.* **2014**, *2*, 810–818. [[CrossRef](#)]
179. Pérez-Martínez, P.; García-Ríos, A.; Delgado-Lista, J.; Pérez-Jiménez, F.; López-Miranda, J. Mediterranean diet rich in olive oil and obesity, metabolic syndrome and diabetes mellitus. *Curr. Pharm. Des.* **2011**, *17*, 769–777. [[CrossRef](#)] [[PubMed](#)]
180. Van Deursen, D.; van Leeuwen, M.; Akdogan, D.; Adams, H.; Jansen, H.; Verhoeven, A.J.M. Activation of hepatic lipase expression by oleic acid: Possible involvement of USF1. *Nutrients* **2009**, *1*, 133–147. [[CrossRef](#)] [[PubMed](#)]
181. Guo, X.; Li, H.; Xu, H.; Halim, V.; Zhang, W.; Wang, H.; Ong, K.T.; Woo, S.-L.; Walzem, R.L.; Mashek, D.G.; et al. Palmitoleate induces hepatic steatosis but suppresses liver inflammatory response in mice. *PLoS ONE* **2012**, *7*, e39286. [[CrossRef](#)] [[PubMed](#)]
182. Nunes, E.A.; Rafacho, A. Implications of palmitoleic acid (palmitoleate) on glucose homeostasis, insulin resistance and diabetes. *Curr. Drug Targets* **2015**, *18*, 619–628. [[CrossRef](#)]

183. Wang, L.; Folsom, A.R.; Zheng, Z.-J.; Pankow, J.S.; Eckfeldt, J.H.; ARIC Study Investigators. Plasma fatty acid composition and incidence of diabetes in middle-aged adults: The Atherosclerosis Risk in Communities (ARIC) Study. *Am. J. Clin. Nutr.* **2003**, *78*, 91–98. [[PubMed](#)]
184. Calder, P.C. Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. *Biochim. Biophys. Acta* **2015**, *1851*, 469–484. [[CrossRef](#)] [[PubMed](#)]
185. Lin, N.; Shi, J.-J.; Li, Y.-M.; Zhang, X.-Y.; Chen, Y.; Calder, P.C.; Tang, L.-J. What is the impact of n-3 PUFAs on inflammation markers in Type 2 diabetic mellitus populations?: A systematic review and meta-analysis of randomized controlled trials. *Lipids Health Dis.* **2016**, *15*, 133. [[CrossRef](#)] [[PubMed](#)]
186. Derosa, G.; Cicero, A.F.G.; D'Angelo, A.; Borghi, C.; Maffioli, P. Effects of n-3 pufas on fasting plasma glucose and insulin resistance in patients with impaired fasting glucose or impaired glucose tolerance. *Biofactors* **2016**, *42*, 316–322. [[PubMed](#)]
187. Chen, C.; Yu, X.; Shao, S. Effects of omega-3 fatty acid supplementation on glucose control and lipid levels in Type 2 diabetes: A meta-analysis. *PLoS ONE* **2015**, *10*, e0139565. [[CrossRef](#)] [[PubMed](#)]
188. Yan, Y.; Jiang, W.; Spinetti, T.; Tardivel, A.; Castillo, R.; Bourquin, C.; Guarda, G.; Tian, Z.; Tschopp, J.; Zhou, R. Omega-3 fatty acids prevent inflammation and metabolic disorder through inhibition of NLRP3 inflammasome activation. *Immunity* **2013**, *38*, 1154–1163. [[CrossRef](#)] [[PubMed](#)]
189. Kato, T.; Shimano, H.; Yamamoto, T.; Ishikawa, M.; Kumadaki, S.; Matsuzaka, T.; Nakagawa, Y.; Yahagi, N.; Nakakuki, M.; Hastay, A.H.; et al. Palmitate impairs and eicosapentaenoate restores insulin secretion through regulation of SREBP-1c in pancreatic islets. *Diabetes* **2008**, *57*, 2382–2392. [[CrossRef](#)] [[PubMed](#)]
190. Pinel, A.; Morio-Liondore, B.; Capel, F. n-3 Polyunsaturated fatty acids modulate metabolism of insulin-sensitive tissues: Implication for the prevention of Type 2 diabetes. *J. Physiol. Biochem.* **2014**, *70*, 647–658. [[CrossRef](#)] [[PubMed](#)]
191. Risérus, U.; Willett, W.C.; Hu, F.B. Dietary fats and prevention of Type 2 diabetes. *Prog. Lipid Res.* **2009**, *48*, 44–51. [[CrossRef](#)] [[PubMed](#)]
192. Hennig, B.; Toborek, M.; Joshi-Barve, S.; Barger, S.W.; Barve, S.; Mattson, M.P.; McClain, C.J. Linoleic acid activates nuclear transcription factor-kappa B (NF-κB) and induces NF-κB-dependent transcription in cultured endothelial cells. *Am. J. Clin. Nutr.* **1996**, *63*, 322–328. [[PubMed](#)]
193. Pischon, T.; Hankinson, S.E.; Hotamisligil, G.S.; Rifai, N.; Willett, W.C.; Rimm, E.B. Habitual dietary intake of n-3 and n-6 fatty acids in relation to inflammatory markers among US men and women. *Circulation* **2003**, *108*, 155–160. [[CrossRef](#)] [[PubMed](#)]
194. Calder, P.C. Functional roles of fatty acids and their effects on human health. *J. Parenter. Enter. Nutr.* **2015**, *39*, 18S–32S. [[CrossRef](#)] [[PubMed](#)]
195. Welter, D.; MacArthur, J.; Morales, J.; Burdett, T.; Hall, P.; Junkins, H.; Klemm, A.; Flicek, P.; Manolio, T.; Hindorf, L.; et al. The NHGRI GWAS Catalog, a curated resource of SNP-trait associations. *Nucleic Acids Res.* **2014**, *42*, D1001–D1006. [[CrossRef](#)] [[PubMed](#)]
196. Kaul, N.; Ali, S. Genes, Genetics, and Environment in Type 2 diabetes: Implication in Personalized Medicine. *DNA Cell Biol.* **2016**, *35*, 1–12. [[CrossRef](#)] [[PubMed](#)]
197. Cornelis, M.C.; Hu, F.B. Gene-environment interactions in the development of Type 2 diabetes: Recent progress and continuing challenges. *Annu. Rev. Nutr.* **2012**, *32*, 245–259. [[CrossRef](#)] [[PubMed](#)]
198. Grant, S.F.A.; Thorleifsson, G.; Reynisdottir, I.; Benediktsson, R.; Manolescu, A.; Sainz, J.; Helgason, A.; Stefansson, H.; Emilsson, V.; Helgadóttir, A.; et al. Variant of transcription factor 7-like 2 (TCF7L2) gene confers risk of Type 2 diabetes. *Nat. Genet.* **2006**, *38*, 320–323. [[CrossRef](#)] [[PubMed](#)]
199. Zeggini, E.; McCarthy, M.I. TCF7L2: The biggest story in diabetes genetics since HLA? *Diabetologia* **2007**, *50*, 1–4. [[CrossRef](#)] [[PubMed](#)]
200. Chandak, G.R.; Janipalli, C.S.; Bhaskar, S.; Kulkarni, S.R.; Mohankrishna, P.; Hattersley, A.T.; Frayling, T.M.; Yajnik, C.S. Common variants in the TCF7L2 gene are strongly associated with Type 2 diabetes mellitus in the Indian population. *Diabetologia* **2006**, *50*, 63–67. [[CrossRef](#)] [[PubMed](#)]
201. Horikoshi, M.; Hara, K.; Ito, C.; Nagai, R.; Froguel, P.; Kadowaki, T. A genetic variation of the transcription factor 7-like 2 gene is associated with risk of Type 2 diabetes in the Japanese population. *Diabetologia* **2007**, *50*, 747–751. [[CrossRef](#)] [[PubMed](#)]

202. Lehman, D.M.; Hunt, K.J.; Leach, R.J.; Hamlington, J.; Arya, R.; Abboud, H.E.; Duggirala, R.; Blangero, J.; Göring, H.H.H.; Stern, M.P. Haplotypes of transcription factor 7-like 2 (TCF7L2) gene and its upstream region are associated with Type 2 diabetes and age of onset in Mexican Americans. *Diabetes* **2007**, *56*, 389–393. [[CrossRef](#)] [[PubMed](#)]
203. Dou, H.; Ma, E.; Yin, L.; Jin, Y.; Wang, H. The association between gene polymorphism of TCF7L2 and Type 2 diabetes in Chinese Han population: A meta-analysis. *PLoS ONE* **2013**, *8*, e59495. [[CrossRef](#)] [[PubMed](#)]
204. Yako, Y.Y.; Guewo-Fokeng, M.; Balti, E.V.; Bouatia-Naji, N.; Matsha, T.E.; Sobngwi, E.; Erasmus, R.T.; Echouffo-Tcheugui, J.B.; Kengne, A.P. Genetic risk of Type 2 diabetes in populations of the African continent: A systematic review and meta-analyses. *Diabetes Res. Clin. Pract.* **2016**, *114*, 136–150. [[CrossRef](#)] [[PubMed](#)]
205. Hindy, G.; Mollet, I.G.; Rukh, G.; Ericson, U.; Orho-Melander, M. Several Type 2 diabetes-associated variants in genes annotated to WNT signalling interact with dietary fibre in relation to incidence of Type 2 diabetes. *Genes Nutr.* **2016**, *11*, 6. [[CrossRef](#)] [[PubMed](#)]
206. Hindy, G.; Sonestedt, E.; Ericson, U.; Jing, X.-J.; Zhou, Y.; Hansson, O.; Renström, E.; Wirfält, E.; Orho-Melander, M. Role of TCF7L2 risk variant and dietary fibre intake on incident Type 2 diabetes. *Diabetologia* **2012**, *55*, 2646–2654. [[CrossRef](#)] [[PubMed](#)]
207. InterAct Consortium. Investigation of gene-diet interactions in the incretin system and risk of Type 2 diabetes: The EPIC-InterAct study. *Diabetologia* **2016**, *59*, 2613–2621.
208. Chimienti, F. Zinc, pancreatic islet cell function and diabetes: New insights into an old story. *Nutr. Res. Rev.* **2013**, *26*, 1–11. [[CrossRef](#)] [[PubMed](#)]
209. Rutter, G.A.; Chimienti, F. SLC30A8 mutations in Type 2 diabetes. *Diabetologia* **2015**, *58*, 31–36. [[CrossRef](#)] [[PubMed](#)]
210. Patel, C.J.; Chen, R.; Kodama, K.; Ioannidis, J.P.A.; Butte, A.J. Systematic identification of interaction effects between genome- and environment-wide associations in Type 2 diabetes mellitus. *Hum. Genet.* **2013**, *132*, 495–508. [[CrossRef](#)] [[PubMed](#)]
211. Kanoni, S.; Nettleton, J.A.; Hivert, M.-F.; Ye, Z.; van Rooij, F.J.A.; Shungin, D.; Sonestedt, E.; Ngwa, J.S.; Wojczynski, M.K.; Lemaitre, R.N.; et al. Total zinc intake may modify the glucose-raising effect of a zinc transporter (SLC30A8) variant: A 14-cohort meta-analysis. *Diabetes* **2011**, *60*, 2407–2416. [[CrossRef](#)] [[PubMed](#)]
212. Sonestedt, E.; Lyssenko, V.; Ericson, U.; Gullberg, B.; Wirfält, E.; Groop, L.; Orho-Melander, M. Genetic variation in the glucose-dependent insulinotropic polypeptide receptor modifies the association between carbohydrate and fat intake and risk of Type 2 diabetes in the Malmo Diet and Cancer cohort. *J. Clin. Endocrinol. Metab.* **2012**, *97*, E810–E818. [[CrossRef](#)] [[PubMed](#)]
213. Nettleton, J.A.; McKeown, N.M.; Kanoni, S.; Lemaitre, R.N.; Hivert, M.-F.; Ngwa, J.; van Rooij, F.J.A.; Sonestedt, E.; Wojczynski, M.K.; Ye, Z.; et al. Interactions of dietary whole-grain intake with fasting glucose- and insulin-related genetic loci in individuals of European descent: A meta-analysis of 14 cohort studies. *Diabetes Care* **2010**, *33*, 2684–2691. [[CrossRef](#)] [[PubMed](#)]
214. Blanco-Rojo, R.; Delgado-Lista, J.; Lee, Y.-C.; Lai, C.-Q.; Perez-Martinez, P.; Rangel-Zuñiga, O.; Smith, C.E.; Hidalgo, B.; Alcalá-Díaz, J.F.; Gomez-Delgado, F.; et al. Interaction of an S100A9 gene variant with saturated fat and carbohydrates to modulate insulin resistance in 3 populations of different ancestries. *Am. J. Clin. Nutr.* **2016**, *104*, 508–517. [[CrossRef](#)] [[PubMed](#)]
215. Park, S.; Zhang, X.; Lee, N.R.; Jin, H.-S. TRPV1 gene polymorphisms are associated with Type 2 diabetes by their interaction with fat consumption in the Korean genome epidemiology study. *J. Nutrigenet. Nutrigenom.* **2016**, *9*, 47–61. [[CrossRef](#)] [[PubMed](#)]
216. Song, Y.; Hsu, Y.-H.; Niu, T.; Manson, J.E.; Buring, J.E.; Liu, S. Common genetic variants of the ion channel transient receptor potential membrane melastatin 6 and 7 (TRPM6 and TRPM7), magnesium intake, and risk of Type 2 diabetes in women. *BMC Med. Genet.* **2009**, *10*, 4. [[CrossRef](#)] [[PubMed](#)]
217. Cho, Y.S.; Chen, C.-H.; Hu, C.; Long, J.; Hee Ong, R.T.; Sim, X.; Takeuchi, F.; Wu, Y.; Go, M.J.; Yamauchi, T.; et al. Meta-analysis of genome-wide association studies identifies eight new loci for Type 2 diabetes in east Asians. *Nat. Genet.* **2011**, *44*, 67–72. [[CrossRef](#)] [[PubMed](#)]
218. Kaido, T.; Yebra, M.; Cirulli, V.; Rhodes, C.; Diaferia, G.; Montgomery, A.M. Impact of defined matrix interactions on insulin production by cultured human beta-cells: Effect on insulin content, secretion, and gene transcription. *Diabetes* **2006**, *55*, 2723–2729. [[CrossRef](#)] [[PubMed](#)]

219. Zheng, J.-S.; Huang, T.; Li, K.; Chen, Y.; Xie, H.; Xu, D.; Sun, J.; Li, D. Modulation of the association between the PEPD variant and the risk of Type 2 diabetes by n-3 fatty acids in Chinese Hans. *J. Nutrigenet. Nutrigenom.* **2015**, *8*, 36–43. [[CrossRef](#)] [[PubMed](#)]
220. Lamri, A.; Abi Khalil, C.; Jaziri, R.; Velho, G.; Lantieri, O.; Vol, S.; Froguel, P.; Balkau, B.; Marre, M.; Fumeron, F. Dietary fat intake and polymorphisms at the PPARG locus modulate BMI and Type 2 diabetes risk in the D.E.S.I.R. prospective study. *Int. J. Obes.* **2012**, *36*, 218–224. [[CrossRef](#)] [[PubMed](#)]
221. Hwang, J.-Y.; Park, J.E.; Choi, Y.J.; Huh, K.B.; Chang, N.; Kim, W.Y. Carbohydrate intake interacts with SNP276G>T polymorphism in the adiponectin gene to affect fasting blood glucose, HbA1C, and HDL cholesterol in Korean patients with Type 2 diabetes. *J. Am. Coll. Nutr.* **2013**, *32*, 143–150. [[CrossRef](#)] [[PubMed](#)]
222. Alsaleh, A.; Crepostnaia, D.; Maniou, Z.; Lewis, F.J.; Hall, W.L.; Sanders, T.A.B.; O'Dell, S.D.; MARINA study team. Adiponectin gene variant interacts with fish oil supplementation to influence serum adiponectin in older individuals. *J. Nutr.* **2013**, *143*, 1021–1027. [[CrossRef](#)] [[PubMed](#)]
223. Zheng, J.-S.; Arnett, D.K.; Parnell, L.D.; Smith, C.E.; Li, D.; Borecki, I.B.; Tucker, K.L.; Ordovás, J.M.; Lai, C.-Q. Modulation by dietary fat and carbohydrate of IRS1 association with Type 2 diabetes traits in two populations of different ancestries. *Diabetes Care* **2013**, *36*, 2621–2627. [[CrossRef](#)] [[PubMed](#)]
224. Zheng, J.-S.; Parnell, L.D.; Smith, C.E.; Lee, Y.-C.; Jamal-Allial, A.; Ma, Y.; Li, D.; Tucker, K.L.; Ordovás, J.M.; Lai, C.-Q. Circulating 25-hydroxyvitamin D, IRS1 variant rs2943641, and insulin resistance: Replication of a gene-nutrient interaction in 4 populations of different ancestries. *Clin. Chem.* **2014**, *60*, 186–196. [[CrossRef](#)] [[PubMed](#)]
225. Fisher, E.; Schreiber, S.; Joost, H.-G.; Boeing, H.; Döring, F. A two-step association study identifies CAV2 rs2270188 single nucleotide polymorphism interaction with fat intake in Type 2 diabetes risk. *J. Nutr.* **2011**, *141*, 177–181. [[CrossRef](#)] [[PubMed](#)]
226. Ortega-Azorín, C.; Sorlí, J.V.; Asensio, E.M.; Coltell, O.; Martínez-González, M.; Salas-Salvadó, J.; Covas, M.-I.; Arós, F.; Lapetra, J.; Serra-Majem, L.; et al. Associations of the FTO rs9939609 and the MC4R rs17782313 polymorphisms with Type 2 diabetes are modulated by diet, being higher when adherence to the Mediterranean diet pattern is low. *Cardiovasc. Diabetol.* **2012**, *11*, 137. [[CrossRef](#)] [[PubMed](#)]
227. Corella, D.; Coltell, O.; Sorlí, J.; Estruch, R.; Quiles, L.; Martínez-González, M.; Salas-Salvadó, J.; Castañer, O.; Arós, F.; Ortega-Calvo, M.; et al. Polymorphism of the transcription factor 7-like 2 gene (TCF7L2) interacts with obesity on Type-2 diabetes in the PREDIMED study emphasizing the heterogeneity of genetic variants in type-2 diabetes risk prediction: time for obesity-specific genetic risk scores. *Nutrients* **2016**, *8*, 793.
228. Dashti, H.S.; Smith, C.E.; Lee, Y.-C.; Parnell, L.D.; Lai, C.-Q.; Arnett, D.K.; Ordovás, J.M.; Garaulet, M. CRY1 circadian gene variant interacts with carbohydrate intake for insulin resistance in two independent populations: Mediterranean and North American. *Chronobiol. Int.* **2014**, *31*, 660–667. [[CrossRef](#)] [[PubMed](#)]
229. Koochakpoor, G.; Hosseini-Esfahani, F.; Daneshpour, M.S.; Hosseini, S.A.; Mirmiran, P. Effect of interactions of polymorphisms in the Melanocortin-4 receptor gene with dietary factors on the risk of obesity and Type 2 diabetes: A systematic review. *Diabet. Med.* **2016**, *33*, 1026–1034. [[CrossRef](#)] [[PubMed](#)]
230. Corella, D.; Qi, L.; Tai, E.S.; Deurenberg-Yap, M.; Tan, C.E.; Chew, S.K.; Ordovas, J.M. Perilipin gene variation determines higher susceptibility to insulin resistance in Asian women when consuming a high-saturated fat, low-carbohydrate diet. *Diabetes Care* **2006**, *29*, 1313–1319. [[CrossRef](#)] [[PubMed](#)]
231. Liu, P.; Yu, D.; Jin, X.; Li, C.; Zhu, F.; Zheng, Z.; Lv, C.; He, X. The association between the FABP2 Ala54Thr variant and the risk of Type 2 diabetes mellitus: A meta-analysis based on 11 case-control studies. *Int. J. Clin. Exp. Med.* **2015**, *8*, 5422–5429. [[PubMed](#)]
232. Gouda, H.N.; Sagoo, G.S.; Harding, A.-H.; Yates, J.; Sandhu, M.S.; Higgins, J.P.T. The association between the peroxisome proliferator-activated receptor-gamma2 (PPARG2) Pro12Ala gene variant and Type 2 diabetes mellitus: A HuGE review and meta-analysis. *Am. J. Epidemiol.* **2010**, *171*, 645–655. [[CrossRef](#)] [[PubMed](#)]
233. Fan, Y.; Wang, K.; Xu, S.; Chen, G.; Di, H.; Cao, M.; Liu, C. Association between ADIPOQ +45T>G polymorphism and Type 2 diabetes: A systematic review and meta-analysis. *Int. J. Mol. Sci.* **2014**, *16*, 704–723. [[CrossRef](#)] [[PubMed](#)]
234. Li, Q.; Qiao, Y.; Wang, C.; Zhang, G.; Zhang, X.; Xu, L. Associations between two single-nucleotide polymorphisms (rs1801278 and rs2943641) of insulin receptor substrate 1 gene and Type 2 diabetes susceptibility: A meta-analysis. *Endocrine* **2016**, *51*, 52–62. [[CrossRef](#)] [[PubMed](#)]

235. Yang, Y.; Liu, B.; Xia, W.; Yan, J.; Liu, H.-Y.; Hu, L.; Liu, S.-M. FTO genotype and Type 2 diabetes mellitus: spatial analysis and meta-analysis of 62 case-control studies from different regions. *Genes* **2017**, *8*, 70. [[CrossRef](#)] [[PubMed](#)]
236. Marcheva, B.; Ramsey, K.M.; Buhr, E.D.; Kobayashi, Y.; Su, H.; Ko, C.H.; Ivanova, G.; Omura, C.; Mo, S.; Vitaterna, M.H.; et al. Disruption of the CLOCK components CLOCK and BMAL1 leads to hypoinsulinaemia and diabetes. *Nature* **2010**, *466*, 627–631. [[CrossRef](#)] [[PubMed](#)]
237. Qi, L.; Zhang, C.; Greenberg, A.; Hu, F.B. Common variations in perilipin gene, central obesity, and risk of Type 2 diabetes in US women. *Obesity* **2008**, *16*, 1061–1065. [[CrossRef](#)] [[PubMed](#)]
238. Yu, D.; Li, C.; Xie, J.; Xu, G.; Li, Y.; Liu, J.; Chen, B.; Pan, J.; Shen, M.; Yang, L.; Hu, D. Association between three genetic variants of the Perilipin Gene (PLIN) and glucose metabolism: Results from a replication study among Chinese adults and a meta-analysis. *Endocr. Res.* **2013**, *38*, 263–279. [[CrossRef](#)] [[PubMed](#)]
239. Marín, C.; Pérez-Jiménez, F.; Gómez, P.; Delgado, J.; Paniagua, J.A.; Lozano, A.; Cortés, B.; Jiménez-Gómez, Y.; Gómez, M.J.; López-Miranda, J. The Ala54Thr polymorphism of the fatty acid-binding protein 2 gene is associated with a change in insulin sensitivity after a change in the type of dietary fat. *Am. J. Clin. Nutr.* **2005**, *82*, 196–200. [[PubMed](#)]
240. Franks, P.W.; McCarthy, M.I. Exposing the exposures responsible for Type 2 diabetes and obesity. *Science* **2016**, *354*, 69–73. [[CrossRef](#)] [[PubMed](#)]
241. Sommese, L.; Zullo, A.; Mancini, F.P.; Fabbri, R.; Soricelli, A.; Napoli, C. Clinical relevance of epigenetics in the onset and management of Type 2 diabetes mellitus. *Epigenetics* **2017**, 1–15. [[CrossRef](#)] [[PubMed](#)]
242. Yagihashi, S. Diabetes and pancreas size, does it matter? *J. Diabetes Investig.* **2016**. [[CrossRef](#)] [[PubMed](#)]
243. Rojas, A.; Carrasco, M.; Delgado, I.; Cobo, N.; Tejado, J.R.; Bedoya, F.J.; Gauthier, B.; Soria, B.; Martín, F. Signaling pathways and transcription factors involved in pancreatic islet development. In *Advances in Experimental Medicine and Biology: The Islets of Langerhans 2*; Islam, S., Ed.; Springer: Berlin, Germany, 2014; pp. 109–128.
244. Volkmar, M.; Dedeurwaerder, S.; Cunha, D.A.; Ndlovu, M.N.; Defrance, M.; Deplus, R.; Calonne, E.; Volkmar, U.; Igoillo-Esteve, M.; Naamane, N.; et al. DNA methylation profiling identifies epigenetic dysregulation in pancreatic islets from type 2 diabetic patients. *EMBO J.* **2012**, *31*, 1405–1426. [[CrossRef](#)] [[PubMed](#)]
245. Dhawan, S.; Georgia, S.; Tschen, S.-I.; Fan, G.; Bhushan, A. Pancreatic  $\beta$  cell identity is maintained by DNA methylation-mediated repression of *Arx*. *Dev. Cell* **2011**, *20*, 419–429. [[CrossRef](#)] [[PubMed](#)]
246. Hall, E.; Dayeh, T.; Kirkpatrick, C.L.; Wollheim, C.B.; Dekker Nitert, M.; Ling, C. DNA methylation of the glucagon-like peptide 1 receptor (GLP1R) in human pancreatic islets. *BMC Med. Genet.* **2013**, *14*, 76. [[CrossRef](#)] [[PubMed](#)]
247. Lenoir, O.; Flosseau, K.; Ma, F.X.; Blondeau, B.; Mai, A.; Bassel-Duby, R.; Ravassard, P.; Olson, E.N.; Haumaitre, C.; Scharfmann, R. Specific control of pancreatic endocrine  $\beta$ - and  $\delta$ -cell mass by class IIa histone deacetylases HDAC4, HDAC5, and HDAC9. *Diabetes* **2011**, *60*, 2861–2871. [[CrossRef](#)] [[PubMed](#)]
248. Tarabra, E.; Pelengaris, S.; Khan, M. A simple matter of life and death—the trials of postnatal beta-cell mass regulation. *Int. J. Endocrinol.* **2012**, *2012*, 516718. [[CrossRef](#)] [[PubMed](#)]
249. Chen, H.; Gu, X.; Su, I.; Bottino, R.; Contreras, J.L.; Tarakhovskiy, A.; Kim, S.K. Polycomb protein Ezh2 regulates pancreatic beta-cell *Ink4a/Arf* expression and regeneration in diabetes mellitus. *Genes Dev.* **2009**, *23*, 975–985. [[CrossRef](#)] [[PubMed](#)]
250. Mosley, A.L.; Corbett, J.A.; Ozcan, S. Glucose regulation of insulin gene expression requires the recruitment of p300 by the beta-cell-specific transcription factor Pdx-1. *Mol. Endocrinol.* **2004**, *18*, 2279–2290. [[CrossRef](#)] [[PubMed](#)]
251. Evans-Molina, C.; Robbins, R.D.; Kono, T.; Tersey, S.A.; Vestermarck, G.L.; Nunemaker, C.S.; Garmey, J.C.; Deering, T.G.; Keller, S.R.; Maier, B.; et al. Peroxisome proliferator-activated receptor gamma activation restores islet function in diabetic mice through reduction of endoplasmic reticulum stress and maintenance of euchromatin structure. *Mol. Cell. Biol.* **2009**, *29*, 2053–2067. [[CrossRef](#)] [[PubMed](#)]
252. Deiuliis, J.A. MicroRNAs as regulators of metabolic disease: Pathophysiologic significance and emerging role as biomarkers and therapeutics. *Int. J. Obes.* **2016**, *40*, 88–101. [[CrossRef](#)] [[PubMed](#)]

253. Jordan, S.D.; Krüger, M.; Willmes, D.M.; Redemann, N.; Wunderlich, F.T.; Brönneke, H.S.; Merkwirth, C.; Kashkar, H.; Olkkonen, V.M.; Böttger, T.; et al. Obesity-induced overexpression of miRNA-143 inhibits insulin-stimulated AKT activation and impairs glucose metabolism. *Nat. Cell Biol.* **2011**, *13*, 434–446. [[CrossRef](#)] [[PubMed](#)]
254. Zhu, H.; Shyh-Chang, N.; Segrè, A.V.; Shinoda, G.; Shah, S.P.; Einhorn, W.S.; Takeuchi, A.; Engreitz, J.M.; Hagan, J.P.; Kharas, M.G.; et al. The Lin28/let-7 axis regulates glucose metabolism. *Cell* **2011**, *147*, 81–94. [[CrossRef](#)] [[PubMed](#)]
255. Jiang, L.Q.; Franck, N.; Egan, B.; Sjögren, R.J.O.; Katayama, M.; Duque-Guimaraes, D.; Arner, P.; Zierath, J.R.; Krook, A. Autocrine role of interleukin-13 on skeletal muscle glucose metabolism in type 2 diabetic patients involves microRNA let-7. *Am. J. Physiol. Endocrinol. Metab.* **2013**, *305*, E1359–E1366. [[CrossRef](#)] [[PubMed](#)]
256. Crépin, D.; Benomar, Y.; Riffault, L.; Amine, H.; Gertler, A.; Taouis, M. The over-expression of miR-200a in the hypothalamus of ob/ob mice is linked to leptin and insulin signalling impairment. *Mol. Cell. Endocrinol.* **2014**, *384*, 1–11. [[CrossRef](#)] [[PubMed](#)]
257. Zhuang, G.; Meng, C.; Guo, X.; Cheruku, P.S.; Shi, L.; Xu, H.; Li, H.; Wang, G.; Evans, A.R.; Safe, S.; et al. A novel regulator of macrophage activation: MiR-223 in obesity-associated adipose tissue inflammation. *Circulation* **2012**, *125*, 2892–2903. [[CrossRef](#)] [[PubMed](#)]
258. He, A.; Zhu, L.; Gupta, N.; Chang, Y.; Fang, F. Overexpression of micro ribonucleic acid 29, highly up-regulated in diabetic rats, leads to insulin resistance in 3T3-L1 adipocytes. *Mol. Endocrinol.* **2007**, *21*, 2785–2794. [[CrossRef](#)] [[PubMed](#)]
259. Kurtz, C.L.; Peck, B.C.E.; Fannin, E.E.; Beysen, C.; Miao, J.; Landstreet, S.R.; Ding, S.; Turaga, V.; Lund, P.K.; Turner, S.; et al. MicroRNA-29 fine-tunes the expression of key FOXA2-activated lipid metabolism genes and is dysregulated in animal models of insulin resistance and diabetes. *Diabetes* **2014**, *63*, 3141–3148. [[CrossRef](#)] [[PubMed](#)]
260. Poy, M.N.; Eliasson, L.; Krutzfeldt, J.; Kuwajima, S.; Ma, X.; Macdonald, P.E.; Pfeffer, S.; Tuschl, T.; Rajewsky, N.; Rorsman, P.; et al. A pancreatic islet-specific microRNA regulates insulin secretion. *Nature* **2004**, *432*, 226–230. [[CrossRef](#)] [[PubMed](#)]
261. Poy, M.N.; Hausser, J.; Trajkovski, M.; Braun, M.; Collins, S.; Rorsman, P.; Zavolan, M.; Stoffel, M. miR-375 maintains normal pancreatic  $\alpha$ - and beta-cell mass. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 5813–5818. [[CrossRef](#)] [[PubMed](#)]
262. Joglekar, M.V.; Joglekar, V.M.; Hardikar, A.A. Expression of islet-specific microRNAs during human pancreatic development. *Gene Expr. Patterns* **2009**, *9*, 109–113. [[CrossRef](#)] [[PubMed](#)]
263. Poyntout, V.; Robertson, R.P. Glucolipototoxicity: Fuel excess and beta-cell dysfunction. *Endocr. Rev.* **2008**, *29*, 351–366. [[CrossRef](#)] [[PubMed](#)]
264. Franco, R.; Schoneveld, O.; Georgakilas, A.G.; Panayiotidis, M.I. Oxidative stress, DNA methylation and carcinogenesis. *Cancer Lett.* **2008**, *266*, 6–11. [[CrossRef](#)] [[PubMed](#)]
265. Yang, B.T.; Dayeh, T.A.; Kirkpatrick, C.L.; Taneera, J.; Kumar, R.; Groop, L.; Wollheim, C.B.; Nitert, M.D.; Ling, C. Insulin promoter DNA methylation correlates negatively with insulin gene expression and positively with HbA(1c) levels in human pancreatic islets. *Diabetologia* **2011**, *54*, 360–367. [[CrossRef](#)] [[PubMed](#)]
266. Yang, B.T.; Dayeh, T.A.; Volkov, P.A.; Kirkpatrick, C.L.; Malmgren, S.; Jing, X.; Renström, E.; Wollheim, C.B.; Nitert, M.D.; Ling, C. Increased DNA methylation and decreased expression of PDX-1 in pancreatic islets from patients with Type 2 diabetes. *Mol. Endocrinol.* **2012**, *26*, 1203–1212. [[CrossRef](#)] [[PubMed](#)]
267. Lovis, P.; Roggli, E.; Laybutt, D.R.; Gattesco, S.; Yang, J.-Y.; Widmann, C.; Abderrahmani, A.; Regazzi, R. Alterations in microRNA expression contribute to fatty acid-induced pancreatic beta-cell dysfunction. *Diabetes* **2008**, *57*, 2728–2736. [[CrossRef](#)] [[PubMed](#)]
268. Guay, C.; Regazzi, R. New emerging tasks for microRNAs in the control of  $\beta$ -cell activities. *Biochim. Biophys. Acta* **2016**, *1861*, 2121–2129. [[CrossRef](#)] [[PubMed](#)]
269. Akirav, E.M.; Lebastchi, J.; Galvan, E.M.; Henegariu, O.; Akirav, M.; Ablamunits, V.; Lizardi, P.M.; Herold, K.C. Detection of beta cell death in diabetes using differentially methylated circulating DNA. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 19018–19023. [[CrossRef](#)] [[PubMed](#)]
270. Hidalgo, B.; Irvin, M.R.; Sha, J.; Zhi, D.; Aslibekyan, S.; Absher, D.; Tiwari, H.K.; Kabagambe, E.K.; Ordovas, J.M.; Arnett, D.K. Epigenome-wide association study of fasting measures of glucose, insulin, and HOMA-IR in the genetics of lipid lowering drugs and diet network study. *Diabetes* **2014**, *63*, 801–807. [[CrossRef](#)] [[PubMed](#)]

271. Yuan, W.; Xia, Y.; Bell, C.G.; Yet, I.; Ferreira, T.; Ward, K.J.; Gao, F.; Loomis, A.K.; Hyde, C.L.; Wu, H.; et al. An integrated epigenomic analysis for Type 2 diabetes susceptibility loci in monozygotic twins. *Nat. Commun.* **2014**, *5*, 5719. [[CrossRef](#)] [[PubMed](#)]
272. Nilsson, E.; Jansson, P.A.; Perfilyev, A.; Volkov, P.; Pedersen, M.; Svensson, M.K.; Poulsen, P.; Ribel-Madsen, R.; Pedersen, N.L.; Almgren, P.; et al. Altered DNA methylation and differential expression of genes influencing metabolism and inflammation in adipose tissue from subjects with Type 2 diabetes. *Diabetes* **2014**, *63*, 2962–2976. [[CrossRef](#)] [[PubMed](#)]
273. Dayeh, T.; Volkov, P.; Salö, S.; Hall, E.; Nilsson, E.; Olsson, A.H.; Kirkpatrick, C.L.; Wollheim, C.B.; Eliasson, L.; Rönn, T.; et al. Genome-wide DNA methylation analysis of human pancreatic islets from Type 2 Diabetic and non-diabetic donors identifies candidate genes that influence insulin secretion. *PLoS Genet.* **2014**, *10*, e1004160. [[CrossRef](#)] [[PubMed](#)]
274. McCann, S.E.; Liu, S.; Wang, D.; Shen, J.; Hu, Q.; Hong, C.-C.; Newman, V.A.; Zhao, H. Reduction of dietary glycaemic load modifies the expression of microRNA potentially associated with energy balance and cancer pathways in pre-menopausal women. *Br. J. Nutr.* **2013**, *109*, 585–592. [[CrossRef](#)] [[PubMed](#)]
275. Nardelli, C.; Iaffaldano, L.; Ferrigno, M.; Labruna, G.; Maruotti, G.M.; Quaglia, F.; Capobianco, V.; Di Noto, R.; Del Vecchio, L.; Martinelli, P.; et al. Characterization and predicted role of the microRNA expression profile in amnion from obese pregnant women. *Int. J. Obes.* **2014**, *38*, 466–469. [[CrossRef](#)] [[PubMed](#)]
276. Thompson, R.F.; Fazzari, M.J.; Niu, H.; Barzilai, N.; Simmons, R.A.; Grealley, J.M. Experimental intrauterine growth restriction induces alterations in DNA methylation and gene expression in pancreatic islets of rats. *J. Biol. Chem.* **2010**, *285*, 15111–15118. [[CrossRef](#)] [[PubMed](#)]
277. Park, J.H.; Stoffers, D.A.; Nicholls, R.D.; Simmons, R.A. Development of Type 2 diabetes following intrauterine growth retardation in rats is associated with progressive epigenetic silencing of Pdx1. *J. Clin. Invest.* **2008**, *118*, 2316–2324. [[CrossRef](#)] [[PubMed](#)]
278. Sandovici, I.; Smith, N.H.; Nitert, M.D.; Ackers-Johnson, M.; Uribe-Lewis, S.; Ito, Y.; Jones, R.H.; Marquez, V.E.; Cairns, W.; Tadayon, M.; et al. Maternal diet and aging alter the epigenetic control of a promoter-enhancer interaction at the Hnf4a gene in rat pancreatic islets. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 5449–5454. [[CrossRef](#)] [[PubMed](#)]
279. Jimenez-Chillaron, J.C.; Isganaitis, E.; Charalambous, M.; Gesta, S.; Pentinat-Pelegrin, T.; Faucette, R.R.; Otis, J.P.; Chow, A.; Diaz, R.; Ferguson-Smith, A.; et al. Intergenerational transmission of glucose intolerance and obesity by in utero undernutrition in mice. *Diabetes* **2009**, *58*, 460–468. [[CrossRef](#)] [[PubMed](#)]
280. Radford, E.J.; Ito, M.; Shi, H.; Corish, J.A.; Yamazawa, K.; Isganaitis, E.; Seisenberger, S.; Hore, T.A.; Reik, W.; Erkek, S.; et al. In utero effects. In utero undernourishment perturbs the adult sperm methylome and intergenerational metabolism. *Science* **2014**, *345*, 1255903. [[CrossRef](#)] [[PubMed](#)]
281. Dunn, G.A.; Bale, T.L. Maternal high-fat diet promotes body length increases and insulin insensitivity in second-generation mice. *Endocrinology* **2009**, *150*, 4999–5009. [[CrossRef](#)] [[PubMed](#)]
282. Ding, G.-L.; Wang, F.-F.; Shu, J.; Tian, S.; Jiang, Y.; Zhang, D.; Wang, N.; Luo, Q.; Zhang, Y.; Jin, F.; et al. Transgenerational glucose intolerance with Igf2/H19 epigenetic alterations in mouse islet induced by intrauterine hyperglycemia. *Diabetes* **2012**, *61*, 1133–1142. [[CrossRef](#)] [[PubMed](#)]
283. Ng, S.-F.; Lin, R.C.Y.; Laybutt, D.R.; Barres, R.; Owens, J.A.; Morris, M.J. Chronic high-fat diet in fathers programs  $\beta$ -cell dysfunction in female rat offspring. *Nature* **2010**, *467*, 963–966. [[CrossRef](#)] [[PubMed](#)]
284. Dunn, G.A.; Bale, T.L. Maternal high-fat diet effects on third-generation female body size via the paternal lineage. *Endocrinology* **2011**, *152*, 2228–2236. [[CrossRef](#)] [[PubMed](#)]
285. Fullston, T.; Ohlsson Teague, E.M.C.; Palmer, N.O.; DeBlasio, M.J.; Mitchell, M.; Corbett, M.; Print, C.G.; Owens, J.A.; Lane, M. Paternal obesity initiates metabolic disturbances in two generations of mice with incomplete penetrance to the F2 generation and alters the transcriptional profile of testis and sperm microRNA content. *FASEB J.* **2013**, *27*, 4226–4243. [[CrossRef](#)] [[PubMed](#)]
286. Ravelli, A.C.; van der Meulen, J.H.; Michels, R.P.; Osmond, C.; Barker, D.J.; Hales, C.N.; Bleker, O.P. Glucose tolerance in adults after prenatal exposure to famine. *Lancet* **1998**, *351*, 173–177. [[CrossRef](#)]
287. De Rooij, S.R.; Painter, R.C.; Phillips, D.I.W.; Osmond, C.; Michels, R.P.J.; Godsland, I.F.; Bossuyt, P.M.M.; Bleker, O.P.; Roseboom, T.J. Impaired insulin secretion after prenatal exposure to the Dutch famine. *Diabetes Care* **2006**, *29*, 1897–1901. [[CrossRef](#)] [[PubMed](#)]

288. Hillier, T.A.; Pedula, K.L.; Schmidt, M.M.; Mullen, J.A.; Charles, M.-A.; Pettitt, D.J. Childhood obesity and metabolic imprinting: The ongoing effects of maternal hyperglycemia. *Diabetes Care* **2007**, *30*, 2287–2292. [[CrossRef](#)] [[PubMed](#)]
289. Finer, S.; Iqbal, M.S.; Lowe, R.; Ogunkolade, B.W.; Pervin, S.; Mathews, C.; Smart, M.; Alam, D.S.; Hitman, G.A. Is famine exposure during developmental life in rural Bangladesh associated with a metabolic and epigenetic signature in young adulthood? A historical cohort study. *BMJ Open* **2016**, *6*, e011768. [[CrossRef](#)] [[PubMed](#)]
290. Heijmans, B.T.; Tobi, E.W.; Stein, A.D.; Putter, H.; Blauw, G.J.; Susser, E.S.; Slagboom, P.E.; Lumey, L.H. Persistent epigenetic differences associated with prenatal exposure to famine in humans. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 17046–17049. [[CrossRef](#)] [[PubMed](#)]
291. Painter, R.C.; de Rooij, S.R.; Bossuyt, P.M.; Simmers, T.A.; Osmond, C.; Barker, D.J.; Bleker, O.P.; Roseboom, T.J. Early onset of coronary artery disease after prenatal exposure to the Dutch famine. *Am. J. Clin. Nutr.* **2006**, *84*, 322–327. [[PubMed](#)]
292. Ludwig, D.S.; Currie, J. The association between pregnancy weight gain and birthweight: A within-family comparison. *Lancet* **2010**, *376*, 984–990. [[CrossRef](#)]
293. Wellen, K.E.; Hatzivassiliou, G.; Sachdeva, U.M.; Bui, T.V.; Cross, J.R.; Thompson, C.B. ATP-citrate lyase links cellular metabolism to histone acetylation. *Science* **2009**, *324*, 1076–1080. [[CrossRef](#)] [[PubMed](#)]
294. Kaati, G.; Bygren, L.O.; Edvinsson, S. Cardiovascular and diabetes mortality determined by nutrition during parents' and grandparents' slow growth period. *Eur. J. Hum. Genet.* **2002**, *10*, 682–688. [[CrossRef](#)] [[PubMed](#)]
295. Veenendaal, M.; Painter, R.; de Rooij, S.; Bossuyt, P.; van der Post, J.; Gluckman, P.; Hanson, M.; Roseboom, T. Transgenerational effects of prenatal exposure to the 1944–1945 Dutch famine. *BJOG Int. J. Obstet. Gynaecol.* **2013**, *120*, 548–554. [[CrossRef](#)] [[PubMed](#)]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).