



**University of  
Zurich**<sup>UZH</sup>

**Zurich Open Repository and  
Archive**

University of Zurich  
Main Library  
Strickhofstrasse 39  
CH-8057 Zurich  
[www.zora.uzh.ch](http://www.zora.uzh.ch)

---

Year: 2013

---

**Pollinator shifts between *Ophrys sphegodes* populations: might adaptation to different pollinators drive population divergence?**

Breitkopf, H; Schlüter, P M; Xu, S; Schiestl, F P; Cozzolino, S; Scopece, G

DOI: <https://doi.org/10.1111/jeb.12216>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-80992>

Accepted Version

Originally published at:

Breitkopf, H; Schlüter, P M; Xu, S; Schiestl, F P; Cozzolino, S; Scopece, G (2013). Pollinator shifts between *Ophrys sphegodes* populations: might adaptation to different pollinators drive population divergence? *Journal of Evolutionary Biology*, 26(10):2197-2208.

DOI: <https://doi.org/10.1111/jeb.12216>

1 Pollinator shifts between *Ophrys sphegodes* populations: might adaptation to different  
2 pollinators drive population divergence?

3

4

5 HENDRIK BREITKOPF<sup>1,3</sup>, PHILIPP M. SCHLÜTER<sup>2</sup>, SHUQING XU<sup>2,4</sup>, FLORIAN P.  
6 SCHIESTL<sup>2</sup>, SALVATORE COZZOLINO<sup>1\*</sup>, GIOVANNI SCOPECE<sup>1</sup>

7

8 <sup>1</sup> *Department of Biology, University of Naples Federico II, Via Cinthia, I-80126, Naples, Italy*

9 <sup>2</sup> *Institute of Systematic Botany, University of Zürich, Zollikerstrasse 107, CH-8008 Zurich,*  
10 *Switzerland*

11 <sup>3</sup> *Institute of Biochemistry and Biology, Biodiversity Research/Systematic Botany, University*  
12 *of Potsdam, Maulbeerallee 1, D-14469, Potsdam, Germany*

13 <sup>4</sup> *Molecular Ecology Department, Max Planck Institute for Chemical Ecology, Hans-Knöll-*  
14 *Straße 8 D-07745 Jena, Germany*

15

16 \* Corresponding author. E-mail: [cozzolin@unina.it](mailto:cozzolin@unina.it), Tel:++39081678186, Fax  
17 ++39081679233

18 Running title: Local pollinator shift in *Ophrys sphegodes*

19

20

## ABSTRACT

21 Local adaptation to different pollinators is considered one of the possible initial stages of  
22 ecological speciation as reproductive isolation is a by-product of the divergence in pollination  
23 systems. However, pollinator-mediated divergent selection will not necessarily result in  
24 complete reproductive isolation, since incipient speciation is often overcome by gene flow.  
25 We investigated the potential of pollinator shift in the sexually deceptive orchids *Ophrys*  
26 *sphogodes* and *O. exaltata* and compared levels of floral isolation versus genetic distance  
27 among populations with contrasting, predominant pollinators. We analysed floral  
28 hydrocarbons as a proxy for floral divergence between populations. Floral adoption of  
29 pollinators and their fidelity was tested using pollinator choice experiments. Inter-population  
30 gene flow and population differentiation levels were estimated using AFLP markers. The  
31 Tyrrhenian *O. sphogodes* population preferentially attracted the pollinator bee *Andrena*  
32 *bimaculata*, whereas the Adriatic *O. sphogodes* population exclusively attracted *A.*  
33 *nigroaenea*. Significant differences in scent component proportions were identified in *O.*  
34 *sphogodes* populations that attracted different preferred pollinators. High inter-population  
35 gene flow was detected, but populations were genetically structured at species level. The high  
36 inter-population gene flow levels independent of preferred pollinators suggest that local  
37 adaptation to different pollinators has not (yet) generated detectable genome-wide separation.  
38 Alternatively, despite extensive gene flow, few genes underlying floral isolation remain  
39 differentiated as a consequence of divergent selection. Different pollination ecotypes in *O.*  
40 *sphogodes* might represent a local selective response imposed by temporal variation in a  
41 geographic mosaic of pollinators as a consequence of the frequent disturbance regimes typical  
42 of *Ophrys* habitats.

43

44 **Key words:** adaptation, ecotypes, floral scent, gene flow, *Ophrys*, orchids, pollinator shift,  
45 sexual deception, speciation

46

47

## INTRODUCTION

48 One important mechanism driving angiosperm diversification is pollinator-mediated selection  
49 due to pollen limitation and floral isolation (Lowry *et al.*, 2008; Xu *et al.*, 2011). Although the  
50 evolutionary role of pollinator-mediated selection remains debated under complex ecological  
51 conditions (Kay & Sargent, 2009), pollinator specialisation has traditionally been considered

52 one of the prime mechanisms for ecological speciation due to the direct relationship between  
53 pollinator specialisation and floral isolation (Grant, 1971; Johnson, 2006; Schiestl, 2012).

54 Pollinator shift frequency in specialised plant lineages depends on the influence of  
55 local selective forces (i.e. pollen limitation and pollen transfer efficiency), and the degree of  
56 floral isolation and its genetic basis (Xu *et al.*, 2012a; Schiestl & Schlüter, 2009, and  
57 references therein). In fact, if the transition to a different pollinator requires changes in several  
58 non-correlated floral traits and attraction signals, then strong and prolonged directional  
59 selection is required to promote those changes (Nosil *et al.*, 2009). Such evolutionary  
60 transition, involving several traits, will probably proceed gradually. Alternatively, some  
61 recently diverged lineages, where pollinator shifts facilitated rapid radiations, are often  
62 characterised by related species that attract different specific pollinators because of subtle  
63 changes in floral advertisement or shape, rather than large floral structural rearrangements and  
64 large changes in signals (Fulton & Hodges, 1999; Schemske & Bradshaw, 1999; Sturman *et*  
65 *al.*, 2004). These observations suggest that in such specialised plant groups only one or a few  
66 floral traits play a prominent role in preventing pollinator sharing, and that even small  
67 alterations in key traits may have substantial effects in attracting distinct pollinators, or  
68 pollinator groups.

69 Sexually deceptive orchids are typified by a combination of traits mediating specific  
70 pollinator attraction, and sexual mimicry of pollinator females achieves pollination specificity  
71 (Kullenberg, 1961; Paulus & Gack, 1990a; Bower, 1996). Several recent studies provide  
72 strong experimental evidence demonstrating that floral scent holds the key to specific  
73 pollinator attraction, while floral display serves a secondary role by increasing floral detection  
74 against vegetation background (Vereecken & Schiestl, 2009; Peakall *et al.*, 2010; Xu *et al.*,  
75 2012a). Additional empirical evidence supports these results by showing that artificial  
76 manipulation of floral display does not significantly alter pollinator attraction (but see  
77 Spaethe *et al.*, 2007). In contrast, floral scent modifications increase or decrease pollinator  
78 visitation rates (Xu *et al.*, 2012b). Therefore, a subtle change in floral scent, even in the  
79 absence of marked changes in floral display, might potentially attract different orchid  
80 pollinators (Xu *et al.*, 2012a).

81 Recently, the genetic and chemical basis of pollinator specificity in sexually deceptive  
82 orchids has been partially elucidated. In *O. sphegodes* and related species, for instance, the  
83 specificity of pollinator attraction is due to quantitative variation in alkenes, which differ in  
84 double-bond position and carbon chain length (Mant *et al.*, 2005a; Schlüter *et al.*, 2011a).  
85 Therefore, even minor variation in gene expression patterns underlying alkene biosynthesis

86 might be sufficient to produce a different odour bouquet, as in *O. sphegodes* versus *O.*  
87 *exaltata* (Schlüter *et al.*, 2011a; Xu *et al.* 2012b). Closely related species of sexually  
88 deceptive orchids in the same phylogenetic lineage attract different pollinators as a result of  
89 qualitative or only quantitative changes in proportions of active odour bouquet compounds.  
90 Alterations in compound proportions provide variability for selection to act upon, and thus  
91 offer the potential for ecological speciation (Schiestl & Ayasse, 2002; Xu *et al.*, 2012a).

92 Pollinator communities and species population sizes are likely to fluctuate through  
93 time, particularly in disturbed and transient habitats as those typically occupied by *Ophrys*  
94 species (Gardiner, 2009; Hutchings, 2010). Indeed, floral specialisation has been considered a  
95 risky strategy due to strict relationships between the survival/extinction of pollinator species,  
96 and the plant species dependent on its exclusive pollination service (Johnson & Steiner,  
97 2000). Thus, potential interaction plasticity with the specialised pollinator might allow the  
98 plant to attain reproductive success, even under limited presence of the main pollinator,  
99 through temporary exploration of locally more abundant secondary pollinators (Waser &  
100 Ollerton, 2006). Under these conditions, local selection on plants imposed by a variable  
101 geographic and temporal mosaic of potential pollinators could lead to multiple pollination  
102 ecotypes (Harder & Johnson, 2009). This adaptive strategy might facilitate pollinator-  
103 specialised species survival when short-term pollinator community fluctuations occur as a  
104 consequence of habitat alterations, such as those caused by anthropogenic disturbance  
105 (Petanidou, 2008; Potts, 2010).

106 Whether *Ophrys* pollination ecotypes are populations, or incipient or actual ecological  
107 species (Van Valen, 1976) has been hotly debated among evolutionary biologists (*e.g.*  
108 Vereecken *et al.*, 2011; Bateman *et al.*, 2011). The crucial difference between adaptation to  
109 local pollinators (pollination ecotypes) and progenitor-derivative speciation due to a pollinator  
110 shift can be identified in the evolution of reproductive isolation associated with the transition  
111 to a different pollinator - a transition that is ultimately dependent on the strength of divergent  
112 selection and the amount of inter-population gene-flow (Schlüter *et al.*, 2011b).

113 Ultimately, biological species are defined by reproductive isolation, which is typically  
114 weak or absent among geographical races (Slatkin, 1987; Coyne & Orr, 1998). Therefore,  
115 reduction or loss of gene flow (as consequence of the insurgence of some forms of  
116 reproductive isolation) between ancestral populations leads to incipient ecological species  
117 from local ecotypes, allowing their gene pools to develop independently (Macnair & Gardner,  
118 1998). Nevertheless, since local adaptation often represents the initial stages of ecological  
119 speciation, a marked distinction between these sequential stages, based on the level of gene

120 flow between different ecotypes, is often difficult to define (Nosil, 2012). Lexer & Widmer  
121 (2008) emphasised the seriousness of this challenge in several ecological species pairs that  
122 displayed a considerable amount of inter-specific gene flow, and where genomic divergence  
123 was only detected at a few loci despite using several genome-wide markers.

124 To date, few studies have examined the nature and level of reproductive isolation  
125 between different ecotypes of a given species (Scopece *et al.*, 2010, and references therein)  
126 and geographic isolation is considered the primary isolating factor among races or local  
127 ecotypes (Lowry *et al.* 2008). However, it does not represent an obvious barrier that prevents  
128 gene flow between geographically close or adjacent ecotypes/populations (Anderson *et al.*,  
129 2010), a situation that can in principle allow constant gene flow between populations  
130 (Räsänen & Hendry, 2008). In such cases, the amount of floral isolation is comparable with  
131 the levels of inter-population gene flow as an indirect estimate of whether different  
132 populations represent locally adapted pollination ecotypes that established a form of incipient  
133 reproductive isolation (Nosil *et al.*, 2009; Thibert-Plante & Hendry, 2010). In fact, although  
134 divergent selection can favour reproductive isolation at a local scale, incipient divergence can  
135 be prevented by the homogenising effect of gene flow between adjacent populations,  
136 maintaining species cohesion (Morjan & Rieseberg, 2004). Investigations into transient stages  
137 in the *continuum* between local adaptation and incipient speciation are integral to speciation  
138 research. Such studies may serve to identify the selective forces acting on floral traits, and  
139 establish the evolutionary outcomes of pollinator shift and subsequent floral isolation (Fenster  
140 *et al.*, 2004).

141 In the present study, with the aim of disclosing the effect of changes in preferred  
142 pollinators on the establishment of reproductive isolation, we investigated the potential for  
143 pollinator shifts in the Mediterranean sexually deceptive orchid *Ophrys sphegodes* MILL., and  
144 compared levels of floral isolation *versus* genetic distance between populations with different,  
145 dominant pollinators. We estimated intra- and inter-specific reproductive isolation and gene  
146 flow in *O. sphegodes s.s.* and the closely related species *O. exaltata* TEN. The two taxa belong  
147 to the same phylogenetic lineage, and Xu *et al.* (2011) only recently inferred that the species  
148 are effectively reproductively isolated.

149

## 150 MATERIALS AND METHODS

151

### 151 STUDY SYSTEM

152 The sexually deceptive orchid species *Ophrys sphegodes* and *O. exaltata* were chosen for this  
153 study. *Ophrys sphegodes* is a widespread species that inhabits sunny meadows or calcareous

154 grasslands from the British Isles to the southern Mediterranean countries (Delforge, 2005)  
155 whereas *O. exaltata* is more restricted to the Mediterranean region, inhabiting sandy soils in  
156 southern France and the central part of the Mediterranean region. In a broad survey of the  
157 entire genus, Devey *et al.* (2008) showed that the two species are phylogenetically closely  
158 related. In peninsular Italy, *O. sphegodes* and *O. exaltata* frequently occupy similar ecological  
159 niches - primarily coastal sandy areas (Delforge, 2005) - and the taxa often occur in ‘mosaic  
160 sympatry’ (*sensu* Mallet *et al.*, 2009). Due to severe habitat fragmentation, particularly where  
161 the Apennine mountain chain separates the Tyrrhenian and Adriatic coasts, both species  
162 accommodate a large number of local varieties, either interpreted as local ecotypes or  
163 endemic species (*cf.* Delforge, 2005; Pedersen & Faurholdt, 2007). Overall, the two species  
164 exhibit well-established reproductive isolation via different primary pollinators (*i.e.* *Andrena*  
165 *nigroaenea* for *O. sphegodes* and *Colletes cunicularis* for *O. exaltata*). However, the species  
166 are interfertile and can produce hybrids in the wild (Xu *et al.*, 2011).

167

168

#### GENETIC ANALYSES

169 We investigated four *O. sphegodes* populations and three *O. exaltata* populations along the  
170 Italian peninsula (Table 1). For each plant sampled, a piece of leaf tissue was field collected  
171 and placed in a plastic bag filled with silica gel. Genomic DNA was extracted using GenElute  
172 Plant Genomic DNA Miniprep Kit (Sigma-Aldrich, Italy). The amplified fragment length  
173 polymorphism (AFLP) procedure followed Vos *et al.* (1995), albeit with modifications  
174 (Moccia *et al.* 2007) and using fluorescent dye-labelled primers. Six primer combinations  
175 were chosen from Xu *et al.* (2011): FAM-*EcoRI*-AGC/*MseI*-ACAC, NED-*EcoRI*-ACC-  
176 /*MseI*-ACTG, HEX-*EcoRI*-AGC/*MseI*-ATCG, FAM-*EcoRI*-ATG/*MseI*-CGG, NED-*EcoRI*-  
177 AAC/*MseI*-CGC, and HEX-*EcoRI*-AGC/*MseI*-CCAA. Fragment separation and detection  
178 were conducted on a 3130 Genetic Analyzer (Applied Biosystems, Foster City, USA).  
179 GeneScan-500 LIZ (Applied Biosystems, Foster City, USA) was used as the internal  
180 standard. Raw data alignment and fragment-size detection were performed using GeneMapper  
181 3.7 software (Applied Biosystems, Foster City, USA). Presence or absence of AFLP bands  
182 was scored visually. Artefacts and missing data were avoided by only including informative  
183 AFLP markers in the binary matrix that could be unambiguously scored for all samples.

184

185

186

187

GenAlEx (Peakall & Smouse, 2006) was run as the macro in Microsoft Excel to  
calculate genetic distances as the basis for generating a principle coordinate analysis (PCoA)  
scatter plot. An analysis of molecular variance (AMOVA) was conducted using a matrix of  
genetic distances between all haplotype pairs. Genetic differentiation was estimated using  $\Phi$ -

188 statistics, an  $F$ -statistics analogue for binary data. Significance of  $\Phi$ -statistics and variance  
189 components were assessed with 999 permutations (Peakall & Smouse, 2006). Pairwise  $\Phi_{ST}$   
190 values based on Jaccard's similarity were calculated from FAMD (Schlüter & Harris, 2006).

191 STRUCTURE 2.2 (Pritchard *et al.*, 2000) was applied to investigate population  
192 structure. The model implemented in STRUCTURE set the posterior probability ( $q$ ) to  
193 describe the proportion of an individual genotype originating from each of  $K$  categories.  
194 Following the method described in Evanno *et al.* (2005), we tested  $K$  from 1 to 7 (*i.e.* the  
195 number of populations sampled) with a burn-in of 50 000 steps, followed by 300 000 MCMC  
196 iterations and 10 replicates to confirm stabilisation of summary statistics. Estimates were  
197 carried out under the admixture model, allowing for correlated allele frequencies on all  
198 sampled individuals, while ignoring sampling localities. The output obtained from  
199 STRUCTURE was graphically displayed with DISTRUCT (Rosenberg, 2004). The genomic  
200 basis of differentiation between *O. sphegodes* populations from Cuma and Gargano was  
201 analysed using a genome scan. Accordingly, to identify putative genomic targets of divergent  
202 selection, an  $F_{ST}$  outlier scan was conducted using the Dfdist package (Beaumont & Nichols,  
203 1996), which applies the Bayesian allele frequency estimate method developed by  
204 Zhivotovsky (1999). The analysis was conducted as in other studies (*e.g.* Minder & Widmer,  
205 2008; Pérez-Figueroa *et al.*, 2010), excluding loci with an allele frequency over 0.99, and  
206 using mean  $F_{ST}$  trimmed at the 30% level. This trimmed mean  $F_{ST}$  was chosen as the target  
207 average for Dfdist to simulate  $F_{ST}$  null distribution values (50000 realisations), assuming a  $\theta$   
208 parameter of 0.05; variation in this  $\theta$  parameter was reported to have very little impact on  
209 results (*e.g.* Minder & Widmer, 2008, and references therein).  $F_{ST}$  outliers were identified as  
210 data points outside the 95% confidence interval (outside the 0.025 and 0.975 quartiles, and at  
211  $P < 0.025$ ).

212 The potential effects of geography and pollinator differentiation on genetic structure  
213 within *O. sphegodes* were separated using a generalised linear model (GLM). Pairwise  $\Phi_{ST}$   
214 values were modelled with the explanatory variables geographic distance and shared  
215 pollinator, as well as an interaction term between the two variables. A geographic distance  
216 matrix was computed with ArcGIS 9.2. The analysis was performed in R 2.15.2 (R  
217 Development Core Team, 2012), as described in Schlüter *et al.* (2011b).

218

#### 219 POLLINATOR CHOICE EXPERIMENTS

220 Pollinator fidelity was tested through pollinator choice experiments between *O. sphegodes*  
221 and *O. exaltata* individuals. Plant inflorescences were collected from natural populations



222 along the Tyrrhenian (Cuma, Campania) and Adriatic (Gargano, Puglia) coasts where *O.*  
223 *sphegodes* and *O. exaltata* are sympatric. The study populations co-flower at the end of  
224 March/beginning of April. Pollinator choice plots with individuals of both species from the  
225 two coasts were established at Cuma and Gargano. We established 12 choice-plots from 17<sup>th</sup>  
226 to 25<sup>th</sup> of March 2011 available during a two hour period from 09.00 to 11.00 h; five choice-  
227 plots were established in Cuma, seven plots in Gargano, intermixed with natural populations.  
228 Each plot consisted of four inflorescences, with one individual at a time of each species from  
229 both coasts. The inflorescences were randomly placed, 30 cm apart, in flowering bushes along  
230 sandy paths. Inflorescences in the choice plots were replaced with new ones after every  
231 pollination event or, in the absence of any visit, after 30 min.  
232 Male bees patrolled along sandy footpaths, where numerous nesting places of female solitary  
233 bees were observed; the male bees also checked for females in flowering shrubs (*e.g.*  
234 *Rosmarinus officinalis*, *Spartium junceum*, *Emerus major*) where the female bees foraged for  
235 nectar. Pollination events were recorded only when the bee was successfully caught after an  
236 observed pseudo-copulation leading to pollinarium removal. The bees were later identified by  
237 comparison to an *Ophrys* pollinator reference collection at the University of Zürich,  
238 Switzerland.  
239 An additional pollinator-baiting experiment was performed from 10.00 to 13.00 h on the 21<sup>st</sup>  
240 of March 2012 on the Tyrrhenian coast by exposing freshly cut inflorescences of *O.sph*<sub>CUMA</sub>  
241 during local pollinator hours. Visiting bees were caught only after observation of pseudo-  
242 copulation with pollinia removal, and inflorescences were replaced following each pollination  
243 event.

244

245

#### SCENT ANALYSES

246 Floral hydrocarbons produced by the two species were analysed for plants from all  
247 populations for which pollinator choice experiments were performed, *i.e.* *O. sphegodes* from  
248 Cuma, Tyrrhenian Coast (*O.sph*<sub>CUMA</sub>) and Gargano, Adriatic Coast (*O.sph*<sub>GAR</sub>), and *O.*  
249 *exaltata* from Cuma, Tyrrhenian Coast (*O.exa*<sub>CUMA</sub>) and Gargano, Adriatic Coast (*O.exa*<sub>GAR</sub>).  
250 For each sampled individual plant (the same plant used for genetic analyses, plus additional  
251 individuals bearing up to 20 flowers), one labellum of an unpollinated flower was placed in a  
252 2 ml glass vial (Supelco) and rinsed in 500 µl hexane (HPLC grade, Fluka) for one minute  
253 while gently shaken. The labellum was subsequently removed from the vial; all scent samples  
254 were stored at -20° C until analysis. Gas chromatography (GC) was performed following  
255 Mant *et al.* (2005b) with minor modifications as detailed in Xu *et al.* (2011). Several samples

256 were re-analysed for compound identification with a mass selective detector (GC/MSD;  
257 Agilent 5975) using the same oven and column parameters. It is noted that (Z)-11 and (Z)-12  
258 alkenes cannot be discriminated with the parameters used. Compound spectrum and retention  
259 time were compared with those of a synthetic standard, as reported by Xu *et al.* (2011). The  
260 relative amount of each odour compound was calculated as the proportion of total alkene and  
261 alkane amounts with a chain length between 18 and 30 carbons. Principal component analysis  
262 (PCA) was used for analysis of inter-species floral scent variation based on scaled relative  
263 amount of hydrocarbons. The within species differences in floral scent bouquet between  
264 populations were analysed using distance-based tests for homogeneity of multivariate  
265 dispersions (Anderson, 2006), with 999 permutations. The differences for each individual  
266 compound and total alkenes between different populations within species were analysed using  
267 student t-test or Mann–Whitney U test, depending upon the results of testing for normality  
268 and heteroscedasticity of variance using Leven’s and Shapiro-Wilk tests. All statistical  
269 analyses for floral scent were performed in R 2.15.2 (R Development Core Team, 2012).

270

## 271 RESULTS

271

### 272 GENETIC ANALYSES

272

273 AFLP analysis yielded 322 variable markers. Genetic divergence between population pairs  
274 was assessed via  $\Phi_{ST}$  values, yielding the following results: the lowest  $\Phi_{ST}$  values were  
275 detected between *O.sph<sub>ARG</sub>* and *O.sph<sub>CLA</sub>* (0.07) and the highest between *O.sph<sub>ARG</sub>* and  
276 *O.exaltatus* (0.27); *O.sph<sub>CUMA</sub>* and *O.sph<sub>GAR</sub>* populations showed a  $\Phi_{ST}$  of 0.10. Average  $\Phi_{ST}$   
277 was 0.14 between *O. sphegodes* populations, and 0.18 between *O. exaltata* populations.  
278 Interspecific  $\Phi_{ST}$  between *O. sphegodes* and *O. exaltata* populations averaged 0.21 (Table 2,  
279 Fig.1). AMOVA showed that 87% of genetic variance was partitioned among individuals  
280 within a population, while the remaining 13% was explained by variance among populations  
281 (Table 3).

282 PCoA analysis grouped allopatric and sympatric populations based on taxonomic  
283 classification (the first two axes explained 25.9% and 19.4% of the variation, respectively;  
284 Fig. 2). Only a few individuals from sympatric Cuma and Gargano populations were  
285 intermixed between the two groups. PCoA showed that all *O. sphegodes* populations from the  
286 Tyrrhenian Coast (*O.sph<sub>CLA</sub>*, *O.sph<sub>ARG</sub>* and *O.sph<sub>CUMA</sub>*) formed a cohesive group, but the  
287 species populations also largely overlapped with the *O.sph<sub>GAR</sub>* Adriatic population.

288 The most probable number of genetic clusters ( $K$ ) present in the data, (determined  
289 following Evanno *et al.* 2005) was  $K=2$ , corresponding to the assumption that only two

290 species contributed to the sample gene pool. Individuals of the two species exhibited strong  
291 assignment to their respective cluster, with the exclusion of a few putative hybrid individuals  
292 (Fig. 3).

293 A genome scan for  $F_{ST}$  outliers was performed for *O. sphegodes* populations from  
294 Cuma and Gargano by using a trimmed mean  $F_{ST}$  of 0.027 (Fig. 4). Overall, 269 loci were  
295 included in the genome scan, which identified four outliers ( $P < 0.025$ ) below or above the  
296 respective 0.025 or 0.975 quartiles of the expected  $F_{ST}$  distribution. All four outlier loci were  
297 more strongly differentiated than expected. However, among the four loci, only two were  
298 unique to the comparison between *O. sphegodes* populations from Cuma and Gargano, while  
299 the other two loci were found as outliers also in other pairwise population comparisons.

300 Our GLM analysis modelled genetic pair-wise population differentiation ( $\Phi_{ST}$ ) as a  
301 function of geography and shared pollinators. Geography and pollinators were both  
302 significant ( $P < 0.05$ ) factors in explaining population differentiation (Table 4).

303

304

#### POLLINATOR CHOICE EXPERIMENTS

305 Pollinator activity was negligible in the afternoon, and overall activity in Gargano was  
306 notably higher than that in Cuma. The captured pollinator summary is given in Table 5.  
307 *O. exa<sub>GAR</sub>* attracted 10 *C. cunicularius* individuals and *O. sph<sub>GAR</sub>* attracted four *A. nigroaenea*  
308 individuals; both taxa from Gargano attracted their legitimate pollinators independent of  
309 where the plots were located (Gargano and Cuma). Results differed for the two Cuma species.  
310 *O. exa<sub>CUMA</sub>* attracted four *C. cunicularius* individuals, and two individuals of a yet  
311 unidentified bee species in the genus *Eucera*. The *Eucera* bees were only captured in the *O.*  
312 *exaltata* Cuma population, and it is the first time a *Eucera* bee has been reported to pollinate  
313 individuals from the *O. sphegodes/exaltata* lineage. *O. sph<sub>CUMA</sub>* attracted five *A. nigroaenea*  
314 individuals and 46 *A. bimaculata* individuals. The pollinator-baiting experiment performed in  
315 2012 confirmed that *O. sph<sub>CUMA</sub>* attracts *A. bimaculata* and *A. nigroaenea*. These species were  
316 caught six times and twice, respectively, on freshly cut inflorescences.

317

318

#### SCENT ANALYSES

319 The PCA plot of *O. exaltata* and *O. sphegodes* from the Tyrrhenian and Adriatic  
320 coasts showed a clear separation between the two species (Fig. 5), consistent with previous  
321 studies (Xu *et al.* 2011). The distance-based tests for homogeneity of multivariate dispersions  
322 showed that *O. sphegodes* populations from the Tyrrhenian and Adriatic coast were  
323 significantly different in their floral scent bouquet ( $p=0.023$ ; 999 permutations). In contrast,

324 the floral scent bouquet of *O. exaltata* was not significantly different between two populations  
325 ( $p=0.209$ ; 999 permutations). These floral scent differences were mainly due to a higher  
326 proportion of total alkenes in Tyrrhenian populations (Fig. 6). For *O. sphegodes*, the  
327 *O.sph*<sub>CUMA</sub> population produced  $62.8 \pm 5.2$  % total alkenes, whereas the *O.sph*<sub>GAR</sub> population  
328 only produced  $38.7 \pm 15.0$  % total alkenes ( $p < 2.2 \cdot 10^{-16}$ , Mann–Whitney U test). For *O.*  
329 *exaltata*, the *O.exa*<sub>CUMA</sub> and *O.exa*<sub>GAR</sub> populations produced  $69.3 \pm 8.0$  % and  $59.3 \pm 16.2$  %  
330 alkenes respectively ( $p=0.019$ , Mann–Whitney U test) (Fig. 6). It is noticeable that the  
331 *O.sph*<sub>CUMA</sub> population not only produced a higher proportion overall of alkenes that are active  
332 compounds to *A. nigroaenea* (the pollinator of *O. sphegodes*) but also produced three  
333 compounds, (*Z*)-7-C<sub>21</sub>, (*Z*)-7-C<sub>23</sub> and (*Z*)-7-C<sub>25</sub>, that are active compounds to the pollinator of  
334 *O. exaltata*, *C. cunicularius* (Mant *et al.*, 2005a), but were absent from the *O.sph*<sub>GAR</sub>  
335 population.

336

337

## DISCUSSION

338 Overall, our results indicated a pollinator shift, and showed significant differences in floral  
339 scent between populations of *O. sphegodes*. However, significant intra-specific genetic  
340 structuring was not observed between populations. These results suggest the following: (a) the  
341 Cuma and Gargano *O. sphegodes* populations have reached an early stage of divergence, and  
342 are adapting to different pollinator species; or (b) plant-pollinator relationships in some  
343 sexually deceptive orchid species are more geographically variable than has traditionally been  
344 proposed (*i.e.* Kullenberg, 1961).

345 Our previous pollinator choice experiments with *O. sphegodes* and the sympatric *O.*  
346 *exaltata* in semi-natural/disturbed habitats along the Tyrrhenian and Adriatic Italian coasts  
347 indicated the absence of pollinator sharing between these two closely related species (Xu *et*  
348 *al.*, 2011). However, our genetic analyses, even if with a small data set, supported previous  
349 findings showing that hybridisation can sporadically occur (Xu *et al.*, 2011). Interestingly, our  
350 experiments also revealed that the Tyrrhenian coast *O. sphegodes* (*O.sph*<sub>CUMA</sub>) was pollinated  
351 primarily by the bee species *A. bimaculata*. *Andrena nigroaenea* is the typical *O. sphegodes*  
352 pollinator throughout most of the species range (Mant *et al.*, 2005b), but this species was only  
353 responsible for approximately 10% of pollinia removal at Cuma (Table 5). More importantly,  
354 *O. sphegodes* from the Tyrrhenian coast (*O.sph*<sub>CUMA</sub>) maintained its attractiveness to *A.*  
355 *bimaculata* when transferred to Gargano (Adriatic Coast), indicating that *A. bimaculata*  
356 attraction is based on specific floral traits - probably floral scent - and not merely on the

357 (potentially) more frequent occurrence of this bee species at Cuma. To date, *A. bimaculata*  
358 has only been reported as a pollinator of the typically abdomen-pollinated *O. sicula* (Gaskett,  
359 2010), and as a possible pollinator of *O. creticola* on Crete (Paulus & Gack, 1990b; Paulus &  
360 Schlüter, 2007).

361 Floral scent analysis results indicated stronger differentiation between the *O.*  
362 *sphogodes* populations than between the *O. exaltata* populations from the two coastal regions  
363 (Figs. 5, 6) and such differences were mainly due to a higher proportion of alkenes produced  
364 by the *O.sph<sub>CUMA</sub>* population. Three compounds [*i.e.* (*Z*)-7-C<sub>21</sub>, (*Z*)-7-C<sub>23</sub> and (*Z*)-7-C<sub>25</sub>] that  
365 are active to *C. cunicularius* (preferred pollinator of *O. exaltata*) were exclusive to the  
366 *O.sph<sub>CUMA</sub>* population (Fig. 6). These three (*Z*)-7 alkenes have not previously been recorded  
367 as an EAD-active compound for *A. nigroaenea* (Stökl *et al.*, 2005), and the addition of a (*Z*)-7  
368 alkene mix [made up of (*Z*)-7-C<sub>21</sub>, (*Z*)-7-C<sub>23</sub> and (*Z*)-7-C<sub>25</sub>] rendered *O. sphogodes* flowers  
369 less attractive to *A.* possible role played by these three compounds in *A. bimaculata* attraction  
370 has yet to be tested. Interestingly, the Tyrrhenian *O. sphogodes* (*O.sph<sub>CUMA</sub>*) scent continued  
371 to attract *A. nigroaenea*, even though its scent bouquet was different both qualitatively and  
372 quantitatively from that of the Adriatic *O. sphogodes* (*O.sph<sub>GAR</sub>*). Therefore, the two *O.*  
373 *sphogodes* populations share *A. nigroaenea* as pollinator, and no substantial floral isolation  
374 should be expected between them.

375 In spite of the small genetic differences, our data showed that between the seven study  
376 populations, levels of intra-specific gene flow were higher than inter-specific levels. Overall,  
377 lower genetic divergence was detected between allopatric *O. sphogodes* populations than  
378 between sympatric *O. sphogodes* and *O. exaltata* populations. Despite the different pollinators  
379 attracted by each species, the Adriatic *O. sphogodes* (*O.sph<sub>GAR</sub>*) population exhibited a lower  
380 genetic distance from the geographically closer *O.sph<sub>CUMA</sub>* population than from the more  
381 distant *O.sph<sub>CLA</sub>* and *O.sph<sub>ARG</sub>* Tyrrhenian populations. *O.sph<sub>CLA</sub>* and *O.sph<sub>ARG</sub>* probably  
382 attract the same pollinator as the Adriatic *O. sphogodes*, *i.e.* *A. nigroaenea*. Nevertheless,  
383 GLM results suggested that geography and shared pollinators were significant factors in  
384 differentiation between *O. sphogodes* populations. Despite the difficulties in resolving  
385 ancestral polymorphisms due to hybridisation in very recently diverged populations/species,  
386 the low genetic differentiation estimates (*i.e.* low  $\Phi_{ST}$  values; Table 2) observed between  
387 different *O. sphogodes* populations were comparable with, or even lower than, the values  
388 previously reported between other populations of *Ophrys* species using, as in our study, AFLP  
389 markers (Devey *et al.*, 2009; Schlüter *et al.*, 2011b). Therefore, data suggest that *O.*  
390 *sphogodes* populations continue exchanging alleles despite an apparent change in preferred

391 pollinator species. Nevertheless, the genome scan for  $F_{ST}$  outliers identified four loci that  
392 were more strongly differentiated than expected by chance. Among these four loci, two were  
393 consistent with a scenario of divergent selection between *O. sphegodes* populations from  
394 Cuma and Gargano, even if the small sample set and the absence of multiple pairwise  
395 comparisons between populations with the same pollinator combination strongly limit the  
396 power of this analysis.

397         Based on high inter-population gene flow in *O. sphegodes*, independent of pollinators,  
398 we can infer that local adaptation by pollinator shift in the Tyrrhenian *O. sphegodes*  
399 populations has not (yet) generated detectable genome-wide separation from the Adriatic  
400 populations. In an alternative (but not mutually exclusive) scenario, the Tyrrhenian and  
401 Adriatic populations represent a step towards incipient ecological speciation driven by genic  
402 divergence (*i.e.* loci linked to  $F_{ST}$  outliers). In this case, even if the entire genome is  
403 experiencing gene flow, as long as the one or few genes underlying the floral isolation trait  
404 remain differentiated as a consequence of divergent selection, these populations have the  
405 potential to maintain reproductive isolation, regardless of gene flow between them (*i.e.* genic  
406 ecological speciation model and porous genomes; Wu, 2001; Lexer & Widmer, 2008).

407         Our results match studies by Paulus & Gack (1990b), Lorella *et al.*, (2002) and  
408 Claessens & Kleynen (2011) that demonstrate a (narrow) range of related *Ophrys* species  
409 (particularly those with a wide distribution) pollinated by different bees, and suggest that  
410 *Ophrys* - pollinator interactions are flexible rather than a static one-to-one relationship.  
411 Vereecken *et al.* (2011) observed this flexibility in *Ophrys* - pollinator relationships, and  
412 Bower (1996) reported that so-called “minor responders” often accompany the main  
413 pollinator(s) in the sexually deceptive Australian orchid genus *Chiloglottis*. Our finding that a  
414 *Eucera* bee species serves as a minor *O. exaltata* pollinator in the Cuma population suggests  
415 that even bees of other genera can act as secondary pollinators.

416         The different pollination ecotype resolved in the Tyrrhenian *O. sphegodes* population  
417 may represent a response to local selection imposed by variability in the geographic mosaic of  
418 pollinators. Genetic drift is an alternative explanation for the observed pattern in floral odour  
419 bouquet evolution, and its attractiveness to a novel pollinator in the Tyrrhenian populations.  
420 However, the low between-population genetic differentiation (Table 3) is indicative of  
421 extensive gene flow. In addition, the large effective population size typical found in these  
422 terrestrial deceptive orchids (Cozzolino & Widmer, 2005 and reference therein), including our  
423 study system (Mant *et al.*, 2005b), indicates that a drift scenario is less likely compared with  
424 an adaptive process of pollinator shift.

425           A classical question in the evolution of specialised pollination mechanisms is how  
426 much stringency in the relationship between a plant species and its pollinators can limit the  
427 ability of a plant species to adapt to and survive local and temporal changes in the pollinator  
428 community. Interestingly, a few studies on the spatial and temporal breadth of pollinator of  
429 *Ophrys* taxa (including this study) have revealed other, minor pollinators. An imperfect match  
430 in odour between the plant mimic and its model (the female insect) may represent a form of  
431 pre-adaptation for a local or temporary shift in specialised pollinators (Bower, 1996; Peakall  
432 *et al.*, 2010). Indeed, bioassays have shown that sexual deception is a type of imperfect  
433 mimicry, the pollinator bees actively preferring “novel” signals over the more commonly  
434 encountered cues as adaptive responses to promote outbreeding (Vereecken *et al.*, 2008).  
435 Therefore, compound variability involving compounds inactive to some pollinators but active  
436 to others, and negative frequency-dependent selection, might provide an advantage to the  
437 uncommon odour phenotypes by increasing pollination success. Such selection for imperfect  
438 mimicry may facilitate more relaxed signal refinement in mimics to optimally match the  
439 signals released by the mimic’s specific insect models. It would thereby maintain a source of  
440 natural inter-individual variation in floral scent, which could represent the standing genetic  
441 variation necessary for rapid local adaptive responses to a fluctuating pollinator community in  
442 disturbed habitats.

443           Although compelling evidence supports floral adaptation as the basis for ecological  
444 speciation (Kay & Sargent, 2009), a local shift to a different pollinator does not necessarily  
445 lead to speciation, particularly if the selective pressure is transient or fluctuating. *Ophrys*  
446 populations adapted to different pollinators may well represent a very interesting case-study  
447 of incipient speciation by local adaptation. How easily this transient change in preferred  
448 pollinators between locally adapted populations can turn into a “permanent” form of  
449 reproductive isolation and speciation probably depends on the tempo and mode of divergent  
450 selective forces working on local populations, and on the degree and duration of the  
451 disturbances generating the local ecological differences.

452

453

#### FUNDING

454 This work was supported by the PhD program of University of Naples, Salerno Camera  
455 Commercio grant to SC, ETH Zurich (TH0206-2) to FPS, and Swiss National Science  
456 Foundation (SNF 31003A\_130796) to PMS.

457

458

459

## ACKNOWLEDGEMENTS

460 We thank Donata Cafasso and Rosita Rinaldi for help with molecular analyses. Giuseppe  
461 Pavarese gave support with the pollinator experiments in Cuma and Nicolas Vereecken  
462 helped with the identification of pollinating bee species. The authors also thank Michael  
463 Hutchings for his constructive comments on the manuscript and the Write Science  
464 Rightcompany for language editing. The authors are very grateful to Richard Bateman and  
465 another anonymous referee for their extremely constructive comments on an earlier version of  
466 the MS.

467

468

## LITERATURE CITED

469

- 470 Anderson, B., Alexandersson, R. & Johnson, S.D. 2010. Evolution and coexistence of  
471 pollination ecotypes in an African *Gladiolus* (Iridaceae). *Evolution* **64**: 960-972.
- 472 Anderson, M.J. 2006. Distance-based tests for homogeneity of multivariate dispersions.  
473 *Biometrics* **62**: 245-253.
- 474 Bateman, R.M., Bradshaw, E., Devey, D.S., Glover, B.J., Malmgren, S., Sramkó, G. *et al.*  
475 2011. Species arguments: clarifying competing concepts of species delimitation in the  
476 pseudo-copulatory orchid genus *Ophrys*. *Bot. J. Linn. Soc.* **165**: 336-347.
- 477 Beaumont, M.A. & Nichols, R.A. 1996. Evaluating loci for use in the genetic analysis of  
478 population structure. *Proc. R. Soc. London Ser. B* **263**: 1619-1626.
- 479 Bower, C.C. 1996. Demonstration of pollinator-mediated reproductive isolation in sexually  
480 deceptive species of *Chiloglottis* (Orchidaceae: Caladeniinae). *Aust. J. Bot.* **44**: 15-33.
- 481 Butlin, R., Debelle, A., Kerth, C., Snook, R.R., Beukeboom, L.W., Castillo, R.F. *et al.* 2012.  
482 What do we need to know about speciation? *Trends Ecol. Evol.* **27**: 27-39.
- 483 Claessens, J. & Kleynen, J. 2011. The flower of the European orchid: form and function.  
484 Geulle, Netherlands: Published by the authors
- 485 Coyne, J.A. & Orr, H.A. 1998. The evolutionary genetics of speciation. *Philos. Trans. R. Soc.*  
486 *London Ser. B* **353**: 287-305.
- 487 Cozzolino, S. & Widmer, A. 2005. Orchid diversity: an evolutionary consequence of  
488 deception? *Trends Ecol. Evol.* **20**: 487-494.



- 489 Delforge, P. 2006. *Orchids of Europe, North Africa, and the Middle East*. A & C Black,  
490 London.
- 491 Devey, D.S., Bateman, R.M., Fay, M.F. & Hawkins, J.A. 2008. Friends or relatives?  
492 Phylogenetics and species delimitation in the controversial European orchid genus *Ophrys*.  
493 *Ann. Bot.* **101**: 385-402.
- 494 Devey, D.S., Bateman, R.M., Fay, M.F. & Hawkins, J.A. 2009. Genetic structure and  
495 systematic relationships within the *Ophrys fuciflora* aggregate (Orchidaceae: Orchidinae):  
496 high diversity in Kent and a wind-induced discontinuity bisecting the Adriatic. *Ann. Bot.* **104**:  
497 483-495.
- 498 Evanno, G., Regnaut, S. & Goudet, J. 2005. Detecting the number of clusters of individuals  
499 using the software STRUCTURE: a simulation study. *Mol. Ecol.* **14**: 2611-2620.
- 500 Fenster, C.B., Armbruster, W.S., Wilson, P., Dudash, M.R. & Thomson, J.D. 2004.  
501 Pollination syndromes and floral specialization. *Annu. Rev. Ecol. Evol. Syst.* **35**: 375-403.
- 502 Fulton, M. & Hodges, S.A. 1999. Floral isolation between *Aquilegia formosa* and *A.*  
503 *pubescens*. *Proc. R. Soc. London Ser. B* **266**: 2247-2252.
- 504 Gardiner, T. & Vaughan, A. 2009. Shrub clearance and soil disturbance increases bee orchid  
505 *Ophrys apifera* frequency in calcareous grassland at Norton Heath roadside verge, Essex,  
506 England. *Conserv. Evi.* **6**: 39-41.
- 507 Gaskett, A.C. 2010. Orchid pollination by sexual deception: pollinator perspectives. *Biol.*  
508 *Rev.* **86**: 33-75.
- 509 Grant, V. 1971. *Plant speciation*, 2nd edn. Columbia University Press, Columbia.
- 510 Harder, L.D. & Johnson, S.D. 2009. Darwin's beautiful contrivances: evolutionary and  
511 functional evidence for floral adaptation. *New Phytol.* **183**: 530-534.
- 512 Hutchings, M.J. 2010. The population biology of the early spider orchid *Ophrys sphegodes*  
513 Mill. III. Demography over three decades. *Ecology* **98**: 867-878.
- 514 Johnson, S.D. 2006. Pollinator driven speciation in plants. In: *Ecology and evolution of*  
515 *flowers* (Harder, L.D. & Barrett, S.C.H., eds), pp. 295–310. Oxford University Press, New  
516 York.

- 517 Johnson, S.D. & Steiner, K.E. 2000. Generalization versus specialization in plant pollination  
518 systems. *Trends Ecol. Evol.* **15**: 140-143.
- 519 Kay, K.M. & Sargent, R.D. 2009. The role of animal pollination in plant speciation:  
520 integrating ecology, geography, and genetics. *Annu. Rev. Ecol. Evol. Syst.* **40**: 637-656.
- 521 Kullenberg, B. 1961. Studies in *Ophrys* pollination. *Zool. Bidr. Upps.* **34**: 1-340.
- 522 Lexer, C. & Widmer, A. 2008. The genic view of plant speciation: recent progress and  
523 emerging questions. *Philos. Trans. R. Soc. London Ser. B* **363**: 3023-3036.
- 524 Lorella, B., Mahè, G. & Sèitè, F. 2002. Pollinisateurs d'*Ophrys* en Bretagne. *L'Orchidophile*  
525 **151**: 91-96.
- 526 Lowry, D.B., Modliszewski, J.L., Wright, K.M., Wu, C.A. & Willis, J.H. 2008. The strength  
527 and genetic basis of reproductive isolation in flowering plants. *Philos. Trans. R. Soc. London*  
528 *Ser. B* **363**: 3009-3021.
- 529 Macnair, M.R. & Gardner, M. 1998. The evolution of edaphic endemics. In: *Endless forms:*  
530 *Species and speciation* (Howard, D.J. & Berlocher, S.H., eds), pp. 157–171. Oxford  
531 University Press, New York.
- 532 Mallet, J., Meyer, A., Nosil, P. & Feder, J.L. 2009. Space, sympatry and speciation. *J. Evol.*  
533 *Biol.* **22**: 2332-2341.
- 534 Mant, J., Brändli, C., Vereecken, N.J., Schulz, C.M., Francke, W. & Schiestl, F.P. 2005a.  
535 Cuticular hydrocarbons as sex pheromone of the bee *Colletes cunicularius* and the key to its  
536 mimicry by the sexually deceptive orchid, *Ophrys exaltata*. *J. Chem. Ecol.* **31**: 1765-1787.
- 537 Mant, J., Peakall, R. & Schiestl, F.P. 2005b. Does selection on floral odor promote  
538 differentiation among populations and species of the sexually deceptive orchid genus *Ophrys*?  
539 *Evolution* **59**: 1449-1463.
- 540 Minder, A.M. & Widmer, A. 2008. A population genomic analysis of species boundaries:  
541 neutral processes, adaptive divergence and introgression between two hybridizing plant  
542 species. *Mol. Ecol.* **17**: 1552-1563.
- 543 Moccia, M.D., Widmer, A. & Cozzolino, S. 2007. The strength of reproductive isolation in  
544 two hybridizing food-deceptive orchid species. *Mol. Ecol.* **16**: 2855-2866.

- 545 Morjan, C.L. & Rieseberg, L.H. 2004. How species evolve collectively: implications of gene  
546 flow and selection for the spread of advantageous alleles. *Mol. Ecol.* **13**: 1341-1356.
- 547 Nosil, P. 2012. *Ecological speciation*. Oxford University Press, New York.
- 548 Nosil, P., Harmon, L.J. & Seehausen, O. 2009. Ecological explanations for (incomplete)  
549 speciation. *Trends Ecol. Evol.* **24**: 145-156.
- 550 Paulus, H.F. & Gack, C. 1990a. Pollinators as pre-pollination isolation factors. *Isr. J. Bot.* **39**:  
551 43-79.
- 552 Paulus, H.F. & Gack, C. 1990b. Untersuchungen zur Pseudokopulation und  
553 Bestäuberspezifität in der Gattung *Ophrys* im östlichen Mittelmeerraum (Orchidaceae und  
554 Insecta, Hymenoptera, Apoidea). *Jahresber. Nat.wiss. Ver. Wupp.* **43**: 80-118.
- 555 Paulus, H.F. & Schlüter, P.M. 2007. Neues aus Kreta und Rhodos: Bestäubungsbiologie und  
556 molekular-genetische Trennung in der *Ophrys fusca* - Gruppe, mit Neubeschreibungen von  
557 *Ophrys phaidra* Paulus nov.sp., *O. pallidula* Paulus nov.sp. und *O. kedra* Paulus nov.sp. aus  
558 Kreta (Orchidaceae und Insecta, Apoidea). *Jahresber. Nat.wiss. Ver. Wupp.* **60**: 101-151.
- 559 Peakall, R. & Smouse, P.E. 2006. GENALEX 6: genetic analysis in Excel. Population genetic  
560 software for research and teaching. *Mol. Ecol. Notes* **6**: 288-295.
- 561 Peakall, R., Ebert, D., Poldi, J., Barrow, R.A., Francke, W., Bower, C.C. *et al.* 2010.  
562 Pollinator specificity, floral odour chemistry and the phylogeny of Australian sexually  
563 deceptive *Chiloglottis* orchids: implications for pollinator-driven speciation. *New Phytol.* **188**:  
564 437-450.
- 565 Pedersen, H.A. & Faurholdt, H. 2007. *Ophrys: The bee orchids of Europe*. Kew Publishing,  
566 Royal Botanic Gardens, London.
- 567 Pérez-Figueroa, A., García-Pereira, M.J., Saura, M., Rolán-Alvarez, E. & Caballero, A. 2010.  
568 Comparing three different methods to detect selective loci using dominant markers. *J. Evol.*  
569 *Biol.* **23**: 2267-2276.
- 570 Petanidou, T., Kallimanis, A.S., Tzanopoulos, J., Sgardelis, S.P. & Pantis, J.D. 2008. Long-  
571 term observation of a pollination network: fluctuation in species and interactions, relative  
572 invariance of network structure and implications for estimates of specialization. *Ecol. Lett.*  
573 **11**: 654-575.

- 574 Potts, S.G., Biesmeijer, J.C., Kremen, C., Neumann, P., Schweiger, O. & Kunin, W.E. 2010.  
575 Global pollinator declines: trends, impacts and drivers. *Trends Ecol. Evol.* **25**: 345-353.
- 576 Pritchard, J.K., Stephens, M. & Donnelly, P. 2000. Inference of population structure using  
577 multilocus genotype data. *Genetics* **155**: 945-959.
- 578 R Development Core Team. 2012. *R: A language and environment for statistical computing*.  
579 R Foundation for Statistical Computing, Vienna, Austria.
- 580 Räsänen, K. & Hendry, A.P. 2008. Disentangling interactions between adaptive divergence  
581 and gene flow when ecology drives diversification. *Ecol. Lett.* **11**: 624-636.
- 582 Rosenberg, N.A. 2004. DISTRUCT: a program for the graphical display of population  
583 structure. *Mol. Ecol. Notes* **4**: 137-138.
- 584 Schemske, D.W. & Bradshaw, H.D. Jr. 1999. Pollinator preference and the evolution of floral  
585 traits in monkeyflowers (*Mimulus*). *Proc. Natl. Acad. Sci. USA* **96**: 11910-11915.
- 586 Schiestl, F.P. 2012. Animal pollination and speciation in plants: general mechanisms and  
587 examples from the orchids. In: *Evolution of plant-pollinator relationships* (Patiny, S., ed), pp.  
588 263–278. Cambridge University Press, Cambridge.
- 589 Schiestl, F.P. & Ayasse, M. 2002. Do changes in floral odor cause speciation in sexually  
590 deceptive orchids? *Plant Syst. Evol.* **234**: 111-119
- 591 Schiestl, F.P. & Schlüter, P.M. 2009. Floral isolation, specialized pollination, and pollinator  
592 behavior in orchids. *Annu. Rev. Entomol.* **54**: 425-446.
- 593 Schlüter, P.M. & Harris, S.A. 2006. Analysis of multilocus fingerprinting data sets containing  
594 missing data. *Mol. Ecol. Notes* **6**: 569-572.
- 595 Schlüter, P.M., Ruas, P.M., Kohl G., Ruas, C.F., Stuessy, T.F. & Paulus, H.F. 2011b.  
596 Evidence for progenitor-derivative speciation in sexually deceptive orchids. *Ann. Bot.* **108**:  
597 895-906.
- 598 Schlüter, P.M., Xu, S., Gagliardini, V., Whittle, E., Shanklin, J., Grossniklaus, U. *et al.*  
599 2011a. Stearoyl-acyl carrier protein desaturases are associated with floral isolation in sexually  
600 deceptive orchids. *Proc. Natl. Acad. Sci. USA* **108**: 5696-5701.

- 601 Scopece, G., Cozzolino, S., Johnson, S.D. & Schiestl, F.P. 2010. Pollination efficiency and  
602 the evolution of specialized deceptive pollination systems. *Am. Nat.* **175**: 98-105.
- 603 Slatkin, M. 1987. Gene flow and the geographic structure of natural populations. *Science* **236**:  
604 787-792.
- 605 Spaethe, J., Moser, H.W. & Paulus, H.F. 2007. Increase of pollinator attraction by means of a  
606 visual signal in the sexually deceptive orchid, *Ophrys heldreichii* (Orchidaceae). *Plant Syst.*  
607 *Evol.* **264**: 31-40.
- 608 Stökl, J., Paulus, H.F., Dafni, A., Schulz, C., Francke, W. & Ayasse, M. 2005. Pollinator  
609 attracting odour signals in sexually deceptive orchids of the *Ophrys fusca* group. *Plant Syst.*  
610 *Evol.* **254**: 105-120.
- 611 Stuurman, J., Hoballah, M.E., Broger L., Moore, J., Basten, C. & Kuhlemeier, C. 2004.  
612 Dissection of floral pollination syndromes in *Petunia*. *Genetics* **168**: 1585-1599.
- 613 Thibert-Plante, X. & Hendry, A.P. 2010. When can ecological speciation be detected with  
614 neutral loci? *Mol. Ecol.* **19**: 2301-2314.
- 615 Van Valen, L. 1976. Ecological species, multispecies, and oaks. *Taxon* **25**: 233-239.
- 616 Vereecken, N.J. & Schiestl, F.P. 2008. The evolution of imperfect floral mimicry. *Proc. Natl.*  
617 *Acad. Sci. USA* **105**: 7484-7488.
- 618 Vereecken, N.J. & Schiestl, F.P. 2009. On the roles of colour and scent in a specialized floral  
619 mimicry system. *Ann. Bot.* **104**: 1077-1084.
- 620 Vereecken, N.J., Streinzer, M., Ayasse, M., Spaethe, J., Paulus, H.F., Stökl, J. *et al.* 2011.  
621 Integrating past and present studies on *Ophrys* pollination – a comment on Bradshaw *et al.*  
622 *Bot. J. Linn. Soc.* **165**: 329-335.
- 623 Vos, P., Hogers, R., Bleeker, M., Reijans, M., van de Lee, T., Hornes, M. *et al.* 1995. AFLP:  
624 a new technique for DNA fingerprinting. *Nucleic Acids Res.* **23**: 4407-4414.
- 625 Waser, N.M. & Ollerton, J. 2006. Plant-pollinator interactions: from specialization to  
626 generalization. In: *Geographical variation in diversity and specificity of pollination systems*  
627 (Ollerton, J., Johnson, S.D. & Hingston, A.B., eds), pp. 283-308. University of Chicago Press,  
628 Chicago.

- 629 Wu, C-I. 2001. The genic view of the process of speciation. *J. Evol. Biol.* **14**: 851-865.
- 630 Xu, S., Schlüter, P.M. & Schiestl, F.P. (2012a) Pollinator-driven speciation in sexually  
631 deceptive orchids. *Int. J. Ecol.*, doi:10.1155/2012/285081.
- 632 Xu, S., Schlüter, P.M., Grossniklaus, U. & Schiestl, F.P. 2012b. The genetic basis of  
633 pollinator adaptation in a sexually deceptive orchid. *PLoS Genet.*,  
634 doi:10.1371/journal.pgen.1002889.
- 635 Xu, S., Schlüter, P.M, Scopece, G., Breitkopf, H., Gross, K., Cozzolino, S. *et al.* 2011. Floral  
636 isolation is the main reproductive barrier among closely related sexually deceptive orchids.  
637 *Evolution* **65**: 2606-2620.
- 638 Zhivotovsky, L.A. 1999. Estimating population structure in diploids with multilocus  
639 dominant DNA markers. *Mol. Ecol.* **8**: 907-913.

640 TABLE 1. Examined populations of *O. exaltata* and *O. sphegodes*.

641

Species	Sample Size	Origin	Date	Collector
<i>O. exaltata</i> (O.exa <sub>GAR</sub> )	17	Capoiale, Gargano, Puglia, Italy Marina di	Mar-09	H. Breitkopf
<i>O. exaltata</i> (O.exa <sub>TUS</sub> )	10	Castagneto, Tuscany, Italy	Apr-09	H. Breitkopf
<i>O. exaltata</i> (O.exa <sub>CUMA</sub> )	15	Cuma, Campania, Italy	Mar-09	H. Breitkopf
<i>O. sphegodes</i> ssp. <i>classica</i> (O.sph <sub>CLA</sub> )	12	Porto San Stefano, Tuscany, Italy	Mar-10	R. Souche
<i>O. sphegodes</i> (O.sph <sub>CUMA</sub> )	15	Cuma, Campania, Italy	Mar-09	H. Breitkopf
<i>O. sphegodes</i> (O.sph <sub>GAR</sub> )	17	Capoiale, Gargano, Puglia, Italy	Mar-09	H. Breitkopf
<i>O. sphegodes</i> ssp. <i>argentaria</i> (O.sph <sub>ARG</sub> )	9	Caldine-Fiesole, Tuscany, Italy	Apr-09	H. Breitkopf

642

643

644 TABLE 2. Pairwise  $\Phi_{ST}$  values (Coefficient: Standard Jaccard. Distance Transformation:  $d=1-$   
 645  $s$ ) for the examined populations.

646  
647

648	O.sph <sub>ARG</sub>	O.sph <sub>CLA</sub>	O.exa <sub>TUS</sub>	O.sph <sub>GAR</sub>	O.exa <sub>GAR</sub>	O.sph <sub>CUMA</sub>	O.exa <sub>CUMA</sub>
649	0.00						
650	0.07	0.00					
651	0.28	0.24	0.00				
652	0.17	0.16	0.23	0.00			
653	0.27	0.25	0.20	0.13	0.00		
654	0.18	0.17	0.24	0.10	0.19	0.00	
655	0.26	0.25	0.16	0.21	0.16	0.17	0.00

656

657

658

659

660

661

662 **Table 3** Analysis of molecular variance (AMOVA) for AFLP markers (d.f.: degree of  
 663 freedom; SSD: sum of squared deviations; %: proportion of variance components, standard  
 664 error  $\leq 2.93\%$ ;  $\Phi$ : genotypic variation; \*  $P \leq 0.001$ ).

665

Source of variation	d.f.	SSD	%	$\Phi$
Among all populations	6	603.9	13	0.133*
Within all populations	87	2861.7	87	
Among <i>O.sphogodes</i> populations	3	231.2	10	0.098*
Within <i>O. sphogodes</i> populations	48	1548.7	90	
Among <i>O.exaltata</i> populations	2	193.9	12	0.121*
Within <i>O.exaltata</i> populations	39	1313.0	88	

666

667



668 TABLE 4. Generalised linear model of pairwise  $\Phi_{ST}$  values among *O. sphegodes* populations  
 669 as explained by the factors indicated. To facilitate the analysis, the Cuma population was  
 670 assumed to have a different pollinator than the remaining *O. sphegodes* populations.

<i>Factor</i>	<i>Estimate</i>	<i>S.E.</i>	<i>t-value</i>	<i>p-value</i>	<i>Significance</i>
<i>Intercept</i>	0.12636	0.00642	19.672	0.00257	**
<i>Geography</i>	0.00639	0.00088	7.232	0.01859	*
<i>Shared Pollinator</i>	-0.53657	0.10196	-5.263	0.03426	*
<i>Geography × Shared Pollinators</i>	0.09420	0.01817	5.184	0.03525	*

671 Significance: \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .

672

673

674 TABLE 5. Pollinator choice experiment: Bee counts. CUMA, Campania, Tyrrhenian Coast;

675 GARGANO, Puglia, Adriatic Coast.

		<i>O. sphegodes</i>	<i>O. sphegodes</i>	<i>O. exaltata</i>	<i>O. exaltata</i>
		CUMA	GARGANO	CUMA	GARGANO
Cuma choice-plots	<i>A. bimaculata</i>	4			
	<i>A. nigroaenea</i>	1	2		
	<i>C. cunicularius</i>				2
	<i>Eucera</i> sp.			2	
Gargano choice-plots	<i>A. bimaculata</i>	42			
	<i>A. nigroaenea</i>	4	2		
	<i>C. cunicularius</i>			4	8
	<i>Eucera</i> sp.				
all choice-plots (Cuma+Gargano)	<i>A. bimaculata</i>	46 (4+42)			
	<i>A. nigroaenea</i>	5 (1+4)	4 (2+2)		
	<i>C. cunicularius</i>			4 (0+4)	10 (2+8)
	<i>Eucera</i> sp.			2 (2+0)	

676

677

## FIGURE LEGENDS

678

679

680 FIG. 1. Map of Central Italy with  $\Phi_{ST}$  values plotted for pairwise population comparisons.  
681 Black dots indicate the locations of sympatric populations of *O. sphegodes* and *O. exaltata* in  
682 the Italian regions of Tuscany, Campania and Apulia.

683

684 FIG. 2. PCoA plot based on individual genetic distances calculated from 322 polymorphic  
685 AFLP markers. The first two axes explained 25.9% and 19.4% of the variation, respectively.

686

687 FIG. 3. Summary of population structure in *O. exaltata* and *O. sphegodes* using Bayesian  
688 assignment analysis for a  $K=2$  model. Most individuals from *O. sphegodes* populations  
689 showed strong assignment probabilities associated with cluster 1 (dark grey), whereas  
690 specimens from *O. exaltata* were classified with cluster 2 (light grey). Two putative hybrid  
691 individuals showing admixed proportions were allied with the *O. sphegodes* population from  
692 Cuma.

693

694 FIG. 4. Dfdist plot of  $F_{ST}$  values against heterozygosity estimates for the *O. sphegodes*  
695 population pair (*O.sph*<sub>CUMA</sub> and *O.sph*<sub>GAR</sub>). Each circle indicates an AFLP marker. The  
696 lower, intermediate and higher lines represent 5%, 50% and 95% confidence intervals,  
697 respectively. Outlier loci ( $P < 0.025$ ) are labelled and displayed as black dots.

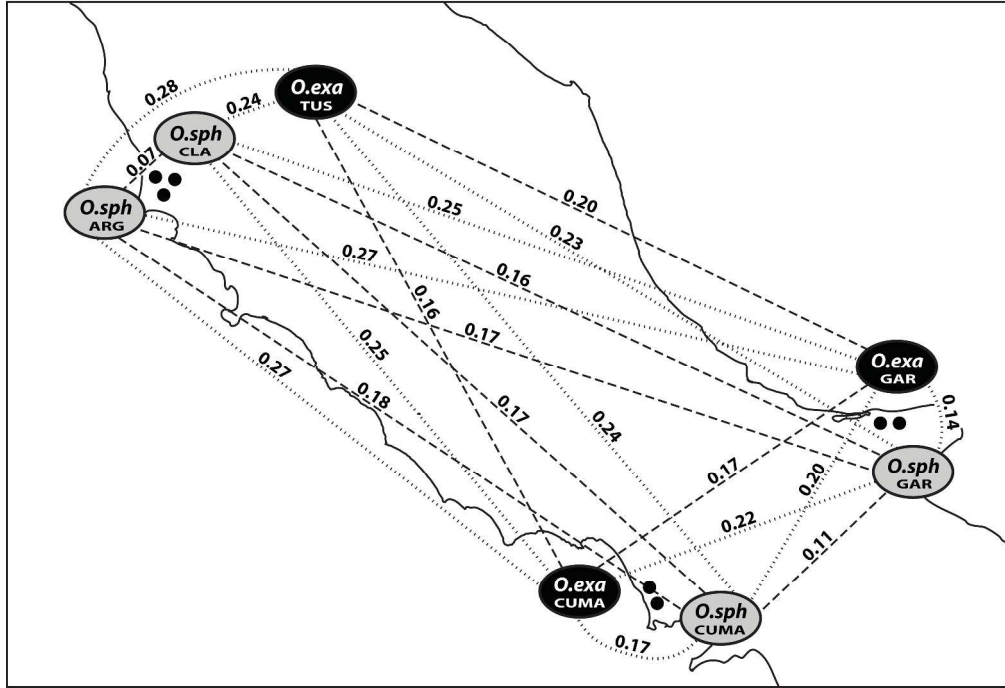
698

699 FIG. 5. PCA plot of floral scent derived from sympatric *O. sphegodes* and *O. exaltata*  
700 populations.

701

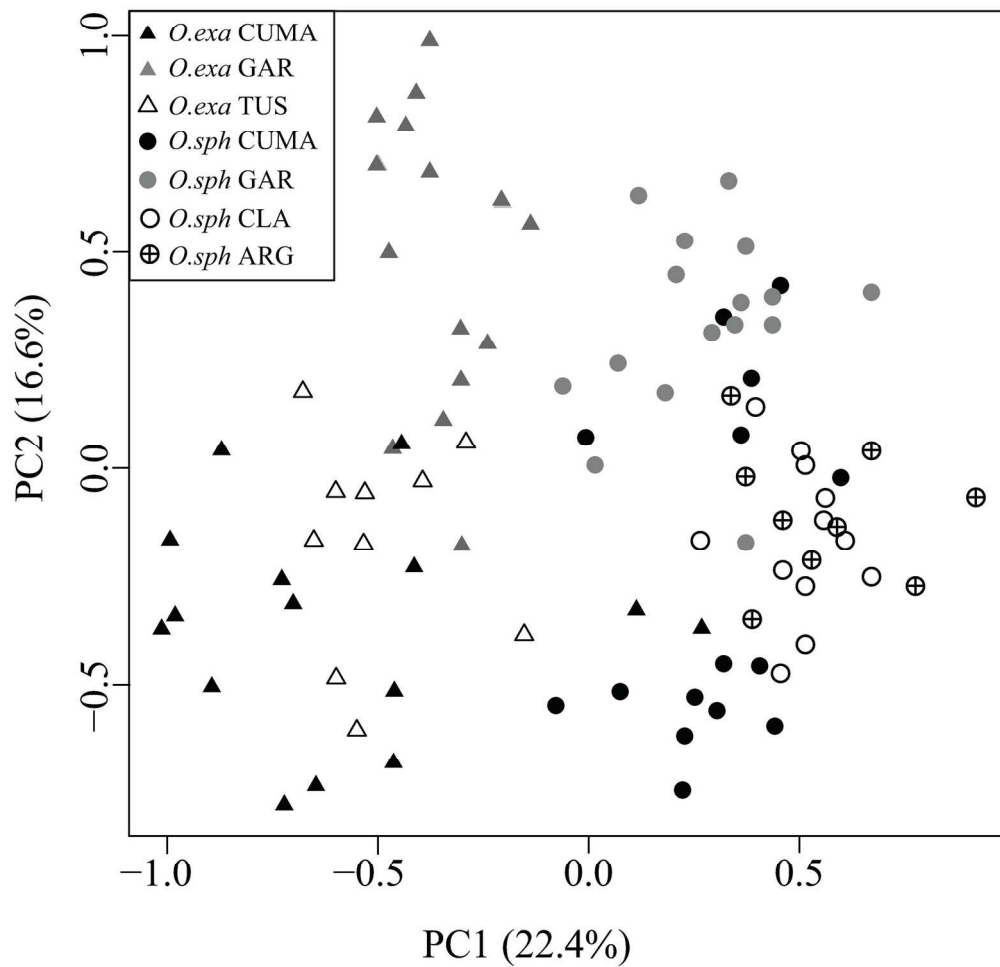
702 FIG. 6. Scent profiles, from sympatric *O. exaltata* (*O.exa*<sub>CUMA</sub> and *O.exa*<sub>GAR</sub>), and *O.*  
703 *sphegodes* (*O.sph*<sub>CUMA</sub> and *O.sph*<sub>GAR</sub>) populations.

704

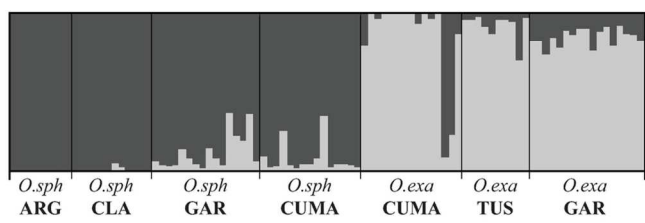


341x293mm (300 x 300 DPI)

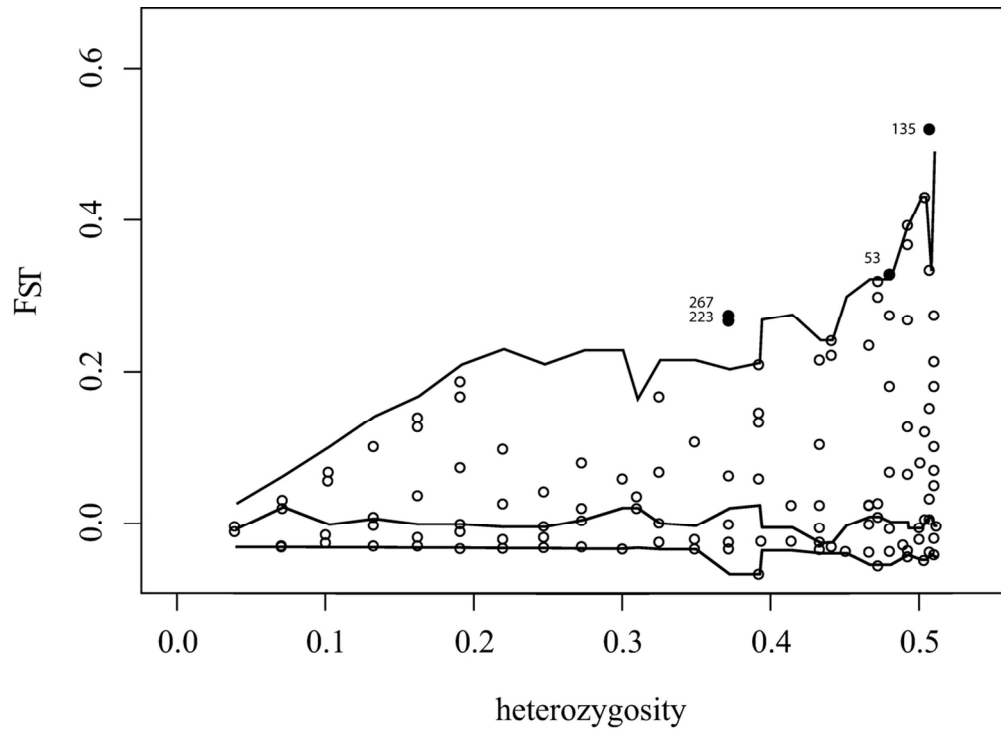
AFLP's



166x173mm (300 x 300 DPI)

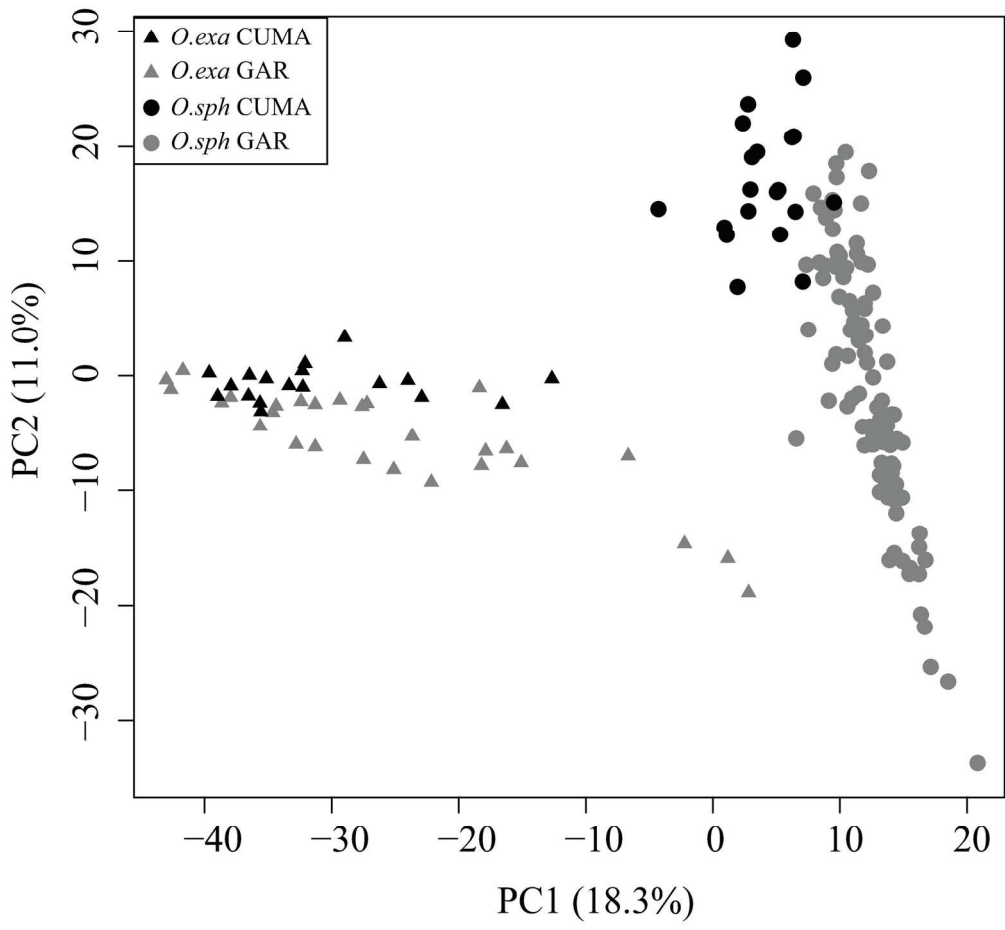


121x112mm (300 x 300 DPI)



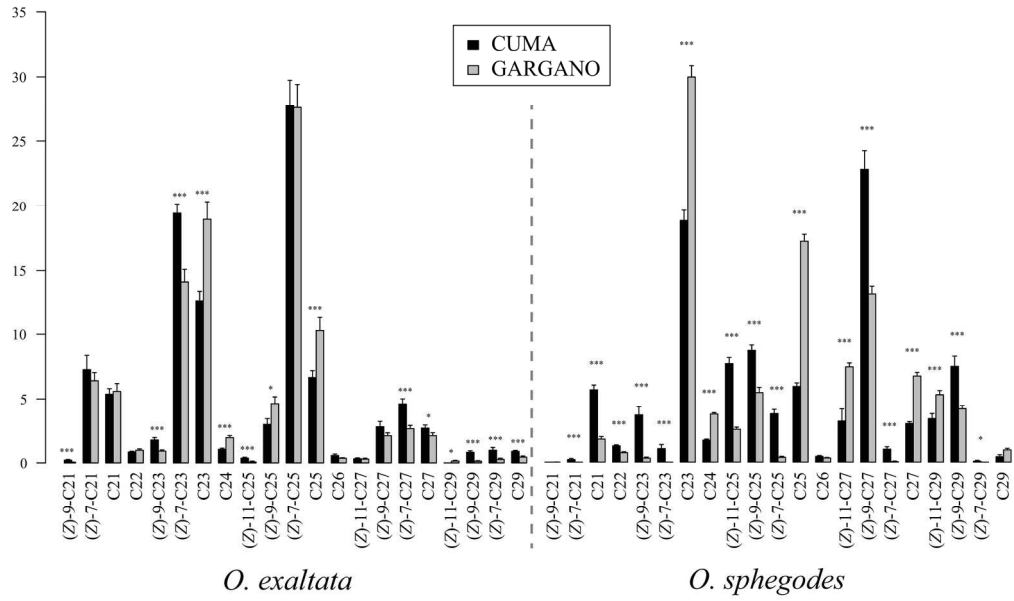
110x79mm (300 x 300 DPI)

### Floral Scent



166x164mm (300 x 300 DPI)





174x104mm (300 x 300 DPI)