

Prognostic Value of VEGF in Human Pancreatic Ductal Adenocarcinoma

Yun Jeong Lim, M.D.*, Jong Kyun Lee, M.D., Cheol Keun Park, M.D.[†], Sang Yong Song, M.D.[†], Woo Young Jang, M.D.[†], Hye Young Ha, Dong Il Park, M.D., Kyu Taek Lee, M.D., Seung Woon Paik, M.D., Byung Chul Yoo, M.D. and Jong Chul Rhee, M.D.

*Center for Health promotion**, *Departments of Internal Medicine and Pathology[†]*, *Samsung Medical Center, Sungkyunkwan University School of Medicine, Seoul, Korea*
Department of Pathology[‡], *College of Medicine, Hallym University, Seoul, Korea*

Background : Since pancreatic cancer metastasizes early regardless of the size of the primary tumor, it is suggested that angiogenic factor is upregulated in this disease. Among the angiogenic factors, vascular endothelial growth factor (VEGF) is the most potent and specific growth factor. The aim of this study is to elucidate the prognostic value of VEGF expression in pancreatic cancers.

Methods : We analyzed the VEGF expression using immunohistochemistry in 72 resected pancreatic ductal adenocarcinomas. We examined the prognostic value of the VEGF expression along with its relationship with the clinicopathological features.

Results : VEGF expression and mutant p53 expression were not associated with microvessel density. VEGF expression was positively associated with mutant p53 expression. There were no statistically significant relationships between the VEGF expression and other clinicopathological features, such as age, sex, CA19-9, tumor size, location, tumor differentiation, and stage. VEGF expression was not associated with patient survival.

Conclusion : VEGF expression was not associated with the microvessel density and patient survival in pancreatic ductal adenocarcinoma.

Key Words : VEGF, Pancreatic neoplasm, Prognosis

INTRODUCTION

Pancreatic cancer is one of the most common and lethal cancers in the world with a median survival time of approximately 3 months¹⁾. Although combined chemoradiation results in improvement of local control, it has only a modest impact on survival due to the development of distal metastases¹⁾. Pancreatic cancer can entail the substantial development of new blood vessels within the tumor tissue, and it is known that the growth and progression of solid tumors depend on such angiogenesis²⁾. Among the proangiogenic factors, the vascular endothelial growth factor (VEGF) is the most potent and specific growth factor^{3, 4)}. There are discrepancies about the prognostic value of the VEGF in pancreatic cancer. A

previous report demonstrated that VEGF expression was an independent prognostic factor in pancreatic cancer⁵⁾. There were somewhat contradictory results that the VEGF expression did not correlate with various clinicopathological parameters such as the vessel count, metastasis, recurrence and survival⁶⁾. To estimate the usefulness of the prognostic factor, many molecular and biological markers were investigated. This study was conducted to elucidate the prognostic value of VEGF expression as well as evaluate its relationship with clinicopathological variables in pancreatic ductal adenocarcinoma.

• Received : September 15, 2003

• Accepted : November 13, 2003

• Correspondence to : Jong Kyun Lee, M.D., Department of Internal Medicine, Samsung Medical Center, 50, Irwonbon-dong, Gangnam-gu, Seoul, 135-710, Korea Tel : 82-2-3410-3407, Fax : 82-2-3410-3849, E-mail : jklee@smc.samsung.co.kr

Table 1. The relationship between VEGF immunostaining and other clinicopathological features of pancreatic ductal adenocarcinomas (Spearman's rank correlation)

| | No. of cases | VEGF staining | | <i>p</i> value |
|------------------------|--------------|---------------|-------------|----------------|
| | | Negative | positive | |
| Age (mean±S.D.) | 72 | 61.68±9.59 | 56.95±13.81 | NS |
| Sex | | | | |
| Man | 43 | 28 | 15 | NS |
| Woman | 29 | 21 | 8 | |
| CA19-9 (mean±S.D.) | 72 | 54.16±12.5 | 58.45±9.5 | NS |
| Tumor size (mean±S.D.) | 72 | 3.86±2.94 | 4.28±5.14 | NS |
| Location | | | | |
| Head | 47 | 33 | 14 | NS |
| Body / tail | 25 | 16 | 9 | |
| Tumor differentiation | | | | |
| Well | 14 | 6 | 8 | NS |
| Moderate | 44 | 34 | 10 | |
| Poor | 14 | 9 | 5 | |
| Stage | | | | |
| 1 | 13 | 6 | 7 | NS |
| 2 | 21 | 15 | 6 | |
| 3 | 15 | 9 | 6 | |
| 4 | 23 | 19 | 4 | |

NS, not significant

MATERIALS AND METHODS

1. Patients and tissue samples.

Samples of 72 pancreatic ductal adenocarcinomas were obtained at the time of surgical resection from Samsung Medical Center (Seoul, Korea) between January 1995 and September 1999. The age of patients ranged from 16 to 79 years with a median age of 59.8 years. There were 43 male and 29 female patients. The median period of follow-up was 12.6 months (range, 1–66.2 months). The median survival time was 353 days. Follow-up data were gathered by reviewing of the patient's charts. The patients' outcomes were verified and updated through the medical record departments of Samsung Medical Center and the telephone. Fifty-six (77.8%) patients died, and 16 patients were still alive during the follow-up periods. Twenty-six patients received adjuvant chemotherapy. Tissues were fixed in 10% formalin for 12 hours and embedded in paraffin. Tissues were stained with hematoxylin and eosin. The histopathologic features of the pancreatic ductal adenocarcinoma were examined for the following: tumor size, location, tumor differentiation, and stage. The tumor stage was determined according to the staging manual of the American Joint Committee on Cancer⁷⁾.

2. Immunohistochemistry

The representative tissue sections including both an adequate amount of tumor tissue and the adjacent non-tumor epithelial cells were selected and sectioned in 4- μ m-thickness.

Immunohistochemical study was performed using the strept-
tavin-biotin complex method. The primary antibodies used and working dilutions employed were as follows: VEGF (Santa Cruz Biotechnology, Inc, Santa Cruz, CA, USA, 1:2000), CD 34 (Immunotech, Marseille Cedex, France, 1:50), and p53 (Zymed, San Francisco, CA, USA, 1:80 dilution). For antigen retrieval, the sections were pretreated in an 800-W microwave oven in a 10 mM citrate buffer of pH 6.0 for 10 minutes. The sections were incubated with the primary antibody for 10 minutes, and then, the secondary antibody and the strept-
tavin-peroxidase complex (LSAB kit, Dako, CA, USA) were applied, sequentially. DAB (3,3'-diaminobenzidine tetrahydrochloride) was used as a chromogen, and Mayer's hematoxylin counterstain was applied. Negative controls were run simultaneously with

Table 2. Cox proportional hazard model for Survival

| | <i>p</i> value |
|-----------------------|----------------|
| Age | NS |
| Sex | NS |
| CA 19-9 | NS |
| Size | NS |
| Location | NS |
| Tumor differentiation | NS |
| Stage | NS |
| CD34 | NS |
| Mutant p53 | NS |
| VEGF | NS |

NS, not significant

an omission of the primary antibody. The proportion of VEGF positive tumor cells was quantified and was assigned to one of two categories; 0, 10% : 1, >10%³⁾. Microvessel density was recorded by counting the CD 34 positive vessels in the highest vascularized area in X200 fields (a X10 ocular and a X20 objective lens)⁸⁾. Vessels of a caliber larger than approximately eight RBCs, vessels with thick muscular walls, and vessels in sclerotic areas were excluded from the count. The counts were expressed as the total number of microvessels per mm² in X200 fields using the image analysis system (CIRES program, IBAS model, Zeiss, Germany). Mutant p53 immunostaining was scored as follows. The proportion of positive tumor cells was quantified and was assigned to one of two categories; 0, 10% : 1, >10%⁹⁾. The stained slides were independently evaluated by two authors (W.O.J. and S.Y.S), and the difference in interpretation was resolved by a consensus.

3. Statistical analysis

The correlation between VEGF expression and clinicopathological features (age, sex, CA19-9, tumor size, location, tumor differentiation, stage, CD 34 and mutant p53) was determined by the Spearman's rank correlation. A Cox pro-

portional hazards model for the risk ratio was used to assess the simultaneous contribution of the following baseline covariates: age, sex, CA19-9, tumor size, location, tumor differentiation, stage, CD 34, mutant p53, and VEGF. Survival curves were constructed using the method of Kaplan-Meier. A probability of $p < 0.05$ was considered statistically significant.

RESULTS

VEGF immunoreactivity was present in 23/72 tumors (32%) (Figure 1A), and The mutant p53 expression was present in 44/72 tumors (61%) (Figure 1C). The VEGF expression was not associated with microvessel density ($p=0.059$) (Figure 1B, 2), and microvessel density was not associated with the mutant p53 expression ($p=0.09$). VEGF expression had a positive correlation with the p53 mutation ($p=0.049$). When the relationship between the VEGF expression and other clinicopathological parameters were evaluated, the statistical analysis revealed no significant correlation with other clinicopathological parameters (Table 1).

Multivariate analysis was performed according to the Cox proportional hazard model in order to evaluate the prognostic value of the VEGF expression, microvessel density, and mutant p53 expression. VEGF expression ($p=0.239$), microvessel density ($p=0.398$), and mutant p53 expression ($p=0.772$) did not predict

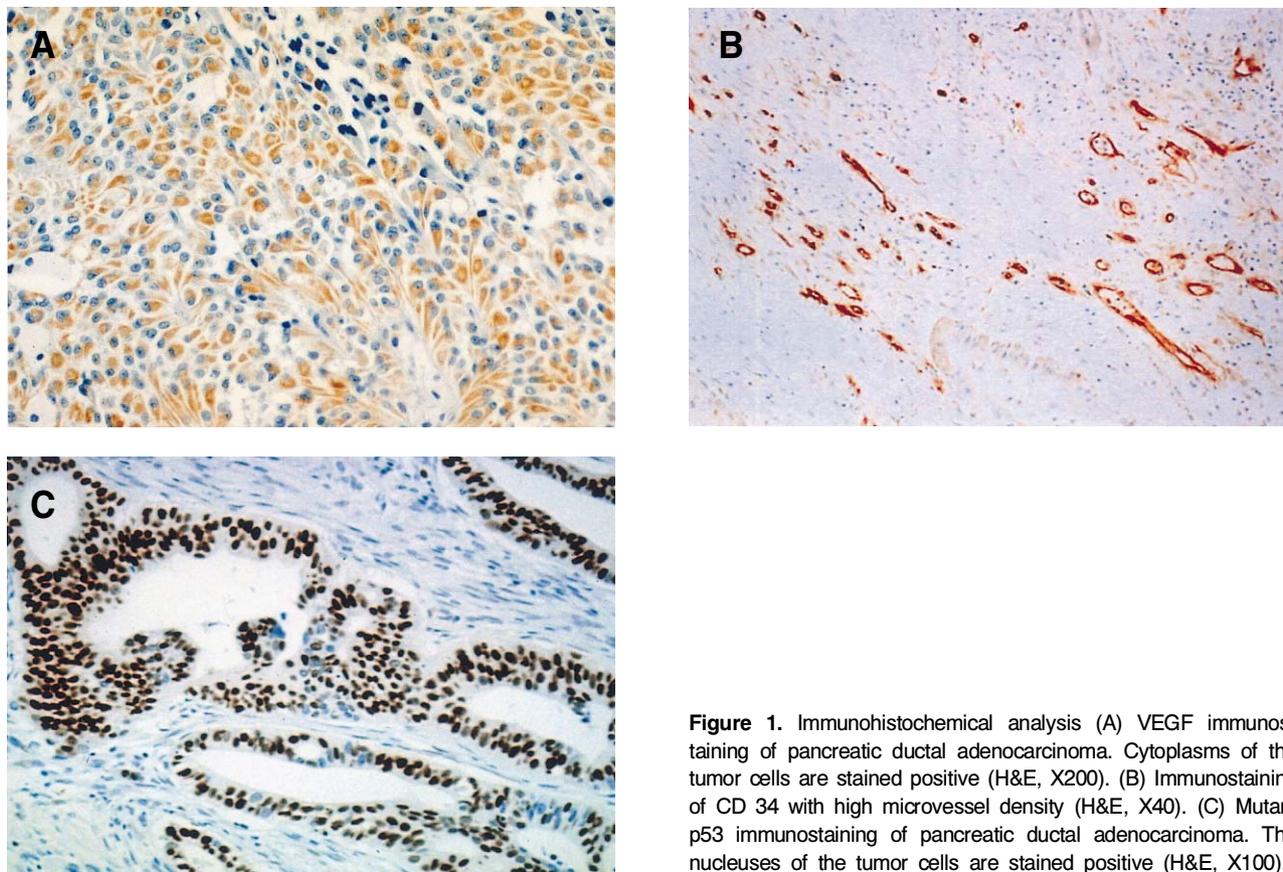


Figure 1. Immunohistochemical analysis (A) VEGF immunostaining of pancreatic ductal adenocarcinoma. Cytoplasm of the tumor cells are stained positive (H&E, X200). (B) Immunostaining of CD 34 with high microvessel density (H&E, X40). (C) Mutant p53 immunostaining of pancreatic ductal adenocarcinoma. The nuclei of the tumor cells are stained positive (H&E, X100).

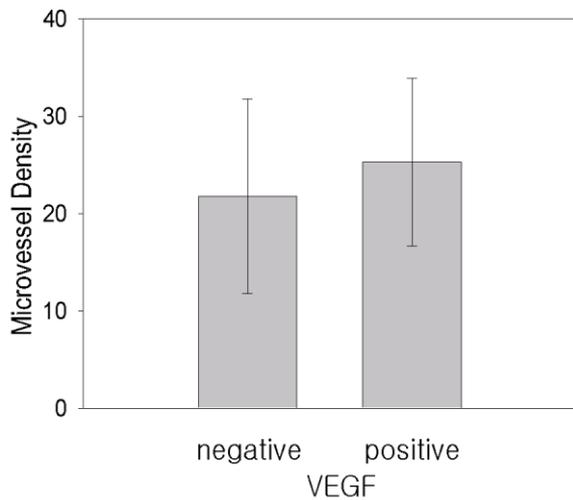


Figure 2. The correlation between VEGF expression and microvessel density. VEGF expression was not associated with microvessel density ($p=0.059$).

unfavorable prognosis independently of other clinicopathological features (Table 2). The survival curves for the patients as a function of the VEGF expression are shown in Figure 3. It was illustrated using the Kaplan–Meier survival curve ($p=0.448$) that VEGF expression was not associated with patient survival.

DISCUSSION

A reason for the poor prognosis in pancreatic cancer is the development of early metastasis regardless of the primary tumor growth¹. Angiogenesis is an integral part of the cascade of biologic events involved in tumor metastasis¹⁰. If angiogenesis is essential for tumor growth and metastasis, there may be differences in the quantitative assessment of vascular proliferation between tumors with different prognostic features^{10, 11}. Increased angiogenesis in pancreatic cancer is related to cancer aggressiveness^{2, 5}. To date, many angiogenic factors for pancreatic cancers have been reported, such as the transforming growth factors (TGF)- α , TGF- β , aFGF (acidic fibroblast growth factor), bFGF (basic fibroblast growth factor), angiogenin, VEGF, and PD-ECGF (platelet-derived endothelial cell growth factor)^{5, 12, 13}. Among the angiogenic factors, VEGF is the most potent and the most specific growth factor¹³. Since pancreatic cancer metastasizes early regardless of the size of the primary tumor, it was expected that VEGF was upregulated in this disease³. But, there are many controversies about the prognostic value of VEGF in pancreatic cancer. The previous report showed that strong VEGF immunoreactivity was present in the cancer ceu in 64% of the pancreatic cancer tissues, and the presence of VEGF was

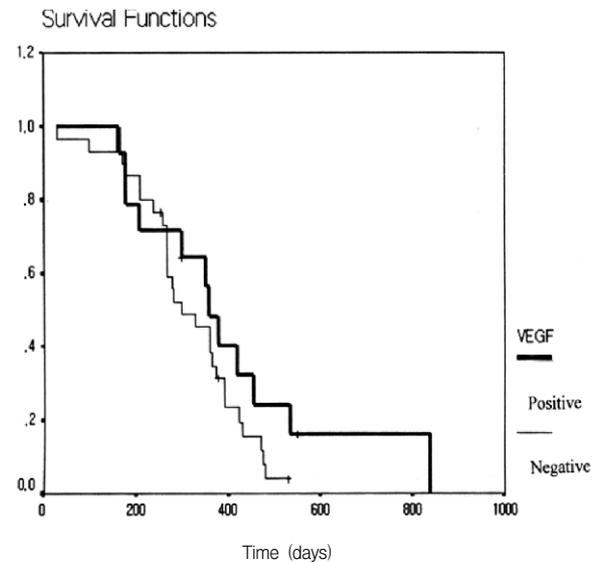


Figure 3. The effect of VEGF immunostaining on cumulative survival rate after the surgical resection of pancreatic ductal adenocarcinomas. Positive VEGF expression did not predict short survival ($p=0.448$).

associated with an increased blood vessel number, large tumor size, and enhanced local spread but was not associated with a reduction in patient survival time³. In our study, the VEGF expression was neither associated with an increased blood vessel number nor patient survival. Dilution, incubation time, and method of interpretation were somewhat different in the previous reports^{3-5, 13}. Most of all, the analysis of the large sample size by immunohistochemistry combined with other objective experimental methods, such as Northern blot and real time PCR, was needed to elucidate the exact prognostic value of the VEGF in pancreatic ductal adenocarcinoma.

p53 mutations might be acquired in the later stages associated with metastatic progression and higher proliferation activity^{14, 15}. It was suggested that the mutant p53 gene was a potent stimulant of VEGF and was related to tumor angiogenesis¹⁴. Mutant p53 expression may be a beneficial prognostic factor in pancreatic cancers⁹. The previous report showed that mutations in the p53 gene occurred in approximately 60% of the tumors and appeared to be an independent prognostic factor for patient survival^{1, 9}. The present study demonstrated that mutations in the p53 gene occurred in 61% of the tumors and was not a beneficial prognostic factor for patients with pancreatic cancer. The positive VEGF expression was associated with a poor prognostic factor such as mutant p53.

The only limitation of our study are the immunohistochemical results. To know the exact prognostic value of VEGF, analysis of large sample size by immunohistochemistry, as

well as the quantitative assay, such as the Northern blot, needs to be elucidated. The present study demonstrated that VEGF expression was neither associated with microvessel density nor patient survival in pancreatic ductal adenocarcinoma. This study suggests that it is difficult for the VEGF expression to be used as a prognostic marker in clinical practice.

ACKNOWLEDGMENTS

The authors thank Dr. Seonwoo Kim for the statistical analysis.

REFERENCES

- 1) van Riel JM, Giaccone G, Pinedo HM. *Pancreaticobiliary cancer: the future aspects of medical oncology*. *Ann Oncol* 10:296-299, 1999
- 2) Folkman J. *What is the evidence that tumors are angiogenesis dependent?* *J Natl Cancer Inst* 82:4-6, 1990
- 3) Itakura J, Ishiwata T, Friess H, Fujii H, Matsumoto Y, Buchler MW, Korc M. *Enhanced expression of vascular endothelial growth factor in human pancreatic cancer correlates with local disease progression*. *Clin Cancer Res* 3:1309-1316, 1997
- 4) Chaffey KP, Robinson GS. *Regulation of VEGF/NPF expression in tumor cells: consequences for tumor growth and metastasis*. *Cancer Metastasis Rev* 15:165-176, 1996
- 5) Ikeda N, Adachi M, Taki T, Huang C, Hashida H, Takabayashi A, Shomura M, Nakajima Y, Kanehiro H, Hatanaga M, Nakano H, Miyake M. *Prognostic significance of angiogenesis in human pancreatic cancer*. *Br J Cancer* 79:1553-1563, 1999
- 6) Fujimoto K, Hosotani R, Wada M, Lee JU, Koshba T, Miyamoto Y, Tsuji S, Nakajima S, Doi R, Imamura M. *Expression of two angiogenic factors, vascular endothelial growth factor and platelet-derived endothelial cell growth factor in human pancreatic cancer, and its relationship to angiogenesis*. *Eur J Cancer* 34:1439-1447, 1998
- 7) Fleming ID, Cooper JS, Henson DE, Hutter RVP, Kennedy BJ, Murphy GP, O'Sullivan B, Sobin LH, Yabro JW. *American Joint Committee on Cancer, Cancer Staging Manual*. pp.121-126, Philadelphia, Lippincott-Raven, 1997
- 8) Weidner N, Semple JP, Welch WR, Folkman J. *Tumor angiogenesis and metastasis: correlation in invasive breast carcinoma*. *N Engl J Med* 324:1-8, 1991
- 9) Barton CM, Staddon SL, Hughes CM, Hall PA, O'Sullivan C, Koppel G, Theis B, Russell RC, Neoptolemos J, Williamson RC. *Abnormalities of the p53 tumor suppressor gene in human pancreatic cancer*. *Br J Cancer* 64:1076-1082, 1991
- 10) Ozer E, Ozkal S, Karademir S, Sagol O, Sokmen S, Coker A, Kpeloglu A, Astarcılu I. *Angiogenesis and p53 and H-ras mutations in pancreatic ductal adenocarcinoma*. *Anal Quant Cytol Histol* 21:473-476, 1999
- 11) Shimoyama S, Kamishima M. *Increased angiogenic expression in gastric cancer correlated with cancer progression*. *J Cancer Res Clin Oncol* 126:468-474, 2000
- 12) Yamataka Y, Friess H, Buchler M, Beger HG, Uchida E, Onda M, Kobrin MS, Korc M. *Overexpression of acidic and basic fibroblast growth factors in human pancreatic cancer correlates with advanced tumor stage*. *Cancer Res* 53:5289-5296, 1993
- 13) Mineta H, Miura K, Ogino T, Takebayashi S, Misawa K, Ueda Y, Suzuki I, Dector M, Borg A, Wennerberg J. *Prognostic value of vascular endothelial growth factor (VEGF) in head and neck squamous cell carcinomas*. *Br J Cancer* 83:775-781, 2000
- 14) Bouvet M, Ellis LM, Nishizaki M, Fujiwara T, Liu W, Bucana CD, Fang B, Lee JJ, Roth JA. *Adenovirus-mediated wild type p53 gene transfer downregulates vascular endothelial growth factor expression and inhibits angiogenesis in human colon cancer*. *Cancer Res* 58:2288-2292, 1998
- 15) Dameron KM, Voelpert OV, Tainsky MA, Bouck N. *The p53 tumor suppressor gene inhibits angiogenesis by stimulating the production of thrombospondin*. *Cold Spring Harb Symp Quant Biol* 59:483-489, 1994