
Campylobacter Antimicrobial Drug Resistance among Humans, Broiler Chickens, and Pigs, France

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We describe isolates from human *Campylobacter* infection in the French population and the isolates' antimicrobial drug resistance patterns since 1986 and compare the trends with those of isolates from broiler chickens and pigs from 1999 through 2004. Among 5,685 human *Campylobacter* isolates, 76.2% were *C. jejuni*, 17.2% *C. coli*, and 5.0% *C. fetus*. Resistance to nalidixic acid increased from 8.2% in 1990 to 26.3% in 2004 ($p < 10^{-3}$), and resistance to ampicillin was high over time. Nalidixic acid resistance was greater for *C. coli* (21.3%) than for *C. jejuni* (14.9%, $p < 10^{-3}$). *C. jejuni* resistance to ciprofloxacin in broilers decreased from 31.7% in 2002 to 9.0% in 2004 ($p = 0.02$). The patterns of resistance to quinolones and fluoroquinolones were similar between 1999 and 2004 in human and broiler isolates for *C. jejuni*. These results suggest a potential benefit of a regulation policy limiting use of antimicrobial drugs in food animals.

Campylobacter infections are, along with *Salmonella* infections, the most common cause of bacterial diarrhea in humans worldwide (1–6). A recent study on illness and death due to foodborne infections in France estimated an isolation rate of 27–37/100,000 persons/year for *Campylobacter* infection (7).

Campylobacter are part of normal enteric flora in animals (poultry, pigs, and cattle) and can be transmitted to humans through contaminated foods (8). Several studies identified chicken as the main source of infection (9,10). Most *Campylobacter* enteric infections are self-limited and do not require antimicrobial drug treatment. However,

severe or long-lasting *Campylobacter* infections do occur and may justify antimicrobial drug therapy. Macrolides as first-line therapy and fluoroquinolones as alternative therapy are recommended (2,11). Resistance of *Campylobacter* to antimicrobial agents has increased substantially during the past 2 decades and has become a matter of concern in severe human *Campylobacter* infections (12–14). Combined studies in humans and poultry have implicated the use of fluoroquinolones in poultry in the emergence of drug resistance (15–17). As a consequence, in 2004 the US Food and Drug Administration withdrew the 1995/1996 approval for the new animal drug application to use enrofloxacin for prophylaxis treatment or growth promotion in poultry (18). Veterinary licensing of enrofloxacin in poultry was approved by the European Union (EU) in 1991, and in 1999 the EU recommended limiting the use of fluoroquinolones in poultry.

In this article, we describe characteristics of human *Campylobacter* isolates in France and trends of antimicrobial resistance in such isolates from 1986 to 2004. Trends of *Campylobacter* antimicrobial drug resistance in human isolates were compared with those of isolates from broiler chickens and pigs between 1999 and 2004.

Materials and Methods

National Surveillance System for Human *Campylobacter* Infections

Surveillance for *Campylobacter* infections in France is based on a network of laboratories that send their isolates to the National Reference Center for *Campylobacter* and *Helicobacter* (CNRCH). The network of participating laboratories, limited to hospital laboratories from 1986 to 2001, was complemented by private (which usually cared

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for outpatients) and additional hospital laboratories in 2002 to be more representative of the whole French territory (19–21). The network is currently composed of 325 private laboratories (9% of the 3,444 registered private laboratories in France), located in 90 of the 95 districts in mainland France, and 92 hospital laboratories (25% of the 409 registered hospital laboratories). Participating laboratories perform a systematic screening for *Campylobacter* in stools. Each isolate recovered is sent to CNRCH in a transport medium (medium for storage of bacteria, Bio-Rad, Marnes-la-Coquette, France) with information on the type of specimen; date and district of isolation; patient's age, sex, and history of travel abroad; and eventual context of an outbreak.

On reception at CNRCH, isolates are tested for viability, confirmed as *Campylobacter* by standard phenotypic identification, and identified at the species level with phenotypic methods and real-time PCR to differentiate between *C. jejuni*, *C. coli* and *C. fetus* (2,22). The other species are identified by comparing their 16S rDNA sequences to those of DNA databases by using the BLAST program (23). Identification at the species level is considered correct when at least 99% identity occurs with only 1 species.

Antimicrobial Drug Resistance Monitoring of *Campylobacter* in Humans

Campylobacter isolates from all species were evaluated for susceptibility to 7 antimicrobial drugs (nalidixic acid, ciprofloxacin [since 2000], erythromycin, amoxicillin, gentamicin, tetracycline, and doxycycline [since 2003]) by the agar diffusion method on Mueller-Hinton agar enriched with 5% sheep blood by using antibiotic disks, according to recommendations for *Campylobacter* of the Antibiogram Committee of the French Society for Microbiology (CA-SFM) (24). Since the hospital laboratory network set up in 1986 was extended to private laboratories in 2002, antimicrobial susceptibility trends were analyzed exclusively for hospital laboratory isolates between 1986 and 2004. Multidrug resistance was defined as resistance to ≥ 2 antimicrobial drugs.

Antimicrobial Drug Resistance Monitoring in Broilers and Pigs

Surveillance of *Campylobacter* antimicrobial drug resistance was implemented in France in 1999 for broilers in conventional and free-range broiler farms and in 2000 for pigs as part of a surveillance program on resistance in sentinel bacteria (*Escherichia coli* and *Enterococcus* spp.) and zoonotic bacteria (*Salmonella* spp. and *Campylobacter* spp.) in animal products for human consumption. Thus, data collection began just after the ban of 4 antimicrobial growth promoters (bacitracin zinc, spiramycin, virginini-

amycin, and tylosin phosphate) by the European Community (EC) Council Regulation (No. 2821/98, December 1998). Conventional broiler flocks are characterized by an indoor rearing period of 6 weeks, and free-range broiler flocks have an indoor rearing period of 6 weeks followed by 6 additional weeks with access to an open-air area.

From 200 to 600 broiler cecal samples or pig fecal samples were collected each year in 10 broiler and 10 pig slaughterhouses representative of French production of these animals for human consumption (25). Strain isolation was performed in a central laboratory (Agence Française de Sécurité Sanitaire des Aliments [AFSSA], Ploufragan, France) for the first 2 years and then in district veterinary laboratories, except for antimicrobial susceptibility testing. After identifying isolates by using multiplex PCR (26), the MIC of ampicillin, nalidixic acid, enrofloxacin or ciprofloxacin, tetracycline, erythromycin, and gentamicin were determined by agar dilution. As for human isolates, susceptibility to antimicrobial drugs was categorized according to the 2004 statement of the CA-SFM (24). The study of antimicrobial resistance of *Campylobacter* from animal sources was supported by the French Ministry of Agriculture.

Statistical Analysis

Differences between proportions and isolation rates were tested by χ^2 and Fisher exact tests. Means were compared with Student and Fisher tests. Patterns of antimicrobial resistance were analyzed by 4-year increments from 1986 to 2004.

Results

Surveillance for Human *Campylobacter* Infections

From April 2002 to December 2004, CNRCH received 5,685 presumptive *Campylobacter* isolates. Among the 5,112 (89.9%) viable isolates, 3,896 (76.2%) were *C. jejuni*, 878 (17.2%) *C. coli*, 257 (5.0%) *C. fetus*, 21 (0.4%) *C. lari*, 40 (0.8%) *Arcobacter butzleri*, and 13 (0.25%) other species of *Campylobacter*. Seven strains (0.1%) were *Helicobacter* spp. A seasonal increase during the warmer months was noted and was more pronounced for *C. jejuni*.

The median age of patients was 29.4 years (range 5 days–100 years). Thirteen (0.2%) were newborns (5–30 days), 258 (4.5%) infants (1–11 months), 1,907 (33.5%) children (1–10 years), 2,555 (44.9%) ages 11–65 years, and 767 (13.5%) >65 years (Figure 1). Isolation of *Campylobacter* was more frequent among male than female patients (male/female ratio = 1.2, $p = 0.04$), except for young adults (16–30 years), with a male/female ratio = 0.9 ($p < 10^{-3}$). The ratio of *C. jejuni* to *C. coli* varied between

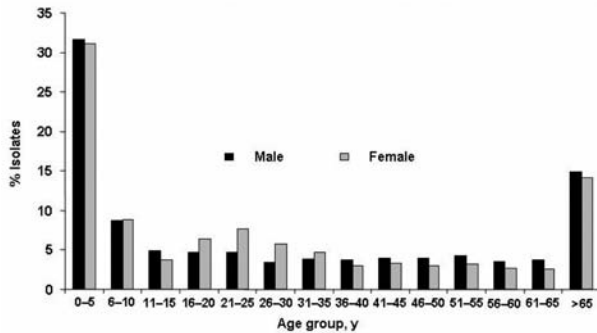


Figure 1. Distribution of *Campylobacter* isolates according to age and sex of patient, France, 2002–2004.

4.5 and 7.2 in those <30 years of age and decreased thereafter. *C. fetus* was isolated among adults >30 years of age and peaked in the elderly ($p < 10^{-3}$, Figure 2).

Among the 5,620 isolates with a known clinical source, 5,253 (93.4%) were isolated from stools, 308 (5.5%) from blood, and 50 (0.9%) from other sites presumably seeded as a result of bacteremic infections. Both *C. jejuni* and *C. coli* were isolated essentially from stools, whereas 158 (63.5%) of 249 *C. fetus* isolates were from blood. Patients with blood isolates were older than those with stool isolates (median age 69 years vs. 19.3 years, $p < 10^{-3}$).

Travel history was available for 1,370 (24.1%) case-patients; 184 (3.2%) reported traveling outside France during the 2 weeks before onset of illness. The country of travel was specified for 169 (91.8%) case-patients (Africa, 98 persons; Asia, 26; Europe, 16; South America, 10; and other countries, 19).

Antimicrobial Drug Resistance of Human *Campylobacter* Isolates

Resistance to nalidixic acid and tetracycline/doxycycline increased from 1986–1989 to 2002–2004 ($p < 10^{-3}$, Figure 3). Resistance to ampicillin, although frequent, decreased from 49.2% (1,027/2,087) in 1986–1989 to 42.4% (501/1,198) in 2002–2004 ($p < 10^{-3}$). Resistance to erythromycin remained low, and no isolate was resistant to gentamicin.

Nalidixic acid resistance increased from 8.2% (26/315) in 1990 to 26.3% (115/438) in 2004 ($p < 10^{-3}$). Resistance was greater for *C. coli* (21.3%) than *C. jejuni* (14.9%, $p < 10^{-3}$, Figure 4). Nalidixic acid resistance increased for *C. jejuni* >4-fold from 1995 to 1997 and for *C. coli* >3-fold from 1994 to 1996. Then, resistance decreased for both *C. coli* and *C. jejuni* in 1999 but remained higher than before 1995 (Figure 4). Ciprofloxacin resistance, tested since 2000, followed the same pattern (Table).

Fifty-eight percent of *Campylobacter* isolates were resistant to ≥ 1 drug, 34.7% to ≥ 2 drugs, and 20.0% to ≥ 3 drugs. The most common multidrug resistance (≥ 2 drugs) patterns included resistance to nalidixic acid or ciprofloxacin, to doxycycline, and to ampicillin.

Among the case-patients ≤ 15 years of age, 28.0% (618/2,207) had a *Campylobacter* strain resistant to nalidixic acid compared with 37.6% (1,029/2,736) of the case-patients >15 years of age ($p < 10^{-3}$). The proportion of resistance to ciprofloxacin did not vary according to age (27.3% of case-patients ≤ 15 years and 27.9% >15 years). For ampicillin, 41.9% (925/2,207) of case-patients ≤ 15 years had a resistant strain compared with 37.3% (1,024/2,736) of the case-patients >15 years ($p = 0.001$).

Of the case-patients who traveled abroad, for which strain resistance was available, 40.3% (67/166) had a strain resistant to ciprofloxacin, compared with 27.0% (294/1,090) of case-patients who did not travel abroad ($p < 10^{-3}$). For nalidixic acid, 42% (70/166) of case-patients who traveled abroad compared with 34.7% (378/1,090) of case-patients who did not have a resistant strain ($p = 0.06$). Resistance to ampicillin was present for 28.3% (47/166) who had traveled abroad compared with 31.1% (339/1,090) for those who had not ($p = 0.01$).

Antimicrobial Resistance in Broilers and Pigs

Between 1999 and 2004, a total of 544 *C. jejuni* and 374 *C. coli* isolates were recovered from poultry, and 871 *C. coli* were recovered from pigs by the antibiotic resistance surveillance system. Among the broiler isolates, the proportion of *C. jejuni* from animals raised in standard and export production facilities gradually decreased from 83.5% (279/334) in 1999 to 43% (28/65) in 2004 ($p < 10^{-3}$), while the proportion of *C. jejuni* decreased from 32% (18/57) to 10% (4/40) in the free-range production facilities ($p = 0.01$).

Campylobacter isolates were inconstantly sensitive to ampicillin, and a high proportion of isolates resistant to tetracycline was recorded in poultry and pigs, but all strains remained sensitive to gentamicin (Table). Isolates from pigs were less frequently resistant to ampicillin but more often resistant to tetracycline. For erythromycin,

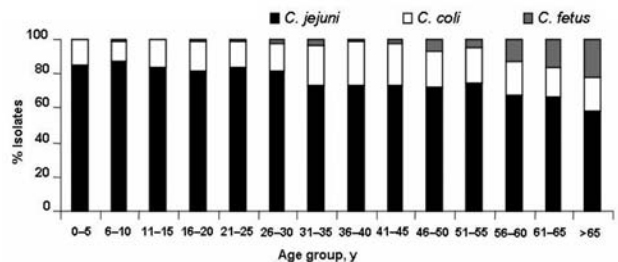


Figure 2. Distribution of human *Campylobacter* strains by species and patient age group, France, 2002–2004.

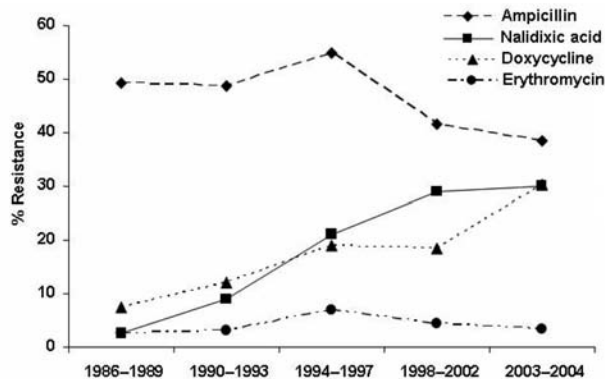


Figure 3. Resistance of human *Campylobacter* spp. isolates to antimicrobial agents, France, 1986-2004.

resistance was rare among *C. jejuni* strains (1.8%), but much more frequent for *C. coli* (21.1% of broiler isolates and 58.9% of pig isolates, $p < 10^{-3}$).

In broilers, *C. coli* strains were more often resistant to ciprofloxacin (37.4%, 140/374) than were *C. jejuni* strains (19.5%, 106/544, $p < 10^{-3}$) (Table, Figure 5). *C. coli* resistance to ciprofloxacin increased in pigs from 12.3% (39/316) in 2000 to 26.9% (18/67) in 2004 ($p = 0.002$). For *C. jejuni* in broilers, after an increase in resistance from 16.8% (50/297) in 1999 to 31.7% (13/41) in 2002 ($p = 0.02$), resistance to ciprofloxacin decreased to 9.0% (3/32) in 2004 ($p = 0.02$) (Table, Figure 5). Similar trends were observed for quinolone resistance. The trend of resistance to nalidixic acid and ciprofloxacin was similar for *C. jejuni* isolated from humans and broilers between 1999 and 2004 (Table, Figure 5).

Discussion

Our surveillance of *Campylobacter* isolates in France indicates some differences with findings from other western countries, i.e., a greater proportion of *C. coli* (17.0%). The epidemiologic characteristics of *Campylobacter* infections were, however, similar. *Campylobacter* is predominant in the summer (27), the isolation rate was much greater in children <5 years of age, and *C. jejuni* was predominant. The rate of resistance to certain antimicrobial drugs increased substantially from 1990 to 2004, and the proportion of resistant isolates varied according to *Campylobacter* sp. Resistance to quinolones was greater for *C. coli* (21.3%) than for *C. jejuni* (14.9%). Quinolone and fluoroquinolone resistance for *C. coli* was high in broilers and pigs. Resistance to quinolone and fluoroquinolone for *C. jejuni* had the same pattern over time in broilers and human isolates.

The proportion of *C. coli* was higher in France (17%) than in the United States (<1%) or Belgium (11%) (2,28). Methods for characterization of the species vary by coun-

try and by laboratory within a country (29,30). In France, CNRCH routinely characterizes the species using a combination of phenotypic and molecular methods (specific PCR) with verification of discrepant results (22). In some other countries, *Campylobacter* are not routinely characterized at the species level and could be incorrectly identified as *C. jejuni* or *Campylobacter* spp. This may account for an underestimation of species other than *C. jejuni* in some countries and therefore some distortion of the proportion of antimicrobial drug resistance by species. In France, the high proportion of *C. coli* isolates is probably real, as an increasing proportion of *C. coli* is colonizing broilers (31). This trend may be related to the use of different isolation and identification methods, to a recent increase in the ratio of *C. coli* to *C. jejuni*, or both. The ban of antimicrobial growth promoters and of animal protein-based feed may have influenced the digestive bacterial flora equilibrium of chickens. Udayamputhoor et al. showed that the ceca of birds that receive plant protein-based feed are less likely to be colonized with *C. jejuni* than the ceca of birds that receive other types of feed (32). Another hypothesis is that because 100% of pigs are colonized by *C. coli* in France, the proximity of pig and poultry farms in the main producing regions may result in cross-contamination. However, this explanation is unlikely because *C. coli* strains isolated from broilers and pigs had different antimicrobial resistance patterns, and *C. coli* poultry strains clustered separately from those of porcine origin (33). Nonetheless, strains may undergo different selection pressures.

Resistance to ampicillin is of clinical interest because this drug may be used for the treatment of severe *Campylobacter* infections. The proportion of resistance to ampicillin was higher among patients who did not travel than among patients who did and in children ≤ 15 years. In addition, resistance of *Campylobacter* isolates in humans did not follow the same patterns over time as resistance in broiler and porcine isolates. These results suggest that resistance to ampicillin is more frequently domestically acquired and may be related to the use of ampicillin in human therapy because ampicillin is widely prescribed for infections in children.

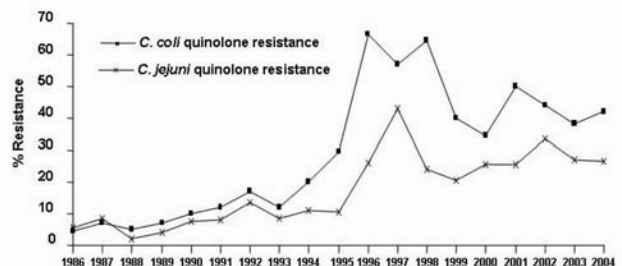


Figure 4. Quinolone resistance of human *Campylobacter jejuni* and *C. coli*, France, 1986-2004.

Table. Resistance of *Campylobacter jejuni* and *C. coli* in humans, broilers, and pigs to ampicillin, erythromycin, tetracycline/doxycycline, and ciprofloxacin/enrofloxacin, France, 1999–2004

| Antimicrobial agent/year | <i>C. jejuni</i> | | <i>C. coli</i> | | Pigs, no. (%) |
|-----------------------------------|------------------|-------------------|-----------------|-------------------|---------------|
| | Humans, no. (%) | Broilers, no. (%) | Humans, no. (%) | Broilers, no. (%) | |
| Ampicillin | | | | | |
| 1999 | 123 (42.3) | 297 (22.6) | 20 (70.0) | 96 (29.2) | * |
| 2000 | 189 (40.2) | 67 (31.3) | 35 (37.1) | 35 (31.4) | 317 (12.3) |
| 2001 | 137 (47.4) | 61 (13.1) | 26 (53.8) | 44 (25.0) | 291 (11.0) |
| 2002 | 184 (45.6) | 41 (29.3) | 43 (48.8) | 64 (28.1) | 101 (14.9) |
| 2003 | 479 (46.1) | 46 (34.8) | 68 (28.1) | 60 (30.0) | 101 (17.8) |
| 2004 | 438 (38.1) | 32 (28.1) | 88 (14.9) | 74 (24.3) | 67 (7.5) |
| Erythromycin | | | | | |
| 1999 | 123 (0.8) | 297 (0.3) | 20 (10.0) | 96 (31.3) | * |
| 2000 | 187 (3.2) | 67 (3.0) | 35 (8.6) | 36 (11.1) | 317 (65.3) |
| 2001 | 136 (3.7) | 61 (4.9) | 26 (7.7) | 44 (4.5) | 289 (49.1) |
| 2002 | 184 (2.7) | 40 (5.0) | 44 (9.0) | 64 (17.2) | 101 (58.4) |
| 2003 | 478 (0.6) | 46 (4.3) | 68 (7.3) | 61 (31.1) | 97 (78.4) |
| 2004 | 437 (1.4) | 32 (0.0) | 88 (12.5) | 74 (17.6) | 67 (43.3) |
| Tetracycline/doxycycline | | | | | |
| 1999 | 118 (23.7) | 297 (56.6) | 6 (25.0) | 96 (69.8) | * |
| 2000 | 188 (12.8) | 67 (55.2) | 35 (14.3) | 35 (60.0) | 317 (82.6) |
| 2001 | 137 (9.5) | 61 (65.6) | 26 (26.9) | 45 (80.0) | 289 (88.9) |
| 2002 | 184 (22.8) | 41 (67.5) | 43 (41.8) | 64 (84.4) | 101 (86.1) |
| 2003 | 479 (26.7) | 46 (60.9) | 68 (63.2) | 61 (96.7) | 97 (95.9) |
| 2004 | 438 (28.8) | 32 (40.6) | 88 (53.4) | 74 (71.6) | 67 (61.2) |
| Ciprofloxacin/enrofloxacin | | | | | |
| 1999 | † | 297 (16.8) | † | 96 (39.6) | * |
| 2000 | 185 (23.8) | 68 (23.5) | 35 (31.4) | 35 (28.6) | 316 (12.3) |
| 2001 | 137 (21.9) | 61 (29.5) | 26 (34.6) | 45 (37.8) | 292 (12.3) |
| 2002 | 184 (31.7) | 41 (31.7) | 44 (43.2) | 63 (41.3) | 101 (21.8) |
| 2003 | 479 (25.9) | 45 (13.3) | 68 (38.2) | 61 (41.0) | 99 (24.2) |
| 2004 | 438 (25.3) | 32 (9.4) | 88 (42.0) | 74 (32.4) | 67 (26.9) |

*Strains isolated from poultry in 1999 and 2000 were tested with enrofloxacin; strains isolated between 2001 and 2004 were tested with ciprofloxacin; all pig strains were tested with ciprofloxacin.

†Human strains have been tested for ciprofloxacin resistance since 2000.

Nalidixic acid resistance increased 5-fold from 1990 (5.3%) to 2004 (26.3%), consistent with trends observed in other countries (16,17). The use of fluoroquinolones in animal feed was approved in Europe in 1990. Studies have shown the development of ciprofloxacin-resistant *Campylobacter* in treated chickens and the spread of ciprofloxacin-resistant *Campylobacter* from animal food sources to humans (17,34,35). Australia, where fluoroquinolones have never been licensed for use in food-producing animals, did not experience fluoroquinolone resistance in human *Campylobacter* isolates (36).

The high proportions of resistance to nalidixic acid and ciprofloxacin in broilers and pigs are consistent with the findings of Desmots et al. in France (31). In this study, quinolone and fluoroquinolone resistance increased between 1992–1996 and 2001–2002 among isolates from standard chicken flocks, while resistance remained low for free-range flock isolates. In France, antimicrobial growth promoters have never been authorized in the production of free-range chickens, contrary to standard methods of production of chicken flocks, and antimicrobial therapy is limited (31).

From 2002 to 2004, ciprofloxacin resistance dropped substantially in *C. jejuni* isolated from broilers; nalidixic acid resistance decreased as well, although not significantly. The decrease in broilers may be related to the restriction in the use of fluoroquinolones in animal feed after the 1999 EU recommendation. Similarly, in Denmark, resistance to macrolides of *C. coli* declined after the prophylactic and growth-promoting use of macrolides was banned (37). However, the decrease in ciprofloxacin resistance occurred 2–3 years after the EU recommendation, which suggests that EU recommendations were not followed immediately by application or, alternatively, that the effect of the restriction in the use of fluoroquinolones in animal feed is not immediate. Unfortunately, no resistance data in broilers and pigs were available before 1999, which is a limitation to interpret recent trends in relation to the EU recommendations. According to the French food security agency (AFSSA), global sales of antimicrobial agents decreased consistently from 2001 through 2002, but information on species-specific consumption was not available (38). Specific data from veterinary prescriptions and livestock consumption

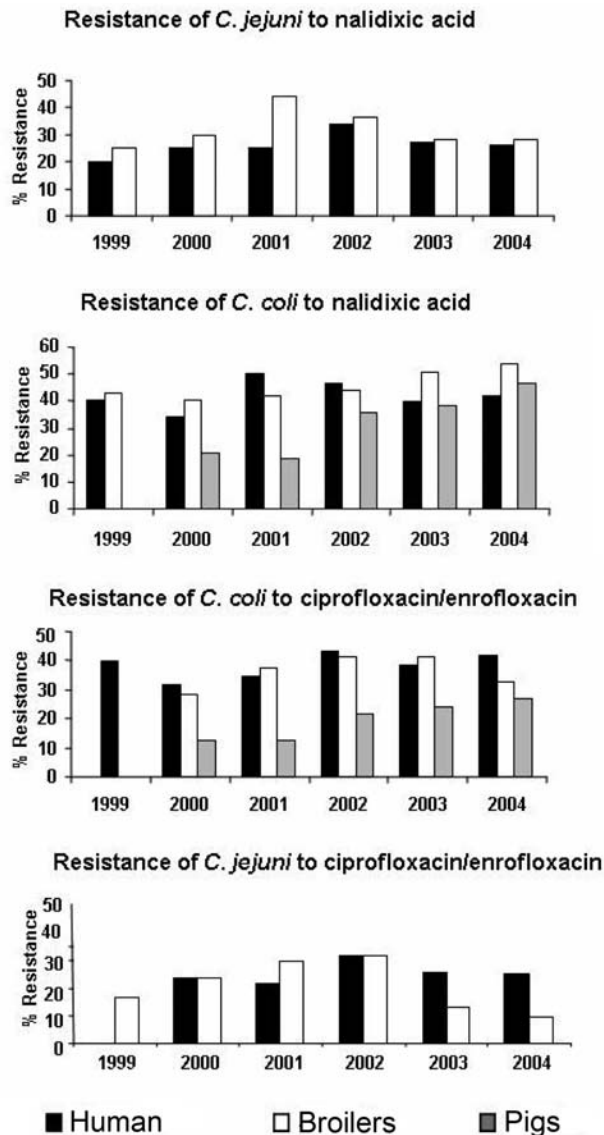


Figure 5. Resistance of *Campylobacter jejuni* and *C. coli* to nalidixic acid and ciprofloxacin in humans, broilers, and pigs, France, 1999–2004. Human strains were tested with ciprofloxacin since 2000. Strains isolated from poultry and pigs in 1999 and 2000 were tested with enrofloxacin, strains isolated between 2001 and 2004 were tested with ciprofloxacin.

are necessary to quantify the amount of antimicrobial agents consumed by animals.

C. jejuni nalidixic acid and ciprofloxacin resistance decreased concomitantly in humans and broilers from 2002 to 2004. Because the decrease was less pronounced in humans than in broilers, a longer period is needed to detect an effect of the restriction in the use of antimicrobial agents in animal feeds or resistance may be also related to other exposure. Fluoroquinolones are the first drugs of choice for the empiric treatment of human diarrhea or

prophylactic treatment associated with travel in France and may be responsible for a part of resistance in humans (39). However, >80% of patients infected with a ciprofloxacin-resistant strain did not travel to a foreign country before onset of illness, which indicates that a substantial proportion of fluoroquinolone resistance was domestically acquired (40). The resistance rate to ciprofloxacin was not higher in adults compared with children, as could be expected if treatment of cases was contributing to resistance (39), because fluoroquinolone treatment is not used in children ≤ 15 years of age.

In contrast to *C. jejuni*, we observed no decrease in quinolone and fluoroquinolone resistance in *C. coli* in pigs, broilers, or human isolates. The use of these antimicrobial agents in pigs may not have changed, and a part of human *C. coli* infection may be related to other sources. Alternatively, unknown mechanisms could be implicated in *C. coli* resistance, such as a high number of point mutations.

Our study has several limitations. Laboratories participate voluntarily in the surveillance network, which may result in a selection of strains that is not representative of the general population. However, all districts of mainland France were included. In addition, this limitation, if it exists, may not affect antimicrobial drug resistance because participating laboratories routinely screen stools for *Campylobacter* and send all their isolates to CNRCH. Since the laboratory network was extended to private and additional hospitals laboratories in 2002, this may have had an effect on the trends in resistance. However, trends of human antimicrobial susceptibility were analyzed exclusively among strains from hospital laboratories from 1986 to 2004. In addition, characteristics of *Campylobacter* isolates sent to CNRCH did not change for age, sex, seasonality, and species after the network extension in 2002 (20,21). Comparison of human and animal data was not based on a continuum between human isolates and contaminated food consumption (isolates from retail chicken). However, broiler chicken and pig data were representative of French livestock and were consistent with those of another recent survey done in France (31).

The extension of the surveillance of human *Campylobacter* allowed the epidemiologic characteristics of *Campylobacter* infections that occurred in the general French population to be better understood. *Campylobacter* resistance to antimicrobial agents increased to a high level among humans in France from 1990 through 2004. Comparison of antimicrobial resistance patterns in humans, broilers, and pigs from 1999 to 2004 showed similar patterns of quinolone and fluoroquinolone resistance for *C. jejuni* isolates from broilers and humans. These results suggest that a limitation of the use of fluoro-

quinolones in broilers may reduce fluoroquinolone resistance of *Campylobacter* in humans. Other studies, however, are needed to further quantify the effect of restricted use of antimicrobial drugs in animals on bacterial resistance in human isolates. Ongoing national surveillance of *Campylobacter* in humans, livestock, and animal feeds at the retail level and antimicrobial susceptibility testing are necessary to evaluate the effects of implementing European policies. Further research is also needed to better understand the relationship between antimicrobial use in animals and humans and bacterial resistance in humans.

Acknowledgments

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References

- Allos BM. *Campylobacter jejuni* Infections: update on emerging issues and trends. *Clin Infect Dis*. 2001;32:1201–6.
- Nachamkin I, Blaser MJ. *Campylobacter*. 2nd ed. Washington: ASM Press; 2000.
- Wheeler JG, Sethi D, Cowden JM, Wall PG, Rodrigues LC, Tompkins DS, et al. Study of infectious intestinal disease in England: rates in the community, presenting to general practice, and reported to national surveillance. The Infectious Intestinal Disease Study Executive. *BMJ*. 1999;318:1046–50.
- de Wit MA, Koopmans MP, Kortbeek LM, van Leeuwen NJ, Vinje J, van Duynhoven YT. Etiology of gastroenteritis in sentinel general practices in the Netherlands. *Clin Infect Dis*. 2001;33:280–8.
- Hall G, Kirk MD, Becker N, Gregory JE, Unicomb L, Millard G, et al. Estimating foodborne gastroenteritis, Australia. *Emerg Infect Dis*. 2005;11:1257–64.
- Samuel MC, Vugia DJ, Shallow S, Marcus R, Segler S, McGivern T, et al. Epidemiology of sporadic *Campylobacter* infection in the United States and declining trend in incidence, FoodNet 1996–1999. *Clin Infect Dis*. 2004;38(Suppl 3):S165–74.
- Vaillant V, de Valk H, Baron E, Ancelle T, Colin P, Delmas MC, et al. Foodborne infections in France. *Foodborne Pathog Dis*. 2005;2:221–32.
- Stern NJ, Hiatt KL, Alfredsson GA, Kristinsson KG, Reiersen J, Hardardottir H, et al. *Campylobacter* spp. in Icelandic poultry operations and human disease. *Epidemiol Infect*. 2003;130:23–32.
- Friedman CR, Hoekstra RM, Samuel M, Marcus R, Bender J, Shiferaw B, et al. Risk factors for sporadic *Campylobacter* infection in the United States: a case-control study in FoodNet sites. *Clin Infect Dis*. 2004;38(Suppl 3):S285–96.
- Neimann J, Engberg J, Molbak K, Wegener HC. A case-control study of risk factors for sporadic *Campylobacter* infections in Denmark. *Epidemiol Infect*. 2003;130:353–66.
- Kuschner RA, Trofa AF, Thomas RJ, Hoge CW, Pitarangsi C, Amato S, et al. Use of azithromycin for the treatment of *Campylobacter enteritis* in travelers to Thailand, an area where ciprofloxacin resistance is prevalent. *Clin Infect Dis*. 1995;21:536–41.
- Endtz HP, Ruijs GJ, van KB, Jansen WH, van der RT, Mouton RP. Quinolone resistance in *Campylobacter* isolated from man and poultry following the introduction of fluoroquinolones in veterinary medicine. *J Antimicrob Chemother*. 1991;27:199–208.
- Lucey B, Cryan B, O'Halloran F, Wall PG, Buckley T, Fanning S. Trends in antimicrobial susceptibility among isolates of *Campylobacter* species in Ireland and the emergence of resistance to ciprofloxacin. *Vet Rec*. 2002;151:317–20.
- Nachamkin I, Ung H, Li M. Increasing fluoroquinolone resistance in *Campylobacter jejuni*, Pennsylvania, USA, 1982–2001. *Emerg Infect Dis*. 2002;8:1501–3.
- Engberg J, Aarestrup FM, Taylor DE, Gerner-Smidt P, Nachamkin I. Quinolone and macrolide resistance in *Campylobacter jejuni* and *C. coli*: resistance mechanisms and trends in human isolates. *Emerg Infect Dis*. 2001;7:24–34.
- Luber P, Wagner J, Hahn H, Bartelt E. Antimicrobial resistance in *Campylobacter jejuni* and *Campylobacter coli* strains isolated in 1991 and 2001–2002 from poultry and humans in Berlin, Germany. *Antimicrob Agents Chemother*. 2003;47:3825–30.
- Gupta A, Nelson JM, Barrett TJ, Tauxe RV, Rossiter SP, Friedman CR, et al. Antimicrobial resistance among *Campylobacter* strains, United States, 1997–2001. *Emerg Infect Dis*. 2004;10:1102–9.
- Davidson DJ. In the matter of enrofloxacin for poultry: withdrawal of approval of Bayer Corporation's new animal drug application 1 (NADA) 140–828 (Baytril). *FDA Docket no. 00N–1571*. 2004.
- Gally A, Simon F, Megraud F. Surveillance of human *Campylobacter* infections in France—part 2—implementation of national surveillance. *Euro Surveill*. 2003;8:218.
- Mégraud F. Les Infections à *Campylobacter* en France (1986–2000). *Surveillance National des Maladies Infectieuses*. 1998–2000;2003:133–5.
- Mégraud F, Prouzet-Mauleon V. Evolution de la résistance des *Campylobacters* aux antibiotiques en France (1986–2002). *Bull Epidémiol Heb*. 2004;32–33:156–8.
- Menard A, Dachet F, Prouzet-Mauleon V, Oleastro M, Megraud F. Development of a real-time fluorescence resonance energy transfer PCR to identify the main pathogenic *Campylobacter* spp. *Clin Microbiol Infect*. 2005;11:281–7.
- National Center for Biotechnology Information. Basic Local Alignment Search Tool (BLAST) program. National Center for Biotechnology Information 2006 May 7 [cited 2006 Oct 17]. Available from <http://www.ncbi.nlm.nih.gov/blast/>
- Société Française de Microbiologie. Recommandations du Comité de l'Antibiogramme de la Société Française de Microbiologie. Société Française de Microbiologie 2006 Feb 24 [cited 2006 Oct 17]. Available from <http://www.sfm.asso.fr/nouv/general.php?pa=2>
- Avrain L, Humbert F, L'Hospitalier R, Sanders P, Vernozy-Rozand C, Kempf I. Antimicrobial resistance in *Campylobacter* from broilers: association with production type and antimicrobial use. *Vet Microbiol*. 2003;96:267–76.
- Denis M, Soumet C, Rivoal K, Ermel G, Blivet D, Salvat G, et al. Development of a m-PCR assay for simultaneous identification of *Campylobacter jejuni* and *C. coli*. *Lett Appl Microbiol*. 1999;29:406–10.
- Nylen G, Dunstan F, Palmer SR, Andersson Y, Bager F, Cowden J, et al. The seasonal distribution of *Campylobacter* infection in nine European countries and New Zealand. *Epidemiol Infect*. 2002;128:383–90.

28. Vandenberg O, Dediste A, Houf K, Ibeqwem S, Souayah H, Cadranel S, et al. *Arcobacter* species in humans. *Emerg Infect Dis.* 2004;10:1863–7.
29. Takkinen J, Ammon A, Robstad O, Breuer T. European survey on *Campylobacter* surveillance and diagnosis 2001. *Euro Surveill.* 2003;8:207–13.
30. Gallay A, Simon F, Megraud F. Surveillance of human *Campylobacter* infections in France—part 1—which data? A study of microbiological laboratories, 2000. *Euro Surveill.* 2003;8:213–7.
31. Desmots MH, Dufour-Gesbert F, Avrain L, Kempf I. Antimicrobial resistance in *Campylobacter* strains isolated from French broilers before and after antimicrobial growth promoter bans. *J Antimicrob Chemother.* 2004;54:1025–30.
32. Udayamputhoor RS, Hariharan H, Van Lunen TA, Lewis PJ, Heaney S, Price L, et al. Effects of diet formulations containing proteins from different sources on intestinal colonization by *Campylobacter jejuni* in broiler chickens. *Can J Vet Res.* 2003;67:204–12.
33. Hopkins KL, Desai M, Frost JA, Stanley J, Logan JM. Fluorescent amplified fragment length polymorphism genotyping of *Campylobacter jejuni* and *Campylobacter coli* strains and its relationship with host specificity, serotyping, and phage typing. *J Clin Microbiol.* 2004;42:229–35.
34. Hooper DC. Emerging mechanisms of fluoroquinolone resistance. *Emerg Infect Dis.* 2001;7:337–41.
35. Threlfall EJ, Ward LR, Frost JA, Willshaw GA. Spread of resistance from food animals to man—the UK experience. *Acta Vet Scand Suppl.* 2000;93:63–8.
36. Unicomb L, Ferguson J, Riley TV, Collignon P. Fluoroquinolone resistance in *Campylobacter* absent from isolates, Australia. *Emerg Infect Dis.* 2003;9:1482–3.
37. Iovine NM, Blaser MJ. Antimicrobial resistance in *Campylobacter*. *Emerg Infect Dis.* 2004;10:1346.
38. Moulin G, Roux S. Suivi des ventes de médicaments vétérinaires contenant des antibiotiques en France en 2002. Agence Française de Sécurité Sanitaire des Aliments. Report 30 Nov 2004. Available from <http://www.afssa.fr/Ftp/Afssa/28169-28170.pdf>
39. Smith KE, Besser JM, Hedberg CW, Leano FT, Bender JB, Wicklund JH, et al. Quinolone-resistant *Campylobacter jejuni* infections in Minnesota, 1992–1998. Investigation Team. *N Engl J Med.* 1999;340:1525–32.
40. Kassenborg HD, Smith KE, Vugia DJ, Rabatsky-Ehr T, Bates MR, Carter MA, et al. Fluoroquinolone-resistant *Campylobacter* infections: eating poultry outside of the home and foreign travel are risk factors. *Clin Infect Dis.* 2004;38(Suppl 3):S279–84.

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