

## Prevalence of *Toxoplasma gondii* Infection in Rabbits of Korea by Serological Tests and Nested Polymerase Chain Reaction

Hyun-Guk SHIN<sup>1</sup>, Sang- Eun LEE<sup>2</sup>, Sung-Hee HONG<sup>2</sup>, Se-Mi KIM<sup>3</sup>, Young-Ki CHOI<sup>3</sup>, Hyung-Jin PARK<sup>1</sup>, Kyoung-Won SEO<sup>1</sup> and Kun-Ho SONG<sup>1</sup>\*

<sup>1</sup>Laboratory of Veterinary Internal Medicine, College of Veterinary Medicine, Chungnam National University, Daejeon 305–764, Korea

<sup>2</sup>Division of Malaria & Parasitic Diseases, Korea National Institute of Health, Korea Centers for Disease Control and Prevention, Osong-eup, Cheongwon-gun, Chungbuk 363–951, Korea

<sup>3</sup>Laboratory of Microbiology, College of Medicine, Chungbuk National University, Cheongju 361–763, Korea

(Received 14 July 2013/Accepted 30 July 2013/Published online in J-STAGE 13 August 2013)

**ABSTRACT.** This study surveyed the *Toxoplasma (T.) gondii* infection prevalence in the Korean rabbit population. Rabbits (n=142) were obtained from two breeding farms in the Gongju area, Chungnam Province, and in the Kochang area, Junbuk Province, Korea. Of 142 sera samples analyzed by enzyme-linked immunosorbent assay (ELISA), 15 (10.6%) exhibited *T. gondii*-specific IgG antibodies, and 1 (0.7%) rabbit harbored *T. gondii*-specific IgM. Female rabbits (9/84; 10.7%) had a similar *T. gondii* prevalence to males (6/58; 10.3%). When stratified by age, rabbits aged >1 year had a similar prevalence of *T. gondii* infection (7/66; 10.6%) to rabbits aged <1 year (8/76; 10.5%). Immunoblotting detected 6 major antigenic bands corresponding to *T. gondii*-positive sera at 20, 28, 30, 35, 63 and 77 kDa. Nested polymerase chain reaction (PCR) of whole-blood samples detected the *T. gondii* B1 gene in 23 rabbits (16.2%). All PCR-positive samples corresponded to partial *T. gondii* B1 gene sequences with 99% homology to a *T. gondii* sequence deposited in GenBank (accession number EU340874). Female rabbits (13/84; 15.5%) harbored a similar prevalence of *T. gondii* DNA to males (10/58; 17.2%). Rabbits aged >1 year had a similar prevalence (12/66; 18.2%) of *T. gondii* infection to rabbits aged <1 year (11/76; 14.5%). No statistically significant differences were observed regarding the prevalences of infection according to sex or age using molecular or serological tests. This study is the first survey using serological tests and nested PCR to analyze the *T. gondii* prevalence in rabbits in Korea.

**KEY WORDS:** ELISA, immunoblotting, nested PCR, rabbit, *Toxoplasma gondii*.

doi: 10.1292/jvms.13-0360; *J. Vet. Med. Sci.* 75(12): 1609–1613, 2013

*Toxoplasma gondii* is an obligate intracellular parasite that infects all warm-blooded vertebrates [14, 15]. It is generally known that cats are a major contributor to *T. gondii* transmission via fecal contamination of soil, food and water, because they can excrete millions of oocysts over a period of 1–2 weeks [4]. Although the prevalence of *T. gondii* infection in many kinds of animals has been described in many reports in the literature, there have been relatively few reports of the prevalence of *T. gondii* infection in the rabbit.

Rabbits are potential reservoirs for *T. gondii* transmission, and the detection of *T. gondii* from rabbits is a public health concern, as human consumption of rabbit meat continues to increase [15]. High titers of anti-*Toxoplasma* antibodies have been reported among rabbit hunters and workers at rabbit farms, and rabbits have been suggested to be a potential reservoir of *Toxoplasma* infection in humans [3, 13]. Fetal toxoplasmosis was found in three domestic rabbits in the U.S.A., and the most striking lesions in these rabbits were necrotic foci in the spleen and liver associated with massive presence of multiplying *T. gondii* tachyzoites [5]. In Korea, rabbits are raised on farms, and some rabbits are sold as food, while

others are distributed to pet shops. Also, in Korea, reports of *T. gondii* infection in dogs, cats, cattle and pigs and in the human population have been continuously published [10]. However, little is known regarding the prevalence of anti-*T. gondii* antibodies and PCR analysis among rabbits in Korea. The purpose of this study was to survey *T. gondii* infection among breeding farm rabbits in Korea using ELISA, immunoblotting and nested PCR.

### MATERIALS AND METHODS

**Animals:** One hundred forty-two rabbits (84 females, 58 males; age, 3 months–3 years; weight, 2.5–4 kg) were obtained from two collective breeding farms, one of which was located in the Gongju area of Chungnam Province in central Korea (100 rabbits), and one of which was located in the Kochang area of Junbuk Province in southwestern Korea (42 rabbits). All animals were raised communally and were asymptomatic for *Toxoplasma* infection. The rabbit breeds were as follows: crossbreed (n=34), Flemish giant (n=28), chinchilla (n=42) and New Zealand white (n=38). All rabbits were vaccinated for viral hemorrhagic disease. Whole blood samples were collected from auricular veins and were centrifuged to obtain sera for serological tests or combined with EDTA for nested polymerase chain reaction (PCR). All blood samples were stored at –80°C prior to use. This study was conducted in accordance with the Guide for the Care and Use of Laboratory Animals as approved by Chungnam

\*CORRESPONDENCE TO: SONG, K. H., Laboratory of Veterinary Internal Medicine, College of Veterinary Medicine, Chungnam National University, Daejeon 305–764, South Korea.  
e-mail: songkh@cnu.ac.kr

National University (No. CNU-00283).

**ELISA for *T. gondii* infection:** *Toxoplasma* lysate antigen (TLA) was prepared from *T. gondii* tachyzoites (RH strain). Briefly, tachyzoites were obtained from BALB/c mice that had been infected intraperitoneally with *T. gondii* 4 days earlier. Tachyzoites were washed several times with PBS and were homogenized by 5 sonications on ice for 30 sec each. Homogenates were centrifuged at  $15,000 \times g$  at  $4^{\circ}\text{C}$ , and supernatants were retained. The protein concentrations of the lysates were measured using a protein assay kit (Bio-Rad, Richmond, VA, U.S.A.), and lysates were stored at  $-70^{\circ}\text{C}$ . ELISA was performed using a previously reported method [12]. Plates were read at 490 nm using an automatic ELISA plate reader (Sunrise, Tecan, Salzburg, Austria). This cutoff value was calculated by method of Kimbita *et al.* [9]. Briefly, the mean optical densities of negative samples plus three standard deviations. ELISA results were considered positive for *T. gondii* infection when optical density values  $>0.39$  were obtained.

**Immunoblotting assay for *T. gondii* infection:** Immunoblotting was performed using the modified method of Quan *et al.* [12]. Briefly, TLA was boiled with sample buffer at  $100^{\circ}\text{C}$  for 5 min, and antigens were resolved through 12% acrylamide gels (Bio-Rad). Antigens were electro-transferred to nitrocellulose membranes (Millipore, Billerica, MA, U.S.A.) at a constant voltage of 100 V for 1 hr at  $4^{\circ}\text{C}$ . Membranes were blocked overnight with 5% skim milk in PBS at  $4^{\circ}\text{C}$ . Each serum sample was diluted 1:100 in 1% BSA/PBS and was incubated with nitrocellulose strips for 2 hr at room temperature. After 3 washes with PBST, the strips were incubated for 2 hr in HRP-conjugated anti-rabbit IgG (Abcam, Cambridge, MA, U.S.A.) diluted 1:5,000 in 1% BSA/PBS at room temperature. After 3 washes, the strips were incubated with 4-chloro-1-naphthol solution for 1 hr at room temperature.

**Nested PCR of *T. gondii* and sequencing analysis:** Genomic DNA was extracted from whole blood isolates using a PrimePrep Genomic DNA Isolation Kit (GeNet Bio, Daejeon, Korea). Samples were stored at  $4^{\circ}\text{C}$  until analysis by nested PCR. The following two PCR primer pairs were used to amplify the B1 gene [7]: S1 (5'-TGTTCTGTCCTATCGCAACG-3') and AS1 (5'-ACGGATG-CAGTTCCTTTCTG-3'), which amplify a 580-bp fragment, and S2 (5'-TCTTCCCAGACGTGGATTTC-3') and AS2 (5'-CTCGACAATACGCTGCTTGA-3'), which amplify a 530-bp fragment. Nested PCR was performed using a previously reported method [1]. PCR products were resolved through an ethidium bromide-stained 2% agarose gel in TAE buffer, and amplicons were visualized under UV light. Upon confirmation of the band of interest (530 bp), PCR products were purified using a Gene All Elution kit (GenAll, Seoul, Korea) according to the manufacturer's instructions.

The purified PCR products were sequenced by Cosmo Genetech (Seoul, Korea). The Lasergene sequence analysis software package (DNASTAR, Madison, WI, U.S.A.) was used to edit DNA sequences. Sequence alignment was conducted using ClustalV.

**Statistical analysis:** Statistical analyses of *T. gondii* preva-

lence among age and gender groups were performed using a Chi-squared ( $\chi^2$ ) test in Predictive Analytics Software (PASW<sup>®</sup>) Statistics 18 (Release 18.0 standard version, SPSS Inc., Chicago, IL, U.S.A.). A *P*-value  $<0.05$  was considered statistically significant.

## RESULTS

**ELISA to detect *T. gondii* infection:** Fifteen sera samples were positive for IgG antibodies against *T. gondii*, and one sample was positive for *T. gondii*-specific IgM antibodies. Variable optical densities were measured among the samples (Table 1).

Nine of the 84 female rabbits (10.7%) harbored *T. gondii*-positive sera compared with 6 of the 58 males (10.3%). Among animals aged  $>1$  year, 7 of 66 (10.6%) were *T. gondii* positive, versus 8 of 76 (10.5%) in the  $<1$ -year-old group. There were no statistically significant differences regarding the prevalences of infection according to sex or age. In the Gongju area of Chungnam Province, of 100 sera samples analyzed by ELISA, 9 (9.0%) exhibited *T. gondii*-specific IgG antibodies, and 1 (1.0%) contained *T. gondii*-specific IgM. Female rabbits (6/59; 10.2%) had a similar *T. gondii* prevalence to males (3/41; 7.3%). Rabbits aged  $>1$  year had a similar prevalence of *T. gondii* infection (5/47; 10.6%) to rabbits aged  $<1$  year (4/53; 7.5%). No statistically significant differences were detected regarding the prevalences of infection according to sex or age. In the Kochang area of Junbuk Province, of 42 sera samples analyzed by ELISA, 6 (14.3%) exhibited *T. gondii*-specific IgG antibodies. Female rabbits (3/25; 12.0%) had a similar prevalence to males (3/17; 17.6%). Rabbits aged  $>1$  year had a similar prevalence of *T. gondii* infection (2/19; 10.5%) to rabbits aged  $<1$  year (4/23; 17.4%). No significant differences were observed regarding the prevalences of infection according to sex or age. In the ELISA assay, the prevalence of *T. gondii* infection according to rabbit breeds was as follows: 14.3% (4/28) in the Flemish giant, 14.2% (6/42) in the chinchilla, 8.8% (3/34) in the crossbreed and 5.3% (2/38) in the New Zealand white.

**Immunoblotting assay to detect *T. gondii* infection:** Immunoblotting was performed in three *T. gondii*-negative samples and six *T. gondii*-positive samples by ELISA. The positive samples corresponded to antigenic bands at 20, 28, 30, 35, 63 and 77 kDa (Fig. 1).

**Nested PCR of *T. gondii* and sequencing analysis:** Of 142 DNA samples, 23 (16.2%) were positive for the *T. gondii* B1 gene as detected by nested PCR (Table 2). A similar prevalence of female rabbits (13/84, 15.5%) harbored the B1 gene compared with male rabbits (10/58, 17.2%). When the population was stratified by age, rabbits aged  $>1$  year (12/66, 18.2%) exhibited a similar *T. gondii*-positive prevalence to rabbits aged  $<1$  year (11/76, 14.5%). We detected no statistically significant differences regarding the prevalences of infection according to sex or age.

In the Gongju area of Chungnam Province, of 100 DNA samples analyzed by nested PCR, 14 (14.0%) were positive for the *T. gondii* B1 gene (Table 2). A total of 9 of 59 (15.3%) female rabbits were positive for the B1 gene versus 5 of 41

Table 1. The prevalence of *T. gondii* infection in rabbits by ELISA (IgG antibody) assay

	Gongju area (Chungnam Province)			Kochang area (Junbuk Province)			Total		
	NE	NP	PR (%)	NE	NP	PR (%)	NE	NP	PR (%)
Sex									
Females	59	6	10.2	25	3	12	84	9	10.7
Males	41	3	7.3	17	3	17.6	58	6	10.3
Age (years)									
<1	53	4	7.5	23	4	17.4	76	8	10.5
>1	47	5	10.6	19	2	10.5	66	7	10.6
Total	100	9	9	42	6	14.3	142	15	10.6

NE, numbers of examined; NP, numbers of positives; PR, positive rate.

Table 2. The prevalence of *T. gondii* infection in rabbits by nested PCR assay

	Gongju area (Chungnam Province)			Kochang area (Junbuk Province)			Total		
	NE	NP	PR (%)	NE	NE	PR (%)	NE	NE	PR (%)
Sex									
Females	59	9	15.3	25	4	16	84	13	15.5
Males	41	5	12.2	17	5	29.4	58	10	17.2
Age (years)									
<1	53	6	11.3	23	5	21.7	76	11	14.5
>1	47	8	17	19	4	21.1	66	12	18.2
Total	100	14	14	42	9	21.4	142	23	16.2

NE, numbers of examined; NP, numbers of positives; PR, positive rate.

(12.2%) male rabbits. When the population was stratified by age, rabbits aged >1 year (8/47, 17.0%) had a similar *T. gondii*-positive prevalence to rabbits aged <1 year (6/53, 11.3%). No statistically significant differences were observed regarding the prevalences of infection according to sex or age.

In the Kochang area of Junbuk Province, of 42 DNA samples analyzed by nested PCR, 9 (21.4%) were positive for the *T. gondii* B1 gene (Table 2). Then, 4 of 25 (16.0%) female rabbits were positive for the B1 gene versus 5 of 17 (29.4%) male rabbits. Rabbits aged >1 year (4/19, 21.1%) had a similar *T. gondii*-positive prevalence to rabbits aged <1 year (5/23, 21.7%). No significant differences were observed regarding the prevalences of infection according to sex or age. In nested-PCR analysis, the prevalence of *T. gondii* infection according to rabbit breeds was as follows: 21.4% (6/28) in the Flemish giant, 21.4% (9/42) in the chinchilla, 11.8% (4/34) in the crossbreed and 10.5% (4/38) in the New Zealand white.

PCR amplicons corresponding to *T. gondii*-positive samples were sequenced. All PCR-positive samples corresponded to partial *T. gondii* B1 gene sequences with 99% homology to a *T. gondii* sequence deposited in GenBank (accession number EU340874). In the present study, we deposited data for 16 partial sequences that showed 99% homology to each other and to the corresponding B1 gene of *T. gondii* (accession numbers KF038116, KF038117, KF038118, KF038119, KF038120, KF038121, KF038122, KF038123, KF038124, KF038125, KF038126, KF038127, KF038128, KF038129, KF413760 and KF143761).

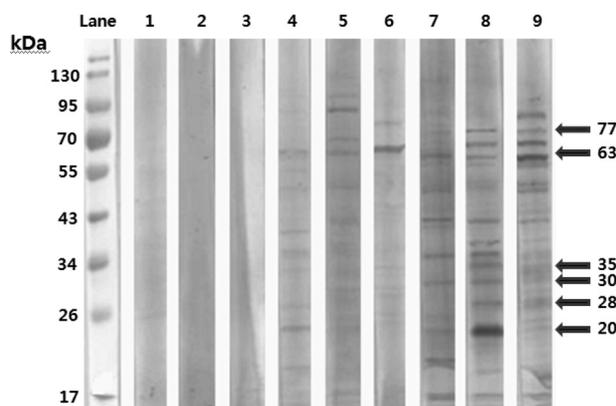


Fig. 1. Antigenic bands of *T. gondii*-positive samples in rabbits (lanes 1, 2 and 3 are negative samples by ELISA; lanes 4, 5, 6, 7, 8 and 9 are positive samples by ELISA) by immunoblotting.

## DISCUSSION

The rabbit population in Korea has increased recently with more people keeping rabbits as pets. We examined rabbits housed at two breeding farms in a mountainous region in the Gongju area of Chungnam Province and the Kochang area of Junbuk Province. Rabbits were reared outdoors, so they could potentially contact stray cats living in the area. It has been reported that the main factor affecting the prevalence of *T. gondii* infection is contact with *T. gondii*-shedding cats

[2]. Almeria *et al.* [2] reported that rabbits could be infected by *T. gondii* after ingesting food or water contaminated with *T. gondii* oocysts that had been excreted by felids. The rabbits examined in the present study may have consumed food or water contaminated with the feces of wild cats harboring *T. gondii* oocysts. However, no rabbits in the study population displayed clinical symptoms of toxoplasmosis (e.g., diarrhea, weight loss), and all rabbits were observably healthy. Using ELISA, Figueroa-Castillo *et al.* [6] detected serum antibodies to *T. gondii* in rabbits at three farms in Mexico. The farm with higher rearing standards carried a *T. gondii* seroprevalence of 18.7%, whereas the farm with mid-level standards and a family-managed farm carried seroprevalences of 39.7 and 33.3%, respectively. In the present study, *T. gondii*-specific IgG antibodies were detected in 10.6% of rabbits surveyed from 2 family-managed farms. These results are lower than those reported by Figueroa-Castillo *et al.* [6]. We observed no significant differences regarding the prevalences of *T. gondii* infection when rabbits were stratified by sex or age, consistent with a report by Almeria *et al.* [2]. The presence of IgG antibodies in blood samples suggests chronic infection with *T. gondii*.

Quan *et al.* [12] reported that IgM antibodies increase from day 4 post experimental infection with *T. gondii* to day 14 post infection and then decreased on day 16 post infection. In contrast, these researchers found that IgG antibodies increased from day 10 post infection to the end of the experiment. In the present study, a rabbit was identified by ELISA as harboring both IgM and IgG antibodies recognizing *T. gondii*. This result suggests an overlap of increasing IgM and IgG antibodies. This same rabbit was positive for *T. gondii* DNA by nested PCR. Of 142 rabbits in the study population, only this rabbit was shown to be *T. gondii*-positive by both ELISA and nested PCR. We also performed immunoblotting experiments on the *T. gondii* infected rabbits and detected major antigenic bands corresponding to 20, 28, 30, 35, 63 and 77 kDa. The results of our immunoblotting analyses were similar to those of described by Quan *et al.* [12].

Ishikawa *et al.* [8] reported a case of cervical toxoplasmosis transmitted from a rabbit to a man. The B1 genomic sequence is the primary molecular target for diagnosing toxoplasmosis [9], and several primer sets have been designed against various regions on the gene [11]. The primer set S1-AS1/S2-AS2 has been used for differentiation of *T. gondii* strains, and these primers are sufficiently sensitive and specific to be used as a diagnostic tool for cerebral toxoplasmosis [7].

Nested PCR amplified the *T. gondii* B1 gene in 23 of the 142 rabbits (16.2%). DNA samples derived from whole blood. No significant differences were observed in the prevalence of infection according to sex or age, consistent with the serological results. The presence of *T. gondii* DNA in whole blood indicates parasitemia, most likely related to acute *T. gondii* infection.

In the present study, there was a higher *T. gondii* prevalence by nested PCR (16.2%) than by ELISA (10.6%). We suggest that the nested PCR-positive/ELISA-negative rabbits might have harbored prepatent or latent infections with *T. gondii*,

because nested PCR is an extremely sensitive method. All PCR-positive samples corresponded to partial *T. gondii* B1 gene sequences with 99% homology to a *T. gondii* sequence deposited in GenBank (accession number EU340874).

We deposited data for 16 of 23 partial sequences that showed 99% homology to each other and to the corresponding B1 gene of *T. gondii* (accession numbers KF038116, KF038117, KF038118, KF038119, KF038120, KF038121, KF038122, KF038123, KF038124, KF038125, KF038126, KF038127, KF038128, KF038129, KF413760 and KF143761). In the present study, the rabbit breeds included the crossbreed (n=34), Flemish giant (n=28), chinchilla (n=42) and New Zealand white (n=38). Higher prevalence of *T. gondii* infection according to rabbit breed was found for the Flemish giant and chinchilla than the crossbreed and New Zealand white in the ELISA and nested PCR assay.

This study is the first survey to use serological tests and nested PCR to assess *T. gondii* prevalence in rabbits in Korea. A further study is needed to characterize the infective pathways from wild cats to rabbits and from rabbits to humans.

**ACKNOWLEDGMENT.** This work was supported by funding (4847-302-210-13, 2012) from the Korea National Institute of Health, Korea Centers for Disease Control and Prevention.

## REFERENCES

- Alfonso, Y., Fraga, J., Jimenez, N., Fonseca, C., Dorta-Contreras, A. J., Cox, R., Capo, V., Bandera, F., Pomier, O. and Ginorio, D. 2009. Detection of *Toxoplasma gondii* in cerebrospinal fluid from AIDS patients by nested PCR and rapid identification of type I allele at B1 gene by RFLP analysis. *Exp. Parasitol.* **122**: 203–207. [Medline] [CrossRef]
- Almeria, S., Calvete, C., Pages, A., Gauss, C. and Dubey, J. P. 2004. Factors affecting the seroprevalence of *Toxoplasma gondii* infection in wild rabbits (*Oryctolagus cuniculus*) from Spain. *Vet. Parasitol.* **123**: 265–270. [Medline] [CrossRef]
- Beverley, J. K. A., Beattie, C. P. and Roseman, C. 1954. Human *Toxoplasma* infection. *J. Hyg.* **52**: 37–46. [Medline] [CrossRef]
- Dubey, J. P. 1994. Toxoplasmosis. *J. Am. Vet. Med. Assoc.* **205**: 1593–1598. [Medline]
- Dubey, J. P., Brown, C. A., Carpenter, J. L. and Moore, J. J. 1992. Fatal toxoplasmosis in domestic rabbits in the USA. *Vet. Parasitol.* **44**: 305–309. [Medline] [CrossRef]
- Figueroa-Castillo, J. A., Duarte-Rosas, V., Juarez-Acevedo, M., Luna-Pasten, H. and Correa, D. 2006. Prevalence of *Toxoplasma gondii* antibodies in rabbits (*Oryctolagus cuniculus*) from Mexico. *J. Parasitol.* **92**: 394–395. [Medline] [CrossRef]
- Grigg, M. E. and Boothroyd, J. C. 2001. Rapid identification of virulent type I strain of the protozoan pathogen *Toxoplasma gondii* by PCR-restriction fragment length polymorphism analysis at the B1 gene. *J. Clin. Microbiol.* **39**: 398–400. [Medline] [CrossRef]
- Ishikawa, T., Nishino, H., Ohara, M., Shimosato, T. and Nanba, K. 1990. The identification of a rabbit-transmitted cervical toxoplasmosis mimicking malignant lymphoma. *Am. J. Clin. Pathol.* **94**: 107–110. [Medline]
- Kimbita, E. N., Xuan, X., Huang, X., Miyazawa, T., Fukumoto, S., Mishima, M., Suzuki, H., Sugimoto, C., Nagasawa, H., Fuji-

- saki, K., Suzuki, N., Mikami, T. and Igarashi, I. 2001. Serodiagnosis of *Toxoplasma gondii* infection in cats by enzyme-linked immunosorbent assay using recombinant SAG1. *Vet. Parasitol.* **102**: 35–44. [[Medline](#)] [[CrossRef](#)]
10. Lee, J. Y., Lee, S. E., Lee, E. G. and Song, K. H. 2008. Nested PCR-based detection of *Toxoplasma gondii* in German shepherd dogs and stray cats in South Korea. *Res. Vet. Sci.* **85**: 125–127. [[Medline](#)] [[CrossRef](#)]
11. Pujol-Rique, M., Derouin, F., Garcia-Quintanilla, A., Valls, M. E., Miro, J. M. and Jimenez de Anta, M. T. 1999. Design of a one tube hemi-nested PCR for the detection of *Toxoplasma gondii* and comparison of three DNA purification methods. *J. Med. Microbiol.* **48**: 857–862. [[Medline](#)] [[CrossRef](#)]
12. Quan, J. H., Hassan, H. A., Cha, G. H., Shin, D. W. and Lee, Y. H. 2009. Antigenemia and specific IgM and IgG antibody response in rabbits infected with *Toxoplasma gondii*. *Korean J. Parasitol.* **47**: 409–412. [[Medline](#)] [[CrossRef](#)]
13. Sroka, J. 2001. Seroepidemiology of toxoplasmosis in the Lublin region. *Ann. Agric. Environ. Med.* **8**: 25–31. [[Medline](#)]
14. Sugi, T., Kato, K., Kobayasi, K., Kurokawa, H., Takemae, H., Gong, H., Recuenco, F. C., Iwanaga, T., Horimoto, T. and Akashi, H. 2011. 1NM-PP1 treatment of mice infected with *Toxoplasma gondii*. *J. Vet. Med. Sci.* **73**: 1377–1379. [[Medline](#)] [[CrossRef](#)]
15. Zhou, Y., Zhang, H., Cao, J., Gong, H. and Zhou, J. 2013. Isolation and genotyping of *Toxoplasma gondii* from domestic rabbits in China to reveal the prevalence of type III strains. *Vet. Parasitol.* **193**: 270–276. [[Medline](#)] [[CrossRef](#)]