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# Polyamine analysis for chemotaxonomy of thermophilic eubacteria: Polyamine distribution profiles within the orders *Aquificales*, *Thermotogales*, *Thermodesulfobacteriales*, *Thermales*, *Thermoanaerobacteriales*, *Clostridiales* and *Bacillales*

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Cellular polyamines of 45 thermophilic and 8 related mesophilic eubacteria were investigated by HPLC and GC analyses for the thermophilic and chemotaxonomic significance of polyamine distribution profiles. Spermidine and a quaternary branched penta-amine, *N*<sup>4</sup>-bis(aminopropyl)norspermidine, were the major polyamine in *Thermocrinis*, *Hydrogenobacter*, *Hydrogenobaculum*, *Aquifex*, *Persephonella*, *Sulfurihydrogenibium*, *Hydrogenothermus*, *Balnarium* and *Thermovibrio*, located in the order *Aquificales*. *Thermodesulfobacterium* and *Thermodesulfatator* belonging to the order *Thermodesulfobacteriales* contained another quaternary penta-amine, *N*<sup>4</sup>-bis(aminopropyl)spermidine. In the order *Thermotogales*, *Thermotoga* contained spermidine, norspermidine, caldopentamine and homocaldopentamine. The latter two linear penta-amines were not found in *Marinitoga* and *Petrotoga*. In the order *Thermales*, *Thermus* and *Marinithermus* contained homospermidine, norspermine and the linear penta-amines. *Meiothermus* lacked penta-amines. *Vulcanithermus* contained linear penta-amines and hexa-amines but not homospermidine. *Oceanithermus* contained spermine alone. Within the order *Thermoanaerobacteriales*, the two quaternary branched penta-amines were found in *Thermanaeromonas* and *Thermoanaerobacter*. *Caldanaerobacter* contained *N*<sup>4</sup>-bis(aminopropyl)spermidine. *Thermoanaerobacterium* lacked penta-amines. *Thermaerobacter* of the order *Clostridiales* contained *N*<sup>4</sup>-bis(aminopropyl)spermidine and agmatine. *Thermosyntropha*, *Thermanaerovibrio*, *Thermobrachium* (the order *Clostridiales*), *Sulfobacillus*, *Alicyclobacillus*, *Anoxybacillus*, *Ureibacillus*, *Thermicanus* (the order *Bacillales*), *Desulfotomaculum*, *Desulfitobacterium* and *Pelotomaculum* (the family *Peptococcaceae*) ubiquitously contained spermine. Some thermophiles of *Bacillales* added linear and branched penta-amines.

**Key Words**—*Alicyclobacillaceae*; *Aquificales*; *polyamine*; *Thermales*; *Thermoanaerobacteriales*; *Thermodesulfobacteriales*; thermophile; *Thermotogales*

## Introduction

Aliphatic cellular polyamines, linear diamines, triamines, tetra-amines, penta-amines and hexa-amines, and quaternary branched penta-amines, were found in eubacteria (domain Bacteria) and archaebacteria (do-

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main Archaea), and are important to stabilize cellular nucleic acid structure as a major function (Hamana, 2002; Hamana and Matsuzaki, 1992). The chemotaxonomic and phylogenetic significance of the distribution of long and branched polyamines, possibly associated with their thermophily, has been proposed (Hamana and Itoh, 2001; Hamana and Kato, 2000; Hamana et al., 1994, 1995, 1996a, b, 1998, 1999, 2001, 2003).

The three deeply branching lineages of eubacteria, the orders *Aquificales*, *Thermotogales* and *Thermodesulfobacteriales*, are phylogenetically located in early eubacterial evolution and comprise entirely thermophiles. We have attempted to analyze the polyamines of hydrogen-oxidizing *Hydrogenothermus*, *Hydrogenobacter* (Stohr et al., 2001; Takai et al., 2001a), *Persephonella*, *Sulfurihydrogenibium*, *Thermocrinis* and *Balnearium*, and nitrate-reducing *Thermovibrio* (Eder and Huber, 2002; Huber et al., 1998, 2002; Nakagawa et al., 2003; Takai et al., 2003a, b), belonging to *Aquificales*. Polyamines of new members of *Thermotoga*, *Petrotoga* and *Marinitoga* (Alain et al., 2002; Balk et al., 2002; L'Haridon et al., 2002; Takahata et al., 2001) of *Thermotogales*, and thermophilic sulfate-reducing, *Thermodesulfatator* belonging to the order *Thermodesulfobacteriales* (Moussard et al., 2004), were analyzed.

The thermophilic order *Thermales* currently comprises *Thermus*, *Meiothermus*, *Marinithermus* and *Oceanithermus* (Miroshnichenko et al., 2003a; Sako et al., 2003). A novel member, *Vulcanithermus*, was isolated as the fifth genus (Miroshnichenko et al., 2003b). Thus, polyamine profiles of new species of the five genera were determined, in relation to their growth temperature and their phylogenetic distance.

Thermophiles are distributed also in the Gram-positives of the orders *Thermoanaerobacteriales*, *Clostridiales* and *Bacillales*. Polyamines of new species of *Thermaerobacter* (Nunoura et al., 2002; Spanevello et al., 2002), *Caldanaerobacter* (Fardeau et al., 2000, 2004; Xue et al., 2001), *Thermanaeromonas* (Mori et al., 2002) and *Thermoanaerobacterium* (Cann et al., 2001) were determined. The thermophilic anaerobes, *Gelria* (Plugge et al., 2002), *Thermanaerovibrio* (Baena et al., 1999b), *Thermosyntropha* (Svetlitsnyi et al., 1996), *Thermicanus* (Gossner et al., 1999) and *Thermobrachium* (Engle et al., 1996), and the mesophiles, *Aminobacterium* and *Aminomonas* (Baena et al., 1999a, 2000), and several anaerobes located in the *Desulfotomaculum* lineage I (Brauman

et al., 1998; Fardeau et al., 1995; Imachi et al., 2002; Qiu et al., 2003; Robertson et al., 2001), and the alkaliphilic clostridia, *Alkaliphilus* (Cao et al., 2003; Takai et al., 2001b) were available in the present study. Polyamines of thermophilic *Anoxybacillus* and *Ureibacillus* (Belduz et al., 2003; Fortina et al., 2001; Yumoto et al., 2004), as well as thermoacidophilic *Sulfobacillus* and *Alicyclobacillus* (Dufresne et al., 1996; Goto et al., 2002a, b, 2003; Matsubara et al., 2002; Tsuruoka et al., 2003), were also determined.

Distribution catalogues of long and branched polyamines in the 53 newly validated species were presented here to elucidate their chemotaxonomic significance. Their distribution profiles, at different culture temperatures under acidic, neutral and alkaline conditions, were also described.

## Materials and Methods

The culture media designated by the culture collections were used according to the Catalogue of Strains (2004) of ATCC, JCM, NBRC and NCIMB. The medium pH and culture temperature used for optimum growth are given in Table 1. *Thermocrinis*, *Persephonella*, *Sulfurihydrogenibium*, *Balnearium* and *Thermovibrio* were grown under  $N_2$ - $O_2$ - $H_2$  or  $H_2$ - $CO_2$ . *Hydrogenothermus* was aerobically grown. *Marinitoga*, *Petrotoga* and *Thermotoga* were cultivated under  $N_2$  or  $N_2$ - $CO_2$ . *Thermodesulfatator* was grown under  $H_2$ - $CO_2$ . *Thermus*, *Marinithermus* and *Meiothermus* were aerobically grown whereas *Vulcanithermus* and *Oceanithermus* were grown under a  $N_2$  atmosphere. *Thermanaeromonas*, *Thermoanaerobacterium*, *Caldanaerobacter*, *Gelria*, *Thermosyntropha*, *Thermanaerovibrio*, *Thermobrachium*, *Aminobacterium* and *Aminomonas* were grown anaerobically. *Thermaerobacter*, *Alkaliphilus*, *Desulfotomaculum*, *Desulfosporosinus*, *Pelotomaculum* and *Sporotomaculum* were grown under  $N_2$ - $CO_2$ . *Sulfobacillus*, *Alicyclobacillus*, *Anoxybacillus* and *Ureibacillus* were grown aerobically and *Thermicanus* anaerobically.

The organisms at stationary phase were harvested and homogenized in equal volumes of cold 1 M  $HClO_4$ . The whole  $HClO_4$  extract was analyzed by high-performance liquid chromatography (HPLC) on a column of cation-exchange resin in a Hitachi L6000 high-speed liquid chromatograph (Hamana et al., 1998). Polyamines were detected with the *o*-phthalaldehyde reagent. The whole extract was subjected to a Dowex

Table 1. Cellular concentration of polyamines of thermophilic eubacteria.

		Polyamines ( $\mu\text{mol/g}$ wet wt. cell)												
		°C	pH	3	4	5	33	34	333	333/334/333	334/343/334	333/334/333	343/334/333	
<b>Order Aquificales</b>														
<b>Family Aquificaceae</b>														
	<i>Thermocrinis albus</i>	80	7.0	—	0.02	—	0.10	0.56	*	0.02	*	*	0.02	0.30
	<i>Hydrogenobacter hydrogenophilus</i>	(c)	7.0	—	0.01	—	0.60	0.50	—	*	0.06	—	0.03	2.10
	<i>Hydrogenobacter subterraneus</i>	75	7.0	0.01	—	—	0.06	1.20	—	*	0.03	0.02	—	2.40
	<i>Hydrogenobacter thermophilus</i>	(c)	7.0	—	—	—	—	1.50	—	*	0.03	0.05	0.02	2.60
	<i>Hydrogenobaculum acidophilum</i>	(c)	65	3.0	—	—	—	0.08	—	*	—	—	—	1.12
	<i>Aquifex pyrophilus</i>	(b)	85	6.5	—	—	0.02	—	—	—	—	0.02	—	1.60
<b>Family Hydrogenothermaceae</b>														
	<i>Persephonella hydrogeniphila</i>	70	7.0	—	0.02	0.10	—	0.03	*	0.02	—	*	—	1.60
	<i>Sulfurhydrogenibium subterraneum</i>	62	7.5	—	0.40	0.20	—	1.50	—	—	*	—	*	0.01
	<i>Hydrogenothermus marinus</i>	65	7.0	0.01	—	—	0.18	0.40	—	*	—	0.08	—	3.10
<b>Family Desulfurobacteriaceae</b>														
	<i>Balnearium lithotrophicum</i>	75	5.5	—	0.04	—	—	0.30	—	—	—	—	—	0.60
	<i>Thermovibrio ammonificans</i>	75	5.5	—	—	—	—	0.08	—	—	—	—	—	0.80
	<i>Thermovibrio ruber</i>	75	6.0	—	0.02	—	—	0.10	—	—	*	—	*	0.10
<b>Order Thermotogales</b>														
<b>Family Thermotogaceae</b>														
	<i>Marinitoga piezophila</i>	65	6.0	—	0.15	0.02	0.35	0.57	—	0.03	0.05	—	—	—
	<i>Petrotoga miotherma</i>	(d)	55	7.2	—	0.57	0.16	0.55	—	0.02	0.17	—	—	—
	<i>Petrotoga olearia</i>	55	7.0	—	0.90	0.15	—	1.20	—	—	0.30	—	—	—
	<i>Petrotoga sibirica</i>	55	7.0	—	0.25	—	—	0.80	—	—	0.25	—	—	—
	<i>Thermotoga elfii</i>	(a)	65	8.0	—	0.08	—	0.36	—	—	0.66	0.04	—	—
	<i>Thermotoga lettingae</i>	65	6.9	—	0.10	0.05	1.15	0.40	0.04	—	0.90	—	—	—
	<i>Thermotoga maritima</i>	JCM 10099 <sup>T</sup>	60	6.5	—	0.02	0.60	0.79	—	0.02	0.10	0.03	—	—
		(a)	70	6.5	—	0.04	0.60	0.80	—	0.10	0.20	0.05	—	0.02
			80	6.5	0.02	0.02	0.20	0.15	—	0.40	0.20	1.25	—	0.59

Table 1. (Continued.)

		Polyamines ( $\mu\text{mol/g wet wt. cell}$ )																					
		343/																					
		°C	pH	3	4	5	33	34	333	343	334	3333	3343	3334	33333	33334	333333						
<i>Thermotoga naphthophila</i>	JCM 10882 <sup>T</sup>	80	6.5	—	0.25	0.05	0.80	1.20	—	1.30	0.02	0.01	0.01	0.01	—	—	0.01						
<i>Thermotoga neapolitana</i>	ATCC 49049 <sup>T</sup>	(a)	80	6.5	0.20	0.15	0.02	1.35	1.15	—	0.15	0.14	0.16	0.05	0.02	0.02	0.02						
<i>Thermotoga petrophila</i>	JCM 10881 <sup>T</sup>	80	6.5	—	0.19	0.10	0.65	1.40	—	1.52	0.02	0.04	0.02	—	—	—	0.02						
<i>Thermosipho japonicus</i>	JCM 10495 <sup>T</sup>	(f)	70	7.0	—	0.27	—	1.05	1.45	—	0.02	0.40	—	—	—	—	—						
<i>Fervidobacterium islandicum</i>	ATCC 4964 <sup>T</sup>	(d)	70	7.0	0.02	0.14	0.02	0.85	1.42	—	0.12	0.22	0.10	0.12	—	—	—						
<i>Fervidobacterium nodosum</i>	ATCC 35602 <sup>T</sup>	(d)	70	7.0	0.04	0.10	0.05	0.60	1.40	—	0.15	0.45	—	—	—	—	—						
		°C	pH	3	4	33	34	333	343	334	3333	3343	3334	3(3)4	3(3)3	3(3)4	3(3)3						
Order Thermodesulfobacteriales																							
Family Thermodesulfobacteriaceae																							
<i>Thermodesulfobacterium commune</i>	ATCC 33708 <sup>T</sup>	(b)	70	7.5	—	0.02	0.02	0.18	0.05	0.08	—	0.04	—	—	0.03	1.00	—						
<i>Thermodesulfobacterium thermophilum</i>	NCIMB 12254 <sup>T</sup>	(b)	65	7.4	—	0.02	0.02	0.77	0.05	1.05	0.20	0.50	—	0.02	0.08	1.50	—						
<i>Thermodesulfatator indicus</i>	JCM 11887 <sup>T</sup>	70	6.7	—	0.10	—	0.90	—	0.01	—	—	—	—	—	—	—	0.80						
		°C	pH	3	4	33	3(3)3	333	343	334	334	3333	3343	3334	33333	33334	333333	3(3)3					
Order Thermales																							
Family Thermaceae																							
<i>Thermus antranikianii</i>	ATCC 700961 <sup>T</sup>	70	7.6	—	0.10	0.01	0.25	1.00	0.32	—	*	0.10	0.30	*	0.05	0.02	0.01	0.02	*	—	—		
<i>Thermus aquaticus</i>	ATCC 25104 <sup>T</sup>	(a)	70	7.6	—	0.26	0.14	0.68	1.38	1.06	—	*	0.21	0.80	*	0.05	0.16	0.10	0.10	*	—	—	
<i>Thermus brockianus</i>	NCIMB 12676 <sup>T</sup>	(b)	70	7.6	—	0.10	0.10	0.75	1.30	0.80	—	*	0.20	2.50	*	0.20	0.02	0.02	0.01	*	—	—	
<i>Thermus filiformis</i>	ATCC 43280 <sup>T</sup>	(a)	70	7.6	0.04	0.20	0.09	0.88	1.04	0.16	0.02	*	0.10	0.41	*	0.09	0.21	0.05	0.05	*	0.05	—	0.05
<i>Thermus igniterrae</i>	ATCC 700962 <sup>T</sup>	70	7.6	—	0.15	0.01	0.10	1.00	0.38	—	*	0.05	0.10	*	0.05	0.02	0.02	0.02	*	—	—	—	
<i>Thermus oshimai</i>	NCIMB 13400 <sup>T</sup>	(b)	70	8.2	0.01	0.02	0.05	0.18	0.42	0.60	—	*	0.20	1.40	*	0.06	0.06	0.04	0.05	*	—	—	—

Table 1. (Continued.)

		Polyamines ( $\mu\text{mol/g}$ wet wt. cell)																				
		3(3)3/					343/					3343/										
°C	pH	3	4	33	34	44	333	3(3)3	333	343	334	334	344	3333	3343	3334	33334	33333	333343	3(3)3		
<i>Thermus</i> <i>scotoductus</i>	ATCC 51532 <sup>T</sup>	(a)	65	7.2	—	0.15	—	0.24	1.70	0.30	—	*	0.22	1.50	*	0.45	0.10	0.02	0.03	*	—	—
<i>Thermus</i> <i>thermophilus</i>	ATCC 27634 <sup>T</sup>	(a)	70	7.2	—	0.51	0.14	0.14	0.95	0.53	0.04	*	0.19	1.00	*	0.05	0.11	0.06	0.04	*	0.04	0.04
<i>Thermus</i> <i>thermophilus</i>	JCM 10941 <sup>T</sup>		50	7.2	—	0.05	0.10	0.20	1.25	*	*	0.15	*	*	0.15	—	—	—	*	*	—	—
			60	7.2	—	0.10	0.10	0.30	2.10	*	*	0.40	*	*	0.30	0.05	—	—	*	*	—	—
			70	7.2	—	—	0.10	0.25	1.40	*	*	0.80	*	*	0.09	0.05	0.05	*	*	0.02	—	0.02
		3(3)4/																				
		343/					3334/					3334/										
°C	pH	3	4	5	33	34	44	333	343	334	334	344	3333	3334	3334	3333	3334	33333	33343	33333	33334	
<i>Meiothermus</i> <i>chiarophilus</i>	NCIMB 13439 <sup>T</sup>	(b)	55	8.2	0.02	0.10	—	—	1.54	0.80	0.02	0.15	0.02	*	0.02	—	—	—	*	—	—	—
<i>Meiothermus</i> <i>ruber</i>	ATCC 34958 <sup>T</sup>	(a)	60	7.2	0.01	0.04	—	0.01	0.12	1.81	—	0.02	0.01	*	0.05	—	—	—	*	—	—	—
<i>Meiothermus</i> <i>silvanus</i>	NCIMB 13440 <sup>T</sup>	(b)	55	8.2	0.01	0.15	—	—	0.72	1.14	—	0.07	0.01	*	0.01	0.01	—	—	*	—	—	—
<i>Meiothermus</i> <i>taiwanensis</i>	ATCC BAA-399 <sup>T</sup>		60	7.6	—	0.12	—	0.02	0.85	0.95	—	0.23	0.02	*	0.02	—	—	—	*	—	—	—
<i>Marinithermus</i> <i>hydrothermalis</i>	JCM 11576 <sup>T</sup>		70	7.0	—	0.10	0.15	0.10	0.75	0.90	0.12	*	*	0.15	—	—	—	—	*	0.10	*	—
<i>Oceanithermus</i> <i>profundus</i>	NBRC 100410 <sup>T</sup>		60	7.3	—	—	0.05	—	0.05	—	—	1.00	—	*	—	—	—	—	*	—	—	—
			68	7.3	—	—	0.24	—	0.05	—	—	1.50	—	*	—	—	—	—	*	—	—	—
<i>Vulcanithermus</i> <i>mediatlanticus</i>	JCM 11956 <sup>T</sup>		70	6.5	—	0.02	—	0.02	0.50	—	0.03	0.10	0.40	*	—	—	0.10	0.20	*	0.02	0.04	0.01

Table 1. (Continued.)

	°C	pH	4	5	33	34	44	Polyamines (µmol/g wet wt. cell)														
								3(3)3	3(3)4	343	334	3343	3(3)4	3(3)3	3(3)43							
Order <i>Thermoanaerobacteriales</i>																						
Family <i>Thermoanaerobacteriaceae</i>																						
<i>Thermanaeromonas toyohensis</i>	JCM 11376 <sup>T</sup>	70	6.5	—	0.02	0.01	0.10	—	0.15	*	0.85	0.10	—	—	0.20	0.16	0.05					
<i>Thermoanaerobacter acetoeithylicus</i>	ATCC 33265 <sup>T</sup>	(e) 60	7.2	0.95	—	2.80	—	—	—	*	0.45	—	—	—	—	—	—					
<i>Thermoanaerobacter brockii</i>	ATCC 53556 <sup>T</sup>	(e) 60	6.5	0.35	0.10	—	0.45	—	—	*	0.80	—	—	—	—	—	—					
<i>Thermoanaerobacter ethanolicus</i>	ATCC 31550 <sup>T</sup>	(e) 60	6.5	0.68	0.14	—	1.30	—	—	*	0.50	—	—	—	—	—	—					
<i>Thermoanaerobacter kivui</i>	ATCC 33488 <sup>T</sup>	(d) 65	7.2	0.01	0.03	—	0.50	—	—	*	2.15	—	—	0.04	—	0.40	—					
<i>Thermoanaerobacter thermocopriae</i>	IAM 13577 <sup>T</sup>	(d) 60	7.1	0.61	0.02	—	1.25	—	—	*	2.75	—	—	0.01	—	0.16	—					
<i>Thermoanaerobacter thermohydrosulfuricus</i>	ATCC 35045 <sup>T</sup>	(e) 60	7.2	0.12	0.08	—	1.48	—	—	*	2.15	—	—	—	—	—	—					
<i>Caldanaerobacter subterraneus</i> subsp. <i>subterraneus</i>	ATCC BAA-225 <sup>T</sup>	65	7.5	0.15	0.25	—	0.69	—	—	1.62	*	*	—	*	—	0.24	—					
<i>Caldanaerobacter subterraneus</i> subsp. <i>tengcongensis</i>	JCM 11007 <sup>T</sup>	55	7.5	0.65	0.90	—	1.40	—	—	0.50	*	*	—	*	—	0.06	—					
		65	7.5	0.80	0.75	—	1.15	—	—	0.90	*	*	—	*	—	0.17	—					
		75	7.5	0.15	0.05	0.01	0.51	0.01	—	*	0.55	0.10	0.01	—	0.05	1.20	—					
<i>Thermoanaerobacterium polysaccharolyticum</i>	ATCC BAA-17 <sup>T</sup>	65	6.8	—	0.10	—	0.56	—	—	0.46	*	*	—	*	—	—	—					
<i>Thermoanaerobacterium saccharolyticum</i>	ATCC 49915 <sup>T</sup>	(e) 60	7.0	0.40	0.10	—	1.64	—	—	*	0.20	—	—	—	—	—	—					
<i>Thermoanaerobacterium thermosaccharolyticum</i>	ATCC 7956 <sup>T</sup>	(e) 50	7.0	1.35	0.02	—	1.49	—	—	*	0.35	—	—	—	—	—	—					
<i>Thermoanaerobacterium thermosulfurigenes</i>	ATCC 33743 <sup>T</sup>	(e) 60	7.3	0.64	0.05	—	1.22	—	—	*	2.45	—	—	—	—	—	—					
<i>Thermoanaerobacterium zeae</i>	ATCC BAA-16 <sup>T</sup>	65	6.8	—	0.10	—	0.45	—	—	0.35	*	*	—	*	—	—	—					
<i>Gelria glutamica</i>	ATCC BAA-262 <sup>T</sup>	55	7.0	—	—	—	0.15	—	—	*	0.85	—	—	—	—	—	—					



Table 1. (Continued.)

			Polyamines ( $\mu\text{mol/g}$ wet wt. cell)															
			°C	pH	4	34	343	3343	33343	33433	3(3)4	3(3)(3)4	3(3)(3)4	3(3)(3)4	343	343	Agm	
<i>Alicyclobacillus pomorum</i>			IAM 14988 <sup>T</sup>	55	4.5	—	0.83	0.06	—	—	—	—	—	—	—	—	—	—
				45	4.0	—	0.75	0.15	—	—	—	—	—	—	—	—	—	0.09
				60	4.0	—	0.80	0.15	—	—	—	—	—	—	—	—	—	0.08
<i>Alicyclobacillus sendatensis</i>			JCM 11817 <sup>T</sup>	55	4.5	—	1.04	0.75	—	—	—	—	—	—	—	—	—	0.20
<i>Bacillus schlegelii</i>			ATCC 43741 <sup>T</sup>	(h)	7.0	—	0.60	1.50	0.01	0.01	0.15	0.20	—	—	—	—	—	—
<i>Bacillus tusciae</i>			IFO 15132 <sup>T</sup>	(h)	5.0	0.06	1.90	1.78	—	—	—	—	—	—	—	—	—	0.01
Family <i>Paenibacillaceae</i>																		
<i>Thermicanus aegyptius</i>			ATCC 700890 <sup>T</sup>	55	7.0	0.10	1.20	0.10	—	—	—	—	—	—	—	—	—	—
Family <i>Bacillaceae</i>																		
<i>Bacillus smithii</i>			IFO 15311 <sup>T</sup>	(h)	60	7.3	—	1.57	0.96	—	—	—	—	—	—	—	0.01	0.01
<i>Bacillus thermocloacae</i>			ATCC 49805 <sup>T</sup>	(g)	60	7.0	—	0.74	1.60	—	—	—	—	—	—	—	—	—
<i>Saccharococcus thermophilus</i>			ATCC 43125 <sup>T</sup>	(h)	70	7.0	0.10	0.76	0.93	—	0.02	0.02	0.02	0.03	0.04	0.04	0.05	
<i>Geobacillus stearothermophilus</i>			IAM 11062 <sup>T</sup>	(h)	65	7.5	—	0.45	2.50	—	—	—	—	—	—	0.01	0.05	
<i>Geobacillus thermocatenulatus</i>			IFO 15316 <sup>T</sup>	(h)	65	7.3	0.04	0.14	3.25	—	—	—	—	—	—	0.02	0.04	
<i>Anoxybacillus flavithermus</i>			IFO 15317	(h)	65	7.3	—	1.15	0.50	—	—	—	—	—	—	—	0.02	
<i>Anoxybacillus gonensis</i> (Aero)			NCIMB 13933 <sup>T</sup>	60	7.3	—	0.35	0.90	—	—	—	—	—	—	—	—	—	
			(Anaero)	60	7.3	—	0.50	1.05	—	—	—	—	—	—	—	—	—	
<i>Anoxybacillus</i> (Aero)			JCM 12111 <sup>T</sup>	50	7.5	0.03	0.90	0.70	0.04	—	—	—	—	—	—	—	—	0.05
<i>voinovskiensis</i> (Anaero)				50	7.5	—	0.72	0.60	0.04	—	—	—	—	—	—	—	—	0.10
<i>Ureibacillus thermosphaericus</i>			NCIMB 13819 <sup>T</sup>	60	7.3	—	1.60	0.20	—	—	—	—	—	—	—	—	—	0.04
				°C	pH	4	34	343	343	343	343	343	343	Agm				
<i>Desulfotomaculum</i> lineage I																		
Family <i>Peptococcaceae</i>																		
<i>Desulfotomaculum acetoxidans</i>			ATCC 49208 <sup>T</sup>	(g)	37	7.2	—	—	—	—	0.35	0.77	—	—	—	—	—	
<i>Desulfotomaculum nigrificans</i>			ATCC 7946	(g)	55	7.5	0.04	0.04	0.04	0.74	0.10	0.10	0.52					
			IFO 13698 <sup>T</sup>	(g)	55	7.5	0.02	0.04	0.04	1.20	0.20	0.20	0.45					
			ATCC 23193 <sup>T</sup>	(g)	37	7.5	0.10	—	—	0.05	—	—	—					
<i>Desulfotomaculum ruminis</i>			ATCC 49756 <sup>T</sup>	(g)	60	7.0	—	—	0.25	2.05	0.45	0.02						
<i>Desulfotomaculum thermobenzoicum</i>			NCIMB 13375 <sup>T</sup>	(g)	50	7.4	—	—	—	0.22	0.18	—						
<i>Desulfotomaculum thermosapovorans</i>			ATCC 51507 <sup>T</sup>	(g)	35	6.5	—	—	—	0.57	0.02	—						
<i>Desulfotomaculum dehalogenans</i>			ATCC 700041	(g)	35	7.3	—	—	—	0.74	0.14	—						



Table 1. (Continued.)

	°C	pH	Polyamines (μmol/g wet wt. cell)					
			4	5	34	343	Agm	
<i>Desulfosporosinus meridiei</i>	30	7.4	—	—	0.40	0.10	—	
<i>Desulfosporosinus orientis</i>	30	7.5	—	—	0.97	0.30	—	
<i>Pelotomaculum thermopropionicus</i>	55	7.0	0.05	—	0.05	0.50	—	
Family <i>Thermoanaerobacteriaceae</i>								
<i>Sporotomaculum hydroxybenzoicum</i>	30	7.6	—	—	0.65	0.15	—	
<i>Sporotomaculum syntrophicum</i>	37	6.5	—	—	0.35	0.45	—	
Family <i>Lachnospiraceae</i>								
<i>Acetitomaculum ruminis</i>	37	7.5	0.20	—	0.24	0.06	0.02	

3, diaminopropane; 4, putrescine; 5, cadaverine; 33, norspermidine; 34, spermidine; 44, homospermidine; 333, norspermine; 343, spermine; 334, thermospermine; 344, aminopropylhomospermidine; 3333, caldopentamine; 3334, homocaldopentamine; 33333, caldohexamine; 333334, homocaldohexamine; 33343, thermohehexamine; 33433, homothermohehexamine; 3(3)3, N<sup>4</sup>-aminopropylhomospermidine; 3(3)4, N<sup>4</sup>-aminopropylspermidine; 3(3)(3)3, N<sup>4</sup>-bis(aminopropyl)norspermidine; 3(3)(3)4, N<sup>4</sup>-bis(aminopropyl)spermidine; 3(3)43, N<sup>4</sup>-bis(aminopropyl)spermine; Agm, agmatine; (Aero), aerobically grown; (Anaero), anaerobically grown; —, not detected (<0.005); \*, data not separated; †, type strain; ATCC, American Type Culture Collection, Manassas, Virginia, USA; IAM, IAM Culture Collection, Institute of Molecular and Cellular Biosciences, the University of Tokyo, Tokyo, Japan; JCM, Japan Collection of Microorganisms, RIKEN, Wako, Saitama, Japan; NBRC, Biological Resource Center, National Institute of Technology and Evaluation, Kisarazu, Chiba, Japan; NCIMB, the National Collections of Industrial, Food and Marine Bacteria, Aberdeen, Scotland, UK. (a) Cited from Hamana et al.(1998); (b) Hamana et al.(1999); (c) Hamana et al.(1996a); (d) Hamana et al.(1996a); (e) Hamana et al. (1996c); (f) Hamana et al. (2001); (g) Hamana (1999); (h) Hamana et al. (1993). °C, culture temperature.

50W column to concentrate long and branched polyamines (Hamana et al., 1998). Gas chromatography (GC) was performed after heptafluorobutyrylation of the concentrated polyamine samples (Niitsu et al., 1993). Polyamines were identified by gas chromatography-mass spectrometry (GC-mass) (Niitsu et al., 1993).

## Results and Discussion

HPLC analysis of polyamines has been developed in our laboratory and resolution of many polyamines was improved (Hamana and Itoh, 2001; Hamana et al., 2001). Norspermine [333] and *N*<sup>4</sup>-aminopropyl norspermidine [3(3)3] were co-eluted in the HPLC. Separation of thermospermine [334], spermine [343] and *N*<sup>4</sup>-aminopropylspermidine [3(3)4] was difficult by HPLC. When a concentrated polyamine sample is subjected to GC analysis, they are identified. Thermopentamine

[3343] and homocaldopentamine [3334] were separately eluted in the HPLC and GC. Typical HPLC chromatograms of the acid extracted polyamines are shown in Fig. 1. Some GC chromatograms of the concentrated polyamine samples are shown in Fig. 2. Polyamine peaks were further identified by GC-mass. During GC analysis, a quaternary penta-amine, *N*<sup>4</sup>-bis(aminopropyl)norspermidine [3(3)(3)3], was converted to a tertiary tetra-amine, [3(3)3], and another quaternary penta-amine, *N*<sup>4</sup>-bis(aminopropyl)spermidine [3(3)(3)4] was converted to two tertiary tetra-amines, [3(3)3] and [3(3)4] (Hamana et al. 1992). Cellular concentrations of polyamines estimated from the HPLC and GC analyses are shown in Table 1. A list of trivial names, systematic names and chemical formulae of polyamines has been recorded elsewhere (Hamana, 2002; Hamana and Matsuzaki, 1992; Hamana et al., 1994). Polyamine profiles within each order and lineage are followed.

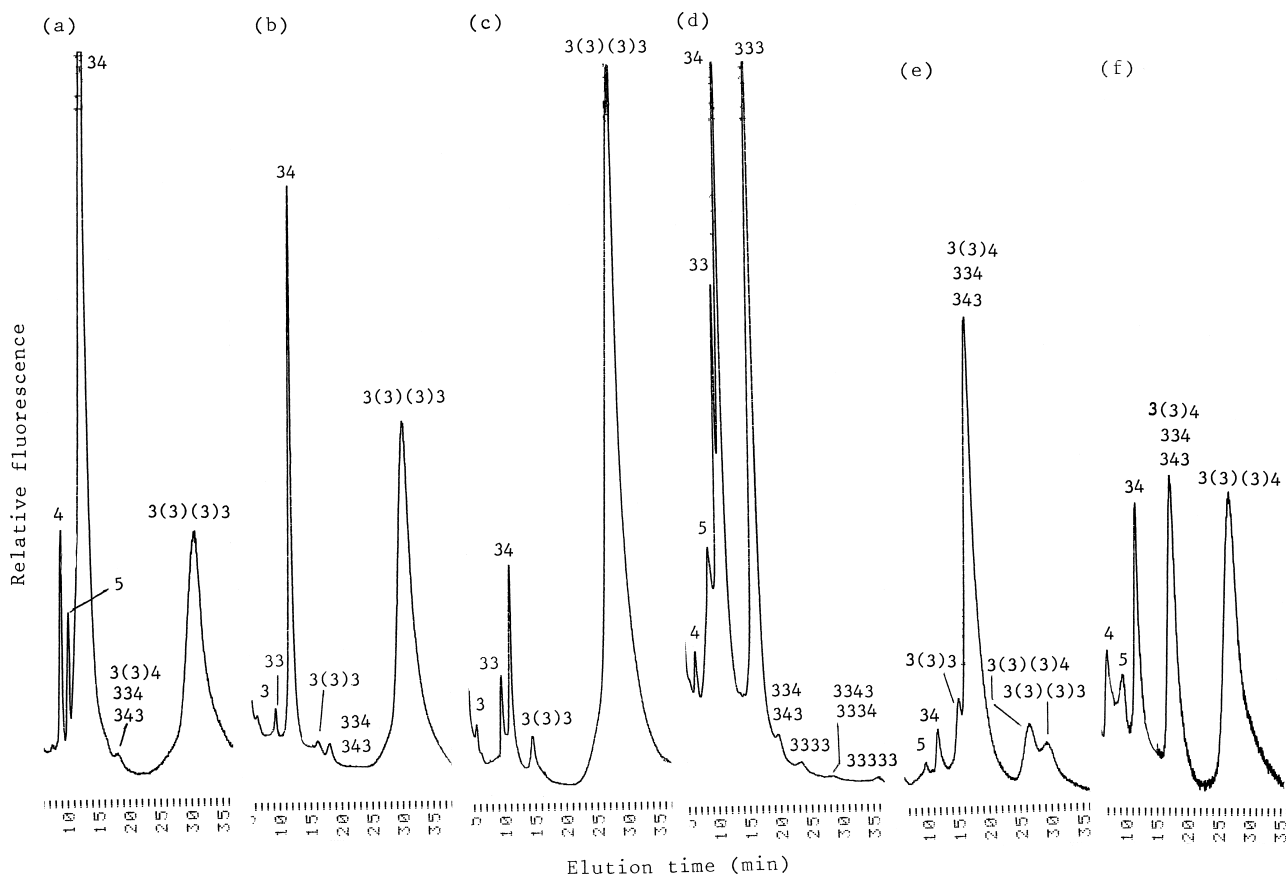


Fig. 1. HPLC analysis of polyamines extracted from (a) *Sulfurihydrogenibium subterraneum* JCM 11477; (b) *Hydrogenobacter subterraneus* JCM 10560; (c) *Hydrogenothermus marinus* JCM 10974; (d) *Thermotoga petrophila* JCM 10881; (e) *Thermanaeromonas toyohensis* JCM 11376; (f) *Caldanaerobacter subterraneus* subsp. *tengcongensis* JCM 11007 (grown at 75°C).

Abbreviations for polyamines are given in Table 1.

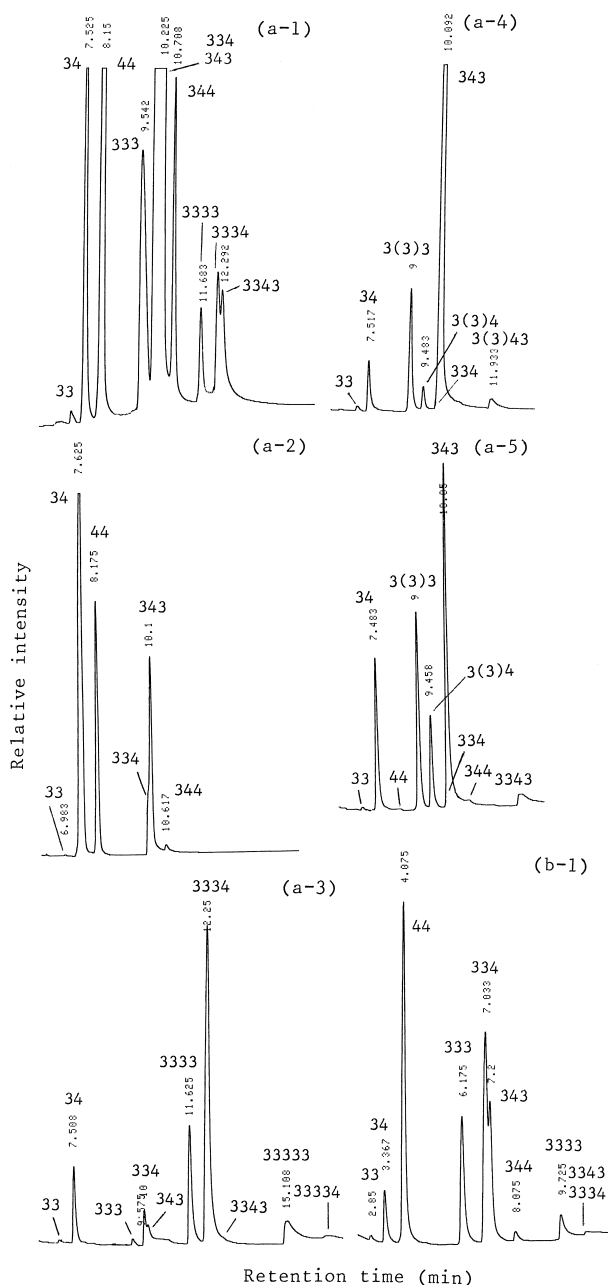


Fig. 2. GC analysis of the concentrated long polyamine fractions from (a-1) *Thermus igniterrae* ATCC 700962; (a-2) *Meiothermus taiwanensis* ATCC BAA-399; (a-3) *Vulcanithermus mediatlanticus* JCM 11956; (a-4) *Thermaeromonas toyohensis* JCM 11376; (a-5) *Caldanaerobacter subterraneus* subsp. *tengcongensis* JCM 11007 (grown at 75°C); (b-1) *Thermus antranikianii* ATCC 700961.

GC-column oven temperature was raised from 120 to 280°C in (a) and from 200 to 280°C in (b) (Niitsu et al., 1993). Abbreviations for polyamines are given in Table 1. Retention times (min) are printed on each peak.

### Aquificales

The genera *Aquifex*, *Thermocrinis*, *Hydrogenobacter* and *Hydrogenobaculum* phylogenetically belong to the family *Aquificaceae* of the order *Aquificales*. *Persephonella*, *Hydrogenothermus* and *Sulfurihydrogenibium* are located in the family *Hydrogenothermaceae* of the order (Eder and Huder, 2002; Takai et al., 2003a). *Thermovibrio* and *Balnearium* together with *Desulfurobacterium*, represent a tentative family *Desulfurobacteriaceae* of the order (Huber et al., 2002; Takai et al., 2003b). We reported the occurrence of a quaternary branched penta-amine, *N*<sup>4</sup>-bis(aminopropyl)norspermidine, as the main polyamine of a species of *Aquifex*, a species of *Hydrogenobaculum* and two *Hydrogenobacter* species (Hamana et al., 1995, 1999). The eight new members of *Aquificales* ubiquitously contained *N*<sup>4</sup>-bis(aminopropyl)norspermidine as the major polyamine.

The ubiquitous and abundant occurrence of *N*<sup>4</sup>-bis(aminopropyl)norspermidine within the moderate-, extreme- and hyper-thermophiles, ranging from 62 to 85°C as the optimum growth temperature, characterizes the order *Aquificales*.

### Thermotogales

Several thermophiles belonging to *Thermotogales*, previously analyzed, contained linear triamines, tetraamines, penta-amines and a hexa-amine (Hamana et al., 1998, 2001). Among the *Thermotoga* species examined, *Thermotoga naphthophila* and *Thermotoga petrophila* growing at 80°C contained norspermidine, norspermine, caldopentamine and thermopentamine, in addition to spermidine and spermine (and/or thermospermine). On the other hand, *Thermotoga littingae* growing at 65°C was devoid of norspermine, caldopentamine and thermopentamine. Existence of homospermidine in *T. littingae* was confirmed by the GC analysis. Such difference in the polyamine profile of *T. littingae*, as well as that of *Thermotoga elfii*, might reflect their distinct phylogenetic position and growth temperature from other *Thermotoga* species. When *Thermotoga maritima* was cultivated at lower temperatures, penta-amine and hexa-amine were decreased. Therefore, the difference in the long polyamine composition may be attributed to the difference of growth temperature. Likewise, *Marinitoga piezophila*, *Petrotoga olearia* and *Petrotoga sibirica* growing at 55–65°C showed polyamine profiles lacking penta-amines and hexa-amines. The level of norspermidine was low in all

the *Petrotoga* species and not significantly affected by the growth temperatures in *Thermotoga maritima*. The low norspermidine level in the genus *Petrotoga* and *T. elfii* may represent a chemotaxonomic feature.

Within other members of the order *Thermotogales* (the family *Thermotogaceae*), linear penta-amines have been detected in *Fervidobacterium islandicum* growing at 70°C, but not in *Fervidobacterium nodosum* and *Thermosipho japonicus* growing at 70°C (Hamana et al., 1996a, 2001).

#### *Thermodesulfobacteriales*

The occurrence of a quaternary branched penta-amine,  $N^4$ -bis(aminopropyl)spermidine, in the two species of *Thermodesulfobacterium* growing at 65–70°C, have been reported in our previous reports (Hamana et al., 1996a, 1999). A new member of the order *Thermodesulfobacteriales* (the family *Thermodesulfobacteriaceae*), *Thermodesulfatator indicus*, growing at 70°C, contained the branched penta-amine indicating the ubiquitous occurrence of the branched penta-amine in the two extremely-thermophilic, sulfate-reducing genera of this order.

#### *Thermales*

Six *Thermus* species previously analyzed contained many triamines, tetra-amines and penta-amines (Hamana et al., 1991, 1998, 1999), as listed in Table 1. A branched tetra-amine,  $N^4$ -aminopropyl-norspermidine, and a branched penta-amine,  $N^4$ -bis(aminopropyl)norspermidine, have been detected in two of the six species. In the two new members of extremely thermophilic *Thermus* growing at 70°C, almost the same polyamine components were identified; however, hexa-amines and branched polyamines were not detected. Penta-amines increased at higher growth temperature of *T. thermophilus*. A new species of the genus *Meiothermus*, *M. taiwanensis* had a polyamine pattern similar to that of the three *Meiothermus* species previously reported (Hamana et al., 1991, 1998, 1999). Spermidine, homospermidine and spermine were found as the major polyamine and norspermidine and norspermine were minor in the moderately thermophilic genus *Meiothermus* growing at 55–60°C. Although levels of norspermine were affected by growth temperature in *T. thermophilus*, a significant amount of norspermine as well as norspermidine was found in the cells grown at the lower temperature 50°C.

A marine species, *Marinithermus hydrothermalis* growing at 70°C, contained norspermidine, spermidine, homospermidine, spermine (and/or thermospermine) and homocaldopentamine (and/or thermopentamine). Two hexa-amines were detected but not homospermidine in *Vulcanithermus mediatlanticus* growing at 70°C. *Oceanithermus profundus* growing at 60–68°C contained spermine alone as the major polyamine and lacked norspermidine and homospermidine as well as long and branched polyamines. Therefore, some genus-specific polyamine profiles among the five genera of the *Thermaceae* were observed.

The family *Thermaceae* (the order *Thermales*) belongs to the phylum *Deinococcus-Thermus*, however, mesophilic *Deinococcus* species belonging to another family *Deinococcaceae* (the order *Deinococcales*) lacked norspermidine, homospermidine, and long and branched polyamines (Hamana, 1994).

#### *Thermoanaerobacteriales*

Two quaternary penta-amines,  $N^4$ -bis(aminopropyl)spermidine and  $N^4$ -bis(aminopropyl)norspermidine, were found as major polyamines in extremely thermophilic *Thermanaeromonas toyohensis* growing at 70°C. In addition, a novel tertiary branched penta-amine,  $N^4$ -aminopropylspermine, was detected by GC analysis of the concentrated polyamine fraction from this organism. This tertiary penta-amine is produced from a quaternary hexa-amine,  $N^4$ -bis(aminopropyl)spermine, during GC analysis (Hamana et al., 1992, 1993). It is strongly suggested that the quaternary branched hexa-amine exists in *T. toyohensis*.

Recently, *Caldanaerobacter subterraneus* subsp. *subterraneus* (formerly *Thermoanaerobacter subterraneus*) and *Caldanaerobacter subterraneus* subsp. *tengcongensis* (formerly *Thermoanaerobacter tengcongensis*) were phylogenetically separated from other *Thermoanaerobacter* species (Fardeau et al., 2004).  $N^4$ -bis(aminopropyl)spermidine was found in the two *Caldanaerobacter* species, as a major polyamine, as well as *Thermoanaerobacter kivui* and *Thermoanaerobacter thermocopriae*, previously reported (1996c). Extremely thermophilic *C. subterraneus* subsp. *tengcongensis* growing at 75°C contained norspermidine, homospermidine, aminopropylhomospermidine and thermopentamine as a minor polyamine component. A minor occurrence of thermospermine was found by GC-mass analysis. When *C. subterraneus* subsp.

*tengcongensis* was cultivated at lower temperatures, 55°C and 65°C, it still had the branched penta-amine in decreased amounts. Moderate thermophilic *Thermoanaerobacterium* species and *Gelria glutamica*, growing at 55–65°C, contained putrescine, spermidine and spermine and lacked long and branched polyamines (Table 1). In *Caprothermobacter* and *Moorella* species growing at 60°C,  $N^4$ -bis(aminopropyl)spermidine was found in some species (Hamana et al., 1996c). Thus, the distribution profiles of long and branched polyamines were variable within the order *Thermoanaerobacteriales*.

#### *Clostridiales*

Strictly-aerobic, thermophilic *Thermaerobacter mari-anensis* contained a quaternary branched penta-amine,  $N^4$ -bis(aminopropyl)spermidine, and agmatine (Hamana et al., 2001). Two new species of *Thermaerobacter*, *T. nagasakiensis* and *T. subterraneus*, also contained  $N^4$ -bis(aminopropyl)spermidine and agmatine. These extreme thermophiles, growing at 70°C, show same polyamine profiles.

Moderately thermophilic anaerobes, belonging to the genera *Thermosyntropha* and *Thermanaerovibrio*, growing at 55–60°C, contained spermidine and spermine, but lacked long and branched polyamines. Two mesophiles, *Aminobacterium* and *Aminomonas*, were rich in putrescine and spermidine and poor in spermine. These findings suggest that the occurrence of long and branched polyamines are coupled with extreme thermophily within *Syntrophomonadaceae*, a family of the order *Clostridiales*. Thermophilic *Caldicellulosiruptor* species (growing at 70°C) of this family contained also linear and branched penta-amines (Hamana et al., 2001).

Moderately thermophilic *Clostridium* species contained spermine as a major polyamine and mesophilic *Clostridium* species lacked a significant amount of spermine, as reported in our previous studies (Hamana, 1999; Hamana et al., 1996c). Two mesophilic anaerobes, *Alkaliphilus transvaalensis* and *Alkaliphilus crotonatoxidans*, growing at 37°C, and a moderately thermophilic anaerobe, *Thermobrachium celere*, growing at 60°C, are located in the family *Clostridiaceae*. The former two strains contained spermidine as the major polyamine and the latter strain contained spermine as a major polyamine. *Caloramator fervidus* of this family, growing at 70°C, also contained spermine but not long branched polyamine

(Hamana et al., 1996c). The genus *Alkaliphilus* contained alkaliphilic and non-alkaliphilic species, and alkaliphily did not affect the polyamine composition in the two species.

#### *Bacillales*

The family *Alicyclobacillaceae* of the order *Bacillales* includes mesophilic, acidophiles and moderately (or slightly) thermophilic acidophiles (Dufresne et al., 1996; Goto et al., 2002a,b, 2003; Matsubara et al., 2002; Tsuruoka et al., 2003). When two *Sulfobacillus* species were cultivated at optimum growth temperature, spermine level was higher at higher temperatures. In the three *Alicyclobacillus* species, spermine level did not change significantly in temperature ranges between the optimal and 15°C below. Nevertheless, the levels of spermine among the different species seemed to be variable in accordance with their optimal growth temperatures. A acidic pH range from 3.0 to 4.0 in *Alicyclobacillus* species did not change significantly in their polyamine profiles. Considering that neutrophilic, moderate thermophiles of the families *Bacillaceae* and *Thermoactinomycetaceae* also contained spermine (Hamana, 1999; Hamana et al., 1993), it seems that acidophily does not affect the polyamine composition and the occurrence of spermine correlates to moderate thermophily. This family also contains thermoacidophilic *Bacillus tusciae* (growing at 50°C) containing spermidine and spermine and extremely thermophilic *Bacillus schlegelii* (growing at 75°C) containing linear penta- and hexa-amines and a branched penta-amine (Hamana et al., 1993) (Table 1).

In the family *Bacillaceae*, the occurrence of a quaternary branched hexa-amine,  $N^4$ -bis(aminopropyl)spermine (detected as  $N^4$ -aminopropylspermine in GC analysis), in addition to spermidine and spermine, was reported in thermophilic *Saccharococcus*, *Geobacillus* (Nazina et al., 2001) and *Bacillus* species growing at 60–75°C (Hamana et al., 1993). New thermophilic *Anoxybacillus* and *Ureibacillus* species (growing at 50–65°C) (Belduz et al., 2003; Fortina et al., 2001; Yumoto et al., 2004), analyzed here, contained spermidine and spermine as the major polyamine. Thermopentamine and agmatine were detected in *Anoxybacillus voinovskiensis*. Some polyamine profiles were observed in the both aerobically and anaerobically grown *Anoxybacillus* species (Table 1). The branched hexa-amine was abundant in the extreme ther-

mophiles containing high spermine levels within *Bacillaceae* and *Thermoanaerobacteriaceae*.

A moderately thermophilic acetogen growing at 55°C, *Thermicanus aegyptius*, belongs to the family *Paenibacillaceae* and is related to the other three families of this order (Gossner et al., 1999). This organism contained spermidine and spermine. In *Paenibacillaceae*, it is known that mesophilic *Paenibacillus* and *Brevibacillus* species lack spermine whereas mesophilic *Aneurinibacillus* species contain a small amount of spermine (Hamana, 1999).

#### *Desulfotomaculum lineage I*

Sulfate-reducing *Desulfotomaculum* and *Desulfo- sporosinus*, sulfite-reducing *Desulfitobacterium*, propionate-oxidizing *Pelotomaculum* and benzoate-degrading *Sporotomaculum*, located in the family *Peptococcaceae* of the order *Clostridiales* or the family *Thermoanaerobacteriaceae* of the order *Thermoanaerobacteriales*, are moderate thermophiles growing at 50–60°C or mesophiles growing at 37–30°C (Brauman et al., 1998; Fardeau et al., 1995; Imachi et al., 2002; Qiu et al., 2003; Robertson et al., 2001). The polyamine pattern of an acetogenic anaerobe, *Acetitomaculum ruminis*, belonging to the family *Lachnospiraceae* of the order *Clostridiales* (Hamana, 1999), was cited in Table 1. Although putrescine, spermidine and/or spermine were found as the major polyamines in the *Desulfotomaculum* lineage I, spermine was relatively abundant in the moderate thermophiles as suggested in our previous report (Hamana, 1999). Some of the thermophiles contained agmatine, suggesting that the cellular occurrence of spermine and agmatine may correlate to their moderate thermophily, as found in the thermophiles of *Clostridiales* and *Bacillales*.

#### *Polyamine profiles found in thermophilic eubacteria*

The thermophiles belonging to *Aquificales*, lacking long linear polyamines, ubiquitously contained a quaternary branched penta-amine. In *Thermodesulfobacteriales*, the thermophiles ubiquitously contained another quaternary branched penta-amine. The occurrence of two different quaternary branched penta-amines is chemotaxonomically significant and was not strictly affected by growth temperature in the two orders. The thermophiles of *Thermotogales* contained long linear polyamines. In the three orders phylogenetically located in early eubacterial evolution, the occur-

rence of the branched polyamines or the linear long polyamines seems to be coupled with their thermophily, and serve for a chemotaxonomy.

Extremely thermophilic *Thermus* contained long linear and branched polyamines whereas other extreme thermophiles, *Marinithermus* and *Vulcanithermus*, contained long linear polyamines. Moderately-slightly thermophilic *Meiothermus* and *Oceanithermus* lacked them. The presence or absence of homospermidine, norspermidine and its derivatives differed in the five genera; however, they serve as a chemotaxonomic marker within the *Thermales*. Linear long polyamines and branched polyamines were simultaneously found in some extreme thermophiles belonging to *Thermoanaerobacteriales*, *Clostridiales* and *Bacillales* located in Gram-positives. Various moderate thermophiles of this taxa contained spermidine and spermine alone.

It is suggested that the synthetic abilities of long linear polyamines and branched polyamines were separately developed in early evolved thermophilic eubacteria and the two polyamine synthetic pathways were mixed during later eubacterial evolution. One of them seems to be essential for extremely thermophilic eubacteria. Norspermidine- and homospermidine-syntheses might be phylogenetically developed or eliminated in eubacteria independently of their thermophily.

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