

Research Paper

Comparison of the efficiency between two sampling plans for aflatoxins analysis in maize

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Abstract

Variance and performance of two sampling plans for aflatoxins quantification in maize were evaluated. Eight lots of maize were sampled using two plans: manual, using sampling spear for kernels; and automatic, using a continuous flow to collect milled maize. Total variance and sampling, preparation, and analysis variance were determined and compared between plans through multifactor analysis of variance. Four theoretical distribution models were used to compare aflatoxins quantification distributions in eight maize lots. The acceptance and rejection probabilities for a lot under certain aflatoxin concentration were determined using variance and the information on the selected distribution model to build the operational characteristic curves (OC). Sampling and total variance were lower at the automatic plan. The OC curve from the automatic plan reduced both consumer and producer risks in comparison to the manual plan. The automatic plan is more efficient than the manual one because it expresses more accurately the real aflatoxin contamination in maize.

Key words: mycotoxins, variance, error, characteristic operation curve, *Zea mays*.

Introduction

Aflatoxins are secondary metabolites from certain fungal lineages of the *Aspergillus* genus, mainly *A. flavus* and *A. parasiticus*. Aflatoxins B₁, B₂, G₁, and G₂ are natural contaminants of seeds and feed, mainly nuts, peanuts, cotton seeds, maize, and other oilseeds (Sweeney and Dobson, 1998; Stroka *et al.*, 2000).

Obtaining trustable results on mycotoxins analysis in feed is important for management and mycotoxicological control purposes (Krska *et al.*, 2005). However, the heterogeneous distribution of mycotoxins in cereals leads to variations on the final results, so called errors, not being possible to establish the real mycotoxins concentration in a determined lot (Whitaker 2004; Miraglia *et al.*, 2005; Vargas *et al.*, 2006). Thus, the efficiency of a sampling plan can be estimated by the evaluation of the variance of the results generated by the quantification procedures applied.

The probabilities of acceptance or rejection of a lot with certain mycotoxin concentration depend on the performance of the sampling plan applied (Johansson *et al.*, 2000a; Johansson *et al.*, 2000c; Miraglia *et al.*, 2008), *i.e.* the better the performance of the sampling plan, safer the acceptance or rejection probabilities of this lot. To achieve that, it is necessary to establish the procedures for the sampling plan, as well as a limit for acceptance/rejection. The quantification procedures can be divided into three stages: 1- Sampling: taking a sample from a lot; 2- Sample preparation: milling the sample to reduce the size of its particles and take a subsample for analysis, and; 3- Analysis: extract the mycotoxin from the subsample and quantify it (Whitaker *et al.*, 2011).

The variance on the quantification results can lead to two kinds of errors: the false positive or producer/seller risk, in which a good lot is labeled as bad, and the false negative or consumer/buyer risk, where a bad lot is labeled as

good. The frequency of occurrence of those two kinds of errors depends on the sampling plan and is given by the design of a characteristic operation curve (OC) (Johansson *et al.*, 2000c; Miraglia *et al.*, 2005; Vargas *et al.*, 2006).

In regards to mycotoxins, an OC curve relates the toxin concentration in the lot (x-axis) with the probability to accept this lot (y-axis). Using a pre-established acceptance/rejection limit, the OC curve shows each seller/producer and buyer/consumer risk (as shown in Figure 1) (Miraglia *et al.*, 2008). For each sampling plan, the risks are defined by the sampling method, sample size, subsample preparation and size, number of analysis, and analyses method (Whitaker *et al.*, 2011).

Due to the lack of information on the identification of the efficiency of different sampling plans, the aim of this research was to evaluate the performance of two different sample collection methods for aflatoxins quantification in maize.

Material and Methods

Sample collection

Maize sample collection was performed in eight trucks with a load ranging from 13 to 32 tons of maize. On each truck (lot) two sampling plans for aflatoxins ($B_1+B_2+G_1+G_2$) quantification were applied: manual and automatic. The experimental design used to determine the variance associated to each stage of the aflatoxins quantification procedure on both sampling plans is presented on Figure 2.

For the manual sampling plan, kernel samples were taken using a sampling spear with 10 collector chambers. The size of each kernel sample followed the EC Regulation

No. 401/2006 (European Commission Regulation-EC No 401, 2006). From three lots with sizes ranging from 13 to 20 tons, a 6 kg collective sample was obtained from each lot through 60 increments taken from six different sampling spots. From five lots with sizes greater than 20 tons, a 10 kg collective sample was obtained from each lot through 100 increments taken from ten different sampling spots. This procedure was performed 10 times to each maize lot.

For the automatic sampling plan, the same eight lots of maize were milled (particles smaller than 9 mesh) in hammer mills and moved along the factory through a screw conveyor. Outside this screw conveyor 10 continuous flow sampling systems were installed using 10 holes with 17.5 mm diameter located on the right-hand side (along the milled maize flow), 45° from the base of the tube. The size of the collective samples for each lot was calculated based on the equation 1, and ranged from 16.1 to 25.3 kg.

$$C_s = \sqrt{20m} \quad (1)$$

where C_s is the collective sample in kg and m is the size of the lot in tons.

After the collection procedure, all the 20 samples from each lot were milled in a hammer mill with a 2 mm strainer. The milled samples were reduced to the size of 1 kg (laboratory sample) using a sample splitter with inverted channels, packed in plastic bags, and stored at -18 °C.

Sample preparation

Each laboratory sample was milled in a Retsch® ZM 200 ultra centrifugal mill (Altmann, São Paulo, SP, Brazil), at 17,000 rpm, with a 1.5 mm strainer, and split into 10 sub-

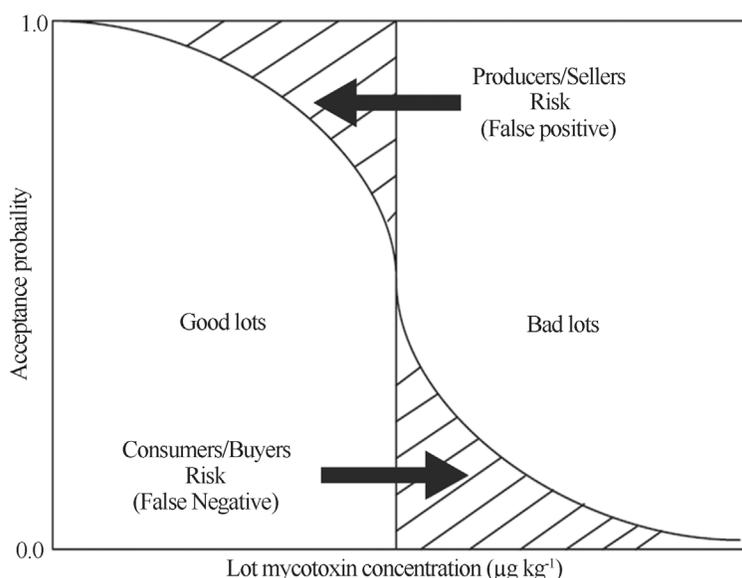


Figure 1 - Operation characteristic (OC) curve to estimate the lot acceptance/rejection probabilities and evaluate producer/seller and consumer/buyer risks, according to Miraglia *et al.* (9).

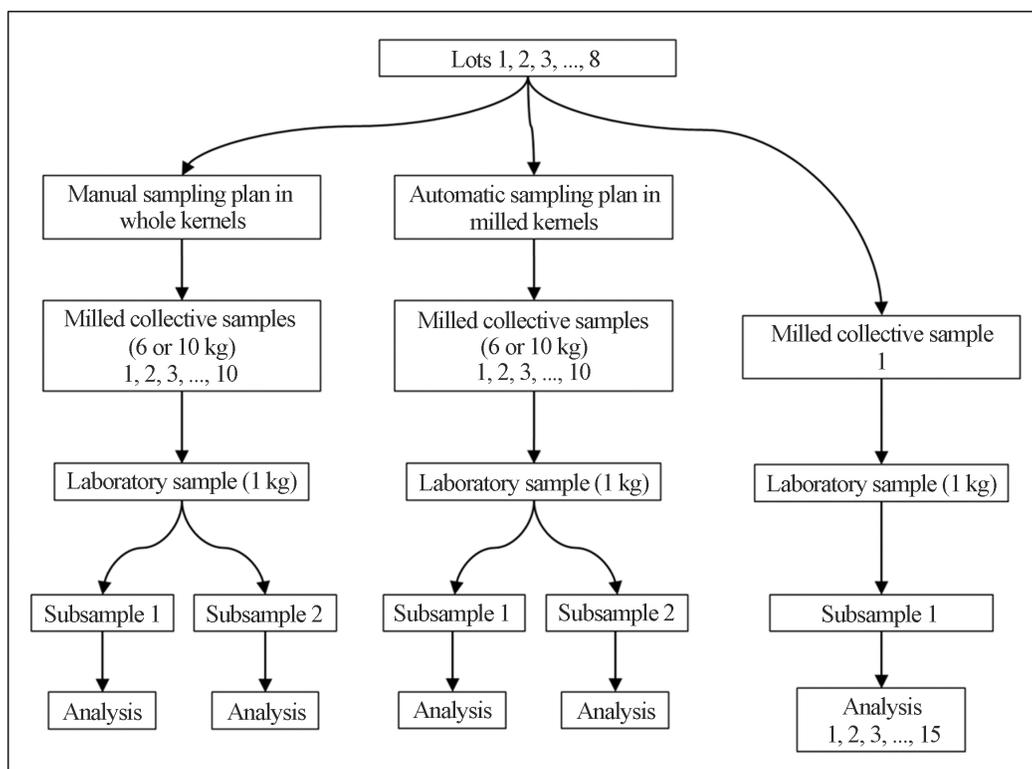


Figure 2 - Experimental design applied to evaluate two sampling plans for aflatoxins analysis in 8 lots of maize.

samples using a Retsch® PT 100 sample divider (Altmann, São Paulo, SP, Brazil).

Aflatoxin analysis

Analysis of aflatoxins ($B_1+B_2+G_1+G_2$) was performed once in two out of ten subsamples from each laboratory sample and 15 times in one extract from one subsample from each lot, totalizing 1,400 analyses. The chromatographic method was applied according to Sulyok *et al.* (2007) and showed quantification limit a recovery coefficient for aflatoxins as follows: $1 \mu\text{g kg}^{-1}$ and 94.5% for B_1 ; $1 \mu\text{g kg}^{-1}$ and 80.0% for B_2 ; $1 \mu\text{g kg}^{-1}$ and 88.5% for G_1 and; $1 \mu\text{g kg}^{-1}$ and 88.1% for G_2 .

Variability of the results on aflatoxins quantification

The variability of the results on aflatoxins quantification was determined by the calculation of the variance. The total variance ($S^2_{(t)}$) associated to the procedure of aflatoxins quantification is composed by the sum of sample variance ($S^2_{(s)}$), preparation variance ($S^2_{(p)}$), and analysis variance ($S^2_{(a)}$) (Cheli *et al.*, 2009; Ozay *et al.*, 2006; Whitaker *et al.*, 2011), according the Eq. (2).

$$S^2_{(t)} = S^2_{(s)} + S^2_{(p)} + S^2_{(a)} \quad (2)$$

The results for $S^2_{(s)}$, $S^2_{(p)}$, and $S^2_{(a)}$ were obtained by replicating results from lot, laboratory sample, and extract, respectively.

Theoretical distributions of the results from aflatoxins concentration

Four theoretical distribution models (all 2-parameter) were tested to simulate the distribution of the results from aflatoxins quantification on each lot of maize: normal, lognormal, gamma, and Weibull. Each theoretical distribution was observed from the results of 20 subsamples per lot and per sampling plan, totalizing 64 distributions.

Observed distributions of the results from aflatoxins concentration

The observed distributions were determined from the 20 results of aflatoxins quantification from each lot at each sampling plan, totalizing 16 observed distributions. The observed acceptance probabilities were calculated for each lot by counting the number of results less than or equal the acceptance limit used ($15 \mu\text{g kg}^{-1}$) and divided by 20 (number of analyses per lot in each sampling plan). A total of 8 observed points were determined to each sampling plan (one per lot). The higher value attributed to the acceptance probability was 1.0 (20/20) or 100% and the next was 0.95 (19/20).

Selection of the theoretical distribution model

The Chi-square test was used as criteria to evaluate the goodness of fit among the theoretical and observed distributions from aflatoxins concentrations. The adjustment between the theoretical distribution and the observed one

was considered to be acceptable when the statistic test did not exceed the critical value 95% ($p \geq 0.05$).

Statistical analysis

The components of the total variance were estimated for the eight lots at both sampling plans using ANOVA. The individual variances from each stage (sampling, preparation, and analysis) were submitted to a multi-factor ANOVA, using the average aflatoxins concentration ($B_1+B_2+G_1+G_2$) as covariate on the variance from each stage. Variances from each stage were compared between sampling plan using the F test at 95% of significance ($p \leq 0.05$). The sampling variances for each sampling plan, as well as the variances of preparation and analysis (regardless the sampling plan) were submitted to regression analysis against aflatoxins concentration ($B_1+B_2+G_1+G_2$) to obtain the equations. Models that showed the less value of p were used.

The estimated acceptance probabilities associated with the sampling plans described above were calculated using the lognormal distribution. Parameters for the lognormal distribution were determined using the average aflatoxin concentration (X) and the variances $S^2_{(t) \text{ manual}}$ and $S^2_{(t) \text{ automatic}}$. The acceptance/rejection limit used was $15 \mu\text{g kg}^{-1}$.

Variance estimates, regression analyses, and evaluation of the theoretical distributions were performed using

the statistical program Statgraphics Plus 5.0. The acceptance/rejection probabilities calculation and the OC curves development were performed using Microsoft Excel 2010.

Results and Discussion

Variability

Table 1 shows the variance of aflatoxins concentrations from the stages of sampling, preparation, analysis, and the total variance. Total aflatoxin concentration results from 320 subsamples analyzed ranged from 1.0 to $37.1 \mu\text{g kg}^{-1}$.

Variance associated with the sampling procedure and the total variance were respectively 5.6 and 3.9 times lower at the automatic sampling plan than at the manual one ($p \leq 0.05$). As the procedures for the preparation and analysis stages were the same, there were no differences between their variances ($p = 0.57$ and $p = 0.80$) (Figure 3).

Johansson *et al.* (2000a) estimated the total variance components of the results from aflatoxins quantification in lots of maize. Thus, they took a 45 kg collective samples from each lot, reduced them to 1.13 kg and milled them. They evaluated lots of maize with similar aflatoxins concentration found in this research. However, the variance associated with each stage of the aflatoxins quantification procedure was lower in this study. Regarding the sampling

Table 1 - Average and median of aflatoxins concentration, coefficient of variation and sampling, preparation, analysis, and total variances by lot of maize and sampling plan.

Lot	Sampling plan	Average ¹	Median ¹	CV ²	Variances ³			
					$S^2_{(s)}$	$S^2_{(p)}$	$S^2_{(a)}$	$S^2_{(t)}$
1	Manual	19.42	16.13	36.10	46.118	0.616	2.427	49.161
2	Manual	21.71	21.11	33.52	48.809	2.456	1.699	52.964
3	Manual	20.05	20.25	23.37	18.988	2.924	0.058	21.970
4	Manual	14.33	13.67	44.57	38.693	1.989	0.122	40.804
5	Manual	5.56	5.87	30.67	2.662	0.108	0.135	2.905
6	Manual	17.43	16.25	38.23	42.084	0.464	1.838	44.386
7	Manual	1.69	1.20	63.25	1.114	0.014	0.019	1.147
8	Manual	17.97	17.77	27.99	21.269	2.705	1.315	25.289
1	Automatic	19.96	19.72	12.72	3.391	0.632	2.427	6.449
2	Automatic	19.55	19.16	17.58	6.812	3.306	1.699	11.818
3	Automatic	14.64	14.15	17.67	3.083	3.547	0.058	6.689
4	Automatic	14.48	14.22	11.88	0.557	2.279	0.122	2.958
5	Automatic	5.34	5.45	15.50	0.485	0.066	0.135	0.686
6	Automatic	17.11	17.37	15.62	4.828	0.484	1.838	7.150
7	Automatic	1.59	1.56	24.12	0.125	0.004	0.019	0.148
8	Automatic	17.30	16.28	24.08	13.422	2.609	1.315	17.346

¹Average aflatoxins concentration ($B_1+B_2+G_1+G_2$) in $\mu\text{g kg}^{-1}$ from 20 analyzed subsamples per lot.

²Coefficient of variation (%).

³ $S^2_{(s)}$, $S^2_{(p)}$, $S^2_{(a)}$ e $S^2_{(t)}$: Sampling, preparation, analysis, and total variances in $(\mu\text{g kg}^{-1})^2$.

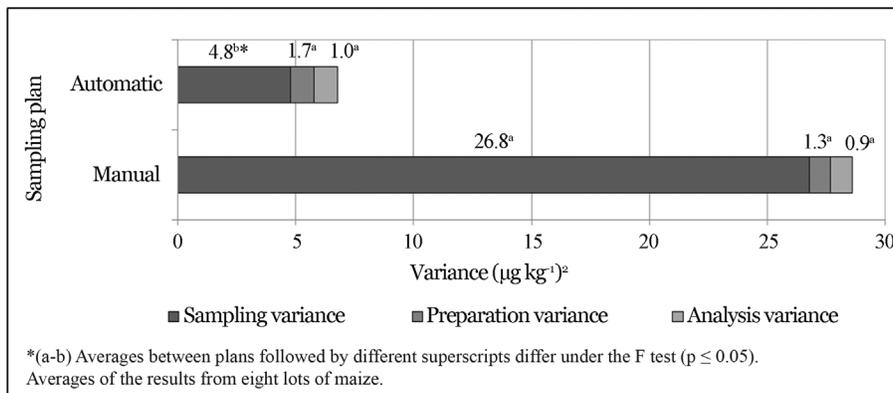


Figure 3 - Average of the variances from each stage of the aflatoxins quantification procedure in maize (sampling, preparation, and analysis) from the manual and automatic sampling plan.

stage, the lower variability found in this research is probably due to the fact that collective samples were milled before the laboratory sample was taken. Milling the collective sample raises the number of particles per unit of mass, lowering the variability among the analysis results (FAO 1993). Preparation and analysis stages also showed lower variability, because samples were prepared using different equipments and methods for milling and sub sampling. The analysis performed with high performance liquid chromatography tandem mass spectrometry (LC-MS/MS) is more specific and has simpler extraction and analyses procedures (Sforza *et al.*, 2006), than those applied to the thin layer chromatography (TLC) used by Johansson *et al.* (2000a).

Shotwell *et al.* (1975) found that if a lot is properly mixed, contaminated particles will probably be equally distributed along its mass. Under this condition, the spot where the sample is taken is probably not relevant. However, if the lot is not previously mixed, mycotoxin contaminated particles can be located in some delimited areas of the lot. Thus, if sampling is done by taking samples from only one spot on the lot, contaminated particles may not be sampled, or, many contaminated particles may be harvested (Whitaker *et al.*, 2011). Those situations can occur more frequently when using the manual sampling system approached in this research.

A sample is much more significant if it is harvested from milled materials due to the highest number of particles per unit of volume or per unit of mass when compared to whole kernels (Tittlemier *et al.*, 2011). In several studies, Whitaker *et al.* (1976, 1994, 1998, 2011) found that the sample size infers on the variability of the results, in other words, as bigger the sample, the smaller the variance. The collection of increments while a lot is being moved from one side to other (dynamic lot) provides better representativity to the sample when compared to the sampling applied to a static lot (Miraglia *et al.*, 2005; Whitaker *et al.*, 2011). In this research, the reduction on the results variability from aflatoxins concentration at the automatic sampling is probably associated to the fact that the whole

lot is milled and conducted through a screw conveyor thus, both the milling process and the transportation help on the lot homogeneity. Besides, the collective sample size at the automatic sampling plan is bigger and the collection of increments is uninterrupted, which means there is always a small portion being harvested giving a better randomness on particles collection and thus, better representativity of the sample.

The $S^2_{(s)}$ estimate was performed using the regression analysis for each sampling plan, where the equation between sampling variance and aflatoxins concentration (X) was:

$$S^2_{(s) \text{ manual}} = 2718^{(-0.952 + 1.514 \cdot \text{NL}(X))} \quad (3)$$

$$S^2_{(s) \text{ automatic}} = 2718^{(-3.0332 + 1.531 \cdot \text{NL}(X))} \quad (4)$$

under a determination coefficient (R^2) of 0.887 and 0.730 respectively, and where NL is the natural logarithm.

The $S^2_{(p)}$ and the $S^2_{(a)}$ were obtained by performing only one regression analysis using both sampling plans, where the equation between preparation variance and aflatoxins concentration (X) was:

$$S^2_{(p)} = 2718^{(-5.997 + 2.232 \cdot \text{NL}(X))} \quad (5)$$

and the equation between analysis variance and aflatoxins concentration (X) was:

$$S^2_{(a)} = \frac{1}{(-1.207 + 85.741 / X)} \quad (6)$$

under $R^2 = 0.849$ and 0.875 respectively.

The $S^2_{(t)}$ was estimated by the sum of $S^2_{(s)}$, $S^2_{(p)}$, and $S^2_{(a)}$:

$$S^2_{(t) \text{ manual}} = 2718^{(-0.952 + 1.514 \cdot \text{NL}(X))} + 2718^{(-5.997 + 2.232 \cdot \text{NL}(X))} + \frac{1}{(-1.207 + 85.741 / X)} \quad (7)$$

$$S_{(t) \text{ automatic}}^2 = 2.718^{(-3.033 + 1.531 \cdot \text{NL}(X))} + \frac{1}{2.718^{(-5.997 + 2.232 \cdot \text{NL}(X))} + (-1.207 + 85.741/X)} \quad (8)$$

The variance estimate in each stage from both sampling plans rose as the aflatoxins concentration rose ($r > 0.85$). Similar results were observed by Johansson *et al.* (2000a) in a research with aflatoxins in maize, Whitaker *et al.* (1994) with aflatoxins in peanuts, and Whitaker *et al.* (1998) with fumonisins in maize.

The $S_{(t)}$ estimate associated to a lot of maize with $15 \mu\text{g kg}^{-1}$ of aflatoxins is $24.57 (\mu\text{g kg}^{-1})^2$ at the manual sampling plan and $4.31 (\mu\text{g kg}^{-1})^2$ at the automatic one. The $S_{(s) \text{ manual}}^2$, $S_{(s) \text{ automatic}}^2$, $S_{(p)}^2$, and $S_{(a)}^2$ are 23.30, 3.04, 1.05, and 0.22 $(\mu\text{g kg}^{-1})^2$ respectively. The $S_{(s)}$, $S_{(p)}$, and $S_{(a)}$ represent about 95%, 4%, and 1% of the total variance from the manual sampling plan and 71%, 24%, and 5% of the total variance from the automatic sampling plan. The percentage of $S_{(t)}$ that corresponds to preparation and analysis was bigger at the automatic sampling plan due to a reduced $S_{(s)}$ in this sampling plan. At both sampling plans, the $S_{(s)}$ had the major contribution to the $S_{(t)}$, followed by the $S_{(p)}$ and $S_{(a)}$. This is consistent with observations on other mycotoxins in several seeds and feeds (Whitaker *et al.*, 1976, 1998; FAO 1993; Johansson *et al.*, 2000c; Ozay *et al.*, 2006).

Distribution of aflatoxins results

A summary of the Chi-square test for each theoretical distribution is described at the end of Table 2. Despite the fact that similar models have been used to precisely simulate the distribution of the results from mycotoxins quantification in seeds and feeds (Sharkey *et al.*, 1994; Giesbrecht *et al.*, 1998; Johansson *et al.*, 2000b; Vargas *et al.*, 2006),

lognormal distribution was chosen because it obtained the major number of lots with acceptable distribution and highest average probability. Thus, this distribution was used to calculate the OC curve and evaluate the performance of both sampling plans for aflatoxins quantification in maize.

Operation characteristic curve

The OC curves of both sampling plans are illustrated on Figure 4 with their observed and estimated acceptance probabilities. The difference on shape from both OC curves is only due to the sampling stage, as the other stages were the same for both sampling plans. Comparing both OC curves, it is possible to verify that the automatic sampling plan reduces both producer/seller and consumer/buyer risks (indicated on Figure 1). As an example: a lot of maize with a theoretical aflatoxins contamination of $20 \mu\text{g kg}^{-1}$ and a limit of acceptance/rejection of $15 \mu\text{g kg}^{-1}$. If this lot were sampled using the manual sampling plan described in this research, it would have a 21% chance of being accepted under the limit established. On the other hand, if the same lot were sampled using the automatic sampling plan described in this research, it would have only a 3% chance of being accepted under the limit established.

Conclusions

The total variance associated to the procedures for aflatoxins quantification in maize at both the manual and automatic sampling plan raises as the aflatoxins concentration raises. The sampling stage contributes with the highest variability, followed by the preparation and analysis stages. Sampling variance and total variance are reduced at the automatic sampling plan, because it is applied in milled grains in movement, in a continuous way, and with a bigger collective sample, proving have a better performance against

Table 2 - Chi-square test probabilities on normal, lognormal, gamma, and Weibull distributions of the results from aflatoxins quantification in eight lots of maize at the two sampling plans evaluated.

Lot	Manual sampling plan				Automatic sampling plan			
	Normal	Lognormal	Gamma	Weibull	Normal	Lognormal	Gamma	Weibull
1	0.0188*	0.0724	0.0725	0.0093*	0.3973	0.5078	0.3044	0.1060
2	0.4233	0.9131	0.7974	0.6661	0.0922	0.0922	0.0922	0.0283*
3	0.4233	0.3910	0.5004	0.1365	0.1365	0.5379	0.3262	0.4232
4	0.3910	0.5004	0.5004	0.3909	0.3262	0.6661	0.4230	0.2301
5	0.5004	0.1239	0.1239	0.5004	0.5379	0.6661	0.6661	0.6661
6	0.1264	0.2301	0.3048	0.0343*	0.0028*	0.0336*	0.0058*	0.0470*
7	0.0015*	0.1365	0.0373*	0.0093*	0.9131	0.6661	0.6661	0.9131
8	0.9131	0.5380	0.2996	0.9131	0.1264	0.3973	0.1715	0.0029*
# acceptable lots	6	8	7	5	7	7	7	5
# best fit lots	0	3	3	2	0	5	1	1
Average probability	0.3497	0.3632	0.3295	0.3325	0.3165	0.4459	0.3319	0.3021

* $p < 0.05$, distribution does not fulfill the 95% confident.

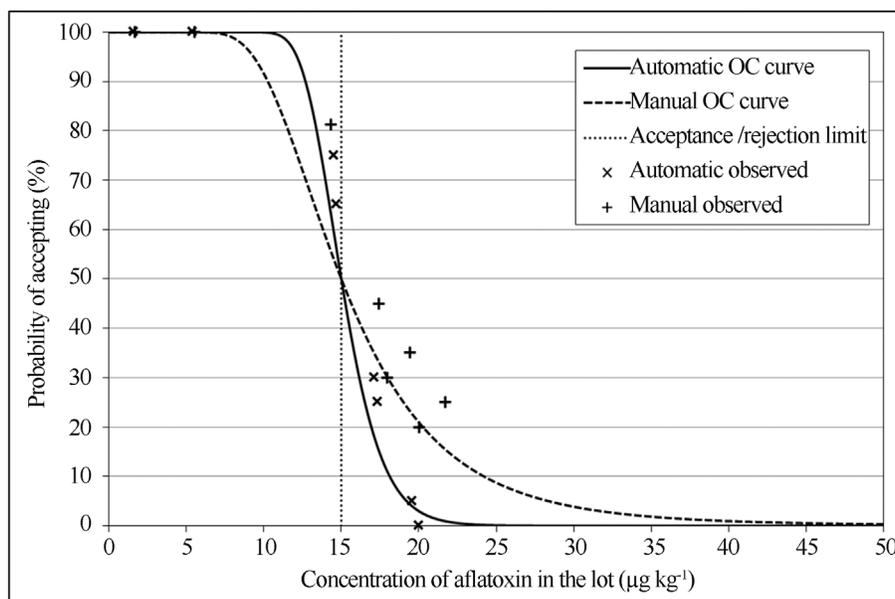


Figure 4 - Operational characteristic (OC) curves from manual and automatic sampling plans for aflatoxins quantification in maize, with observed acceptance probabilities and an acceptance/rejection limit of $15 \mu\text{g kg}^{-1}$.

the manual sampling plan. The OC curve from the automatic sampling plan reduces both producer/seller and consumer/buyer risks when compared to the OC curve from the manual sampling plan, thus, reducing the chances of misclassification of maize lots and providing more safety in commercial trades.

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References

- Cheli F, Campagnoli A, Pinotti L, Fusi E, Dell'orto V (2009) Sampling feed for mycotoxins: acquiring knowledge from food. *Ital J Food Sci* 8:5-22.
- European Commission Regulation (2006) No 401/2006 of 23 February 2006 laying down the methods of sampling and analysis for the official control of the levels of mycotoxins in foodstuffs. *Official Journal of European Union L70*:12-34.
- FAO - Food and Agriculture Organization (1993) Sampling plans for aflatoxin analysis in peanuts and corn. *FAO Food and Nutrition. Paper 55*, FAO, Viale Della Terme di Caracalla 00100, Rome, Italy, p. 27-56.
- Giesbrecht FG, Whitaker TB (1998) Investigations of the problems of assessing aflatoxin levels in peanuts. *Biometrics* 54:739-753.
- Johansson AS, Whitaker TB, Hagler WM, Giesbrecht FG, Young JH, Bowman DT (2000) Testing shelled corn for aflatoxin, part I: estimation of variance components. *J AOAC Int* 83:1264-1269.
- Johansson AS, Whitaker TB, Giesbrecht FG, Hagler WM, Young JH (2000) Testing shelled corn for aflatoxin, part II: modeling the distribution of aflatoxin test results. *J AOAC Int* 83:1270-1278.
- Johansson AS, Whitaker TB, Giesbrecht FG, Hagler WM, Young JH (2000) Testing shelled corn for aflatoxin, part III: evaluating the performance of aflatoxin sampling plans. *J AOAC Int* 83:1279-1284.
- Krska R, Welzig E, Berthiller F, Molinelli A, Mizaikoff B (2005) Advances in the analysis of mycotoxins and its quality assurance. *Food Addit Contam* 22:345-353.
- Miraglia M, De Santis B, Minardi V, Debegnach F, Brera C (2005) The role of sampling in mycotoxin contamination: an holistic view. *Food Addit Contam: part A* 22:31-36.
- Miraglia M, De Santis B, Pannunzi E, Debegnach F, Brera C (2008) Mycotoxin concentration data quality: the role of sampling. In: Leslie, F.J.; Bandyopadhyay, R.; Visconti, A. (eds). *Mycotoxins: detection methods, management, public health and agricultural trade*. Wallingford, UK, Cabi, p. 101-168.
- Ozay G, Seyhan F, Yilmaz A, Whitaker TB, Slate AB, Giesbrecht F (2006) Sampling hazelnuts for aflatoxin: uncertainty associated with sampling, sample preparation, and analysis. *J AOAC Int* 89:1004-1011.
- Sforza S, Dall'asta C, Marchelli R (2006) Recent advances in mycotoxin determination in food and feed by hyphenated chromatographic techniques/mass spectrometry. *Mass Spectrom Rev* 25:54-76.
- Sharkey AJ, Roch OG, Coker RD (1994) A case-study on the development of a sampling and testing protocol for aflatoxin levels in edible nuts and oil-seeds. *J R Soc Series D (The Statistician)* 43:267-75.
- Shotwell OL, Goulden ML, Bothast RJ, Hesseltine CW (1975) Mycotoxins in hot spots in grains. 1. Aflatoxin and zearalenone occurrence in stored corn. *Cereal Chem* 52:687-697.
- Stroka J, Anklam E, Jörissen U, Gilbert J (2000) Immunoaffinity column clean-up with liquid chromatography using post-column bromination for determination of aflatoxins in pea-

- nut butter, pistachio paste, fig paste, and paprika powder: collaborative study. *J AOAC Int* 83:320-340.
- Sulyok M, Krska R, Schuhmacher R (2007) A liquid chromatography/tandem mass spectrometric multi-mycotoxin method for the quantification of 87 analytes and its application to semi-quantitative screening of moldy food samples. *Anal Bioanal Chem* 389:1505-1523.
- Sweeney MJ, Dobson ADW (1998) Mycotoxin production by *Aspergillus*, *Fusarium* and *Penicillium* species. *Int J Food Microbiol* 43:141-158.
- Tittlemier SA, Varga E, Scott PM, Krska R (2011) Sampling of cereals and cereal-based foods for the determination of ochratoxin A: an overview. *Food Addit Contam: part A* 28:775-778.
- Vargas EA, Whitaker TB, Dos Santos EA, Slate AB, Lima FB, Franca RCA (2006) Design of a sampling plan to detect ochratoxin A in green coffee. *Food Addit Contam.: part A* 23:62-72.
- Whitaker TB, Whitten ME, Monroe RJ (1976) Variability associated with testing cottonseed for aflatoxin. *J Am Oil Chem Soc* 53:502-505.
- Whitaker TB, Dowell FE, Hagler WM, Giesbrecht FG, Wu J (1994) Variability associated with sampling, sample preparation, and chemically testing for aflatoxin in farmers' stock peanuts. *J AOAC Int* 77:107-116.
- Whitaker TB, Trucksess MW, Johansson AS, Giesbrecht FG, Hangler WM, Bowman DT (1998) Variability associated with testing corn for fumonisin. *J AOAC Int* 81:1162-1168.
- Whitaker TB, Sampling for mycotoxins (2004) In: Magan, N.; Olsen, M. (eds). *Mycotoxins in food: detection and control*. Cambridge, UK, Woodhead Publishing Ltd., p.69-87.
- Whitaker TB, Slate A, Doko B, Maestroni B, Cannavan A (2011) *Sampling procedures to detect mycotoxins in agricultural commodities*. Springer, New York, USA.

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